

Effects of leaf maturity and wind stress on the nutrition of a generalist caterpillar, *Lymantria dispar*, feeding on poplar

RAYMOND V. BARBEHENN^{1,2}, NOLA HAUGBERG¹, JOSEPH KOCHMANSKI¹ and BRANDON MENACHEM¹

¹Department of Molecular, Cellular and Developmental Biology and ²Department of Ecology and Evolutionary Biology University of Michigan, Ann Arbor, Michigan, U.S.A.

Running title: *Effects of leaf maturity and wind on herbivores*

Correspondence: Raymond V. Barbehenn, Department of Molecular, Cellular and Developmental Biology, University of Michigan, Ann Arbor, Michigan, 48109-1048, U.S.A.

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Tel.: +1 734 764 2770; email: rvb@umich.edu

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1 **Abstract.** The growth rates of insect herbivores commonly decrease when they feed on mature
2 leaves, a result of the combined effects of several nutritional and physiological mechanisms.
3 Environmental stresses during leaf development may also decrease herbivore performance. This
4 study tests two main hypotheses to help clarify the importance of these factors for the nutrition
5 and growth of an insect herbivore: (i) that decreases in nutrient levels, consumption rates, and
6 nutrient assimilation efficiencies impact negatively on herbivores feeding on mature leaves, and
7 (ii) that wind stress has a negative impact on herbivores feeding on mature leaves. The results
8 show that mature poplar (*Populus alba* x *P. tremula*) leaves have decreased levels of protein and
9 increased levels of fibre, and that growth rates of gypsy moth (*Lymantria dispar* L.) are
10 decreased on mature leaves in association with decreased consumption rates. However, contrary
11 to the first hypothesis, protein and carbohydrate are assimilated efficiently (74-82% and 84-87%,
12 respectively) from immature and mature poplar leaves. The larvae are able to chew mature
13 leaves as efficiently as immature leaves, potentially maximizing nutrient extraction. Contrary to
14 the second hypothesis, wind-stressed leaves have no significant detrimental effects on nutrient
15 assimilation efficiencies, and the lower growth rates of *L. dispar* larvae feeding on mature wind-
16 stressed leaves can be explained by lower consumption rates. Therefore, the availability of
17 nutrients to herbivores feeding on mature tree leaves is not necessarily impacted by lower
18 assimilation efficiencies, even when leaves develop under wind stress. These results help
19 explain some of the large variation between the nutritional qualities of trees for forest
20 Lepidoptera.

21
22
23 **Key words:** assimilation, carbohydrates, digestion, insect herbivore, larva, nutrients, protein,

24 **Introduction**

25 The nutritional quality of plants has a major impact on the fitness of insect herbivores.
26 Caterpillar growth rate and body mass (fitness-related parameters) are generally greatest for
27 larvae that feed on immature leaves (Feeny, 1970; Hough & Pimentel, 1978; Schweitzer, 1979;
28 Raupp & Denno, 1983; Raupp *et al.*, 1988; Schroeder, 1986; Hunter & Lechowicz, 1992; Parry
29 *et al.*, 1998; Kursar & Coley, 2003).

30
31 As leaves mature, their nutritional quality typically declines because of the lower
32 availability of some nutrients and increased fibre and toughness. Decreased levels of foliar
33 protein, carbohydrates, and/or water in mature tree leaves can each impact negatively on the
34 growth rates of insects (Scriber, 1979; Mattson, 1980; Coley *et al.*, 2006). Nutrient availability
35 is also affected by how efficiently nutrients are extracted, digested and absorbed (or
36 “assimilated”) by herbivores. Increased fibre is associated with increased leaf toughness, which
37 commonly decreases the consumption rates of insect herbivores (Kursar & Coley, 2003, Clissold
38 *et al.*, 2009). In turn, decreased nutrient assimilation rates have a negative impact on herbivore
39 growth rates.

40
41 Abiotic stresses, such as wind, during leaf development can cause plant responses that
42 potentially decrease foliar nutritional quality for insect herbivores. Wind stress on some plants
43 causes them to produce more flexible and fibrous leaves, and to develop foliar characteristics
44 that minimize water loss from transpiration (Jaffe & Forbes, 1993; Anten *et al.*, 2010). A major
45 study on the effects of wind stress on leaf composition shows increased levels of lignin and
46 peroxidase activity in stressed bean leaves (Cipollini, 1997). However, few studies have

47 examined foliar nutritional components other than fibre. Under certain conditions, the
48 mechanical stimulation of a grass lowers foliar nitrogen levels significantly, but does not
49 consistently affect carbohydrates (Kraus *et al.*, 1994). To characterize the effects of a major
50 abiotic stress on foliar nutritional quality better, this study examines the effects of wind stress on
51 foliar fibre components, as well as nutrients such as protein and carbohydrate.

52
53 No previous work, of which the authors are aware, examines the impact of wind stress on
54 the nutritional physiology of insect herbivores. Previous work on mite populations shows that
55 wind-stressed bean leaves support lower reproductive rates (Cipollini, 1997). However, wind-
56 stressed tomato leaves have no significant effect on the growth rates of *Manduca sexta*
57 caterpillars (Cipollini & Redman, 1999). In these cases, it is unclear whether the herbivores are
58 affected because of decreased feeding (mites), or are unaffected because of compensatory
59 responses, such as increased consumption rates (*M. sexta*). A major purpose of this study is to
60 examine the mechanisms underlying the impact of wind stress on the nutrition of a tree leaf-
61 feeding insect.

62
63 The availability of nutrients such as protein can decline from mature leaves not only
64 because of a decrease in their levels, but also because of decreased extractability. Decreased
65 protein extractability is largely responsible for the decline in protein assimilation efficiency
66 (PAE) by *Lymantria dispar* L. larvae on the mature leaves of red oak (*Quercus rubra*) and
67 presumably also on sugar maple (*Acer saccharum*) (Barbehenn *et al.*, 2013a, 2014). Caterpillars
68 feeding on lush, immature oak and maple leaves assimilate protein with efficiencies of 70-80%,
69 which decline after leaf maturation to efficiencies of less than 50%. By contrast, when *L. dispar*

70 larvae feed on several species in the willow family (Salicaceae), there appears to be little
71 decrease in protein or carbohydrate assimilation efficiencies (Barbehenn *et al.*, 2015). In this
72 study, it is expected that PAE remains high in *L. dispar* that feed on the mature leaves of hybrid
73 poplar (*Populus alba* x *P. tremula*), and that high PAE is associated with high protein
74 extractability from ingested leaf tissues. Previous work on carbohydrate assimilation efficiency
75 (CAE) shows that sugar assimilation efficiencies remain high (ca. 90%) from mature leaves,
76 whereas starch assimilation efficiencies are consistently lower (ca. 30-50%) (Horie *et al.*, 1985;
77 Barbehenn *et al.*, 2014, 2015). Nevertheless, it is expected that overall CAE remains high on
78 mature leaves, based on the predominance of sugars in the carbohydrate budgets of tree-feeding
79 caterpillars (Horie *et al.*, 1985; Barbehenn *et al.*, 2014, 2015). The balance of protein and
80 carbohydrate assimilated by insect herbivores can affect their growth rates (Clissold *et al.*, 2006;
81 Behmer, 2009). Therefore, this study examines the effects of leaf maturation and wind stress on
82 the balance of protein and carbohydrates that are assimilated by an herbivore.

83
84 *Lymantria dispar* is a highly polyphagous European species that was introduced into the
85 eastern United States. It has become a model tree-feeding insect species because of its economic
86 importance and wide host range. Among the hundreds of tree and shrub species on which *L.*
87 *dispar* feeds, poplars are among the most favourable for its growth (Liebhold *et al.*, 1995).
88 Poplar has become a model deciduous tree because of its economic value, rapid growth, and the
89 utility of *Populus* species for genetic research. In this study, rapid tree growth and large leaf size
90 are essential for examining treatment effects in a brief (2 month) period with late-stadium larvae.
91 Late-stadium larvae produce sufficient frass to quantify nutrient assimilation efficiencies for
92 each individual. The use of a continuously growing (indeterminate) tree species such as poplar is

93 important to perform combined experiments on leaf maturity and wind effects because immature
94 and mature leaves are available together. The examination of wind stress in a greenhouse is
95 expected to increase the sensitivity of its detection, given that poplar trees are able to be grown in
96 the absence of wind for comparison with trees grown under wind stress.

97
98 The first main hypothesis tested is that leaf maturation has negative impacts on caterpillar
99 nutrition and growth due to decreased nutrient levels, decreased consumption rates, and
100 decreased nutrient assimilation efficiencies. To compare nutritional factors that could directly or
101 indirectly impact nutrient assimilation efficiencies and rates, levels of foliar protein,
102 carbohydrates, water, and fibre are measured. The second main hypothesis tested is that wind
103 stress has a negative impact on caterpillar nutrition and growth. A 2 x 2 factorial study is
104 performed on the immature and mature leaves from control and wind-blown poplar saplings,
105 examining leaf nutritional quality, nutrient assimilation efficiencies, and larval assimilation and
106 growth rates. Two factors that potentially affect nutrient assimilation efficiencies are also
107 examined: chewing efficiency (i.e., food particle size) and protein extraction efficiency from
108 ingested leaf tissues. If mature leaves are chewed less thoroughly than immature leaves, nutrient
109 assimilation efficiencies could be decreased. To determine whether ingested mature leaf tissues
110 retain higher levels of unextracted protein than do immature leaf tissues, protein levels are
111 measured in leaf particles from the midguts and frass of larvae that consume immature or mature
112 leaves. Together, these physiological measurements are aimed at a better understanding of the
113 impact of leaf quality on the nutritional ecology of forest Lepidoptera.

114

115

116 **Materials and methods**

117 *Insect-plant system*

118 Eggs of *L. dispar* were obtained from the USDA (Otis Air Force Base, Massachusetts).
119 Larvae were reared in Petri dishes in incubators, primarily at 23 °C (16 h light). They were fed
120 artificial diet (Addy, 1969) from egg hatch through the third or fourth stadium. The diet was
121 modified by using linseed oil instead of wheat germ oil.

122
123 Poplar saplings of clone 717 ($n = 20$) were grown in a greenhouse in tall 8-L pots
124 designed for tree growth (Barbehenn *et al.*, 2007). They were watered daily and fertilized with
125 Osmocote 15-9-12 slow-release fertilizer (Hummert International, Earth City, Missouri) (50 mL
126 per pot). Saplings attained a height of over 2 m before their leaves were used in the feeding
127 experiment on leaf maturity (July, 2012). For the experiment on the effects of leaf maturation
128 and wind stress (July, 2013), poplar saplings were grown as in the above experiment, with the
129 exception that wind stress was applied to one group. Box fans were positioned to produce a
130 wind speed averaging 2.1 m sec^{-1} on 9 saplings. The force of the air produced a constant
131 swaying of the tree tops and movement of their leaves for more than a 2-month period before
132 they were used. Saplings were moved to new positions daily. Wind speed was measured with
133 an R.M. Young Wind Monitor-AQ anemometer (R.M. Young Co., Traverse City, Michigan).
134 Control saplings ($n = 9$) were grown in the same room, but on the back side of the fans, where no
135 measurable air speed was recorded. Average wind speeds outside the greenhouse were 1.3 m
136 sec^{-1} (Barbehenn *et al.*, 2015). Daily minimum and maximum temperatures in the greenhouse
137 averaged 20.0 ± 0.6 and 29.7 ± 0.6 °C, respectively. All saplings produced over 40 leaves under
138 the experimental conditions deployed.

139 Leaf position was determined using the leaf plastochron index (LPI), with the first
140 uncurled leaf greater than 2 cm being defined as leaf “0”, and older leaves counted radially
141 around the tree (Larson & Isebrands, 1971). Very young leaves (less than LPI 10) were excluded
142 from this study because they may contain elevated levels of toxins, feeding deterrents, and/or
143 antinutritional compounds (e.g., salicylates and protease inhibitors), and are often avoided by
144 foraging caterpillars (Meyer & Montgomery, 1983; Haruta *et al.*, 2001). For the first
145 experiment, leaves were cut from LPI 10-12 (immature), LPI 20-22 (recently mature), and LPI
146 30-32 (fully mature). Immature leaves retained the same light green colour found in leaves from
147 LPI 0-9, and they were not yet fully expanded. By comparison, mature leaves had become dark
148 green. Leaves were cut with a razor blade through their petioles, and were harvested each
149 morning during the experiments. This method was expected to avoid the induction of defences
150 during the course of the experiment (Osier *et al.*, 2000). A single leaf was cut from each position
151 from 9 trees each day. Leaf turgor was maintained by keeping the leaf petioles in tubes of water
152 until they were used. Leaf discs (2.5 cm-diameter) were cut with a cork borer, avoiding the
153 midrib. The discs were mixed within age groups to eliminate the potential effect of between-tree
154 variation in food quality. The time from leaf harvest to larval feeding ranged between 2-3 hours.
155 Leaf discs were kept turgid in a humidified Petri dish at ambient temperature before they were
156 fed to larvae. The use of leaf disks permitted accurate measurements of consumption by
157 providing amounts of food that would be largely consumed (Schmidt & Reese, 1986).

158

159 Immature leaves were selected for the wind stress experiment from a zone centred near
160 LPI 15, and mature leaves were cut near LPI 25. Again, immature leaves were a light green
161 colour and were not fully expanded. Leaf discs were obtained as described above, with the

162 exception that the leaves were first washed in a mild detergent solution, rinsed by soaking in tap
163 water, and dried with paper towels before being cut into discs.

164

165 *Effect of leaf maturity on larval nutrition and growth*

166 Newly moulted fourth-stadium larvae were assigned randomly to one of three leaf age
167 groups. Individual larvae were fed for a three-day period on immature, newly mature, or fully-
168 mature leaves ($n = 15$ per treatment) in 35-mL snap-cap plastic cups. Groups of freshly cut leaf
169 discs were weighed for each larva daily. A moistened paper filter was placed in the bottom of
170 each cup, and hydrated daily, to keep the leaf disks turgid. Uneaten food was dried ($70\text{ }^{\circ}\text{C}$) each
171 day to determine the amount eaten. Representative leaf discs were weighed fresh and after
172 drying to estimate the dry weight of food given to each larva on each day. Additional leaf discs
173 were frozen ($-80\text{ }^{\circ}\text{C}$) daily and lyophilized. Frass was collected daily, and kept separately for
174 each larva in screw-cap centrifuge tubes at $-80\text{ }^{\circ}\text{C}$. After 3 days of feeding, larvae were kept in
175 empty cups for 6 h to collect their final frass pellets. Larvae were then frozen and dried to obtain
176 their final masses. Initial larval masses were estimated from the fresh weight:dry weight ratio of
177 5 representative larvae. Consumption, assimilation and growth rates were relativized by dividing
178 them by the initial dry mass of the larvae. Nutrient analyses are described below. The following
179 nutritional indices were examined for protein:

180 Relative protein consumption rate (RPCR) = mass of protein ingested $\text{day}^{-1} \text{mg}^{-1}$

181 Protein assimilation efficiency (PAE) = (mass of protein ingested - mass of protein
182 egested) / mass of protein ingested x 100

183 Relative protein assimilation rate (RPAR) = (mass of protein ingested - mass of protein
184 egested) / day

185 (Note that $RPCR \times PAE = RPAR$.) Protein assimilation efficiency is identical to the
186 “approximate digestibility” of protein. Protein ingested was calculated as percent protein in
187 leaves \times total mass ingested, and total protein egested was calculated as percent protein in frass \times
188 total mass egested. Carbohydrate assimilation efficiencies and rates were measured in the same
189 manner as described for protein by analyzing carbohydrate (i.e., sugar and starch) levels in the
190 food and frass of the same larvae.

191

192 *Effects of leaf maturity and wind stress on larval nutrition and growth*

193 Newly moulted fifth-stadium larvae were weighed and assigned randomly to one of four
194 treatment groups: immature control leaves, mature control leaves, immature wind-stressed
195 leaves, or mature wind-stressed leaves. Larvae ($n = 14$ per treatment) were fed weighed leaf
196 discs for a two-day period in the same experimental conditions as described above. At the end of
197 the experiment, larvae were kept in empty cups overnight to collect their final frass pellets. A
198 subgroup of larvae was dissected to confirm that their guts were empty after the period of
199 starvation. Final larval weights were measured after they were dried (70°C , 2 days). All leaf
200 and frass samples were lyophilized for chemical analysis (below).

201

202 *Factors affecting nutrient assimilation efficiency*

203 *Chewing.* To examine whether chewing efficiency is impacted by leaf maturity, fifth-stadium
204 larvae ($n = 15$ per treatment) were fed for a 2-day period on leaf disks from immature, recently
205 mature, or fully mature leaves grown in control conditions (July, 2012), as described above.
206 Larvae were chilled individually (6 min, -20°C) and dissected. The contents of the foreguts of

207 larvae were dispersed in 500 μ L of 50% (v/v) methanol. Particles were stained and their sizes
208 measured in micrographs as described previously (Barbehenn *et al.*, 2014).

209 *Mandible size.* To examine the effect of mandible size on food particle size, mandibles were
210 dissected from the head capsules of each larva used for measuring food particle size. Mandible
211 size was measured using an ocular micrometer in a stereo-microscope at a magnification of 25X,
212 with the length between condyles (points of articulation) serving as an indicator of overall size.

213
214 *Protein extractability from ingested leaf particles.*

215 The ability of larvae to extract protein from ingested leaf tissues can decrease as leaves mature.
216 To examine this directly, the protein that remained inside ingested leaf particles was measured in
217 samples from the posterior half of the midguts of fifth-stadium larvae (July, 2012). These
218 samples were obtained from the same larvae used for measuring food particle size. The contents
219 of each posterior midgut ($n = 15$ per treatment) was placed in a separate tube containing 500 μ L
220 of 20 mM HCl, which was flushed with nitrogen. Samples were dispersed by shaking, and were
221 immediately centrifuged (8,000 $\times g$, 3 min). Supernatant solutions were removed (eliminating
222 any protein that was not contained inside the leaf particles), and the pellets were lyophilized.
223 Protein-bound amino acids in 5-mg samples were measured in acid hydrolysates with high
224 performance liquid chromatography (HPLC), as described below.

225
226 Protein levels were also compared in the frass of fifth-stadium larvae ($n = 15$ per
227 treatment) in a separate experiment in which they fed on poplar leaves from one of the three age
228 groups (July, 2012). The experiment followed the same procedures described above for feeding
229 experiments with immature and mature leaves. Protein levels were measured in lyophilized frass

230 as total amino acids (protein-bound plus free) in acid hydrolysates with HPLC. Total amino
231 acids in the frass were expected to be primarily protein-bound because previous work has found
232 negligible levels of free amino acids in the frass of *L. dispar* larvae that fed on immature or
233 mature red oak leaves (Barbehenn *et al.*, 2014).

234

235 *Chemical analyses*

236 All analyses were performed as described previously (Barbehenn *et al.*, 2014). Briefly,
237 protein was measured with HPLC as total amino acids (protein-bound plus free) in 6 M HCl
238 hydrolysates from each leaf age and date and the frass from each larva. Of the total amino acids
239 measured in the leaves, approximately 99% were expected to be protein-bound (Giovannelli,
240 1987; Ruuhola *et al.*, 2003). Sugars (summing glucose, fructose and sucrose) were measured
241 using an enzymatic method (Zhao *et al.*, 2010). Starch was measured in the extracted pellets
242 remaining from leaf and frass samples (Zhao *et al.*, 2010). Sugars and starch were combined as a
243 measure of total nonstructural carbohydrates. Water was measured in each of the feeding
244 experiments by weighing leaf discs before and after they were oven dried (70 °C). Total fibre
245 was measured with the neutral detergent fibre (NDF) assay, modified from Van Soest & Wine
246 (1967). In the experiment to examine the effects of wind stress, cellulose, hemicellulose and
247 lignin were quantified as follows. The NDF assay was followed on the same samples by the acid
248 detergent fibre (ADF) assay (Van Soest & Wine, 1967, as modified in Barbehenn *et al.*, 2014)
249 and then an assay for lignin. Lignin (Klason type) was determined by treating the ADF pellets
250 with 72% sulphuric acid (Dence, 1992). Cellulose was determined as the difference between
251 ADF and lignin. Hemicellulose was determined as the difference between NDF and ADF.

252

253 Waste nitrogen products were quantified in the frass from each larva in the first
254 experiment. The two major waste nitrogen products (uric acid and ammonium salts) were
255 extracted from 5-mg frass samples in 500 μ L of pH 2.1 buffer, and measured with HPLC as
256 described previously (Barbehenn *et al.*, 2014). Allantoic acid and allantoin were below the
257 limits of detection. Of the total amino acids (protein-bound + free) measured in the frass of *L.*
258 *dispar*, approximately 98% were protein-bound, regardless of food quality (Barbehenn *et al.*,
259 2014). Therefore, free amino acids are not considered a form of waste nitrogen for *L. dispar*.

260

261 *Experimental design and statistical analyses*

262 The experimental design in this study emphasizes the measurement of larval nutrition and
263 growth to test the main hypotheses. The number of replicate larvae used ($n = 14-15$) was
264 sufficient to demonstrate whether there are nutritional effects of leaf maturity and/or wind stress
265 in a short period of larval growth (i.e., 2 or 3 days). By contrast, variation between trees was
266 minimized by using a clone and mixing leaf samples from multiple saplings within treatment
267 groups on each experimental day, leading to sample sizes of only $n = 2$ or 3 replicate days per
268 treatment. This low number of replicates limited the statistical power to show treatment effects
269 on the leaves. However, the experimental design was aimed primarily at quantifying the
270 amounts of nutrients that were consumed by each larva.

271

272 The effects of leaf maturation on the levels of foliar protein, carbohydrate, water, fibre,
273 were made between three leaf age groups with one-way ANOVA (SAS Institute, 2010). The
274 effects of leaf maturation and wind stress on foliar nutritional quality were examined with a 2-
275 factorial ANOVA, using leaf age and wind treatment as main factors. Because leaf discs were

276 pooled within treatment groups, replicates for leaf analyses were days within experiments ($n = 2$ -
277 3 per treatment).

278

279 The effects of leaf maturation on consumption rates and assimilation efficiencies were
280 compared between leaf ages with ANCOVA, using ingested masses or egested masses as main
281 factors and initial dry mass or ingested masses as covariates, respectively (SAS Institute, 2010).

282 The assumption of parallel slopes for ANCOVA was confirmed with a separate analysis of the
283 treatment x covariate interaction term (non-significant). Relative growth rate data could not be
284 transformed to meet the assumptions of ANCOVA, and were examined with one-way ANOVA.

285 The effects of leaf maturation and wind on larval nutrition and growth were compared with 2-
286 factorial ANCOVA. One exception was CAE, which was analyzed with two-way ANOVA.

287 Effects of leaf age on food particle protein, faecal protein, waste nitrogen, food particle size, and
288 mandible size were examined between treatment groups with one-way ANOVA. For all

289 analyses, the normality of the residuals was confirmed with the Shapiro-Wilk test (PROC-
290 MIXED). Post hoc multiple comparisons were made using the probabilities of differences

291 between least squares means (PROC MIXED). These comparisons tested *a priori* hypotheses.

292 Therefore, a P -value of 0.05 was used to determine statistical significance. Individual larvae
293 served as replicates in all experiments on insects (see Methods and Results for sample sizes).

294 Pearson correlation analyses were used to examine associations between (1) nutrient levels,
295 consumption rates, assimilation efficiencies, assimilation rates, and growth rates, (2) mandible
296 size and food particle size, and (3) P:C ratios in leaves and RPAR:RCAR by larvae.

297

298

299 **Results**

300 *Effect of maturation on leaf nutritional quality*

301 The levels of protein, carbohydrate, water and fibre were measured in the three ages of
302 leaves ingested by larvae (Table 1). As often occurs in mature leaves, protein and water levels
303 decreased significantly by 12-28% and 4-5%, respectively, relative to levels in immature leaves.
304 However, changes in carbohydrate were not shown to be significant. Carbohydrate was
305 composed of 87-92% sugar, with the remaining fraction being starch. Fibre levels were
306 significantly higher by 20% in fully mature leaves.

307

308 *Effect of leaf maturation on larval nutrition and growth*

309 Consistent with the first hypothesis, larval consumption rates were 29-32% higher on
310 immature leaves than on mature leaves (Table 2). Although assimilation efficiencies for the
311 entire leaf mass decreased when larvae fed on mature leaves, the assimilation efficiencies of two
312 major macronutrients were not impacted negatively by leaf maturation. Protein assimilation
313 efficiency remained high (74-82%) in *L. dispar* caterpillars feeding on mature leaves.
314 Carbohydrate assimilation efficiency also remained high in larvae on all ages of poplar leaves
315 (Table 3). Sugars were assimilated with exceptionally high efficiencies (averaging 94-97%
316 across leaf ages), whereas starch was assimilated less efficiently (averaging 56-60%). However,
317 the relatively small amounts of starch in poplar leaves meant that low starch assimilation
318 efficiency had little effect on overall CAE. Waste nitrogen produced by larvae on mature leaves
319 dropped by 43-60% (Table 4). The total amount of waste nitrogen produced by each larva was
320 associated positively with the total amount of protein it assimilated ($r = 0.74, P < 0.001$). The
321 growth rates of larvae that fed on mature leaves declined by 33-43%, in association with lower

322 consumption rates ($r = 0.74$), lower RPAR ($r = 0.86$), and to a lesser extent, lower RCAR ($r =$
323 0.41).

324

325 *Effect of leaf maturation and wind stress on leaf nutritional quality*

326 During leaf maturation, protein levels were again found to decrease significantly by 24%,
327 whereas the opposite trend in carbohydrates was observed. In contrast to the first experiment, no
328 significant changes in leaf water content or trends in fibre levels were observed in mature leaves.
329 Wind-stressed leaves contained fibre levels that were increased significantly by 3-5% relative to
330 the levels in control trees (Table 5). Among the three fibre components measured, wind stress
331 was associated with a trend towards higher cellulose ($P = 0.112$), but no increases in lignin or
332 hemicellulose were detected (Fig. 1).

333

334 *Effect of leaf maturation and wind stress on larval nutrition and growth*

335 Consistent with the results of the above experiment on leaf maturation, the PAE and CAE
336 of larvae that fed on mature poplar leaves remained high (ca. 70% and 90%, respectively)
337 (Tables 6 and 7). The negative effect of leaf maturation on PAE was small (5-9% decrease), as
338 was the positive effect of leaf maturation on CAE (3-4% increase). Contrary to the wind stress
339 hypothesis, *L. dispar* larvae that fed on wind-stressed leaves did not have significantly decreased
340 PAE or CAE. However, there was a significant 4-7% decrease in total assimilation efficiency in
341 larvae on wind-stressed leaves, which could not be explained by patterns of protein or
342 carbohydrate assimilation. In addition, larvae that fed on mature wind-stressed leaves had
343 significantly lower consumption rates, which led to a trend towards lower growth rates on wind-
344 stressed leaves (Table 6). The importance of protein nutrition for larval growth can be inferred

345 from correlations between nutrient assimilation rates and growth rates. Larval growth rates
346 (Table 6) were closely associated with protein assimilation rates (RPAR; $r = 0.74$), and more
347 strongly associated with protein than with carbohydrate assimilation rates (RCAR; $r = 0.53$). In
348 turn, RPAR and RCAR were strongly associated with protein and carbohydrate consumption
349 rates ($r = 0.99$ and 0.98 , respectively). Protein assimilation efficiency, which contributes to
350 RPAR, was also associated with RPAR ($r = 0.74$).

351
352 Consistent with the efficient assimilation of protein and carbohydrate from immature and
353 mature poplar leaves, foliar protein : carbohydrate (P:C) ratios were strongly associated with
354 larval RPAR : RCAR ratios (Fig. 2, $r = 0.97$, $P < 0.001$ for combined data). The slope of the
355 regression equation was close to 1.0 (0.88), indicating that the ratio of P:C assimilated was
356 modified slightly from the P:C ratio in the leaves.

357

358 *Factors affecting nutrient assimilation efficiency*

359 *Chewing.* Caterpillars chewed immature and mature leaves into a wide range of particle sizes
360 (Fig. 3). Contrary to expectation, median food particle sizes were similar between each leaf age
361 group, ranging from 0.32 - $0.34 \mu\text{m}^2$. Larval mandibles ranged from 0.89 to 1.22 mm wide.
362 However, mandible size was not associated with food particle size ($r = 0.15$ - 0.34 for each leaf
363 age).

364

365 *Protein extractability from ingested leaf particles.* Protein levels in food particles from the
366 posterior midgut did not differ significantly between larvae on immature and mature leaves (Fig.
367 4), consistent with the high PAEs in *L. dispar* on all ages of poplar leaves. Protein levels

368 remaining in the frass of larvae that fed on immature or mature leaves also supports the
369 conclusion that protein does not become more difficult to extract from mature leaves (Fig. 4).
370 The large difference in protein levels between the posterior half of the midgut and the frass
371 suggests that much of the absorption of amino acids occurs in the posterior midgut.

372

373 **Discussion**

374 The larvae of *L. dispar* grow more rapidly on immature than on mature poplar leaves, as
375 would be expected from previous research (Feeny, 1970; Hough & Pimentel, 1978; Schweitzer,
376 1979; Raupp & Denno, 1983; Raupp et al., 1988; Schroeder, 1986; Hunter & Lechowicz, 1992;
377 Parry *et al.*, 1998; Kursar & Coley, 2003). However, by examining a wide range of
378 physiological factors that potentially impact herbivore growth, this study shows that their higher
379 growth rates on immature poplar leaves are due to higher leaf quality, higher consumption rates,
380 and the associated increases in nutrient assimilation rates, but do not result from higher nutrient
381 assimilation efficiencies or greater chewing efficiency by larvae on lush young leaves. Thus, the
382 findings in this study require the rejection of one component of the first hypothesis: PAE and
383 CAE by *L. dispar* larvae are not negatively impacted when larvae feed on mature poplar leaves.
384 Similar, high PAE (77-80%) and CAE (83-87%) are also observed in fifth-stadium larvae on
385 immature or mature poplar leaves (R. V. Barbehenn, unpublished observations). This is contrary
386 to a common belief that nutrient assimilation efficiencies decrease in insect herbivores on mature
387 or more fibrous leaves (e.g., Read & Stokes, 2006). Whereas there is a pattern of decreased PAE
388 from the mature leaves of some trees, such as red oak and sugar maple (Barbehenn *et al.*, 2013a,
389 2014), there is a large amount of variation in this trait among tree species. Several species in the
390 Salicaceae have highly available protein and carbohydrates in their mature leaves (Barbehenn *et*

391 *al.*, 2015). The present study confirms this finding, and offers a more detailed view of the
392 nutritional physiology of *L. dispar* on a wider range of leaf ages.

393
394 A second major finding is that wind-stress during poplar leaf development has only a
395 limited impact on foliar composition and herbivore nutrition. Wind-stressed leaves develop
396 higher levels of fibre, presumably as an adaptive response to avoid mechanical damage. In one
397 of the few previous studies on wind stress that examine foliar chemical composition, wind stress
398 causes bean leaves to produce higher levels of fibre, especially lignin (Cipollini, 1997).
399 However, lignin is not among the fibre components that increase in wind-stressed poplar. Also
400 contrary to the wind stress hypothesis, the changes in wind-stressed poplar leaves do not
401 significantly affect the availability of nutrients to *L. dispar* larvae. The results of this study
402 confirm recent results showing that when poplar is grown in outdoor wind conditions, protein
403 and carbohydrate are assimilated efficiently from mature leaves (i.e., PAE = 74-75% and CAE =
404 78%; Barbehenn *et al.*, 2015). In the only previous study on the effects of wind stressed leaves
405 on a leaf-chewing herbivore, the growth rate of *M. sexta* caterpillars is not affected significantly
406 by feeding on wind-stressed tomato leaves (Cipollini & Redman, 1999). Similarly, in the current
407 study, there are only limited effects of wind stressed leaves on the nutrition of *L. dispar*, and
408 these appear to be attributable to decreased consumption rate rather than decreased nutrient
409 availability.

410
411 To understand the adverse effects of feeding on mature leaves, several nutritional
412 mechanisms are examined in this study, including chewing efficiency, protein extractability, P:C
413 ratios, and waste nitrogen excretion. The ability of *L. dispar* to chew leaves into fine particles is

414 the same on immature and mature poplar leaves. This is unexpected, given that cell wall
415 thickness increases on the order of ten-fold and fibres become extensively crosslinked during
416 maturation (Brisson *et al.*, 1994; Doblin *et al.*, 2003). However, similar chewing abilities are
417 observed in larvae on immature and mature red oak leaves, the latter of which are remarkably
418 tough (Barbehenn *et al.*, 2013a, 2014). Therefore, in each of these studies on *L. dispar*,
419 increased fibre levels and toughness do not affect nutrient assimilation efficiencies as a result of
420 chewing efficiency. In caterpillars on oak, PAE decreases markedly on mature leaves, whereas
421 in caterpillars on poplar, PAE remains high, regardless of the similar sizes of leaf particles that
422 are chewed by larvae on the immature or mature leaves of each host plant.

423
424 Although leaf toughness (or fibre content) does not affect the chewing efficiency of *L.*
425 *dispar* with respect to food particle size, it does appear to impact the rate of food consumption
426 for *L. dispar* and many other insect herbivores (Choong, 1996; Kursar & Coley, 2003;
427 Barbehenn *et al.*, 2013, 2014, 2015). For *L. dispar*, there are strong associations between
428 consumption rate, protein assimilation rate, and growth rate. Importantly, decreased growth rate
429 commonly leads to decreased fitness (Scriber & Slansky, 1981; Awmack & Leather, 2002).

430
431 Consumption rates could also be decreased on mature leaves by decreased levels of
432 feeding stimulants and/or increased levels of deterrent or toxic plant compounds. It is
433 noteworthy that sugars (feeding stimulants) remain at equally high levels in mature and
434 immature poplar leaves. The major deterrents and toxins in *Populus* species are salicylates (or
435 “phenolic glycosides”), but these are found at lower levels in mature than immature leaves (Osier
436 *et al.*, 2000; Kleiner *et al.*, 2003), presumably causing *L. dispar* larvae to avoid feeding on the

437 youngest leaves (Meyer & Montgomery, 1987). Previous work shows that the poplar leaves
438 used in this study have negligible levels of tannins and low levels of other phenolic compounds
439 (Barbehenn *et al.*, 2007). It is also unlikely that some of the putative defences examined
440 previously in wind stressed leaves, e.g., peroxidase and polyphenol oxidase, affect the nutrition
441 of *L. dispar* (Barbehenn *et al.*, 2007, 2010, 2012). Further work would be needed to examine
442 whether other types of feeding stimulants and/or deterrents affect the consumption rates by *L.*
443 *dispar* on mature poplar leaves (e.g., Coyle *et al.*, 2002).

444
445 Protein extractability is a key difference between the digestibility of the leaf tissues of
446 poplar and the leaf tissues of species such as red oak. Whereas protein remains highly
447 extractable from the ingested tissues of mature poplar leaves, protein in mature red oak leaves
448 becomes significantly more difficult to extract (Barbehenn *et al.*, 2014). In contrast to protein,
449 CAE remains equally high in *L. dispar* larvae that feed on the immature or mature leaves of
450 poplar or red oak. Therefore, the ratios of protein : carbohydrate present in immature and mature
451 poplar leaves are tightly associated with the ratios of RPAR:RCAR in *L. dispar* larvae. Similar
452 results are found in *L. dispar* on a several species of trees in the Salicaceae (Barbehenn *et al.*,
453 2015). These findings reiterate the minimal effect that leaf maturation has on macronutrient
454 extractability from certain host plants, in comparison to the large decrease in PAE from species
455 such as red oak and sugar maple (Barbehenn *et al.*, 2013, 2014). Shifts in the balance of protein
456 and carbohydrates that can be assimilated from mature leaves could be important for the growth
457 of *L. dispar*, as found in other insect herbivores on other food sources (Clissold *et al.*, 2006;
458 Behmer, 2009). Together, these studies of *L. dispar* on a variety of tree species show that

459 developmental changes in leaf tissues can have a strong effect on nutrient availability, but that
460 this effect varies greatly between tree species and nutrient types.

461
462 Given that larvae of *L. dispar* appear to maximize protein and carbohydrate assimilation
463 efficiencies, it is likely that post-absorptive mechanisms are important to deal with unneeded
464 nutrients (e.g., Zanutto *et al.*, 1993; Trier & Mattson, 2003). This study examines waste nitrogen
465 production, but not the metabolism of carbohydrates. Waste nitrogen production by *L. dispar* is
466 proportional to the amount of protein metabolized, as found previously in larvae feeding on red
467 oak (Barbehenn *et al.*, 2014). This could either indicate that protein is in excess supply in
468 immature leaves or that waste nitrogen products must be excreted in larger amounts as larvae
469 metabolize more protein. The strong association between protein assimilation rates and growth
470 rates in larvae feeding on all leaf ages suggests that they effectively utilized the protein for
471 growth at all protein levels found in poplar and oak, rather than facing an excess of protein in
472 immature leaves. Instead, the production of waste nitrogen may largely result from the mismatch
473 between the amino acid profiles of foliar and larval protein (Barbehenn *et al.*, 2013b), i.e.,
474 assimilated amino acids that do not match the pattern required for insect protein synthesis must
475 be metabolized, resulting in the excretion of waste nitrogen products in proportion to ingested
476 protein.

477
478 In conclusion, the results of the current study show that protein and carbohydrate can be
479 assimilated efficiently by *L. dispar* caterpillars feeding on the leaves of poplar trees, regardless
480 of leaf maturity. Leaf development under wind stress does not significantly alter these findings.
481 Instead, the negative impacts of feeding on older (basal) leaves are related to the decreased levels

482 of certain nutrients, such as protein, and to decreased consumption rates. Together, these factors
483 limit nutrient assimilation rates, and ultimately growth. Further work is needed to examine the
484 potential importance of leaf toughness and compounds that act as antifeedants or toxins in
485 mature leaves. Finally, the mechanisms that limit protein extractability from the mature leaves
486 of some tree species are unknown. It is speculated that the chemical and structural properties of
487 cell walls may explain the variation between the extractability of protein from leaf tissues, but
488 the authors are unaware of any work that bears on this topic. Additional studies are needed to
489 compare nutrient availabilities from the immature and mature leaves of a wider range of tree
490 species to determine whether species in the Salicaceae are unusual, or whether there are general
491 patterns across taxa, such as between indeterminate and determinate tree species. These studies
492 will provide a better understanding of the physiological mechanisms underlying the variation in
493 host plant quality for insects such as *L. dispar*.

494

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501

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634 **Figure legends**

635 **Fig. 1.** Fibre composition of immature and mature poplar leaves (*Populus alba* x *P. tremula*)
636 grown without or with wind stress. Fibre components are expressed as a percentage of the total
637 dry weight of the leaf, and sum to equal total fibre (NDF). DW = dry weight.

638
639 **Fig. 2.** Association between protein : carbohydrate ratios in immature and mature poplar leaves
640 and the ratios of protein : carbohydrate assimilation rates (PAR:CAR or RPAR:RCAR) in larvae
641 of the gypsy moth *Lymantria dispar*. ●, leaf age experiment; ◆, wind-stress experiment; ▲,
642 R.V. Barbehenn, unpublished observations. $r = 0.97$, $P < 0.001$.

643
644 **Fig. 3.** Particle size distributions in the foreguts of *L. dispar* larvae after feeding on immature,
645 recently mature, and mature poplar leaves.

646
647 **Fig. 4.** Protein levels remaining in rinsed food particles from the posterior half of the midgut and
648 in the frass from *Lymantria dispar* larvae after feeding on immature, recently mature, and mature
649 poplar leaves (separate experiments). Bars show mean \pm standard error, and letters above bars
650 that differ within each series designate $P < 0.05$.

651

Table 1. Nutritional quality of immature and mature poplar leaves (*Populus alba* x *P. tremula*)¹

Leaf age	Protein (% DW)	Carbohydrate ² (% DW)	Water (% FW)	Fibre (% DW)
Immature	24.2 ± 0.7 ^b	8.80 ± 1.41	76.0 ± 0.3 ^b	26.1 ± 1.4 ^a
Recently mature	21.4 ± 1.4 ^a	10.52 ± 0.17	72.6 ± 0.7 ^a	26.1 ± 1.8 ^a
Mature	17.3 ± 0.7 ^a	9.39 ± 0.73	72.1 ± 0.7 ^a	31.2 ± 0.6 ^b
	<i>P</i> = 0.010	<i>P</i> = 0.203	<i>P</i> = 0.006	<i>P</i> = 0.017

¹Leaf ages represent LPI numbers 10-12, 18-20, and 29-30, respectively. Data are presented as mean ± SE, with *n* = 3 replicate days/leaf age for all measurements. Summary statistics followed by different letters are significantly different (*P* < 0.05).

²Carbohydrate includes sugars (87-92% of total) and starch.

Table 2. Consumption and assimilation of protein from immature and mature hybrid poplar leaves (*Populus alba* x *P. tremula*) and growth rates of *Lymantria dispar* larvae feeding on them.

Leaf age	Relative consumption rate (mg mg ⁻¹ day ⁻¹)	Assimilation efficiency (%)	Relative protein consumption rate (mg mg ⁻¹ day ⁻¹)	Protein assimilation efficiency (%)	Relative protein assimilation rate (mg mg ⁻¹ day ⁻¹)	Relative growth rate (mg mg ⁻¹ day ⁻¹)
Immature	3.14 ± 0.12 ^b	40.5 ± 0.6 ^b	0.755 ± 0.043 ^c	81.5 ± 0.8 ^a	0.614 ± 0.036 ^c	0.870 ± 0.054 ^b
Recently mature	2.43 ± 0.10 ^a	34.9 ± 1.2 ^a	0.517 ± 0.007 ^b	78.2 ± 0.7 ^a	0.404 ± 0.024 ^b	0.586 ± 0.030 ^a
Mature	2.37 ± 0.09 ^a	35.5 ± 0.9 ^a	0.408 ± 0.021 ^a	74.4 ± 1.1 ^a	0.304 ± 0.015 ^a	0.497 ± 0.027 ^a
	<i>P</i> < 0.001	<i>P</i> = 0.002	<i>P</i> < 0.001	<i>P</i> = 0.187	<i>P</i> < 0.001	<i>P</i> < 0.001

Leaf ages represent LPI numbers 10-12, 18-20, and 29-30, respectively. Data are presented as mean ± SE, *n* = 13-15 for all measurements. Assimilation efficiency = (mass ingested – mass egested) / mass ingested * 100.

Table. 3 Consumption and assimilation of carbohydrates from immature or mature hybrid poplar leaves by *Lymantria dispar* larvae

Leaf age	Relative carbohydrate consumption rate (mg mg ⁻¹ day ⁻¹)	Carbohydrate assimilation efficiency (%)	Relative carbohydrate assimilation rate (mg mg ⁻¹ day ⁻¹)
Immature	0.288 ± 0.018 ^b	83.8 ± 0.5 ^a	0.241 ± 0.016 ^{ab}
Recently mature	0.268 ± 0.012 ^b	87.1 ± 0.8 ^b	0.234 ± 0.011 ^b
Mature	0.223 ± 0.011 ^a	87.2 ± 0.7 ^b	0.195 ± 0.009 ^a
	<i>P</i> = 0.032	<i>P</i> = 0.004	<i>P</i> = 0.073

Leaf ages represent LPI numbers 10-12, 18-20, and 29-30, respectively. Summary statistics followed by different letters are significantly different (*P* < 0.05). Assimilation efficiency = (mass ingested – mass egested) / mass ingested * 100.

Table 4. Waste nitrogen excretion by *Lymantria dispar* larvae that fed on immature or mature poplar leaves

Leaf age	Uric acid (% DW)	Ammonium salts (% DW)	Total waste nitrogen (%DW)
Immature	0.228 ± 0.017 ^a	1.03 ± 0.11 ^b	1.257 ± 0.116 ^b
Recently mature	0.196 ± 0.017 ^a	0.75 ± 0.08 ^a	0.959 ± 0.097 ^a
Mature	0.219 ± 0.023 ^a	0.60 ± 0.09 ^a	0.879 ± 0.084 ^a
	<i>P</i> = 0.529	<i>P</i> = 0.012	<i>P</i> = 0.034

Data are presented as mean ± SE, *n* = 8-15/date for all measurements. Summary statistics followed by different letters are significantly different (*P* < 0.05).

Table 5. Nutritional quality of immature and mature poplar leaves grown without or with wind stress

Treatment	Leaf age	Protein (%DW)	Carbohydrate (%DW)	Water (% FW)	Fibre (%DW)
Control	Immature	26.1 ± 0.6 ^b	7.2 ± 0.7 ^{ab}	69.7 ± 0.7 ^a	24.4 ± 0.3 ^{ab}
Control	Mature	19.7 ± 1.1 ^a	8.2 ± 0.5 ^b	71.1 ± 1.5 ^a	24.2 ± 0.1 ^a
Wind	Immature	26.3 ± 0.8 ^b	5.8 ± 0.2 ^a	69.7 ± 0.6 ^a	25.1 ± 0.3 ^{ab}
Wind	Mature	19.9 ± 0.2 ^a	7.3 ± 0.6 ^{ab}	71.7 ± 1.6 ^a	25.4 ± 0.4 ^b
Treatment		<i>P</i> = 0.757	<i>P</i> = 0.098	<i>P</i> = 0.845	<i>P</i> = 0.031
Leaf age		<i>P</i> = 0.001	<i>P</i> = 0.079	<i>P</i> = 0.370	<i>P</i> = 0.783
Treatment x leaf age		<i>P</i> = 0.993	<i>P</i> = 0.693	<i>P</i> = 0.862	<i>P</i> = 0.426

Leaf ages represent LPI numbers near 15 and 25, respectively. Data are presented as mean ± SE, with only *n* = 2 replicate dates/treatment and leaf age. Summary statistics followed by different letters are significantly different (*P* < 0.05).

Table 6. Consumption and assimilation of protein and growth of *Lymantria dispar* larvae on immature or mature leaves of poplar grown without or with wind stress

Treatment	Leaf age	Relative consumption rate (mg mg ⁻¹ day ⁻¹)	Total assimilation efficiency (%)	Relative protein consumption rate (mg mg ⁻¹ day ⁻¹)	Protein assimilation efficiency (%)	Relative protein assimilation rate (mg mg ⁻¹ day ⁻¹)	Relative growth rate (mg mg ⁻¹ day ⁻¹)
Control	Immature	1.21 ± 0.04 ^b	36.6 ± 0.7 ^b	0.315 ± 0.010 ^c	76.1 ± 0.5 ^c	0.245 ± 0.010 ^c	0.189 ± 0.011 ^b
Control	Mature	1.18 ± 0.06 ^b	40.2 ± 0.8 ^c	0.232 ± 0.012 ^b	72.1 ± 1.3 ^{ab}	0.161 ± 0.006 ^b	0.179 ± 0.013 ^b
Wind	Immature	1.11 ± 0.04 ^b	34.2 ± 0.7 ^a	0.291 ± 0.010 ^c	75.4 ± 0.8 ^{bc}	0.234 ± 0.009 ^c	0.181 ± 0.009 ^b
Wind	Mature	0.86 ± 0.03 ^a	38.4 ± 1.0 ^{ab}	0.172 ± 0.007 ^a	68.8 ± 0.9 ^a	0.117 ± 0.004 ^a	0.140 ± 0.006 ^a
Treatment		$P < 0.001$	$P = 0.008$	$P < 0.001$	$P = 0.155$	$P = 0.003$	$P = 0.080$
Leaf age		$P < 0.001$	$P < 0.001$	$P < 0.001$	$P = 0.004$	$P < 0.001$	$P = 0.007$
Treatment x leaf age		$P = 0.008$	$P = 0.979$	$P = 0.032$	$P = 0.790$	$P = 0.007$	$P = 0.099$

Leaf ages represent LPI numbers near 15 and 25, respectively. Data are presented as mean ± SE, $n = 10-14$ for all measurements. Summary statistics followed by different letters are significantly different ($P < 0.05$). Assimilation = (mass ingested – mass egested).

Table 7. Consumption and assimilation of carbohydrates by *Lymantria dispar* larvae on immature or mature leaves of poplar grown without or with wind stress

Treatment	Leaf age	Relative carbohydrate consumption rate (mg mg ⁻¹ day ⁻¹)	Carbohydrate assimilation efficiency (%)	Relative carbohydrate assimilation rate (mg mg ⁻¹ day ⁻¹)
Control	Immature	0.085 ± 0.003 ^b	90.0 ± 0.6 ^a	0.076 ± 0.003 ^b
Control	Mature	0.094 ± 0.004 ^c	92.8 ± 0.3 ^b	0.087 ± 0.004 ^c
Wind	Immature	0.061 ± 0.002 ^a	88.7 ± 0.6 ^a	0.054 ± 0.002 ^a
Wind	Mature	0.061 ± 0.003 ^a	92.7 ± 0.3 ^b	0.057 ± 0.003 ^a
Treatment		<i>P</i> < 0.001	<i>P</i> = 0.143	<i>P</i> < 0.001
Leaf age		<i>P</i> = 0.073	<i>P</i> < 0.001	<i>P</i> = 0.005
Treatment x leaf age		<i>P</i> = 0.119	<i>P</i> = 0.210	<i>P</i> = 0.120

Leaf ages represent LPI numbers near 15 and 25, respectively. Data are presented as mean ± SE, *n* = 10-14 for all measurements except carbohydrate assimilation efficiency and P:C assimilation ratios (*n* = 9-10). Summary statistics followed by different letters are significantly different (*P* < 0.05). Assimilation = (mass ingested – mass egested).

Fig. 1

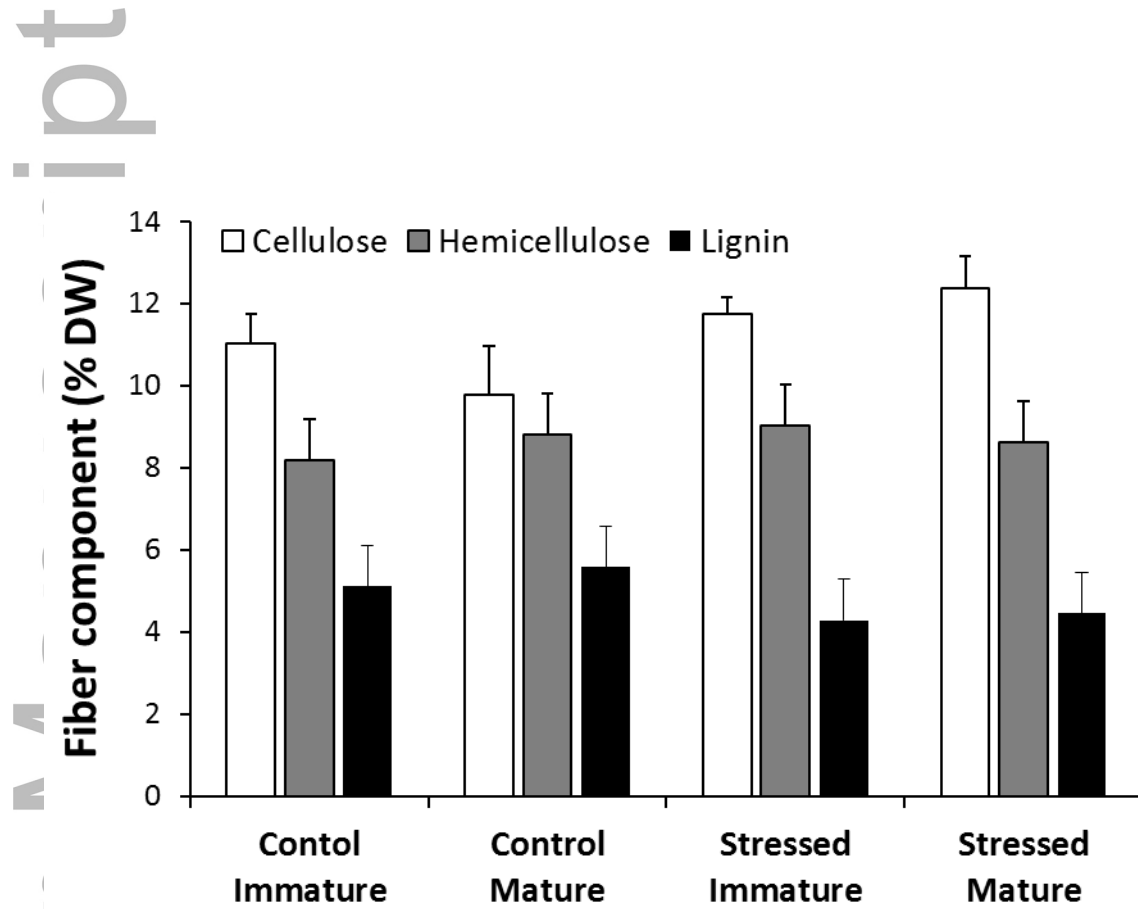


Fig. 2

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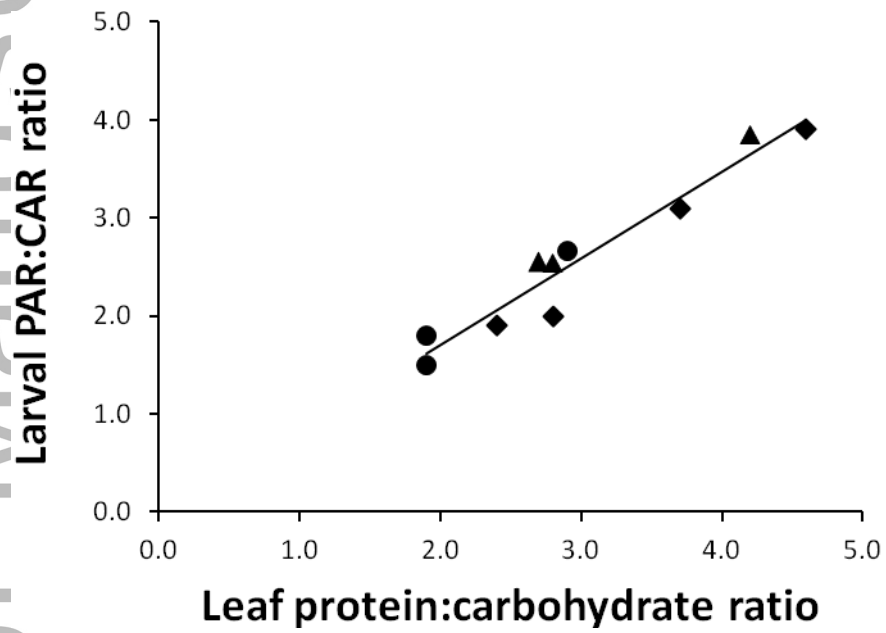


Fig. 3

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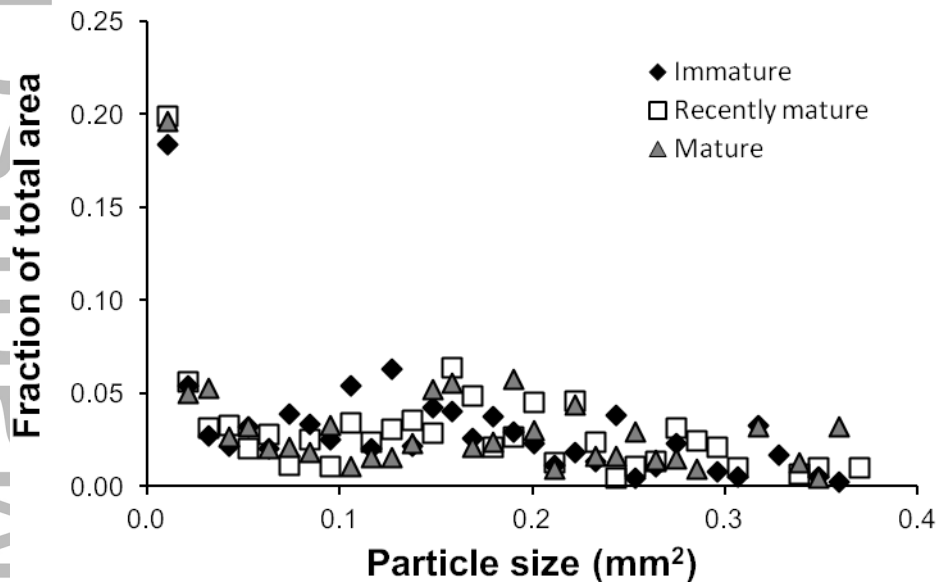


Fig. 4

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