

SHORT PAPER

Prevalence and geographical distribution of *Papio hamadryas papillomavirus 1* (PhPV1) in Kenyan baboons

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Abstract

Papio hamadryas papillomavirus (PhPV) 1, 2, and 3, are Alphapapillomaviruses that have been detected in Kenyan Olive baboons but the distribution is unknown. Therefore, cervical screening for PhPV1 was performed in baboons from various areas in Kenya using a nested polymerase chain reaction. The prevalence rate was 33%.

KEYWORDS

cancer, distribution, PCR, PhPV1

1 | INTRODUCTION

Human papillomavirus (HPV) is an Alphapapillomavirus (α PV) that is the most common sexually transmitted viral infection and main cause of cervical intraepithelial neoplasia (CIN) and cervical cancer in women.¹ In animals, *bovine papillomavirus*² and cotton rabbit papillomavirus³ have been used as model systems to study the biology of HPV while experimental induction of papillomas and their neoplastic progression has been demonstrated and reproduced in cattle⁴ and rabbits.⁵ However, none of these models recapitulate the natural progression

of cervical infection leading over time to neoplastic transformation as seen in HPV-infected women.

In contrast, naturally occurring genital α PVs have been described so far in five nonhuman primate (NHP) species,⁶ offering the opportunity for study in an animal model physiologically and phylogenetically closer to humans. Most notably, cervical infection with the potential for neoplastic transformation has been shown in macaques⁷⁻⁹ and baboons.¹⁰ Twenty-one naturally occurring NHP α PVs have been reported, as reviewed by Rector and Van Ranst.⁶ In Olive baboons, high-grade (HSIL) and low-grade (LSIL) squamous intraepithelial lesions

were associated with infection by PhPV1.¹⁰ In the pygmy chimp and colobus monkey, α PV-associated oral epithelial hyperplasia has been reported.¹¹ As in women, dysplasia or neoplastic transformations are relatively rare outcomes of infection.

The presence of HSIL and LSIL with naturally occurring PhPV1 in Olive baboons (*Papio anubis*) is documented¹⁰ and offers a promising animal model for HPV. The baboon has several advantages and is a unique preclinical model for research in human reproduction.^{12,13}

Apart from our study,¹⁰ previous efforts to identify α PV in other baboon colonies were unsuccessful.¹⁴ We therefore sought to determine the prevalence of PhPV1 infection in baboons from various areas in Kenya.

2 | MATERIALS AND METHODS

A total of 104 baboons were sampled for this study from March 2013 to January 2015. These were a subset of a total IPR population of approximately 200 postadolescent female baboons. Criteria used to select animals for sampling consisted of convenience (eg, sedation for veterinary reasons or for other experiments during the study period), sex (females sampled), and weight (postadolescent or > 6.0 kgs, with heaviest being 26.4 kgs).

Ethical approval for this work was given by the Institute of Primate Research (IPR) Institutional Review committee (IRC/09/13). The animal care and use program at the IPR is based on the Guide for the Care and Use of Laboratory Animals, National Research Council, National Academy Press, Washington D.C., 2011 and in line with the 3R principles. Cervical swabs were collected from each female baboon under ketamine/xylazine anesthesia. DNA extraction was performed using the POWERSOIL[®] DNA Isolation kit (Mo Bio Laboratories Inc., Carlsbad, CA, USA).¹⁵

Polymerase chain reaction (PCR) amplification was performed using primers based on the 8000 bp genomic sequence of PhPV1 which was

TABLE 1 Proportion, geographic origins, and *Papio hamadryas* papillomavirus (PhPV1) status of Olive baboons from the Institute of Primate Research, Kenya

	PhPV1 negative	PhPV1 positive	Percentage positive
Laikipia, ADC Mutara Ranch	27	10	27
Aberdares, Mahinga	17	8	32
Aberdares, Muringa	5	1	17
Aberdares, Country Club	5	4	44
Yatta NYS Field Unit	5	2	29
Aberdares, Lamuria	4	3	42
Amboseli Jean Altman troops	2	0	0
Colony borne-IPR	2	0	0
Ngurumani	1	1	50
Nairobi National Park	1	4	80
Aberdares, Mweiga	1	1	50
Total	70	34	33

TABLE 2 Distribution of the percentage of *Papio hamadryas* papillomavirus 1 (PhPV1) positive baboon by body weight at Institute of Primate Research, Kenya

Weight range	6.0-7.9	8.0-9.9	10.0-11.9	12.0-13.9	14.0-15.9	16.0-17.9	18.0-19.9	20.0-21.9	22.0-23.9	24.0-25.9	26.0-27.9	Unknown weight ^a
Total number	4	11	20	31	27	7	1	0	0	0	1	2
Positive	0	1	4	14	13	1	1	0	0	0	0	0
Percentage of positive cases	0	9	20	45	48	14	1	0	0	0	0	0

^aTwo baboons caught at Amboseli and released had no weights taken.

previously sequenced¹⁰ and is publicly available as Gene bank accession number JF304764 (type 1 isolate Mac2085). Primer sequences were as follows: primary amplification forward primer (Fr2Fout: GGGTATGACGTGAGGCAGTT); primary amplification reverse primer (Fr1Rout: TACGCAACTTTGGTGGTTCA); secondary amplification forward primer (Fr2Fin: TGGCATAGGGTTTCATGAGC); secondary amplification reverse primer (Fr1Rin: TGCAATGTGGCTCAATAAGG).

Nine microliters of each secondary PCR product was run on a 1.5% agarose gel.

Validation of the nested PCR strategy for this study was performed prior to DNA extraction from sample swabs using samples from animals previously demonstrated as positive or negative for PhPV1.¹⁰ Sequencing was performed to the products of this validation samples after purification of the amplicon using GENEJET[®] Genomic DNA Purification Kit (Pittsburgh, PA, USA) and was sequenced using Sanger sequencing with the ABI V3.1 Big dye kit on the ABI 3500XL genetic analyzers at Inqaba Company, South Africa.

3 | RESULTS

The geographical origins of all the animals tested and their respective prevalence of the PhPV1 status are shown in Table 1. A total of 34 of 104 (33%) baboons were positive for PhPV1 using the nested PCR primers.

The highest percentage of PhPV1 positive baboons, by body weight, was between 12.0 kgs and 15.9 kgs, corresponding to sexually mature animals. At this weight range, almost 50% were positive to PhPV1 (Table 2).

4 | DISCUSSION

This study documents the geographical distribution of PV in Kenyan baboons which was previously unknown. The overall prevalence of PhPV1 infection in this study was 34/104 (33%) showing that Kenyan baboons naturally harbor PV.

This current study also revealed that most infections appeared within the weight range of 12–16 kgs. This weight range is consistent with sexual maturity and suggests that this time period is a risk factor for PhPV1 infection. Indeed, baboons menarche occurs between 4 and 5 years of age in wild populations and at about 3.5 years in captivity,¹⁶ corresponding to a body weight 8–10 kgs at IPR. This pattern of distribution of HPV has also been observed in women, where HPV prevalence peaks at younger ages (puberty to <25 years).¹

In conclusion, the findings of this study which showed that there is a natural occurring PV infection, as well as our previous studies showing association of baboon PV with HSIL and LSIL,¹⁰ is a strong support for the baboon model in understanding of aPV pathogenesis as well use in future studies for interventional strategies. However, more studies on genomic diversity and establishment of an experimental PV baboon infection model are recommended.

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There is no conflict of interest from among the authors.

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REFERENCES

- Bruni L, Diaz M, Castellsagué M, Ferrer E, Bosch FX, de Sanjosé S. Cervical *Human Papillomavirus* Prevalence in 5 Continents: Meta-Analysis of 1 Million Women with Normal Cytological Findings. *J Infect Dis*. 2010;202:1789–1799.
- Borzacchiello G, Roperto F. *Bovine Papillomaviruses*, papillomas and cancer in cattle. *Vet Res*. 2008;39:45.
- Breiburd F, Kirnbauer R, Hubbert NL, et al. Immunization with virus like particles from cottontail rabbit Papillomavirus (CRPV) can protect against experimental CRPV infection. *J Virol*. 1995;69:3959–3963.
- Campo MS, O'Neil BW, Barron RJ, Jarrett WF. Experimental reproduction of the papilloma-carcinoma complex of the alimentary canal in cattle. *Carcinogenesis*. 1994;15:1597–1601.
- Campo MS. Animal models of Papillomavirus pathogenesis. *Virus Res*. 2002;89:249–261.
- Rector A, Van Ranst M. Animal papillomaviruses. *Virology*. 2013;445:213–223.
- Chan SY, Bernard HU, Ratterree M, Birkebak TA, Faras AJ, Ostrow RS. Genomic diversity and evolution of Papillomaviruses in rhesus monkeys. *J Virol*. 1997;71:4938–4943.
- Chen Z, van Doorslaer K, DeSalle R, et al. Genomic diversity and interspecies host infection of alpha12 *Macaca fascicularis* papillomaviruses (MfPVs). *Virology*. 2009;393:304–310.
- Roberts JN, Kines RC, Katki HA, Lowry DR, Schiller JT. Effect of Pap smear collection and Carrageenan on cervicovaginal *human papillomavirus* – 16 infection in a macaque model. *J Natl Cancer Inst*. 2011;103:737–743.
- Bergin IL, Bell JD, Chen Z, et al. Novel Genital Alphapapillomaviruses in Baboons (*Papio hamadryas anubis*) With Cervical Dysplasia. *Vet Pathol*. 2013;50:200–208.
- Van Ranst MA, Fuse H, Sobis H, et al. A papillomavirus related to HPV type 13 in oral focal epithelial hyperplasia in the pygmy chimpanzee. *J Oral Pathol Med*. 1991;20:325–331.
- D'Hooghe TM, Kyama CK, Mihalyi AM, Chai D, Falconer H, Mwenda JM. The baboon model for translational research in endometriosis. *Reprod Sci*. 2008;16:152–161.
- Kyama CM, Overbergh L, Mihalyi A, et al. Effect of recombinant human Tumour Necrosis Factor Binding Protein -1 (r-hTBP-1) and GnRH antagonist on mRNA expression of inflammatory cytokines, adhesion and growth factors in endometrium and endometriosis tissues from baboons. *Fertil Steril*. 2007;89:1306–1313.
- Wonderly DE, Chan PJ, Cseh S, Jacobson JD, Bailey L. Analysis of papillomavirus consensus L1 gene in a closed colony of baboons (*Papio anubis*). *Am J Obstet Gynecol*. 2000;182:1016–1017.
- Bell JD, Bergin IL, Harris LH, et al. The effects of a single cervical inoculation of *Chlamydia trachomatis* on the female reproductive tract of the baboon. *J Infect Dis*. 2011;204:1305–1312.
- Altmann J, Hausfater G, Altmann S. Determinants of reproductive success in savannah baboons, *Papio cynocephalus*. In: Clutton-Brock TH, ed. *Reproductive Success*. Chicago: Univ. of Chicago Press; 1988: pp. 403–418.