

# NSUN2-Mediated m5C Methylation and METTL3/METTL14-Mediated m6A Methylation Cooperatively Enhance p21 Translation

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## ABSTRACT

N6-methyladenosine (m6A) and m5C methylation are two major types of RNA methylation, but the impact of joint modifications on the same mRNA is unknown. Here, we show that in p21 3'UTR, NSUN2 catalyzes m5C modification and METTL3/METTL14 catalyzes m6A modification. Interestingly, methylation at m6A by METTL3/METTL14 facilitates the methylation of m5C by NSUN2, and vice versa. NSUN2-mediated m5C and METTL3/METTL14-mediated m6A methylation synergistically enhance p21 expression at the translational level, leading to elevated expression of p21 in oxidative stress-induced cellular senescence. Our findings on p21 mRNA methylation and expression reveal that joint m6A and m5C modification of the same RNA may influence each other, coordinately affecting protein expression patterns. *J. Cell. Biochem.* 118: 2587–2598, 2017. © 2017 Wiley Periodicals, Inc.

**KEY WORDS:** NSUN2; METTL3; METTL14; p21 mRNA METHYLATION; TRANSLATIONAL REGULATION

Methylation is a widespread post-transcriptional modification of RNAs [Dominissini et al., 2012; Meyer et al., 2012] and determines translation efficiency, mRNA turnover, and RNA processing [Ji and Chen, 2005; Das et al., 2008; Seshadri et al., 2009; Schaefer et al., 2010; Zhang et al., 2012; Hussain et al., 2013; Yuan et al., 2014]. Although methylation at all four ribonucleotides (A, U, C, and G) has been reported [Björk et al., 1987; Persson et al., 1992], methylation of an mRNA at other than the 5'-cap occurs predominantly as N6-methyladenosine (m6A), with a small amount of 5-methylcytosine (m5C) identified in mRNA isolated from cultured hamster cells [Dubin and Taylor, 1975; Carroll et al., 1990]. Apart from m5C and m6A, N(1)-Methyladenosine (m1A) and pseudouridine are also prevalent post-transcriptional RNA modification for coding and non-coding RNAs [Li et al., 2016; Zhao et al., 2017]. However, the

functional interconnection between different types of RNA methylation, especially m6A and m5C, has not been studied.

The tRNA methyltransferase NSun2 (Misu) is a typical RNA methyltransferase catalyzing the formation of m5C in coding and non-coding RNAs [Hussain et al., 2013; Tang et al., 2015; Xing et al., 2015; Cai et al., 2016]. Methylation of mRNAs by NSUN2 is found to regulate the translation or turnover of mRNAs encoding CDK1, TP53, SHC, p16 (CDKN2A), and p27 (CDKN1B), thus affecting the expression of these genes in processes such as cell proliferation, oxidative stress, and replicative senescence [Hussain et al., 2013; Tang et al., 2015; Xing et al., 2015; Cai et al., 2016]. METTL3 (methyltransferase like 3), METTL14 (methyltransferase-like 14), and WTAP (Wilm's tumor 1-associated protein) form a methyltransferase complex, which catalyzes m6A formation [Liu et al., 2014; Ping

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et al., 2014]. METTL3/METTL14-mediated RNA methylation is also involved in the regulation of mRNA stability or translation. For example, METTL3/METTL14 synergistically control the formation of m6A in a large number of mRNAs encoding developmental regulators, thereby inhibiting their expression at the levels of mRNA turnover and RNA processing in embryonic stem cells [Lin and Gregory, 2014; Wang et al., 2014]. However, whether NSUN2 and METTL3/METTL14 regulate the expression of an mRNA that is modified both at m5C and m6A has not been reported.

The levels of the broad inhibitor of cyclin-dependent kinases CDKN1A (p21<sup>Cip1</sup>), increases in cells exposed to a wide variety of stress agents, including genotoxins, oxidants, and metabolic perturbations. The increased p21 expression is believed to contribute to the growth arrest that follows exposure to these insults [Deng et al., 1995; Waldman et al., 1996; Gorospe et al., 1996a] and affects profoundly the outcome of the stressed cells, frequently favoring cell survival [Gorospe and Holbrook, 1996; Gorospe et al., 1996b, 1997]. Not unexpectedly, p21 expression is tightly regulated at multiple levels. Transcriptional regulation of p21 levels, particularly by the transcription factor TP53, has been intensively reported [Gartel and Tyner, 1999]. Beyond transcription, p21 levels are also regulated via stabilization of p21 mRNA and protein as well as by controlling translation of p21 mRNA [Timchenko et al., 1996; Gartel and Tyner, 1999; Wang et al., 2000; Iakova et al., 2004; Lal et al., 2004]. The RNA-binding protein (RBP) HuR binds to the 3'UTR of p21 mRNA and stabilizes it in cells exposed to short-wavelength UV light (UVC) [Wang et al., 2000]. AUF1 and HuR competitively associate with the p21 3'UTR and regulate p21 mRNA stability [Lal et al., 2004]. The RBPs CUGBP1 and calreticulin (CRT) competitively interact with the same sequences within the p21 5'UTR and modulate in opposing directions p21 the translation, in turn influencing cell growth arrest and senescence [Iakova et al., 2004]. Another mode of post-transcriptional regulation of p21 is through alteration in the stability of the p21 protein, as illustrated by the extended half-life of p21 through interaction with the transcription factor CEBP $\alpha$  [Timchenko et al., 1996].

In the present study, we set out to investigate the possibility that p21 expression might be modulated by changes in p21 mRNA methylation. We report that NSUN2 catalyzes m5C modifications and METTL3/METTL14 m6A modifications in the 3'UTR of p21 mRNA. Interestingly, methylation by METTL3/METTL14 facilitates the methylation by NSUN2, and vice versa. In addition, joint methylation at m5C and m6A synergistically enhance p21 expression at the level of translation in a model of oxidative stress-induced cellular senescence. Our findings provide a novel mechanism by which p21 expression is regulated.

## MATERIALS AND METHODS

### CELL CULTURE, FACS, TRANSFECTION, AND SA- $\beta$ -GAL STAINING

TP53-deficient human colon carcinoma cells (HCT116 p53<sup>-/-</sup>) and HeLa cells were cultured in Dulbecco's modified Eagle's medium (Invitrogen) supplemented with 10% fetal bovine serum, 100 U/ml penicillin, and 100  $\mu$ g/ml streptomycin, at 37°C in 5% CO<sub>2</sub>. All plasmids were transfected using lipofectamine 2000 (Invitrogen) and cells were collected 48–72 h after transfection for further analysis.

SA- $\beta$ -galactosidase staining was performed as described previously [Tang et al., 2015; Cai et al., 2016].

### KNOCKDOWN OF NSUN2, METT3, AND METTL14

To silence NSUN2, METT3, and METTL14, cells were transfected with siRNA (10 nM) targeting *NSUN2* (GGUGUAGAAAUAACAGCGGUGAAGA), *METT3* (CTGCAAGTATGTCTACTATGA), or *METTL14* (AAGGATGAGTTAATAGCTAAA) mRNAs, or a control siRNA (UUGUUCGAACGUG UCACGUTT) using RNAiMAX (Invitrogen).

### RNA ISOLATION AND REAL-TIME qPCR

Total cellular RNA was isolated by using the RNeasy Mini Kit (Qiagen). For reverse-transcription (RT) followed by real-time, quantitative (q) PCR analysis of the levels of p21 and *GAPDH* mRNAs, primers GACTCTCAGGGTCGAAAACGG and GCGGATTAGGGCTTCTCTT for p21 mRNA and CGAGTCAACGGATTGGTGGTAT and AGCCTTCTCCATGGTGAAGAC for *GAPDH* mRNA were used.

### CONSTRUCTS AND REPORTER GENE ASSAYS

For construction of the pGL3-5'UTR, pGL3-CR, pGL3-3'UTR reporter vectors, p21 5'UTR fragment was amplified by using primer pairs CCAAGCTTGTGTATATCAGGGCCGC and CATGCCATGGG GCGC CTCCTCTGAG and inserted between Hind III and Nco I sites of the pGL3-promoter vector; p21 CR fragment was amplified by using primer pairs GCTCTAGAATGTCAGAACC GGCTGGG and GCTCTAGATTAGGGCTTCTCTTGGAGA and inserted into Xba I site of the pGL3-promoter vector; p21 3'UTR was amplified by using primer pairs GCTCTAGATCCGCCACAGGAAGCCTGCAGT and GCTCTAGATAAAGTCACTAAGAATC and inserted into Xba I site of the pGL3-promoter vector (Promega).

For reporter gene assays, each of the pGL3-derived vectors was co-transfected with pRL-CMV vector by Lipofectamine 2000 (Invitrogen). Forty-eight hours after transfection, cell lysates were collected and the firefly and renilla luciferase activities were measured with a dual luciferase assay system (Promega) following the manufacturer's instructions. All firefly luciferase measurements were normalized to renilla luciferase measurements from the same sample.

### TRANSCRIPT PREPARATION

The fragments of p21 mRNA were amplified by PCR by using cDNA as a template. All 5' primers contained the T7 promoter sequence (CCAAGCTTCTAATACGACTCACTATAGGGAGA). To prepare templates for the p21 5'UTR (positions 1–125), CR (positions 126–620), 3'UTR (positions 621–2158), 3'UTR1 (positions 621–800), 3'UTR2 (positions 778–957), 3'UTR3 (positions 935–1114), 3'UTR4 (positions 1092–1271), 3'UTR5 (positions 1249–1428), 3'UTR6 (positions 1046–1585), 3'UTR7 (positions 1563–1742), 3'UTR8 (positions 1720–1899), 3'UTR9 (positions 1877–2056), 3'UTR10 (positions 1979–2258), the following primer pairs were used: (T7) GTTGTATATCAGGGCCGC and GGCGCTCTCTGAGT for 5'UTR, (T7) ATGTCAGAACC GGCTGGG and TTAGGGCTTCTCTTGGAGA for CR, (T7) TCCGCCACAGGAAGCCTGCAT and TACAAGTAAAGTCACTAAGAATC, (T7) ATGTCAGAACC GGCT and CTCCAGTGGTGTCTCGGTGAC for CR1, (T7) TGTCACCGAGACACCAC and AGGTCCACATGGTCTTCTCTGC for

CR2, (T7) GCAGAGGAAACCAT and TTAGGGCTTCTCTTGGAGAAGA, (T7) TCCGCCACAGGAAG and TAAATAATTCTAATGCAGAGGCTGGGG for 3'UTR1, (T7) GCCTCTGGCATTAGAA and GTGGGGAGGAGGAAGTAGCAGG for 3'UTR2, (T7) GCCAGCTACTCTCTC and CCACTCGGTGAGGCCCTTCAA for 3'UTR3, (T7) TTTGAAGGGGCCTCA and GGACCCACCTTCCC for 3'UTR4, (T7) AAGGCAGGGGGAAGG and CAGCAGAGGGGAATTGCAGAG for 3'UTR5, (T7) GGCTCTGCAATTCCCC and GCTGCTTGAGCTGCCTGAGGTA for 3'UTR6, (T7) CTACCTCAGGCAGCT and ATGAGGAAGGTCGCTGGACGATTG for 3'UTR7, (T7) AATCG TCCAGCGACC and GGCTCAACGTTAGTGCCAGGAAAGAC for 3'UTR8, (T7) TTTCTGGCACTAAC and GCCCTCGAGAGGTTACAGTCTAGG for 3'UTR9, and (T7) GGCCTGGACTGTTTT and TACAAGTAAAGTCACTAAGAATCATT for 3'UTR10.

These PCR products were transcribed *in vitro* following the manufacturer's instructions (Thermo). The *p21* 3'UTR10 mutants (3'UTRcm [bearing C2079G], 3'UTR10m1 [bearing A1986U], 3'UTR10m2 [bearing A2037U], 3'UTR10m3 [bearing A2044U], 3'UTR10m4 [bearing A2061U], 3'UTR10m5 [bearing A2044U and A2061U]) were prepared by overlapping PCR. The *p27* 5'UTR and *SHC* 5'UTR fragments were described previously [Tang et al., 2015; Cai et al., 2016].

#### BISULFITE RNA SEQUENCING

Bisulfite RNA sequencing was performed to identify the methylation site (m5C) within an RNA fragment, as previously described. Briefly, 1  $\mu$ g *in vitro* methylated *p21* 3'UTR fragment (methylated by NSUN2 by using cold SAM [Sigma]) or RNA isolated from cells was dissolved in 10  $\mu$ l of RNase-free water and mixed with 42.5  $\mu$ l of 5 M sodium bisulfite (Epitect) and 17.5  $\mu$ l DNA protection buffer (Epitect), incubated in 70°C for 5 min and 60°C for 60 min, repeating for 3–5 cycles. Samples were desalted by using Micro Bio-spin6 columns and then desulfonated by 1 M Tris (pH 9.0, 1/1, V/V) at 37°C for 1 h, followed by ethanol precipitation. The bisulfite-converted fragments (0.2  $\mu$ g) were reverse-transcribed by RevertAid First-Strand cDNA Synthesis Kit (Thermo) using primer GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACTCTTAACCTTC CTACCATTCC and subjected to PCR analysis by using primer pairs CCAAGCTTCTAATACGACTCATATAGGGAGAGTTTGTGGGAATGGGGCCTGGACTGTTTT and CTTAACCTTCTACCATTCTACAAGTAAAGTCACTAAGAATCATT. To test the m5C formation in the endogenous *p21* mRNA, The bisulfite-converted cellular RNA were reverse-transcribed by RevertAid First Strand cDNA Synthesis Kit (Thermo) using primer GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACTATAATCA and subjected to PCR by using primer pairs GGTTTTGTTTTT TTATT TAGATTGT and GCAGGGTCCGAGGTATTC. The PCR products were inserted into the pGEM-T Easy Vector System (Promega). The plasmids purified from single clones were sequenced, the sequences aligned with the corresponding *p21* mRNA sequence and the cytosines retained were considered to be methylated.

#### IN VITRO TRANSLATION ASSAYS

For *in vitro* translation assays, a cell-free translation system (Promega) in rabbit reticulocyte lysate (RL) was used. Luc-3'UTR fragment was amplified by PCR by using primer pairs (T7) CCATGGAAGACGCCAAAAACATA and TTTTTTTTTTTTTTTTTT

CTAGATACAAGTAAAGTCAC from pGL3-3'UTR reporter vector. The Luc-3'UTR fragment then was *in vitro* transcribed and further methylated by NSUN2, METTL3/METTL14, or by NSUN2 + METTL3/METTL14 *in vitro* or kept untreated. The methylated and non-methylated transcripts (0.01 nM) were used for *in vitro* translation assays. The translation efficiency was determined by measuring the activity of firefly luciferase.

#### LC-MS ANALYSIS

To determine the formation of m5C or m6A in *in vitro* methylated RNA fragments, *in vitro* methylated RNA fragments (1  $\mu$ g) were digested by nuclease P1 (Sigma) and alkaline phosphatase (Thermo). The products were subjected to HPLC-MS analysis at Tsinghua University Mass Spectrum Center (Beijing, China). This method combined High pressure liquid chromatography (HPLC) and mass spectrometry (MS) to measure the mass (by MS) and amount (by HPLC) of m6A or m5C in RNA samples which is digested to single nucleotides. The area value of a peak appeared from a specific nucleotide was calculated by calculus to reflect the amount of methylation.

#### MEASUREMENT OF m6A METHYLATION IN CELLS

To measure the levels of m6A methylation in cells, 1  $\mu$ g of anti-m6A antibody, 20  $\mu$ g of cellular RNA, and 20  $\mu$ l (in 50% slurry) protein-G sepharose were incubated in IPP buffer (150 nM NaCl, 0.1% NP-40, 10 mM Tris.HCl [pH 7.4]) plus 1 U/ $\mu$ l RNasin in 200  $\mu$ l at 4°C for 2 h. The beads were washed with the IPP buffer for five times. RNA isolated from the IP beads were subjected to reverse transcription (RT) followed by real-time, quantitative (q)PCR analysis.

#### IN VITRO METHYLATION ASSAYS

The *in vitro* methylation assay by NSUN2 was performed as described previously (Tang et al., 2015). Briefly, 50  $\mu$ l of reaction mixture containing 0.2 nM purified His-NSun2, 0.01 nM RNA *in vitro*-transcribed RNA fragments, and 1  $\mu$ Ci of <sup>3</sup>H-labeled S-adenosyl-L-methionine (Amersham Bioscience) in reaction buffer (5 mM Tris HCL [pH 7.5], 5 mM EDTA, 10% glycerol, 1.5 mM dithiothreitol, 5 mM MgCl<sub>2</sub>) supplemented with inhibitors (leupeptin [1  $\mu$ g/ml], aprotinin [1  $\mu$ g/ml], 0.5 mM phenylmethylsulfonyl fluoride, and RNasin [5 U/ $\mu$ l]) was incubated for 45 min at 37°C. Unincorporated <sup>3</sup>H S-adenosyl-L-methionine in the reactions were removed by using Qiaquick Spin Columns (Qiagen) and the incorporated radioactivity was measured by liquid scintillation counting.

For *in vitro* methylation assays by METTL3/METTL14, HeLa cells were co-transfected with vectors expressing flag-METTL3 and flag-METTL14. Forty-eight hour later, cell lysates were prepared and subjected to immunoprecipitation assays to enrich flag-METTL3 and flag-METTL14 proteins. The enriched flag-METTL3 and METTL14 in the IP materials (~0.1 nM) were used for *in vitro* methylation assays in a standard 50  $\mu$ l of reaction mixture containing 0.1 nM RNA fragment, 1  $\mu$ Ci of <sup>3</sup>H-labeled S-adenosyl-L-methionine (Amersham Bioscience), 80 mM KCl, 1.5 mM MgCl<sub>2</sub>, 0.2 U/ $\mu$ l RNasin, 10 mM DTT, 4% glycerol, and 15 mM HEPES (pH 7.9). Unincorporated <sup>3</sup>H S-adenosyl-L-methionine was removed by using Qiaquick Spin Columns (Qiagen) and incorporated radioactivity was measured by liquid scintillation counting. RNA fragment ACGAGUCCUGGACUGAAACGGACUUGU was used as a positive control, as described [Liu et al., 2014].

## ANTIBODIES AND WESTERN BLOT ANALYSIS

Monoclonal anti-GAPDH, polyclonal anti-p66SHC was from BD Biosciences. Polyclonal anti-METTL3 was from Proteintech, Monoclonal anti-p21 antibody and monoclonal anti-CDK1 were from Santa Cruz. Polyclonal anti-METTL14 was from Abcam. Monoclonal anti-flag antibody was from Sigma. After secondary antibody incubations, signals were detected by SuperSignal WestPico Chemiluminescent Substrate (Pierce) following the manufacturer's instruction and quantitated by densitometric analysis with ImageMaster VDS software.

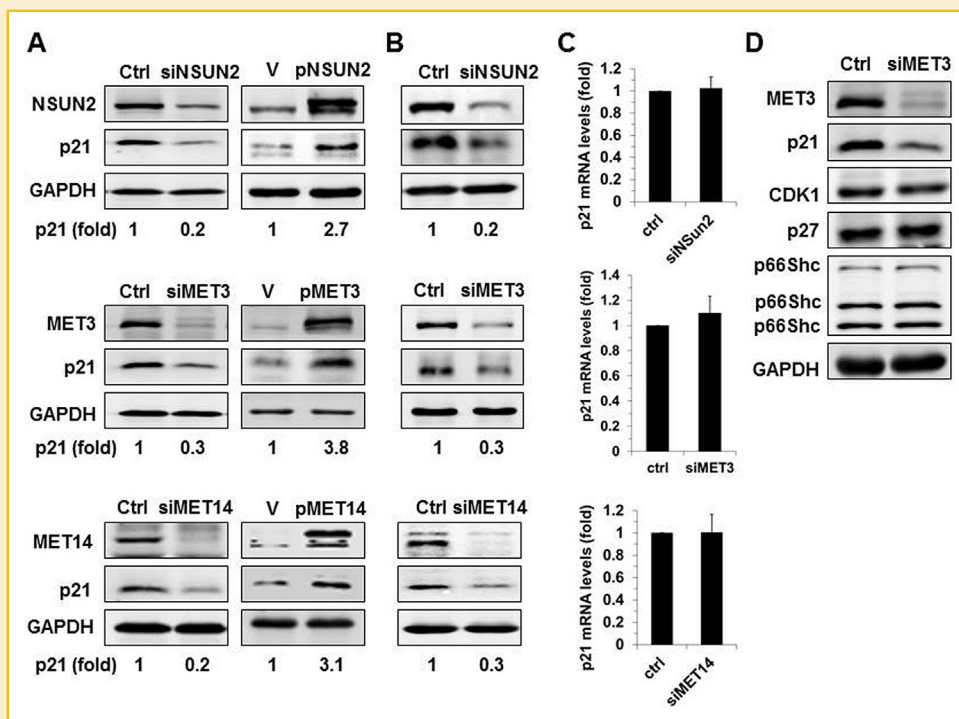
## RESULTS

### NSUN2, METTL3, AND METTL14 REGULATE THE EXPRESSION OF p21

To test if p21 expression might be regulated by methylation, we examined if p21 protein levels were altered in HeLa cells in which these methyltransferases (NSUN2, or METTL3, or METTL14) had been silenced by transfection of specific siRNAs. As shown in Figure 1A (left), Western blot analysis revealed that reducing NSUN2, METTL3, or METTL14 markedly decreased the levels of p21 protein (by ~80% after silencing NSUN2 or METTL14 and by ~70% after silencing METTL3). Conversely, overexpression of NSUN2, METTL3, or METTL14 increased p21 protein levels (by ~2.7, ~3.8, ~3.1 fold, respectively)

(Fig. 1A, Right). Given that p53-mediated transcriptional regulation is a well-known mechanism controlling p21 expression, we further assessed p21 protein levels in p53-deficient human colon carcinoma cells (HCT116 p53<sup>-/-</sup> cells) in which NSUN2, METTL3, or METTL14 were individually silenced. As shown in Figure 1B by Western blot analysis, knockdown of NSUN2, METTL3, or METTL14 in HCT116 p53<sup>-/-</sup> cells decreased p21 protein levels (by ~80% after NSUN2 silencing, by ~70% after silencing METTL3 or METTL14), indicating that NSUN2, METTL3, and METTL14 regulated p21 expression levels in a p53-independent manner. In addition, knockdown of NSUN2, METTL3, or METTL14 did not alter the mRNA levels of *p21* in HCT116 p53<sup>-/-</sup> cells (Fig. 1C). In sum, NSUN2, METTL3, and METTL14 did not appear to affect p21 expression levels by altering mRNA turnover or transcription and instead may affect p21 translation.

NSUN2 has been found to regulate the expression of SHC, CDK1, TP53, p16, and p27 [Hussain et al., 2013; Tang et al., 2015; Xing et al., 2015; Cai et al., 2016]. To ask whether METTL3/METTL14 and NSUN2 share targets other than *p21* mRNA, the levels of CDK1, p27, and SHC were determined by Western blot analysis in cells in which METTL3 was silenced. As shown in Figure 1D, knockdown of METTL3 did not alter the levels of CDK1, p27, or SHC, suggesting that METTL3/METTL14 and NSUN2 specifically share *p21* mRNA as the same target.



**Fig. 1.** NSUN2, METTL3, and METTL14 regulate p21 expression. (A) HeLa cells were transfected with siRNAs targeting NSUN2 (siNSUN2), METTL3 (siMET3), or METTL14 (siMET14) (left), or with a vector expressing flag-tagged NSUN2 (pNSUN2), METTL3 (pMET3), or METTL14 (pMET14) (right). Forty-eight hours later, cell lysates were prepared and subjected to Western blot analysis to assess the levels of proteins NSUN2, METTL3, METTL14, p21, and GAPDH. Data are representative from three independent experiments. (B) HCT116 p53<sup>-/-</sup> cells were transfected with siRNAs targeting NSUN2, METTL3 (siMET3), or METTL14 (siMET14). Forty-eight hours later, cell lysates were prepared and subjected to Western blot analysis to assess the levels of proteins NSUN2, METTL3, METTL14, p21, and GAPDH. Data are representative from three independent experiments. (C) RNA was prepared from cells described in Figure 1B and subjected to real-time qPCR to assess *p21* mRNA levels. Data represent the mean  $\pm$ SD from 3 independent experiments. (D) Cell lysates were prepared from cells silenced with METTL3 (siMET3), as described in Figure 1A. CDK1, p27, p53, and SHC protein levels were determined by Western blot analysis.

## NSUN2 AND METTL3/METTL14 COOPERATIVELY ENHANCE p21 EXPRESSION

To further investigate the impact of NSUN2, and METTL3/METTL14 on p21 expression, the levels of p21 in cells in which NSUN2 and METTL3/METTL14 were silenced individually or jointly were assessed by Western blot analysis. As shown in Figure 2A, combined knockdown of NSUN2 and METTL3 (NSUN2 + METTL3) was more effective in reducing p21 levels than silencing NSUN2 or METTL3 individually (with reductions by ~50% [ $P < 0.05$ ], ~38% [ $P < 0.05$ ], and ~64% [ $P < 0.05$ ], respectively). Similarly, p21 levels in NSUN2 + METTL14 co-silenced cells were much lower than those in cells where only NSUN2 or METTL14 were silenced (with reductions by ~43% [ $P < 0.05$ ], ~42% [ $P < 0.05$ ], and ~66% [ $P < 0.01$ ], respectively) (Fig. 2B). In agreement with the findings shown in Figure 1C, knockdown of NSUN2, METTL3, METTL14, NSUN2, and METTL3 together, or NSUN2 and METTL14 together did not influence the mRNA levels of p21 (Fig. 2C and D). These results suggest that NSUN2, METTL3, and METTL14 may cooperatively enhance p21 expression.

To further test this hypothesis, we constructed pGL3-derived reporters bearing *p21* 5'UTR (pGL3-5'UTR), *p21* CR (coding region, pGL3-CR), and *p21* 3'UTR (pGL3-3'UTR) (Fig. 3A, schematic) and tested the activity of these reporters in HCT116 p53<sup>-/-</sup> cells after silencing NSUN2, METTL3, and METTL14 separately and in combination. As shown in Figure 3B-D, individual knockdown of NSUN2, METTL3, and METTL14 strongly decreased the luciferase activity of reporter pGL3-3'UTR (by ~53.5%, ~40.5%, and ~40.2%, METTL14, respectively), but did not alter significantly luciferase activity from pGL3, pGL3-5'UTR, or pGL3-CR. Notably, simultaneous knockdown of NSUN2 + METTL3 or NSUN2 + METTL14 further decreased the activity of pGL3-3'UTR (by ~64.5% for NSUN2 + METTL3, and by ~82.3% for NSUN2 + METTL14) (Fig. 3E and F). In sum, NSUN2, METTL3, and METTL14 cooperatively regulate the expression of p21; the NSUN2 and METTL3/METTL14 response region is located in the *p21* 3'UTR.

## METTL3/METTL14 AND NSUN2 METHYLATE THE *p21* 3'UTR

The RNA methyltransferase NSUN2 catalyzes the formation of m5C in RNAs. METTL3 and METTL14 form a stable heterodimer that

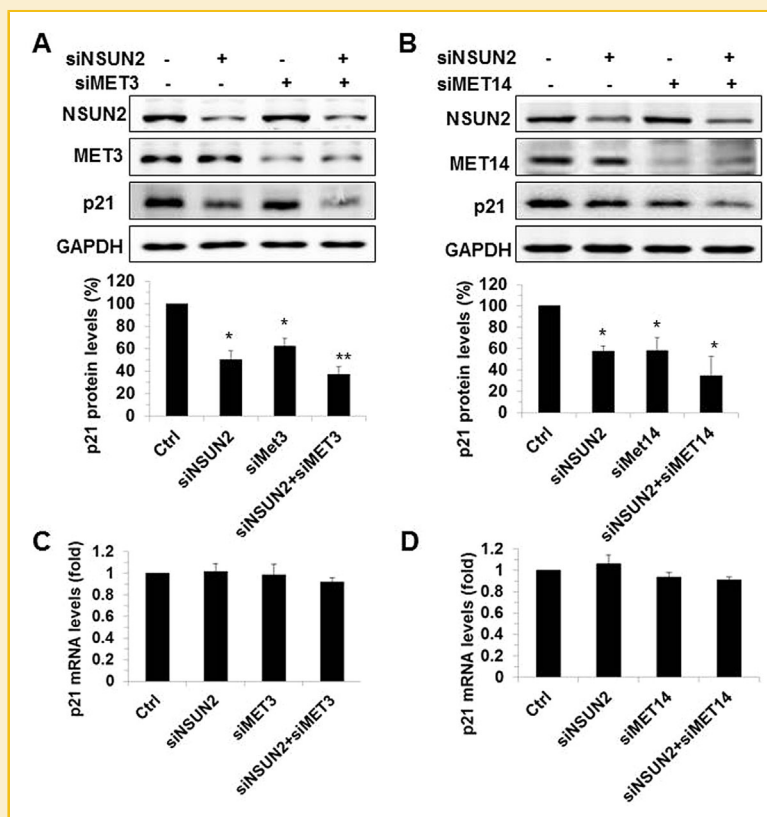
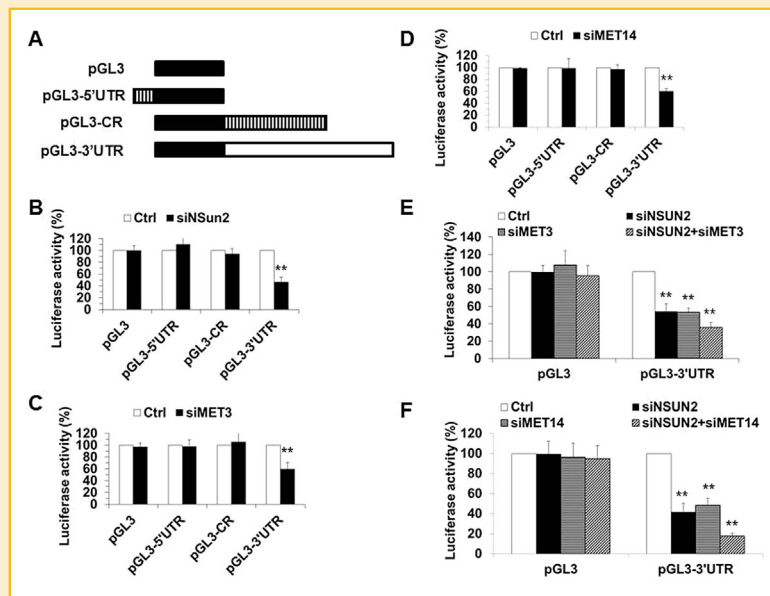


Fig. 2. NSUN2 and METTL3 or METTL14 synergistically enhance p21 expression. (A) Upper, HCT116 p53<sup>-/-</sup> cells were transfected with NSUN2 siRNA, METTL3 siRNA (siMET3), or with both (siNSUN2 + siMET3). Forty-eight hour later, cell lysates were prepared and subjected to Western blot analysis to assess the levels of NSUN2, METTL3, p21, and GAPDH. Bottom, data are means  $\pm$  SD from three independent experiments; significance is analyzed by Student's *t*-test (\*\* $P < 0.01$ ; \* $P < 0.05$ ). Data are representatives of three independent experiments. (B) Upper, HCT116 p53<sup>-/-</sup> cells were transfected with NSUN2 siRNA, METTL14 siRNA (siMET14), or with both (siNSUN2 + siMET14). Forty-eight hour later, cell lysates were prepared and subjected to Western blot analysis to assess the levels of NSUN2, METTL14, p21, and GAPDH. Bottom, data are means  $\pm$  SD from three independent experiments; significance is analyzed by Student's *t*-test (\* $P < 0.05$ ). (C and D) Real-time qPCR analysis was used for analyzing p21 mRNA levels in cells described in Figure 2A and B, respectively. Data are means  $\pm$  SD from three independent experiments.



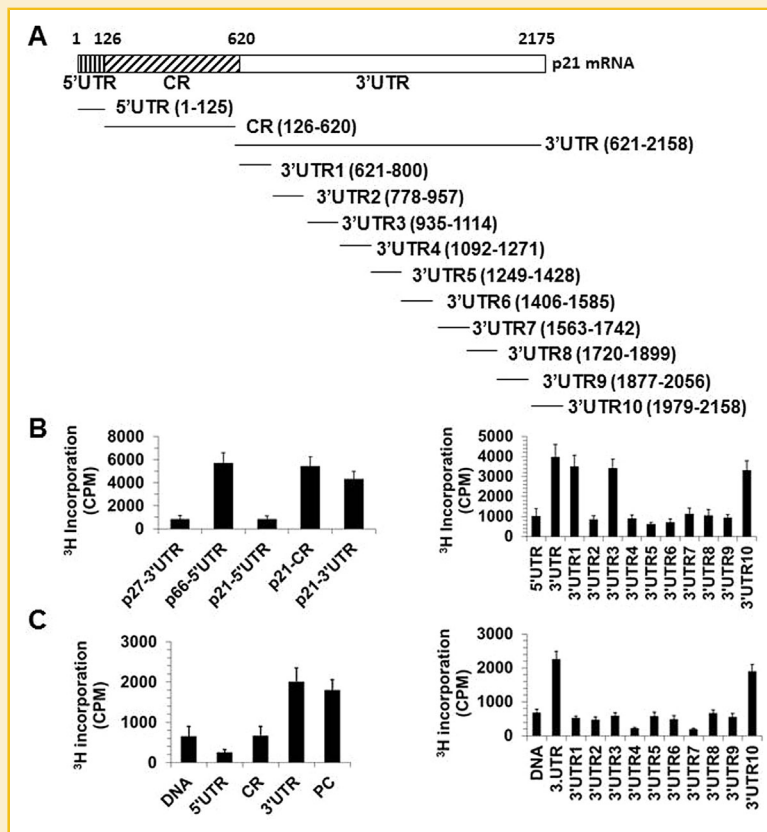
**Fig. 3.** NSUN2 and METTL3 or METTL14 synergistically enhance the activity of pGL3-derived reporter bearing *p21* 3'UTR. (A) Schematic representation depicting the pGL3-derived reporters used for reporter gene assays. (B–D) HCT116 *p53*<sup>-/-</sup> cells were transfected with each of the reporters depicted in Figure 3A together with a pRL-CMV control reporter. Twenty-four hour later, cells were further transfected with a siRNA targeting NSUN2 (B), METTL3 (C), or METTL14 (D), or with a control siRNA (Control) and cultured for an additional 48 h. Firefly luciferase activity against Renilla luciferase activity was analyzed. Data represent the means  $\pm$  SD from three independent experiments; significance was analyzed by Student's *t*-test (\*\**P* < 0.01). (E) HCT116 *p53*<sup>-/-</sup> cells were co-transfected with a pGL3-3'UTR reporter or a pGL3 vector and a pRL-CMV control reporter. Twenty-four hour later, cells were further transfected with NSUN2 siRNA (siNSUN2), METTL3 siRNA (siMET3), or NSUN2 siRNA plus METTL3 siRNA (siNSUN2 + siMET14) and cultured for an additional 48 h. Firefly luciferase activity against Renilla luciferase activity was analyzed. Data represent the means  $\pm$  SD from three independent experiments; significance was analyzed by Student's *t*-test (\*\**P* < 0.01). (F) HCT116 *p53*<sup>-/-</sup> cells were co-transfected with a pGL3-3'UTR reporter or a pGL3 vector and a pRL-CMV control reporter. Twenty-four hour later, cells were further transfected with NSUN2 siRNA (siNSUN2), METTL14 siRNA (siMET14), or NSUN2 siRNA plus METTL14 siRNA (siNSUN2 + siMET14) and cultured for an additional 48 h. Firefly luciferase activity against Renilla luciferase activity was analyzed. Data represent the means  $\pm$  SD from three independent experiments; significance was analyzed by Student's *t*-test (\*\**P* < 0.01).

functions in catalyzing the formation of m6A in RNAs. To test if NSUN2 methylated *p21* mRNA, the *p21* mRNA fragments described in Figure 4A were used for in vitro methylation assays by using purified his-NSUN2 and <sup>3</sup>H-labeled S-Adenosyl methionine (SAM) (Materials and Methods). To test if METTL3/METTL14 methylated *p21* mRNA, HCT116 *p53*<sup>-/-</sup> cells were co-transfected with vectors expressing flag-METTL3 and flag-METTL14. Forty-eight hour later, cell lysates were prepared and subjected to immunoprecipitation analysis to enrich flag-METTL3 and flag-METTL14. The IP materials and <sup>3</sup>H-labeled SAM were used for in vitro methylation assays using as substrates the fragments depicted in Figure 4A. The *p27* 3'UTR was included as a negative control and *SHC* 5'UTR served as a positive control for NSUN2-mediated RNA methylation; *p21* cDNA served as a negative control for the METTL3/METTL14-mediated *p21* mRNA methylation. As shown in Figure 4B, NSUN2 was capable of methylating *SHC* 5'UTR, and a number of *p21* mRNA regions (CR, 3'UTR, 3'UTR1, 3'UTR3, and 3'UTR10), but it did not methylate other *p21* mRNA segments (5'UTR, 3'UTR2, 3'UTR4, 3'UTR5, 3'UTR6, 3'UTR7, 3'UTR8, 3'UTR9) or *p27* 3'UTR (Fig. 4B, left and right). METTL3/METTL14 methylated some *p21* 3'UTR segments (specifically 3'UTR10), but not other areas of *p21* mRNA (5'UTR, CR, 3'UTR1, 3'UTR2, 3'UTR3, 3'UTR4, 3'UTR5, 3'UTR6, 3'UTR7, 3'UTR8, 3'UTR9) or *p21* cDNA (Fig. 4C, left and right). Therefore, the shared region methylated by both NSUN2 and METTL3/METTL14 is *p21* 3'UTR10 (positions 1979–2158).

### METTL3/METTL14 AND NSUN2 COOPERATIVELY METHYLATE *p21* mRNA IN VITRO

To determine the formation of m5C or m6A in the NSUN2- or METTL3/METTL14-methylated fragments, the CR and 3'UTR fragments of *p21* mRNA were methylated in vitro by NSUN2 by using non-isotopic SAM; *p21* 3'UTR was methylated by flag-METTL3/flag-METTL14 in vitro by using non-isotopic SAM. These methylated fragments or unmethylated fragments (same reaction but without adding NSUN2 or METTL3/METTL14) were subjected to MS-HPLC analysis. As shown in Figure 5A, NSUN2 catalyzed the formation of m5C at the CR and 3'UTR of *p21* mRNA. Given that methylation of *p21* 3'UTR, but not *p21* CR, is involved in the regulation of *p21* expression by NSUN2 (Fig. 3B), we did not identify the methylation sites in CR and instead identified the methylation sites in 3'UTR10. As shown in Figure 5B (left) by bisulfite sequencing assays, C2079 was identified as the major methylation site in 3'UTR10 by NSUN2 in vitro. The observation that mutation of C2079 (C-G) greatly reduced the methylation of 3'UTR10 by NSUN2 further supported this view (Fig. 5B, right).

We then examined the formation of m6A in *p21* 3'UTR10 by METTL3 and METTL14 in vitro. As shown, METTL3/METTL14 catalyzes the formation of m6A in *p21* 3'UTR (Fig. 5C, left). The A residue in the middle of motifs AAC and GAC was recognized as the site for m6A methylation. Analysis of the sequence of 3'UTR10



**Fig. 4.** NSUN2 and METTL3/METTL14 methylate *p21* mRNA in vitro. (A) Schematic depiction of the *p21* mRNA fragments used for in vitro methylation assays. (B) In vitro methylation assays by using purified his-NSUN2 and the fragments shown in Figure 4A. Incorporation of <sup>3</sup>H-labeled SAM into *p21* 5'UTR, CR, and 3'UTR fragments (left) as well as 5'UTR, 3'UTR, 3'UTR1, 3'UTR2, 3'UTR3, 3'UTR4, 3'UTR5, 3'UTR6, 3'UTR7, 3'UTR8, 3'UTR9, and 3'UTR10 fragments (right). The incorporation of <sup>3</sup>H-labeled SAM into *p27* 3'UTR and *SHC* 5'UTR (*p66*-5'UTR) served as negative control or positive control, respectively. (C) In vitro methylation assays by using immunoprecipitated flag-METTL3 and flag-METTL14 as well as the fragments depicted in Figure 4A. Incorporation of <sup>3</sup>H-labeled SAM into *p21* cDNA (DNA), 5'UTR, CR, and 3'UTR fragments (left) as well as *p21* cDNA (DNA), 3'UTR, 3'UTR1, 3'UTR2, 3'UTR3, 3'UTR4, 3'UTR5, 3'UTR6, 3'UTR7, 3'UTR8, 3'UTR9, and 3'UTR10 fragments (right). The incorporation of <sup>3</sup>H-labeled SAM into *p21* cDNA (DNA) and RNA fragment ACGAGUCCUGGACUGAAACGGACUUGU served as a negative control and a positive control (PC), respectively.

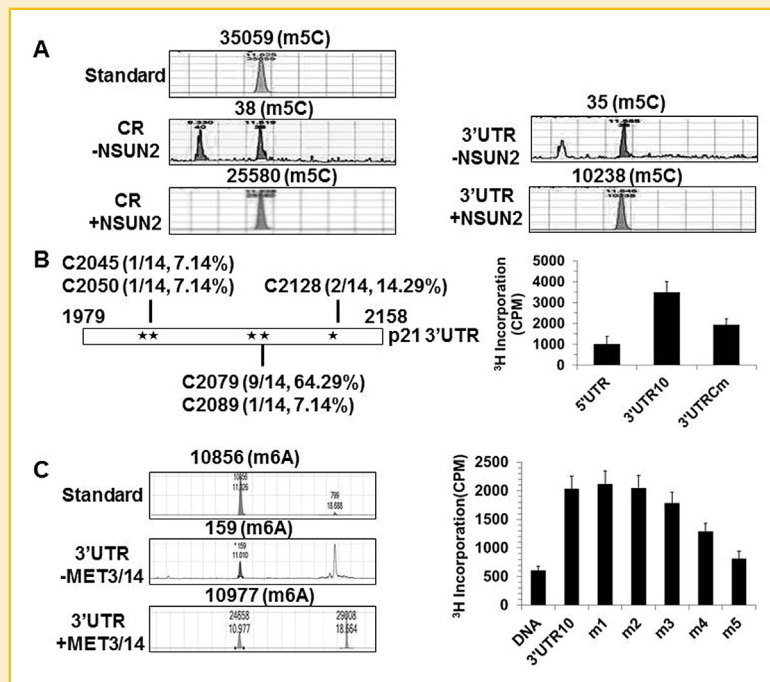
revealed that 3'UTR10 contained four potential sites for m6A formation (A1986, A2037, A2044, and A2061). A-to-U mutations at A2044 (m3) or A2061 (m4), but not at A1986 (m1) or A2037 (m2), reduced the methylation levels of 3'UTR10 by METTL3/METTL14; mutation of A2044 and A2061 together (m5) further reduced the methylation levels of 3'UTR10 by METTL3/METTL14 (Fig. 5C, right). These results suggest that A2044 or A2061 are the methylation sites of METTL3/METTL14.

Based on the findings that NSUN2 and METTL3 or METTL14 cooperatively regulated *p21* expression levels (Fig. 2A, B, E, and 3F) and that *p21* 3'UTR10 was identified as the common target fragment for NSUN2- and METTL3/METTL14-mediated RNA methylation (Fig. 4B and 4C), we tested the interaction between NSUN2-mediated m5C formation and METTL3/METTL14-mediated m6A formation. To this end, fragment 3'UTR10 methylated by NSUN2 and non-isotopic SAM was further subjected to in vitro methylation assays by using METTL3/METTL14 and <sup>3</sup>H-labeled SAM, and vice versa. As shown in Figure 6A and B, pre-methylation by NSUN2 augmented significantly the effect of METTL3/METTL14 in methylating *p21* 3'UTR10 (from 1967 CPM to 3073 CPM) (Fig. 6A); likewise, pre-methylation by METTL3/

METTL14 augmented significantly the effect of NSUN2 in methylating *p21* 3'UTR10 (2978 CPM compared with 4249 CPM) (Fig. 6B).

#### METTL3/METTL14 AND NSUN2 COOPERATIVELY ENHANCE *p21* TRANSLATION

In light of the fact that *p21* mRNA levels were not altered in cells silenced with NSUN2, METTL3, or METTL14, we tested whether methylation by NSUN2 or METTL3/METTL14 influenced the translation of *p21*. To this end, in vitro-transcribed reporter transcripts Luc-3'UTR (transcribed from pGL3-3'UTR) were methylated in vitro individually by NSUN2 or by METTL3/METTL14, or jointly by NSUN2 + METTL3/METTL14; unmethylated RNA was included in control reactions. These transcripts were then used for in vitro translation assays and reporter activities were used as readout of the efficiency of translation. As shown in Figure 6C, methylation by NSUN2 and METTL3/METTL14 led to ~2.5- and 2.2-fold luciferase activities, respectively; methylation by NSUN2 and METTL3/METTL14 showed ~3.3-fold the activity level measured in the control group. Furthermore, methylation of the Luc-3'UTR10Am (mutating A2044 and A2061) and Luc-3-UTR10Cm



**Fig. 5.** Measurement of m5C or m6A in NSUN2 or METTL3/METTL14 methylated fragments. (A) *p21* CR and 3'UTR fragments were in vitro-methylated by purified his-NSUN2 and non-isotopic SAM (+NSUN2) or kept untreated (–NSUN2), whereupon these fragments were subjected to HPLC–MS analysis to determine the formation of m5C. Data represent the peak value of m5C. (B) Left, *p21* 3'UTR10 fragment was in vitro methylated by non-isotopic SAM and subjected to bisulfite RNA sequencing analysis to identify the methylation sites, as described in "Materials and Methods." The percentage of methylation and the number of positive clones from the total clones sequenced are indicated. Right, in vitro methylation assays by using purified his-NSUN2 and fragments 5'UTR, 3'UTR10, and 3'UTR10 mutating C2079 (C–G, 3'UTRCm). (C) Left, *p21* 3'UTR10 fragment was in vitro-methylated by immunoprecipitated flag–METTL3/flag–METTL14 and non-isotopic SAM (+MET3/MET14) or kept untreated (–MET3/MET14), whereupon these fragments were subjected to HPLC–MS analysis to determine the formation of m6A. The peak values of m6A are indicated. Right, in vitro methylation assays by using immunoprecipitated flag–MET3/flag–MET14 as well as 3'UTR10 and its variants m1–m5.

(mutating C2079) by NSUN2, METTL3/METTL14, or NSUN2 and METTL3/METTL14 together was less effective than methylation of Luc-3'UTR10 under same conditions in elevating the translational efficiency (Fig. 6D–F). In conclusion, NSUN2-mediated m5C formation facilitates METTL3/METTL14-mediated m6A formation in *p21* 3'UTR, and vice versa; both types of methylation cooperate to further enhance *p21* translation.

#### NSUN2 AND METTL3/METTL14 COOPERATIVELY METHYLATE *p21* mRNA IN CELLS

To test if NSUN2 and METTL3/METTL14 methylated *p21* mRNA in cells, NSUN2, METTL14, or NSUN2 + METTL14 were silenced in HCT116 *p53*<sup>−/−</sup> cells. RNA was isolated and subjected to IP-based reverse transcription (RT) followed by real-time, quantitative (q)PCR analysis to assess m6A levels in *p21* mRNA, and subjected to bisulfite RNA sequencing analysis to assess the formation of m5C at C2079 of *p21* mRNA. As shown in Figure 7A, the anti-m6A antibody, but not the anti-IgG antibody, effectively immunoprecipitated the *p21* mRNA. As anticipated, knockdown of METTL14 reduced m6A methylation in *p21* mRNA by ~56%. In keeping with the findings that NSUN2-mediated m5C methylation augmented METTL3/METTL14-mediated m6A methylation of *p21* 3'UTR, knockdown of NSUN2 reduced significantly the levels of *p21* mRNA methylation at m6A by ~16%. Knockdown of NSUN2 and

METTL14 together was more effective than individually silencing NSUN2 or METTL14 in reducing the levels of m6A methylation of *p21* mRNA by ~79%. On the other hand, knockdown of NSUN2 or METTL3/METTL14 reduced m5C levels at C2079 (~26.3% vs. ~11.1% in Control vs. siNSUN2 and ~26.3% vs. ~18.8% in Control vs. siMETTL14) in cells expressing LUC-3'UTR (Fig. 7B). Joint knockdown of NSUN2 and METTL14 together further reduced the formation of m5C at C2079 (Control vs. siNSUN2 + siMETTL14, ~26.3% vs. ~7.1%) (Fig. 7B). In sum, METTL3/METTL14 and NSUN2 cooperatively methylate *p21* mRNA in cells.

#### NSUN2 AND METTL3/METTL14 COOPERATIVELY ENHANCE *p21* EXPRESSION LEVELS IN CELLULAR SENSENCE INDUCED BY OXIDATIVE STRESS

By methylating *SHC*, *p53*, and *p16* mRNAs, NSUN2 promotes the expression of *SHC*, *p53*, and *p16* in oxidative stress-induced cellular senescence [15]. Exposure of HCT116 *p53*<sup>−/−</sup> cells to H<sub>2</sub>O<sub>2</sub> (50 μM, 48 h) greatly increased *p21* levels (~4.2 fold) and NSUN2 (~2.5 fold) as well as the percentage of cells expressing senescence-associated β-galactosidase [(SA)–β-gal] (~7.21% in untreated cells, ~85.43% in H<sub>2</sub>O<sub>2</sub>-treated cultures) (Fig. 8A and B). Interestingly, cells exposed to oxidative stress also expressed higher levels of METTL3 (~2.9 fold) and METTL14 (~3.3 fold) (Fig. 8A). To test the impact of NSUN2- and METTL3/METTL14-mediated regulation of *p21* in



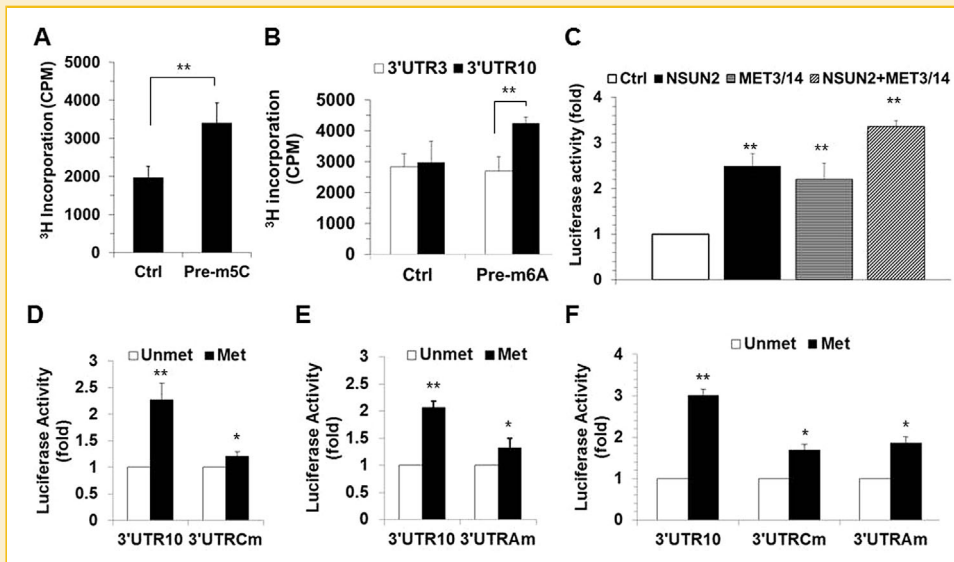


Fig. 6. NSUN2 and METTL3/METTL14 cooperatively methylate *p21* 3'UTR and thereby enhancing the translation of *p21*. (A) *p21* 3'UTR fragment was in vitro-methylated by NSUN2 (Pre-m5C) and non-isotopic SAM or kept untreated (Control). These fragments were further in vitro-methylated by flag-METTL3/METTL14 and <sup>3</sup>H-labeled SAM. Data represent the means ± SD from three independent experiments; significance was analyzed by Student's *t*-test (\*\**P* < 0.01). (B) *p21* 3'UTR3 or 3'UTR10 fragment was in vitro methylated by METTL3/METTL14 and non-isotopic SAM (Pre-m6A) or kept untreated (Control). These fragments were further in vitro methylated by flag-METTL3/METTL14 and <sup>3</sup>H-labeled SAM. Data represent the means ± SD from three independent experiments; significance was analyzed by Student's *t*-test (\*\**P* < 0.01). (C) Luc-3'UTR10 chimeric transcript transcribed from pGL3-3'UTR10 reporter was methylated by NSUN2, METTL3/METTL14 (MET3/14), NSUN2 + METTL3/METTL14 (NSUN2 + MET3/14), or kept untreated (Control). These transcripts then were used for in vitro translation assays. Luciferase activity was represented to reflect the efficiency of translation. (D) Luc-3'UTR10 (3'UTR10) or LUC-3'UTR10Cm (3'UTR10Cm, mutating C2079) was methylated in vitro by NSUN2 (Met) or kept untreated (Unmet), whereupon in vitro translation assays were performed. (E) Luc-3'UTR10 or Luc-3'UTR10Am was methylated by METTL3/METTL14 in vitro (Met) or kept untreated (Unmet), whereupon in vitro translation assays were performed. (F) Luc-3'UTR10, Luc-3'UTR10Am, or Luc-3'UTRCm was methylated by NSUN2 + METTL3/METTL14 in vitro (Met) or kept untreated (Unmet), whereupon in vitro translation assays were performed. Data in C–F represent the means ± SD from three independent experiments; significance was analyzed by Student's *t*-test (\**P* < 0.01; \*\**P* < 0.05).

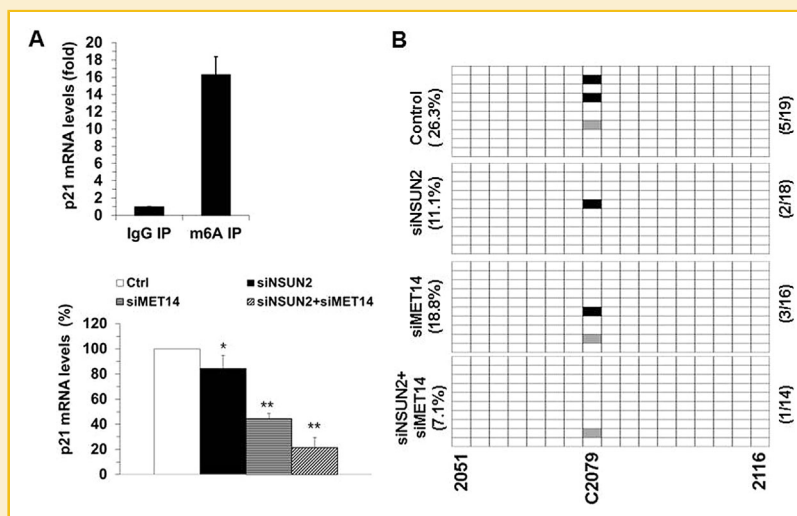
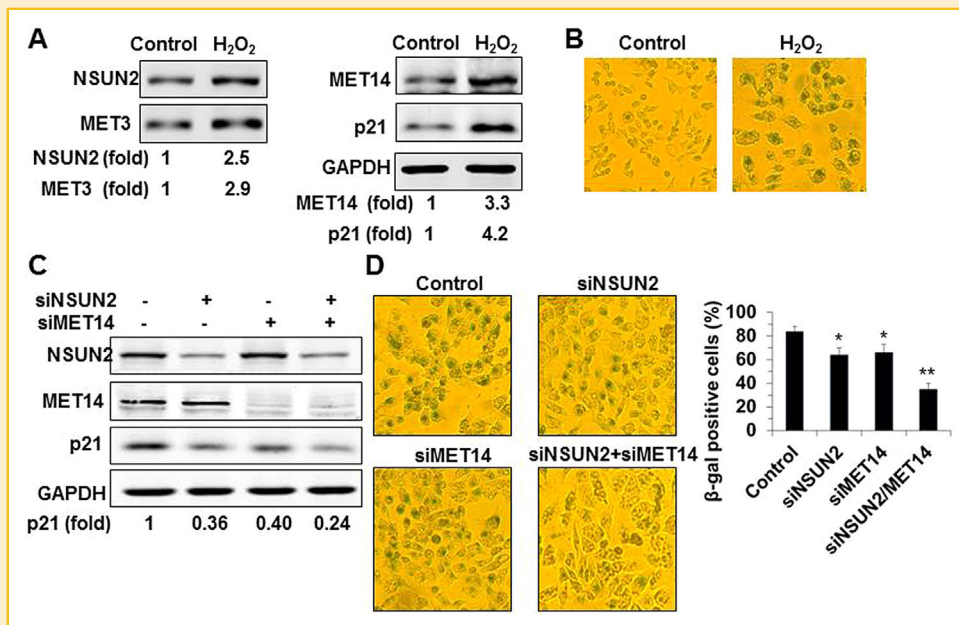


Fig. 7. NSUN2 and METTL3/METTL14 cooperatively methylate *p21* 3'UTR in vivo. (A) top, RNA isolated from HCT116 *p53*<sup>-/-</sup> cells was subjected to IP assays by using an antibody recognizing m6A or a control IgG antibody. The levels of *p21* mRNA in the IP materials normalized against were analyzed by RT-qPCR analysis. Data represent the means ± SD from three independent experiments. Bottom, HCT116 *p53*<sup>-/-</sup> cells were transfected with siRNAs targeting NSUN2 (siNSUN2) or METTL14 (siMET14) or both (siNSUN2 + siMET14). Forty-eight hour later, RNA was prepared and subjected to IP assays by using m6A antibody. The levels of *p21* mRNA were determined by RT-qPCR analysis. Data represent the means ± SD from three independent experiments; significance was analyzed by Student's *t*-test (\**P* < 0.05; \*\**P* < 0.01). (B) HCT116 *p53*<sup>-/-</sup> cells were transfected with a pGL3-3'UTR vector. Twenty four hour later, cells were further transfected with a NSUN2 siRNA, METTL14 siRNA, NSUN2 and METTL14 siRNA, or kept untreated (Control) and cultured for additional 48 h. RNA was isolated and subjected to bisulfite RNA sequencing to assess the methylation of C2079 (m5C). The percent of methylation and the number of positive colons from the total colons sequenced are indicated.



**Fig. 8.** NSUN2 and METTL3/METTL14 synergistically enhance the expression of p21 in oxidative stress-induced cell senescence. (A) HCT116 p53<sup>-/-</sup> cells were exposed to H<sub>2</sub>O<sub>2</sub> (50 μM) for 48 h. The levels of NSUN2, METTL3, METTL14, p21, and GAPDH were analyzed by Western blot analysis. Data are representatives from three independent experiments. (B) The cells described in Figure 8A were subjected to SA-β-gal analysis and the percentage of SA-β-gal-positive cells was counted. Data are representatives of 3 independent experiments. (C) HCT116 p53<sup>-/-</sup> cells were transfected with siRNAs to silence NSUN2 or METTL14 (siMET14) or both proteins. Twenty-four hour later, cells were exposed to H<sub>2</sub>O<sub>2</sub> (50 μM) and cultured for additional 48 h. The levels of NSUN2, METTL14, p21, and GAPDH were analyzed by Western blot analysis. Data are representatives of three independent experiments. (D) Cells described in Figure 8C were subjected to SA-β-gal analysis. Data represent the means ± SD from three independent experiments and statistical significance was analyzed by Student's *t*-test (\**P* < 0.05; \*\**P* < 0.01).

oxidative stress-induced cellular senescence, HCT116 p53<sup>-/-</sup> cells in which NSUN2 and METTL14 were silenced (alone or jointly) were exposed to H<sub>2</sub>O<sub>2</sub> (50 μM, 48 h) and p21 levels were determined by Western blot analysis. As shown in Figure 8C (left and right), H<sub>2</sub>O<sub>2</sub> treatment induced p21 less robustly in cells with silenced NSUN2 or METTL14, but cells in which NSUN2 + METTL14 were jointly silenced showed the weakest expression of p21, in agreement with the results shown in Figure 2B. Moreover, knockdown of NSUN2 or METTL14 lowered the amount of cells expressing senescence-associated β-galactosidase activity following H<sub>2</sub>O<sub>2</sub> treatment, but joint knockdown of NSUN2 and METTL14 most strongly suppressed the number of cells displaying senescence-associated β-galactosidase activity. Together, our findings indicate that modification of *p21* mRNA by NSUN2-mediated m5C and METTL3/METTL14-mediated m6A modification cooperatively enhance p21 translation, in turn promoting oxidative stress-induced cellular senescence.

## DISCUSSION

In the present study, we provide evidence that NSUN2 and METTL3/METTL14 cooperatively methylate *p21* mRNA in vitro and in cells, in turn enhancing p21 production at the level of translation (Figs. 1–7). Although the detailed mechanism whereby translation is regulated by these RNA modifications, m5C and m6A methylation mutually promote the expression of p21 and hence cell senescence following

oxidative damage (Fig. 8). Notably, methylation by NSUN2 in the coding region (CR) of p21 did not significantly influence the expression of p21 (Fig. 3B and 4B). Our previous studies revealed that methylation of the *ICAM1* CR by NSUN2 similarly had no effect on the regulation of *ICAM1* expression [Luo et al., 2016], although methylation of the CR of *SHC* mRNA by NSUN2 did enhance *SHC* translation [Cai et al., 2016]. Therefore, whether or not mRNA methylation influences mRNA fate may depend on the location of the methylated site or on additional factors interacting with the mRNA.

METTL3 and METTL14 form a stable heterodimer core complex that catalyzes the m6A modification of mammalian nuclear RNAs [Liu et al., 2014; Ping et al., 2014]. By interacting with METTL3/METTL14, WTAP regulates the activity of METTL3/METTL14-mediated RNA methylation [Liu et al., 2014; Ping et al., 2014]. Thus far, METTL3/METTL14-mediated RNA methylation was found to regulate the RNA processing and turnover of mRNAs encoding developmental regulators to help sustain embryonic stem cells (ESCs) [Lin and Gregory, 2014; Wang et al., 2014]. Interestingly, NSUN2-mediated RNA methylation is required for balancing stem cell self-renewal and differentiation in skin [Blanco et al., 2011, 2016]. It is plausible to postulate that NSUN2 and METTL3/METTL14 may cooperate to influence stem cell function by regulating p21 or other unidentified targets. In addition, elevation of p21 under stress conditions can promote cell survival. Thus, our findings that H<sub>2</sub>O<sub>2</sub> induce the expression of NSUN2, METTL3, and METTL14 support a function for these methyltransferases to upregulate p21 expression

in order to elicit cell survival. The fact that p21 is elevated in senescent cells while NSUN2 levels are reduced [Tang et al., 2015] suggests that NSUN2 and METTL3/METTL14 do not regulate p21 expression in replicative senescence.

Methylation of m6A is a predominant modification of RNAs. In different RNA species, the levels of m6A have been found to be 1–3 residues per 1,000 nucleotides [Carroll et al., 1990]. In addition to m6A, m5C is also a widespread modification of mRNAs [Squires et al., 2012; Tang et al., 2015; Xing et al., 2015; Cai et al., 2016; Luo et al., 2016; Zhao et al., 2017]. The present study indicates that NSUN2-mediated m5C methylation promotes METTL3/METTL14-mediated m6A methylation and reciprocally m6A methylation by METTL3/METTL14 enhances NSUN2-mediated m5C methylation, jointly inducing p21 translation. Because C2079 is adjacent to A2044 and A2061, the reciprocal influence between m5C and m6A methylation may depend on whether m5C and m6A sites are close enough. Given that NSUN2 and METTL3/METTL14 target numerous coding and non-coding RNAs for methylation [Dominissini et al., 2012; Meyer et al., 2012; Hussain et al., 2013; Liu et al., 2014; Ping et al., 2014], their impact on mRNAs jointly modified at m5C and m6A may not be limited to *p21* mRNA and affect other mRNAs widely.

## ACKNOWLEDGMENTS

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