ORIGINAL ARTICLE

Spectrum of disease associated with partial lipodystrophy: lessons from a trial cohort

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Summary

Context Partial lipodystrophy (PL) is associated with metabolic co-morbidities but may go undiagnosed as the disease spectrum is not fully described.
Objective The objective of the study was to define disease spectrum in PL using genetic, clinical (historical, morphometric) and laboratory characteristics.
Design Cross-sectional evaluation.
Participants Twenty-three patients (22 with familial, one acquired, 78·3% female, aged 12–64 years) with PL and non-alcoholic fatty liver disease (NAFLD).
Measurements Genetic, clinical and laboratory characteristics, body composition indices, liver fat content by magnetic resonance imaging (MRI), histopathological and immunofluorescence examinations of liver biopsies.
Results Seven patients displayed heterozygous pathogenic variants in *LMNA*. Two related patients had a heterozygous, likely pathogenic novel variant of *POLD1* (NM002691·3: c.3199 G>A; p.E1067K). Most patients had high ratios (>1·5) of percentage fat trunk to percentage fat legs (FMR) when compared to reference normals. Liver fat quantified using MR Dixon method was high (11·3 ± 6·3%) and correlated positively with haemoglobin A1c and triglycerides while leg fat by dual-energy X-ray absorptiometry (DEXA) correlated negatively with triglycerides.

In addition to known metabolic comorbidities; chronic pain (78·3%), hypertension (56·5%) and mood disorders (52·2%) were highly prevalent. Mean NAFLD Activity Score (NAS) was 5 ± 1 and 78·3% had fibrosis. LMNA-immunofluorescence staining from select patients (including one with the novel *POLD1* variant) showed a high degree of nuclear atypia and disorganization.
Conclusions Partial lipodystrophy is a complex multi-system disorder. Metabolic parameters correlate negatively with extremity fat and positively with liver fat. DEXA-based FMR may prove useful as a diagnostic tool. Nuclear disorganization and atypia may be a common biomarker even in the absence of pathogenic variants in LMNA.

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Introduction

Partial lipodystrophy (PL) is a cluster of rare heterogeneous diseases characterized by selective loss of adipose tissue, which places patients at risk of common conditions typically associated with metabolic syndrome.[1–3] Commonly associated features are insulin resistance and diabetes, hypertriglyceridaemia and non-alcoholic fatty liver disease (NAFLD).[4] PL may be acquired or inherited and pathogenic variants in multiple genes have been identified as causative for inherited PL including *LMNA*, *PPARG*, *PLIN1*, *AKT2*, *CIDEC*[2] and more recently *LIPE*[5] and *ADRA2A*.[6]

One obstacle in development of novel treatment strategies for these disorders has been the lack of documentation of the differentiating features of these syndromes from more common forms of truncal obesity with metabolic syndrome and Type 2 diabetes. This was noted during the Advisory Meeting of the FDA for

approval of Metreleptin for the treatment of lipodystrophy syndromes (http://www.fda.gov/downloads/AdvisoryCommittees/ CommitteesMeetingMaterials/Drugs/EndocrinologicandMetabolic DrugsAdvisoryCommittee/UCM379648.pdf). Another problem is the lack of objective description of natural history and disease burden. Stated differently, are patients with PL facing different clinical problems, comorbidity risk and mortality compared to patients with more common metabolic disorders such as Type 2 diabetes and truncal obesity? If the answers to these questions are yes, then the pursuit of specific treatments for these disorders will be of high value despite the small number of patients afflicted with PL.

Our study is one of the first in the field to specifically describe the baseline drug-naïve state of patients participating in a drug trial. Accumulation of data such as these presented here may allow more precise definition of the disease state, possibly helping the development of specific diagnostic criteria or disease severity scores. We also hope to increase recognition of similar patients by providing extensive clinical data to endocrinologists.

Methods

We have been recruiting patients with PL for an open-label study conducted at the University of Michigan funded by NIDDK (approved by University of Michigan IRBMED) to evaluate the molecular effects of an investigational drug on treating liver disease associated with PL (ClinicalTrials.gov identifier: NCT01679197, HUM00058708). Here, we describe baseline characteristics of 23 patients who completed screening and study initiation procedures but have not received any investigational drug. Diagnosis of PL was based on physician assessment with evidence of selective loss of body fat.[3] Patients with both inherited and acquired forms of PL were included, excluding lipodystrophy associated with human immunodeficiency virus (HIV). Patients entering this study had a liver ultrasound showing the presence of fatty liver disease or prior biopsy showing evidence of non-alcoholic steatohepatitis (NASH), no evidence of other forms of liver disease and alcohol consumption of <40 g/week.

After informed consent was obtained from all patients or guardians, physical examinations were performed and previous medical records were examined to verify historical details. Blood samples after overnight fast were analyzed for estimating leptin levels using a commercial enzyme linked immunosorbent assay (ELISA) kit (EMDMilipore, Billerica, MA,USA). Haemoglobin A1c, fasting lipids and hepatic function tests were determined in the Clinical Pathology Laboratory of University of Michigan using auto-analyzer equipment.

Clinical and research resources were used to determine the molecular basis of disease in patients (described in Appendix S1). Body composition was evaluated using anthropometric measurements, including skin thickness and waist and hip circumferences using standardized techniques. Fat and lean body mass were estimated using dual X-ray absorptiometry (DEXA; GE Lunar Prodigy, model PA +41744, Madison, WI, USA).[7] Ratio of percent fat mass of the trunk to percent fat mass of the legs (FMR) was calculated.

For determination of hepatic fat content, magnetic resonance (MR) imaging using quantitative multi-echo Dixon method and multi-echo MR spectroscopy have been employed (method described in Appendix S1 and Figure S1). Transcutaneous liver biopsies were performed and total NASH scores were calculated from histopathological examination using the simplified clinical criteria as presented in Table S1, which is a modified and clinically applied version of NIH NASH Clinical Network Criteria.[8] NASH score equals the sum of scores for steatosis (range, 0–3), lobular inflammation (range, 0–3), hepatocellular ballooning (range, 0–2) and fibrosis (range, 0–4). NAFLD activity score (NAS) includes these parameters, but without fibrosis. In addition, selective liver biopsy specimens were processed for lamin A/C immunofluorescence staining and then scored in a blinded fashion (methods described under Appendix S1).

Statistical analyses were performed using GRAPHPAD Prism version 6·05 (LaJolla, CA, USA) and SAS version 9·2 (Cary, NC, USA). Correlations were investigated by calculating Pearson's correlation coefficient. Two-tailed P-value was calculated, and $P < 0.05$ was considered statistically significant. For quantitation of lamin immunofluorescence data, statistical significance was determined via one-way ANOVA followed by Tukey's post-hoc test at a threshold of $P < 0.05$.

Results

Patient characteristics and genetic evaluation

Twenty-three patients with PL were enrolled who were 12–64 years old (median age, 43 years): 18 female and five male. The majority, 20 patients were Caucasian, two Hispanic and one African-American. Summary characteristics are shown in Table 1, and detailed clinical data can be found in Tables S2–S5. Of known genes, *LMNA* variants were quite common, identified in seven patients. Of these seven patients, five had a heterozygous pathogenic variant at position 482, the hot spot. Four of these patients harboured R482Q pathogenic variant (two sisters, patients 13 and 20, and two unrelated patients, 17 and 18). One patient (patient 21) harboured the previously reported R482W pathogenic variant and was also diagnosed with muscular dystrophy.[9] Patient 10 also harboured a previously reported R60G pathogenic variant[10] and had a history of dilated non-ischaemic cardiomyopathy requiring heart transplant at age 25. Patient 22 was heterozygous for *LMNA* R349W and had unique clinical features including loss of fat from the face, hands and feet, a large dorsocervical fat pad, proteinuria, cardiomyopathy (believed to be both primary and ischaemic), infertility, hyperandrogenism and alopecia (Fig. 1, Tables S2-S5). Another interesting laboratory abnormality in patient 22 is low alkaline phosphatase level, typically associated with progeria.[11,12] There have been two other published instances of this mutation, one in a male patient with similar fat distribution, but without the exaggerated dorsocervical fat pad, diabetes, or other features described here.[13] In addition, a large kindred with focal segmental glomerulosclerosis and proteinuria in association with PL has been reported.[14]

Table 1. Demographics and characteristics

Female/Male, n	18/5
Age (years), mean (range)	43 (12–64)
Race, n (%)	
White	20 (87·0)
Black	1 (4·3)
Other	2 (8·7)
Lipodystrophy subtype, n (%)	
Familial partial lipodystrophy	22 (95·7)
Acquired partial lipodystrophy	1 (4·3)
Mutation if known, n	
LMNA	7
POLD1	2, subjects are related
Body weight (kg), mean (SD)	77·2 (21·4)
Body mass index (kg/m^2), mean (SD)	27·3 (5·9)
Waist-to-hip ratio, mean (SD)	0·99 (0·08)
Skin fold thickness, anterior mid-thigh* (mm), mean (SD), [range]	12·1 (7·7) [4–29]
%Fat trunk / %Fat legs*, mean (SD)	1·78 (0·53)
MR spectroscopy liver fat (%), mean (SD)	13·8 (6·1)
Dixon method MR liver fat (%), mean (SD)	11·9 (6·3)
Haemoglobin A1c (%), mean (SD)	8·6 (1·9)
Leptin level (ng/ml), mean (SD)	22·3 (16·6)
Fasting triglycerides (mg/dl), mean (SD)	1058 (1745)
AST (IU/l), mean (SD)	42 (28)
ALT (IU/l), mean (SD)	52 (37)
Haemoglobin (g/dl), mean (SD)	13·8 (1·3)
Platelets (K/μl), mean (SD)	265 (55)
Creatinine (mg/dl),mean (SD)	0·67 (0·22)
CK (IU/l), mean (SD)	145 (101)

MR, magnetic resonance; AST, aspartate aminotransferase; ALT, alanine aminotransferase; CK, creatine kinase.
*Excluding patient with acquired partial lipodystrophy.

Two related patients (mother and daughter, 6 and 7) had a variant in Exon 26 of *POLD1* (NM002691·3: c.3199G>A; p.E1067K, Fig. 1, Figures S2 and S3 and Tables S2–S5) which we classified as a variant of unknown significance, which is most likely pathogenic. Two patients (four and nine) did not harbour mutations in the known 15 LD genes (see Appendix S1) interrogated with whole exome sequencing (WES). WES for the other 11 patients is currently in progress but they did not harbour *LMNA* p.R482 variants using specific sequencing for this locus as described in Appendix S1.

Body composition

All but one patient had a pattern of fat loss consistent with that seen in familial partial lipodystrophy (FPLD) in which there is fat atrophy of the upper and lower extremities as well as the pelvis, with relative fat sparing or accumulation in the neck, trunk and abdomen. One patient had acquired partial lipodystrophy with fat atrophy in the upper body and preservation of fat in the legs. FMR in this patient was 0·84, while in the other 22 patients, FMR was higher, 1·78 ± 0·53. Using the Third National Health and Nutrition Examination Survey (NHANES III) reference data[15] and the conversion to Lunar equipment previously reported,[16] patients were matched for gender, race

and age and the majority had FMR well above 95% confidence interval of reference median values (Fig. 1). We also compared the FMR obtained from our adult female patients with a group of normal and overweight women with a BMI of 20–30 kg/m^2 as well as a group of female diabetic patients with truncal obesity and fatty liver disease (Figure S4) who were age-matched. A cut-off value of 1·5 would be well above the normal for adult female populations from NHANESIII as well as our two comparator groups with a sensitivity of 86·7% and specificity of 96·8%. There were distinctive patients with substantially higher values, most notably patient 22.

Leg fat mass was negatively correlated with log-transformed triglycerides ($P = 0·0498$, Fig. 2). No correlation was found between leg fat mass and haemoglobin A1c ($R^2 = 0·06$, Fig. 2), or between leg fat mass and liver fat ($R^2 = 0·01$). Mid-thigh skin fold thickness (mean: 12·1 ± 7·7; range, 4–29 mm) was low compared to age-, gender- and race-matched data from NHANES data[17] (mean: 26·3 ± 7·2; range, 12·3–32·4 mm).

Leptin levels (mean: 22·3 ± 16·6, range 4·8–67·1 ng/ml) were variable and in general higher than noted in studies that enrolled only patients with leptin deficiency at the NIDDK,[18] but similar to one of our previously published series.[19] Leptin levels correlated with total fat mass ($P = 0·0071$) and trunk fat ($P = 0·0044$; Figure S5) and did not correlate with HbA1c ($R^2 = 0·00$) or triglyceride levels ($R^2 = 0·08$).

Comorbidities

Prevalence of comorbidities and medications used are shown in Fig. 3. All patients had either diabetes (82·6%) or pre-diabetes (17·4%) with variable degree of control. Mean ± SD haemoglobin A1c was 8·6 ± 1·9%. One patient (patient 14) had Type 1 diabetes with undetectable C-peptide and another patient (patient 15) had positive anti-GAD antibody at baseline. Nearly half (11 patients) had very poor control with haemoglobin A1c above 9%. Nineteen patients (82·6%) used a glucose lowering agent. Ten patients (43·5%) used combination therapy with two or more agents. Insulin was used by 12 patients (52·2%) and 9 of these patients required over 100 units/day. Six patients (26·1%) used highly concentrated U-500 insulin.

Microvascular complications of diabetes were common. Twelve patients (52·2%) had peripheral neuropathy. One patient had a history of chronic inflammatory demyelinating polyneuropathy (CIDP). Six patients had known diabetic retinopathy based on patient report, although the prevalence may be higher as we did not have dilated examination results for all patients. Proteinuria or microalbuminuria was known in 10 patients (43·5%). Two patients had kidney biopsies for clinical reasons, one displaying membranoproliferative glomerulonephritis (patient 6, biopsied for massive proteinuria) and another (patient 22, biopsied for haematuria) displaying thin basement membrane disease. One patient had severe gastroparesis.

Twenty-two of 23 patients had hypertriglyceridaemia (mean, 1058 ± 1745 mg/dl). Six had very high triglycerides over 1000 mg/dl. Most (17 patients) used a lipid lowering agent (Fig. 3). Recurrent pancreatitis occurred in seven of 23 patients
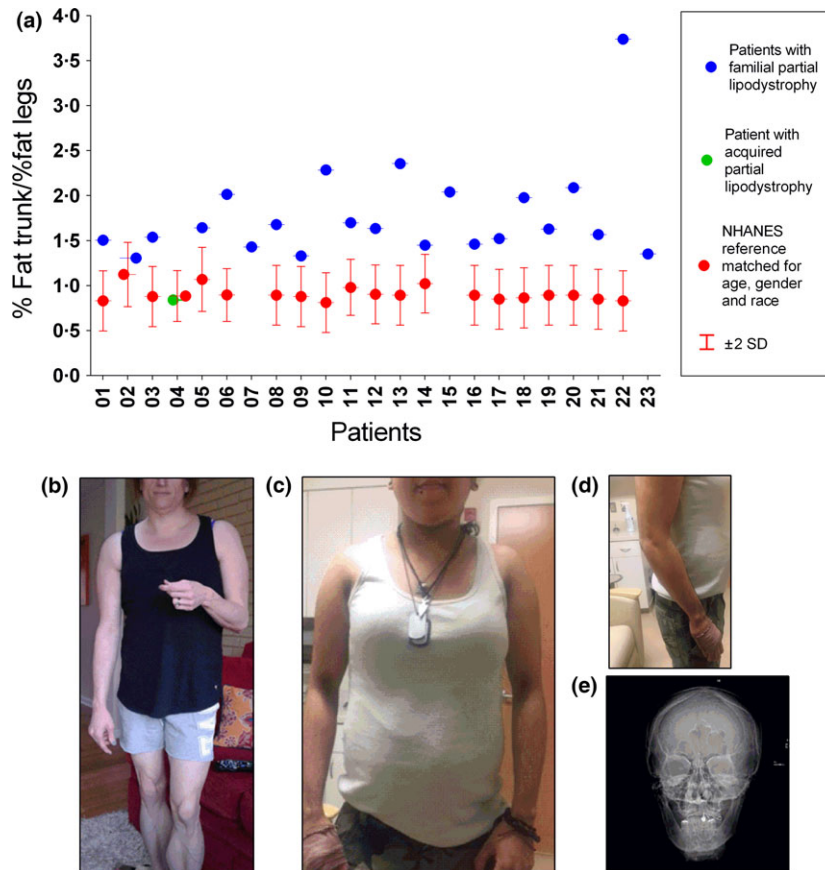
Fig 1 (a) Comparison of %fat trunk/%fat legs (fat mass ratio, FMR) in 23 patients with partial lipodystrophy, one of whom has acquired partial lipodystrophy (patient 4) to age, gender and ethnic group matched reference values (median with 95% confidence interval) from NHANES data.[15] Reference data were not available for paediatric patients (patients 7, 15, 23). (b) Picture of patient 22 who presented with a very high fat mass ratio harbouring a possibly pathogenic variant at position 349 (R349W). Patient displayed loss of fat from face, extremities, including loss of supportive fat from hands and feet, but with excessive fat deposition in the back displaying an exaggerated buffalo hump. Other clinical features included ischaemic and non-ischaemic cardiac disease, PCO-S with severe hyper-androgenism and infertility, alopecia, haematuria (because of thin basement membrane disease on kidney biopsy). Laboratory features included a high testosterone level and a very low alkaline phosphatase level typically associated with progeroid disorders. (c, d) Pictures of patient 7 with a possibly pathogenic *POLD1* variant. This patient and her mother displayed very severe insulin resistance, preservation of subcutaneous fat around the face and neck and even in the abdominal wall. There was a paucity of fat in the extremities. Patients also demonstrated stiffness in joints, elevated CK levels (mother displaying muscle weakness), proteinuria (massive in one leading to kidney biopsy showing membranoproliferative glomerulonephritis). The patient and her mother did not have deafness. (e) X-ray of the narrow mandible for patient 6 with possibly pathogenic *POLD1* variant.

(30·4%). At least three episodes of pancreatitis occurred while triglyceride levels were <800 mg/dl. Two patients who are sisters, with known *LMNA* gene mutations, were found to have pancreatic cystic lesions on magnetic resonance imaging (MRI). One had a single 0·3-cm cystic lesion in the pancreatic neck and the other patient had multiple subcentimeter cystic lesions which may represent side branch intraductal mucinous neoplasms (Figure S6).

Mood disorder requiring medication was identified in 12 patients (52·2%). Depression was the most common psychiatric condition, present in 10 patients (43·5%). Six had anxiety (26·1%), one patient had bipolar disorder, and one had an unidentified psychiatric condition for which he previously used an antipsychotic agent.

Chronic pain was common among our patients. Eighteen patients (78·3%) reported some type of painful condition including neuropathy (52·2%), arthritis (47·8%), chronic back

pain (13·0%), fibromyalgia (17·4%) and myopathy (39·1% had either suspected or confirmed myopathy). One patient had muscular dystrophy.

Other noted conditions include coronary artery disease or myocardial infarction in five patients (22·7%), one patient with ischaemic cardiomyopathy and one patient with *LMNA* mutation and non-ischaemic cardiomyopathy who required a heart transplant. Hypertension was present in 13 patients (56·6%). Seven patients (30·4%) had sleep apnoea. Three (13%) had asthma or emphysema. Nine patients (39·1%) had polycystic ovarian syndrome (PCOS).

Hepatic evaluation

Liver fat quantified using MR spectroscopy and also using MR Dixon method showed fat percentage, mean ± SD of
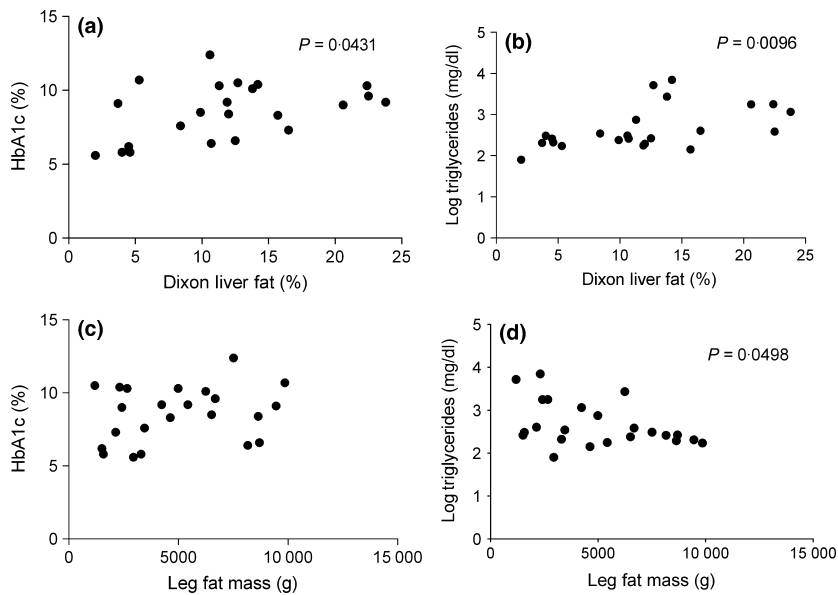
Fig 2 Correlation was noted between metabolic parameters of haemoglobin A1c and liver fat % (a). as well as log transformed triglycerides and liver fat % (b). Haemoglobin A1c was not significantly correlated with leg fat mass (c). A significant correlation was seen between log transformed triglycerides and leg fat mass (d). Leg fat mass and liver fat % did not correlate (data not shown).

$13.8 \pm 6.1\%$ and $11.9 \pm 6.3\%$, respectively (normal <5%).[20] Liver fat was correlated with metabolic parameters. A positive correlation was found between liver fat quantified using MR Dixon method and haemoglobin A1c ($P = 0.0431$) as well as log-transformed triglycerides ($P = 0.0096$) Fig. 2.

Of the 23 patients who underwent liver biopsy, 22 met histopathological criteria for NASH. Total NASH scores of those 22 patients ranged 3–9 (mean, 6 ± 2). NAS in patients who met criteria for NASH ranged 3–8 (mean, 5 ± 1). Eighteen patients (78.3%) had some degree of fibrosis present with variable severity (stage 1a-4). One 12-year-old female patient (patient 15) had stage 4 fibrosis (Fig. 4).

As *LMNA* mutations were common among the FPLD patients, and mice with hepatocyte-specific *LMNA* deletion develop spontaneous steatosis with nuclear alterations and susceptibility to diet-induced steatohepatitis ('unpublished data'; authors: Raymond Kwan, Graham Brady, Maria Brzozowski, Sujith Weerasinghe, Hope Martin, Min-Jung Park, Ram Menon, Xin Tong, Lei Yin, Colin Stewart, M. Bishr Omary), we investigated nuclear envelope integrity from our liver biopsy samples. These early studies were carried out on three patients with *LMNA* pathogenic variants and one patient with a *POLD1* p.E1067K variant. Specimens from two patients with NASH (without lipodystrophy) were used as controls. Immunofluorescence staining of frozen liver sections showed dysmorphic nuclei and lamin disorganization in the patients with *LMNA* pathogenic variants compared with NASH controls (Fig. 5). We also observed abnormal nuclear morphology in the patient with *POLD1* variant, although not to the same extent as the patients with *LMNA* variants.

Discussion

The goal of our study is to provide a complete description of the disease state in a cohort of 23 patients with PL. Lack of clear documentation of clinical manifestations and full appreciation of

disease burden have become an obstacle in therapeutic development for rare metabolic diseases given the significant heterogeneity in presentation.[21] With this report, we are reintroducing the idea of careful morphometric description using DEXA to document the presence of PL. This approach may be adopted widely as a clinical test following further validation. Also, we are reporting a number of important clinical lessons such as the presence of severe disease burden not only because of metabolic disease but also because of neuropsychiatric symptoms, chronic pain and multi-system involvement. It would be useful to collect similar data from a larger number of patients to develop disease severity scores taking into account the entire spectrum of clinical findings.

Our genetic investigations have recapitulated the genetic heterogeneity of PL with multiple patients having different molecular basis for their disease. Several genes have been implicated in various forms of FPLD including *LMNA, PPARG, PLIN1, AKT2 CIDEC, LIPE* and *ADR2A*.[2,5,6,22,23] However, the molecular cause of disease remains unexplained in up to 40% of patients. Lotta *et al.* recently reported enrichment of multiple common variant genes in a cohort of patients formerly classified as FLPD1, suggesting the possibility of polygenic inheritance.[24]

LMNA pathogenic variants are most commonly identified in FPLD and also the most common in our cohort.[22] As novel findings, we have described two unique phenotypes, one of which is associated with an uncommon *LMNA* variant (R349W), characterized by specific body composition and clinical features. We are able to add previously unrecognized clinical characteristics (dorsocervical fat pad, ischaemic cardiac disease and low alkaline phosphatase) to the description of this genotype.[13] As another novel finding, two related patients (mother-daughter) were found to have a heterozygous variant of uncertain significance in *POLD1* that is possibly pathogenic and associated with dysmorphic nuclei in the liver. *POLD1* has been previously implicated in lipodystrophy syndromes. An in-frame deletion mutation in *POLD1* was discovered in two patients with
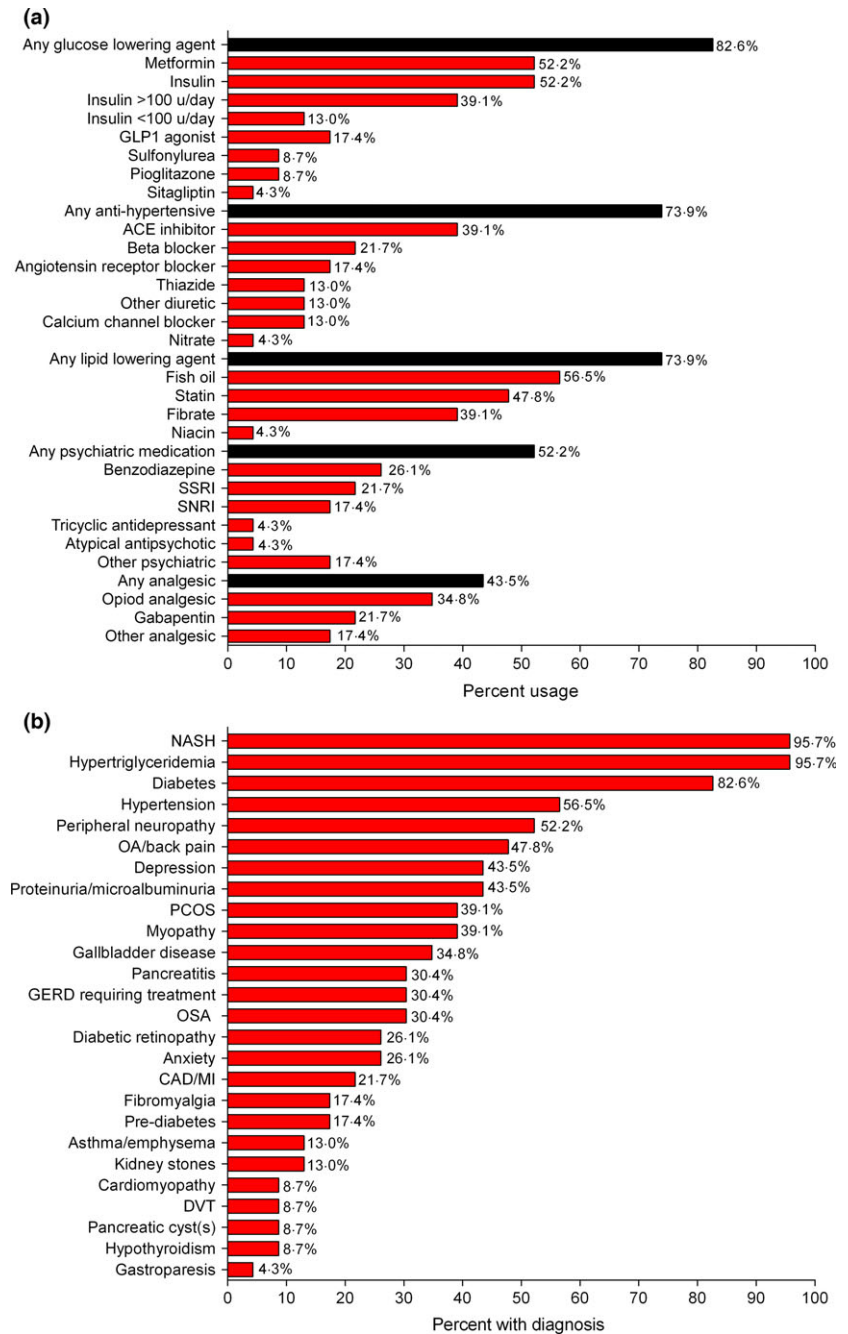
(a)

(b)

Fig 3 (a) Frequency of medication use for diabetes, dyslipidemia, hypertension and mood disorders. (b) Co-morbidity frequency among 23 patients with partial lipodystrophy.

mandibular hypoplasia, deafness and progeroid features (MDP) syndrome with lipodystrophy.[25] Our patients did not have clinical evidence for deafness or progeroid features, but had small mandibles and fluctuating creatine kinase (CK) levels. Our findings need to be complemented with further functional characterization of the observed variant. However, glutamine in position 1067 is highly conserved, within the second zinc module of the zinc finger domain and projected to be pathogenic using *in silico* algorithms (Figure S3).

Another area of interest for us was to determine whether unique morphometric characteristics could be objectively used as diagnostic criteria. Previously, the use of mid-thigh skin thickness has been suggested as a field tool to screen FPLD,[3,26]

and this criterion has been adopted into the consensus statement developed by AACE[3] as well as the most recent multi-society effort.[27] In fact, recently, Guillin-Amarelle *et al.* have described skin-fold characteristics of patients with the Koberling varierty of PL, attributing a Koberling Index, the ratio of subscapular to calf skinfolds (KöB index), with a cut-off value of 3·477 to be good predictor of this subtype of PL (sensitivity, 89%; specificity, 84%).[28] However, accurate measurement of skin thickness may be tricky in some patients in our experience and we have observed high inter-observer variability while collecting data. Body composition in our patients as analyzed using DEXA showed a high FMR when compared to normative data obtained from NHANES reference data,[15] highlighting the loss
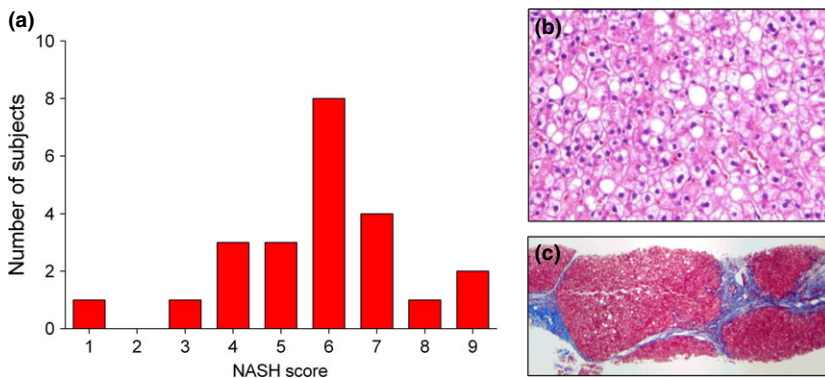
Fig 4 (a) Frequency distribution of NASH scores of liver biopsy specimens. (b) H&E and (c). Trichrome staining of liver biopsy sample in patient 15 (a 12-years-old with atypical PL) showing hepatic injury and steatosis (b) stage 4 fibrosis (c).
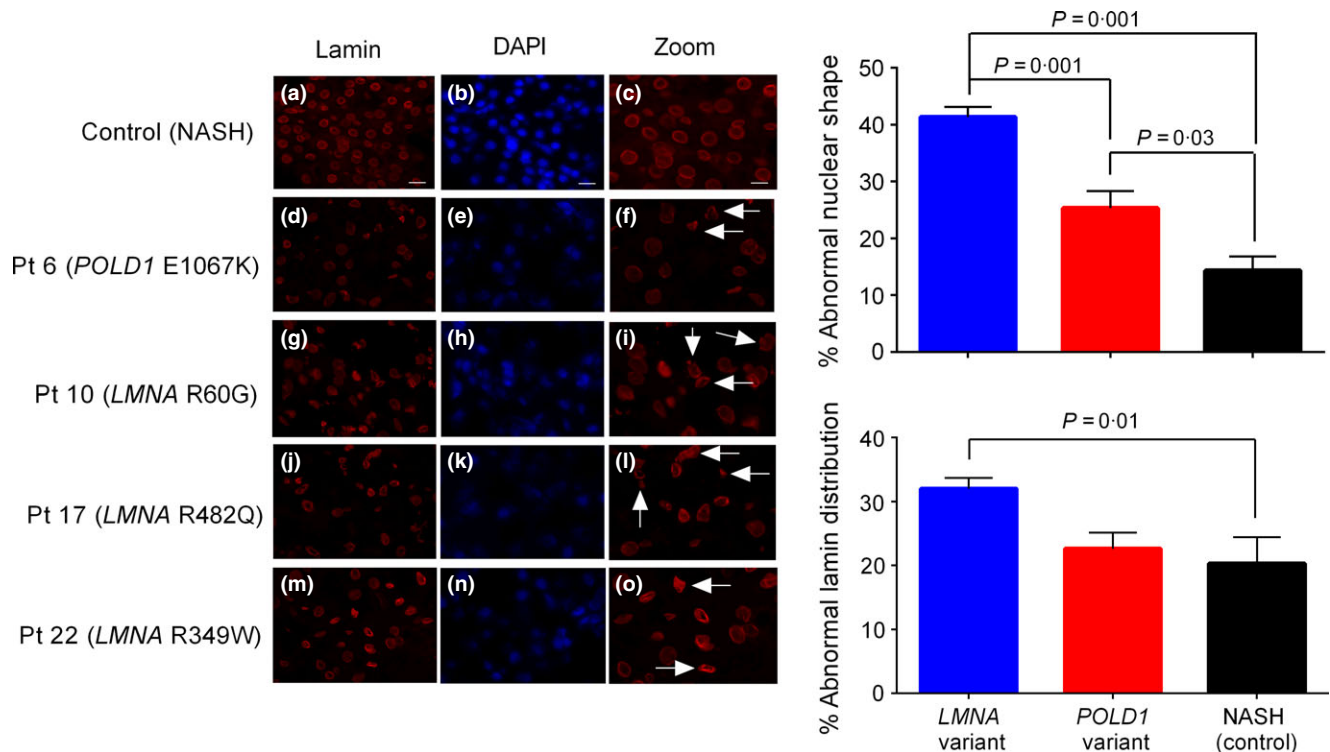

Fig 5 Abnormal nuclear morphology and lamin distribution in livers from patients with *LMNA* mutations. Lamin A/C distribution (red) and nuclei (blue) were visualized by immunofluorescence staining of frozen liver sections from patients with familial partial lipodystrophy (D–O). Liver biopsy specimens from two patients with nonalcoholic steatohepatitis but no history of PL and no pathogenic variants in LMNA were used as controls; representative images from one of these patients are shown (A–C). Dysmorphic nuclei and lamin disorganization are indicated by arrows in high magnification images (right column). Scale bars 20 μm (left and middle column), 10 μm (right column). Right panels show quantification of abnormal nuclear shape (upper right) and abnormal lamin distribution (lower right) in control NASH livers *vs* those from patients with *LMNA* and *POLD1* variants. For each group, 3–4 randomly selected high-power fields were scored for each patient liver section (>150 nuclei per field, >700 nuclei per patient). Statistical significance was determined by one-way ANOVA followed by Tukey's post hoc test at a threshold of $P < 0.05$. Error bars represent the standard error of the mean.

of fat in extremities seen in all of our patients who were classified as having FPLD. Others have proposed this approach previously in both female patients harbouring *LMNA* pathogenic variants and in patients with HIV-associated lipodystrophy.[26,29] In our hands, an FMR above 1·5 (obtained using GE Lunar DEXA systems) in adult females may indicate an abnormal ratio compared to a reference population taking multi-ethnic or age-related variation into account. We are not able to suggest a cut-off value for male patients or children in our cohort. Further

analysis of fat-related parameters in larger groups of lipodystrophy patients and comparison to patients with common metabolic syndrome, and abdominal obesity may help establish objective diagnostic criteria for PL.

We also present novel data showing that low leg fat was correlated with higher triglycerides in our unique population, suggesting a pathogenic link between higher triglycerides and low leg fat. Other researchers have reported a similar association from other cohorts, and our patients possibly display the most

extreme version of this association.[30,31] Further strengthening a pathogenic role for absence of femoral fat, Van Pelt and colleagues have observed worsening lipemia after femoral lipectomy.[32] In contrast, higher fat in the liver in our patients is associated with worse glucose control and higher triglycerides (shown for the first time in a PL cohort), similar to studies in patients with Type 2 diabetes.[33] These findings suggest that the development of hypertriglyceridaemia may be related to absence of peripheral fat depots (and possibly related to changes in their adipokines), but also because of accumulation of ectopic fat, allowing both mechanisms to be exploited for therapeutic development.

As expected in patients with lipodystrophy, our patients had a high prevalence of diabetes, often associated with extreme insulin resistance. The incidence of diabetes was higher in our population (82·6%), compared with that of a previously reported cohort (50%).[34] This discrepancy may be because of recruitment of patients suspected to have NAFLD, and these patients may have more severe metabolic co-morbidities. Microvascular complications of diabetes, particularly peripheral neuropathy and nephropathy were also common. Hypertriglyceridaemia and NAFLD or NASH are known to have associations with lipodystrophy, and most of our patients also demonstrated this.[2] Other frequent conditions included recurrent pancreatitis, hypertension, PCOS, gastroesophagal reflux disease (GERD), sleep apnoea and coronary artery disease or myocardial infarction. Diabetes commonly co-exists with hypertension, hyperlipidaemia, kidney disease and cardiovascular disease.[35] However, we were surprised by the prevalence of chronic pain and mood disorder in this group. Overall, chronic pain was present in 78·3% of patients and was reported to be because of a combination of arthritis, back pain, fibromyalgia and myopathy. Depression has been shown to be prevalent in adults with diabetes to a varying degree (11–32%),[36] but appears higher in our patients (43·5%). It is not clear whether these under-appreciated manifestations are because of primary pathophysiological pathways, a manifestation of ectopic lipid (or other metabolite) deposition in muscles and nerves (or elsewhere) or because of some other adaptive (neurobehavioural, possibly functional) processes.

We further explored the extent of liver disease and NASH by histopathological criteria was present in nearly all patients (22 of 23). As our trial specifically sought patients with fatty liver disease at baseline, the presence of NASH may be overrepresentative of the prevalence of NASH in the entire population of PL. It is noteworthy that significant fibrosis may be present even in young patients without diabetes as was shown in a 12-year-old female patient. Therefore, it would be important to consider a liver biopsy or a careful investigation for clinical fibrosis even in very young patients with PL.

Lastly, we present data that reflect relative nuclear integrity in the liver tissue of PL. Lamin A and lamin C are integral components of the nuclear envelope and are expressed ubiquitously throughout the body. Alterations in the nuclear envelope may lead to premature cell apoptosis. *LMNA* pathogenic variants may increase the risk of arrhythmia or cardiomyopathy, as occurred in two of our patients with non-ischaemic

cardiomyopathy (one requiring cardiac transplantation).[37] Given that patients had multi-system involvement, we examined the degree of nuclear disorganization in liver biopsy tissues of select patients. Patients with *LMNA* pathogenic variants had apparent nuclear disorganization in their liver tissue. We also noted a similar pattern, although not to the same degree, in a patient harbouring a *POLD1* variant of uncertain significance that is likely pathogenic. We posit nuclear atypia and disorganization may be a unifying cellular phenomenon in patients with PL depending on the mutation and the involved gene. This phenomenon may contribute to the multisystem involvement in PL. Further work is warranted in this arena.

Overall, PL was associated with involvement of at least 10 organ systems (with manifestations present in all 10 in nearly 40% of patients). The involvement of multiple organs in PL bears some resemblance to other heterogeneous rare metabolic diseases such as glycogen storage disease. Researchers studying Gaucher disease have developed a disease severity score tool,[38] making use of the number of organ systems affected, severity of signs and symptoms and functional limitations. The diversity of comorbid conditions in our patients has inspired us to propose adoption of a similar approach to define the disease burden more objectively. If such a tool can be developed and validated, the impact of disease modifying treatments can be better measured.

Acknowledgements

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