

REVIEW

WILEY



Different resuscitation strategies and novel pharmacologic treatment with valproic acid in traumatic brain injury

Simone E. Dekker, MD, PhD^{1,2,3} | Vahagn C. Nikolian, MD¹ |

Martin Sillesen, MD, PhD^{4,5} | Ted Bambakidis, MSc¹ | Patrick Schober, MD, PhD³ |

Hasan B. Alam, MD¹ 

¹Department of Surgery, University of Michigan Hospital, Ann Arbor, Michigan

²Department of Neurological Surgery, Case Western Reserve University, Cleveland, Ohio

³Department of Anesthesiology, Institute for Cardiovascular Research, VU University Medical Center, Amsterdam, the Netherlands

⁴Department of Surgical Gastroenterology, Copenhagen University Hospital, Rigshospitalet, Copenhagen, Denmark

⁵Institute for Inflammation Research, Copenhagen University Hospital, Rigshospitalet, Copenhagen, Denmark

Correspondence

Hasan B. Alam, MD, Norman Thompson Professor of Surgery, Head of General Surgery, University of Michigan Hospital, 2920 Taubman Center, 1500 E. Medical Center Dr., Ann Arbor, MI 48109. Email: alamh@med.umich.edu

Funding information

Dr. Alam received numerous federal research grants from the National Institutes of Health (NIH), Office of Naval Research (ONR), Defence Advanced Research Projects Agency (DARPA), and the US Army Medical Research and Materiel Command (USAMRMC, which supported some of the data included in this manuscript).

Abstract

Traumatic brain injury (TBI) is a leading cause of death in young adults, and effective treatment strategies have the potential to save many lives. TBI results in coagulopathy, endothelial dysfunction, inflammation, cell death, and impaired epigenetic homeostasis, ultimately leading to morbidity and/or mortality. Commonly used resuscitation fluids such as crystalloids or colloids have several disadvantages and might even be harmful when administered in large quantities. There is a need for next-generation treatment strategies (especially in the prehospital setting) that minimize cellular damage, improve survival, and enhance neurological recovery. Pharmacologic treatment with histone deacetylase inhibitors, such as valproic acid, has shown promising results in animal studies of TBI and may therefore be an excellent example of next-generation therapy. This review briefly describes traditional resuscitation strategies for TBI combined with hemorrhagic shock and describes preclinical studies on valproic acid as a new pharmacologic agent in the treatment of TBI. It finally discusses limitations and future directions on the use of histone deacetylase inhibitors for the treatment of TBI.

KEYWORDS

histone deacetylase inhibitor, resuscitation, traumatic brain injury, valproic acid

1 | INTRODUCTION

Despite optimization of prehospital and intrahospital treatment strategies, traumatic brain injury (TBI) remains a leading cause of morbidity and mortality in young adults (Faul, Wald, & Coronado, 2010; Centers for Disease Control and Prevention, 2011). TBI is often paralleled by hemorrhagic shock (HS), and this combination is especially lethal, with studies showing that HS can significantly worsen TBI-associated morbidity and mortality (Wald, Shackford, & Fenwick, 1993; McMahon,

Significance

Traumatic brain injury (TBI) is a leading cause of death in young adults, and effective treatment strategies have the potential to save many lives. However, there is considerable debate about the most effective methods for treating blood loss associated with TBI. This article describes different resuscitation strategies and novel drug treatment for TBI using valproic acid. Limitations of current strategies as well as future research opportunities using valproic acid and related drugs are discussed.

Yates, Campbell, Hollis, Woodford, 1999). TBI and HS result in secondary conditions such as inflammation, endothelial dysfunction, coagulopathy, hypoxia, and cerebral edema. Such secondary insults are highly linked, and include cross-talk between endothelial and coagulation pathways as key mediators. The primary insult to the brain occurs in the prehospital setting, involves irreversible destruction of neuronal tissue and cannot be treated. Hence, treatment strategies necessarily focus on prevention of secondary brain injury by maintaining adequate cerebral oxygenation and perfusion (Badjatia 2008). Fluid resuscitation has historically been the mainstay in treating traumatic injuries. It plays a critical role in restoring and maintaining systemic and cerebral circulation in TBI patients (Myburgh et al., 2007). An ideal resuscitation fluid would therefore minimize cerebral edema, as well as attenuate neuronal damage, inflammation, and coagulopathy. However, current resuscitation strategies have failed to prevent this secondary brain injury. For this reason, recent advances in trauma research have focused on next-generation fluids and pharmacologic agents that may attenuate secondary brain injury. The aim of this review is to describe traditional resuscitation strategies for TBI combined with HS, and discuss the novel use of valproic acid (VPA) as a new pharmacologic agent in the treatment of TBI.

2 | CONVENTIONAL RESUSCITATION TREATMENTS

TBI is often associated with hypovolemia, HS, and coagulopathy. In addition, microvascular injury may alter the permeability of the blood-brain barrier (BBB), resulting in intravascular fluid leakage and cerebral edema. The ideal resuscitation strategy should aim to not only replace lost blood but also minimize secondary brain injury. Accordingly, current strategies focus on hemodynamic stabilization, maintaining systemic circulation, and optimizing cerebral perfusion (Hammell & Henning, 2009; Myburgh et al., 2007). However, there is considerable debate about the most effective methods for treating blood loss associated with TBI.

2.1 | Fluid resuscitation

Prompt fluid resuscitation is the first-line therapy to restore the lost intravascular volume. As blood products are often unavailable at the scene of the injury, prehospital management is usually limited to infusion using crystalloids or colloids. Isotonic crystalloids are typically preferred over colloids (Myburgh et al., 2007; Perel & Roberts, 2012) and have served as the standard resuscitation fluid for some time. Yet, traditional resuscitation fluids such as crystalloids have no inherent prosurvival properties (Santry & Alam, 2010), and in severely bleeding patients, aggressive crystalloid resuscitation does not result in any survival benefit (Bickell et al., 1994; Kwan, Bunn, & Roberts, 2003). Moreover, several clinical studies have identified shortcomings of fluid resuscitation, such as hemodilution, hemostatic derangements, brain edema, and inflammation (Grande, Asgeirsson, & Nordstrom, 1997; Peiniger et al., 2011). Such aggressive administration may even worsen outcomes by further exaggerating the cellular damage suffered during shock (Santry & Alam, 2010). Preclinical studies showed that ongoing hemorrhage and coagulopathy are

frequently not corrected by current resuscitation protocols using crystalloids and packed red blood cells (PRBCs) (Selby et al., 1996). In addition, massive fluid resuscitation itself may even result in coagulopathy and hemodilution (Selby et al., 1996; Sondeen, Coppes, & Holcomb, 2013). Compared with other resuscitation fluids such as fresh frozen plasma (FFP) or colloids, normal saline resuscitation is furthermore associated with increased brain swelling (Jin, DeMoya, et al. (2012; Imam et al. 2015); metabolic derangements (Hwabejire, Imam, et al., 2013b); elevated circulating markers of injury (Sillesen, Jin, et al., 2013); and increased activation of the endothelial, coagulation, and anticoagulation systems (Dekker, 2014c; Sillesen et al., 2014).

These findings suggest that traditional fluid resuscitation with crystalloids is mostly supportive, and does not address the specific cellular dysfunction caused by shock and injury. Because of the increased awareness of the negative effects of massive transfusions, trauma care now favors “damage control resuscitation” consisting of minimal use of crystalloids, early hemorrhage control, and early administration of blood products (PRBC, FFP, and platelets in an appropriate ratio) (Kwan et al., 2003).

2.2 | Blood product resuscitation

Commonly used blood products are PRBC, FFP, platelets, and fibrinogen. There is limited evidence concerning the use of PRBC in the setting of isolated TBI. The combination of TBI and HS likely mandates PRBC transfusion, with clinical data suggesting a survival benefit when FFP:PRBC ratios are maintained above 1:2 (Peiniger et al., 2011). The benefits of FFP on the control of acute traumatic bleeding and coagulopathy are furthermore demonstrated in multiple clinical studies (Borgman et al., 2007; Duchesne et al., 2008; Maegele et al., 2008; Scaglia et al., 2008; Wafaisade et al., 2010; Zehtabchi & Nishijima, 2009). In the setting of TBI, however, clinical data are conflicting. Some studies have indicated adverse outcomes associated with FFP administration (Anglin et al., 2013; Etemadzezaie et al., 2007), while others have identified outcome benefits (Peiniger et al., 2011). Preclinical studies demonstrated that early administration of FFP also appears to alter the clinical course following TBI by attenuating the degree of neurologic impairment, improving the rate of recovery, and preserving cognitive functions (Halaweish, 2015a). These effects of blood products can even be detected at the level of gene transcription (Sillesen, Bambakidis, Dekker, Li, & Alam, 2017), affecting the expression of genes involved in metabolism, platelet signaling, and inflammation. These data suggest that the therapeutic benefits of plasma resuscitation might be more extensive than solely through hemodynamic stabilization.

However, FFP transfusion is not without risk (Nascimento et al., 2010; Rossaint et al., 2016). Such risks include transfusion-associated acute lung injury and circulatory overload, allergic reactions, and transmission of infectious diseases (Nascimento et al., 2010). Moreover, the use of FFP and other blood products is challenging in prehospital settings or in underdeveloped areas because of logistical issues such as limited availability, requirement for refrigeration, short life after thawing, and the need for immediately available universal donor plasma. These challenges of FFP resuscitation have fueled the initiative to develop next-generation blood products that are low volume and shelf stable.

2.3 | Future directions of blood products and antifibrinolytics in traumatic bleeding

Lyophilized plasma, a freeze-dried plasma product developed in the 1930s, may solve the shortcomings of FFP resuscitation. It is logistically superior to FFP, as it can be stored as a powder for as long as 30 years, and subsequently reconstituted and administered within minutes following rehydration with water (Fu & Myhre, 1977). Additionally, lyophilized plasma retains its factor function much better than FFP, and can be stored in ambient temperatures for extended periods of time. Freeze-dried plasma has been shown to be as effective as FFP in large animal models of TBI and hemorrhage (Imam et al., 2013a; Halaweish et al., 2016). Freeze-dried plasma is approved for clinical use in Europe and has been used by NATO forces for many years with good results (Alam & Velmahos, 2011; Pusateri et al., 2016). However, it is currently not approved by the Food and Drug Administration (FDA) for use in the United States. In the coming years, there is an urgent need for prospective investigations of blood products such as lyophilized plasma and fibrinogen, and antifibrinolytic drugs such as tranexamic acid (TXA). There is currently a paucity of data concerning the use of TXA in TBI. A recent Cochrane review concluded that TXA may reduce mortality, although the quality of the evidence is low and uncertainties remain (Ker, Roberts, Shakur, & Coats, 2015). Furthermore, studies have suggested a reduction in the incidence of intracranial hemorrhage following TBI associated with administration of TXA (Zehtabchi, Abdel Baki, Falzon, & Nishijima, 2014). Future randomized controlled trials should report on the clinical outcomes of treatment with TXA and fibrinogen to address some of the key unanswered questions in trauma-induced coagulopathy (Wong, Curry, & Stanworth, 2016).

3 | NOVEL RESUSCITATION TECHNIQUES—EPIGENETIC MODULATION USING VPA

At present, there are no proven pharmacologic treatment options for TBI. However, the search for such therapeutic treatment of TBI has received considerable attention in recent years (Fukudome et al., 2012; Ichiyama et al., 2000). A therapeutic strategy for modulating the cellular response to injury may be at the level of the epigenome. One class of promising drugs that affect the epigenome is histone deacetylase inhibitors (HDACIs). In pathologic conditions, such as TBI and HS, there is a decrease in histone acetylation, which limits gene transcription and impairs cellular homeostasis. However, agents such as VPA have been shown to act as HDACIs, thus altering gene transcription and thereby inducing a “prosurvival phenotype.” The precise mechanisms underlying these protective effects are an active area of current research.

3.1 | TBI creates a genomic storm

The genomic response to injury is an important area of current research. Recent studies have revealed that traumatic injuries result in epigenetic changes via DNA methylation, phosphorylation, and acetylation (Wong & Langley, 2016). The so-called “genomic storm” following

injury occurs on an epigenetic level and modulates numerous cellular functions, protein expression, and pathways. These processes are considered “epigenetics,” meaning that they affect the gene expression and resultant phenotype of the cell but do not alter the genome itself (Bernstein, Meissner, & Lander, 2007; Goldberg, Allis, & Bernstein, 2007). Alam et al. showed that during hemorrhage and resuscitation, at least 7% of the genes and downstream pathways are differentially expressed (Alam, Stegalkina, Rhee, & Koustova, 2002). Xiao et al. (2011) previously showed that after traumatic injury, expression of over 80% of the leukocyte transcriptome is altered. These changes occur rapidly after trauma and exhibit a long-lasting effect for months after the injury (Lipponen, Paananen, Puhakka, & Pitkänen, 2016; Xiao, 2011). Modulating these epigenetic changes may thus be an important goal of therapy in patients with TBI.

3.2 | Acetylation status regulates gene expression and protein functions

Briefly, the human genome is organized into chromatin, a highly conserved complex of DNA and histone proteins. DNA transcription, and hence gene expression, is regulated by the acetylation and deacetylation of these histones. Histone deacetylases are enzymes that remove the acetyl group from the histone, making the DNA-histone structure more compact and therefore limiting transcription. In contrast, histone acetyltransferases weaken DNA-histone attraction, thereby unfolding the complex and making DNA more available for transcription (Delcuve, Khan, & Davie, 2012). A homeostatic balance between histone deacetylases and histone acetyltransferases typically exists in cells. However, dysregulation of acetylation homeostasis has been suggested to play a key role in the pathogenesis of cancers and neurodegenerative diseases. Moreover, as noted above, dysregulation of acetylation homeostasis has also been implicated as an important pathologic mechanism in the body's response to traumatic injuries. Thus, pharmacologic agents that can alter histone acetylation may be promising new therapeutic strategies for trauma, as they can rapidly and reversely modify the transcription of desirable genes.

Although histones were considered the main target of HDACIs, we now know that nonhistone proteins involved in key cellular functions such as the cell cycle, stress response, cytoskeleton dynamics, signaling, repair/healing/remodeling, communication, and proliferation are equally involved (Glozak, Sengupta, Zhang, & Seto, 2005). At least 50 such nonhistone proteins have been well characterized (Kim & Bae, 2011), but this list is rapidly growing. In one study, 3,600 acetylation sites (in 1,750 human proteins) were identified, which regulated nearly all nuclear and many key cytoplasmic processes (Choudhary, 2009). This makes acetylation a regulatory mechanism that is as prevalent and important as phosphorylation (Kouzarides, 2000; Norris, Lee, & Yao, 2009). Acetylation of nonhistone proteins is actually even more important than histones in the setting of severe shock and/or TBI. While it takes some time to produce phenotype changes through modulation of gene transcription, direct (nontranscriptional) acetylation of regulatory proteins is extremely fast and thus more relevant for rapidly lethal conditions such as shock or TBI. In short, there are many features that

TABLE 1 Key effects of valproic acid treatment following central nervous system injury, with selected studies

Effect	Model	Reference
Increased cell survival, and decreased apoptosis and necrosis	TBI animal model	Dekker et al., 2014b
	Spinal cord injury animal model	Abdanipour et al., 2012
	Hypoxia-induced neuronal apoptosis in vitro model	Jin et al., 2014; Li et al., 2008
Decreased brain lesion size	TBI animal model	Halaweish et al., 2015b; Imam et al., 2013b; Jepsen et al., 2014; Jin et al., 2012a; Tai et al., 2014; Yu et al., 2013
Improved survival, faster recovery, improved cognitive function	TBI animal model	Dash et al., 2010; Halaweish et al., 2015b; Tai et al., 2014; Yu et al., 2013
	Spinal cord injury animal model	Abdanipour et al., 2012; Darvishi et al., 2014
Reduced inflammation	TBI animal model	Bambakidis et al., 2016; Jin et al., 2012a; Nikolian et al., 2016b; Tai et al., 2014
	Glioma cell in vitro model	Ichiyama et al., 2000
	Spinal cord injury animal model	Abdanipour et al., 2012; Darvishi et al., 2014
Restoration of BBB function	TBI animal model	Nikolian et al., 2016; Yu et al., 2013
	Focal cerebral ischemia animal model	Wang et al., 2011
Attenuated platelet dysfunction	Spinal cord injury animal model	Lee et al., 2012
	TBI animal model	Bambakidis et al., 2016; Dekker et al., 2014a; Sillesen, Johansson, et al., 2013
Decreased metabolism and attenuated mitochondrial dysfunction	TBI animal model	Dekker et al., 2014b; Hwabejire, Jin, et al., 2013b
Modulation of cell signaling	TBI animal model	Dekker et al., 2014b; Nikolian et al., 2016b

BBB = blood-brain barrier; TBI = traumatic brain injury.

make acetylation an attractive therapeutic target: It is *rapid*, completely *reversible*, and can be altered by drugs that are already in clinical use.

3.3 | VPA restores normal cellular acetylation

A growing body of literature suggests that VPA may alter gene expression and protein functions following TBI (Bambakidis et al., 2016; Dekker, 2014b; Halaweish et al., 2015b). VPA has been in clinical use as a mood-stabilizing and antiepileptic drug since the 1970s. In high doses, however, VPA acts as a HDACI. High-dose VPA has been shown to improve survival in otherwise lethal models of HS, polytrauma (Alam et al., 2009, 2011), sepsis, and combined TBI with HS (Halaweish et al., 2015b,c; Jepsen et al., 2014; Jin, Duggan, et al., 2012a). Yet, the precise protective mechanisms of VPA have not been well defined. A conceptual model for the main protective effects of VPA is as follows: TBI and HS result in decreased acetylation of histones and nonhistone proteins, which impairs normal gene expression and alters many homeostatic pathways, leading to cell death. In contrast, VPA acts as an HDACI, which restores normal acetylation homeostasis and restores normal cellular functions, ultimately leading to a pro-survival phenotype. HDACIs, such as VPA, are a promising therapeutic approach as they are already FDA approved for clinical use for a variety of other illnesses. Importantly, VPA is effective as treatment for HS even when

administered in a single bolus in the absence of fluid resuscitation or blood transfusion (Alam et al., 2009). Thus, VPA treatment is very appealing as it can be rapidly administered in the prehospital setting or battlefield environment. Table 1 summarizes the key benefits of pharmacologic treatment with VPA in the setting of central nervous system injury.

3.4 | VPA alters cell survival and cell death pathways

One of the first studies that investigated the effects of hemorrhage and resuscitation on histone acetylation demonstrated that crystalloid resuscitation results in a predominantly deacetylation profile in animal models, and that treatment with HDACIs was able to reverse it (Lin et al., 2006). Our research group sought to better understand this effect of HDACI treatment at the level of gene expression. Using a porcine model of TBI+HS, we performed a high-throughput analysis of cerebral gene profiling following resuscitation with either hextend or hextend+VPA. We hypothesized that treatment with VPA would significantly alter the early transcription of genes in pathways related to cell survival, which may explain its previously observed neuroprotective effects such as reduced brain lesion size and swelling. We found that 1,668 probe sets mapping to 370 known genes were differentially expressed between hextend versus hextend+VPA groups. These genes

were mapped to, among others, pathways related to cell death, apoptosis, and necrosis (Dekker, 2014b). These findings support our hypothesis that VPA alters important genes and pathways that could improve survival.

3.5 | VPA reduces inflammation and BBB dysfunction

As described previously, secondary brain injury includes an acute inflammatory response with BBB disruption, activation of immune cells, and cerebral edema. The anti-inflammatory properties of VPA treatment have been described previously in models of sepsis, hemorrhage, and TBI (Jin, Duggan, et al., 2012; Liu et al., 2014; Shang et al., 2010). In addition, Bambakidis et al. (2016) showed that VPA modulates genes related to inflammation, cell signaling, cell adhesion, and endothelial growth. Dash et al. (2010) furthermore showed that VPA treatment significantly decreased Evans Blue dye extravasation in a TBI rat model, indicating that VPA might restore BBB function following trauma. Protective effects of VPA on BBB function were also demonstrated by Wang Leng, Tsai, Leeds, & Chuang, 2011 and Lee et al., who found a VPA-induced reduction of matrix metalloproteinase-9, a protease that disrupts BBB function (Lee et al., 2012; Wang, et al., 2011). This was furthermore associated with decreased degradation of tight junction and basement membrane-associated proteins, such as zona occludin-1 (ZO-1), and claudin-5 (Lee et al., 2012). This is in line with data from Nikolian and Dekker et al, 2016., who found that VPA significantly increases expression of ZO-1, laminin, and claudin-5. Expression of glucose transporter 1, a marker of barrier-type endothelial cells, was also increased in the VPA treatment group. Moreover, *in vitro* monolayers treated with VPA significantly decreased permeability relative to anoxic controls (Nikolian and Bruhn et al, 2016). Taken together, these results suggest that protective mechanisms of VPA may involve decreasing inflammation and correcting BBB dysfunction.

3.6 | VPA attenuates platelet dysfunction

Coagulopathy plays a major role in the mortality of patients with TBI and HS. A particularly important mechanism of TBI-related coagulopathy is platelet dysfunction. For example, TBI and HS induce a combination of platelet activation but decreased function compared with general trauma patients (no TBI) (Kutcher et al., 2012). The precise mechanisms of this platelet dysfunction remain unclear, but it may be mediated by the so-called "exhausted platelet syndrome." This syndrome involves initial platelet hyperactivation with subsequent depletion of intracellular mediators, ultimately resulting in platelet hypofunction (Pareti, Capitanio, Mannucci, Ponticelli, & Mannucci, 1980). Sillesen et al. showed that VPA may improve platelet function after TBI and HS, but the precise mechanisms remain unknown (Sillesen, Johansson, et al., 2013). One of VPA's protective mechanisms might be its effect on coagulopathy by preventing platelet hyperactivation, which would thereby preserve long-term platelet function. Dekker et al. (2014a) demonstrated that the addition of VPA to FFP resuscitation results in preservation of platelet activation 8 hr after the TBI, compared with FFP alone. This was reflected in both circulatory as well

as cerebral-level platelet activation. However, it remains unclear whether this was a direct effect of VPA on platelets, or the establishment of an overall prosurvival phenotype in animals treated with VPA. Bambakidis et al. (2017) recently conducted *ex vivo* experiments to test the direct effect of VPA on platelet function and coagulation. Results showed that VPA attenuates platelet activation and improves clot dynamics (strength and rate of formation) in blood from animals with TBI and HS. Importantly, VPA did not appear to alter platelet or coagulation function in blood from healthy controls.

3.7 | VPA improves neurological recovery

While recent studies demonstrated that VPA treatment reduces brain lesion size and attenuates damage to tissues, cells, and proteins, understanding longer-term functional outcomes remains an important hurdle to clinical translatability. Halaweish et al. (2015b) recently conducted a 30-day survival model of TBI+HS. Compared with normal saline resuscitation, VPA resuscitation (150 mg/kg) resulted in significantly decreased neurological impairment, significantly faster rate of neurologic recovery, and smaller brain lesion size. Moreover, although NS- and VPA-treated animals reached similar final cognitive function scores, the VPA group reached cognitive normalization significantly faster than the NS controls. In addition, small animal studies showed improved spatial memory (Dash et al., 2010) and functional recovery (Dash et al., 2010; Tai et al., 2014; Yu et al., 2013) when VPA was added to the treatment protocol. In animal models of spinal cord trauma, VPA treatment was associated with reduced secondary damage, improved locomotor scores (Abdanipour, Schluesener, & Tiraihi, 2012; Darvishi, Tiraihi, Mesbah-Namin, Delshad, & Taheri, 2014), and more rapid recovery (Abdanipour et al., 2012) (Table 1).

4 | FUTURE DIRECTIONS

4.1 | VPA treatment shows a promising translation to human patients

One of the challenges with new treatment strategies is the translation of outcomes from animal models to patients in the clinical setting. Animal models are imperfect, and there are several differences between porcine and human species in both physiology and genome. For example, the porcine physiology is hypercoagulable relative to the human coagulation system. One of the main limitations of research in traditional and pharmacological resuscitation is the lack of human studies. Importantly, Sillesen et al. (2016) demonstrated that histone deacetylase gene expression patterns are also associated with outcomes in actual trauma patients. Furthermore, our laboratory is currently conducting a U.S. FDA-approved phase 1, double-blind, placebo-controlled trial to evaluate the safety and tolerability of VPA in healthy volunteers and trauma patients. The first results of this study showed that VPA caused differential expression of a total of 173 proteins. Gene enrichment analysis from these human subjects at 4 hr post infusion showed an upregulation of pathways related to cell death, apoptosis, necrosis, and abnormal morphology of cells and neurons. Eight hours post

infusion, steroid metabolism, lipid synthesis, and vitamin metabolism were also upregulated (Georgoff et al., 2016). In part 2 of the ongoing phase 1 trial, the effects of VPA in trauma patients with HS will be studied (ClinicalTrials.gov identifier NCT01951560). Phase 2 and 3 trials have already been approved for funding and will commence in the next few years.

4.2 | VPA dosing needs further refinement

Most experiments described in this review used a dose of 300 mg/kg, which is 6 times higher than the dose used in humans for the treatment of seizures. While these high doses improve outcomes in animal models, they may be associated with side effects in human patients. For example, several investigators have shown that 300-mg/kg and 400-mg/kg doses of VPA may be toxic, with respiratory and cardiac arrest occurring shortly after infusion (Burns, Baer, Darlington, Dubick, & Wade, 2012). Acute VPA overdose results in hypotension, respiratory depression, thrombocytopenia, and metabolic disorders (Manoguerra et al., 2008). In addition, chronic VPA use (> 4 weeks) has been associated with thrombocytopenia, platelet dysfunction (Davidson et al., 2011; Gesundheit, Kirby, Lau, Koren, & Abdelhaleem, 2002; Gidal et al., 1994; Kis et al., 1999; Manoguerra et al., 2008), and acquired von Willebrand disease (Serदारoglu, Tutuncuoglu, Kavakli, & Tekgul, 2002; Verrotti et al., 1999). A dose as low as 60 mg/kg a day has been found to be biologically active in cancer studies, which led to our hypothesis that lower doses might also be beneficial for trauma patients. As detailed above, our recent test of lower-dose VPA (150 mg/kg) in a TBI+HS survival model showed promising results in terms of neurologic recovery, cognitive outcomes, and lesion size (Halaweish et al., 2015b). A low-dose VPA strategy was further supported by the first results of our phase 1 trial demonstrating that VPA is biologically effective in healthy humans when given at a dose of 120 mg/kg.

4.3 | There are potential problems with pan-HDACI treatment

VPA is cheap, well tested in multiple animal models, and approved worldwide by regulatory agencies. However, it has been shown to affect several different types of histones. Briefly, there are five different histone deacetylase classes: Class I, IIa, IIb, III, and IV. Within these classes, there are 18 different histone deacetylase isoforms (HDAC 1–11 and SIRT 1–7). All isoforms have very different physiological functions, cellular locations, and organ distributions (Halaweish et al., 2015c). As VPA is a nonspecific pan-inhibitor, it may create significant potential for toxicity and side effects. An important area of current research is the development of isoform-specific HDACIs that are both more potent and also target specific organs. Our research group is currently comparing isoform-specific HDACIs and pan-HDACIs, and investigating the synergistic effects between various HDACIs and other cytoprotective strategies.

In particular, the use of HDAC6 inhibition has been shown to be effective in models of sepsis (Li et al., 2015). Previous agents used in such studies have demonstrated poor brain bioavailability and, as such,

have had limited application in the setting of TBI. Recent studies evaluating the use of agents that may provide a higher brain bioavailability (ACY-183, Acetylon Pharmaceuticals) have shown promise, with pre-clinical and in vitro data demonstrating neuroprotective properties that are superior to nonselective inhibitors such as VPA (Nikolian and Bruhn et al, 2016).

5 | CONCLUSION

TBI results in coagulopathy, endothelial dysfunction, inflammation, cell death, and impaired epigenetic homeostasis, ultimately leading to severe injury or death. Traditional resuscitation fluids such as crystalloids and colloids are unable to reverse these imbalances, and might even be harmful when administered in large quantities. There is a need to develop next-generation resuscitation strategies that can minimize cellular damage, improve survival, and be administered in the prehospital setting. A next-generation resuscitation protocol should involve a combination of fluid resuscitation, blood products, and powerful pharmacologic agents. HDACIs, such as VPA, have shown promising results after injury and are therefore excellent examples of these next-generation therapies. However, there is need for further refinement, such as isoform-specific treatment, and further safety and efficacy testing in human patients.

CONFLICT OF INTEREST

None of the authors report any conflicts of interest. Dr. Alam has received numerous federal research grants from the National Institutes of Health (NIH), Office of Naval Research (ONR), Defence Advanced Research Projects Agency (DARPA), and the U.S. Army Medical Research and Materiel Command (USAMRMC).

AUTHOR CONTRIBUTIONS

All authors had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Study concept and design: S.E.D., H.B.A. Drafting of the manuscript: S.E.D., V.C.N., M.S., T.B. Critical revision of the manuscript for important intellectual content: V.C.N., M.S., T.B., P.S., H.B.A. Obtained funding: H.B.A. Study supervision: H.B.A.

REFERENCES

- Abdanipour, A., Schluesener, H. J., & Tiraihi, T. (2012). Effects of valproic acid, a histone deacetylase inhibitor, on improvement of locomotor function in rat spinal cord injury based on epigenetic science. *Iranian Biomedical Journal*, *16*, 90–100.
- Alam, H. B., Stegalkina, S., Rhee, P., & Koustova, E. (2002). cDNA array analysis of gene expression following hemorrhagic shock and resuscitation in rats. *Resuscitation*, *54*, 195–206.
- Alam, H. B., Shuja, F., Butt, M. U., Duggan, M., Li, Y., Zacharias, N., ... Velmahos, G. C. (2009). Surviving blood loss without blood transfusion in a swine poly-trauma model. *Surgery*, *146*, 325–333.
- Alam, H. B., & Velmahos, G. C. (2011). New trends in resuscitation. *Current Problems in Surgery*, *48*, 531–564.

- Alam, H. B., Hamwi, K. B., Duggan, M., Fikry, K., Lu, J., Fukudome, E. Y., ... Velmahos, G. (2011). Hemostatic and pharmacologic resuscitation: Results of a long-term survival study in a swine polytrauma model. *Journal of Trauma*, *70*, 636–645.
- Anglin, C. O., Spence, J. S., Warner, M. A., Paliotta, C., Harper, C., Moore, C., ... Diaz-Arrastia, R. (2013). Effects of platelet and plasma transfusion on outcome in traumatic brain injury patients with moderate bleeding diatheses. *Journal of Neurosurgery*, *118*, 676–686.
- Badjatia, N., Carney, N., Crocco, T. J., Fallat, M. E., Hennes, H. M., Jagoda, A. S., ... Wright, D. W. (2008). Guidelines for prehospital management of traumatic brain injury 2nd edition. *Prehospital Emergency Care*, *12*(Suppl. 1), S1–S52.
- Bambakidis, T., Dekker, S. E., Sillesen, M., Liu, B., Johnson, C. N., Jin, G., ... Alam, H. B. (2016). Resuscitation with valproic acid alters inflammatory genes in a porcine model of combined traumatic brain injury and hemorrhagic shock. *Journal of Neurotrauma*, *33*, 1514–1521.
- Bambakidis, T., Dekker, S. E., Halaweish, I., Liu, B., Nikolian, V. C., Georgoff, P. E., ... Alam, H. B. (2017). Valproic acid modulates platelet and coagulation function ex vivo. *Blood Coagulation & Fibrinolysis*, *28*(6), 479–484. doi: 10.1097/MBC.0000000000000626
- Bernstein, B. E., Meissner, A., & Lander, E. S. (2007). The mammalian epigenome. *Cell*, *128*, 669–81.
- Bickell, W. H., Wall, M. J., Jr., Pepe, P. E., Martin, R. R., Ginger, V. F., Allen, M. K., & Mattox, K. L. (1994). Immediate versus delayed fluid resuscitation for hypotensive patients with penetrating torso injuries. *New England Journal of Medicine*, *331*, 1105–1109.
- Borgman, M. A., Spinella, P. C., Perkins, J. G., Grathwohl, K. W., Repine, T., Beekley, A. C., ... Holcomb, J. B. (2007). The ratio of blood products transfused affects mortality in patients receiving massive transfusions at a combat support hospital. *Journal of Trauma*, *63*, 805–813.
- Burns, J. W., Baer, L. A., Darlington, D. N., Dubick, M. A., & Wade, C. E. (2012). Screening of potential small volume resuscitation products using a severe hemorrhage sedated swine model. *International Journal of Burns and Trauma*, *2*, 59–67.
- Centers for Disease Control and Prevention. (2011). Nonfatal traumatic brain injuries related to sports and recreation activities among persons aged ≤ 19 years—United States, 2001–2009. *MMWR Morbidity and Mortality Weekly Report*, *60*, 1337–1342.
- Choudhary, C., Kumar, C., Gnad, F., Nielsen, M. L., Rehman, M., Walther, T. C., ... Mann, M. (2009). Lysine acetylation target protein complexes and co-regulates major cellular functions. *Science*, *325*, 834–840.
- Darvishi, M., Tiraihi, T., Mesbah-Namin, S. A., Delshad, A., & Taheri, T. (2014). Decreased GFAP expression and improved functional recovery in contused spinal cord of rats following valproic acid therapy. *Neurochemical Research*, *39*, 2319–2333.
- Dash, P. K., Orsi, S. A., Zhang, M., Grill, R. J., Pati, S., Zhao, J., & Moore, A. N. (2010). Valproate administered after traumatic brain injury provides neuroprotection and improves cognitive function in rats. *PLoS One*, *5*, e11383.
- Davidson, D. C., Hirschman, M. P., Spinelli, S. L., Morrell, C. N., Schifitto, G., Phipps, R. P., & Maggirwar, S. B. (2011). Antiplatelet activity of valproic acid contributes to decreased soluble CD40 ligand production in HIV type 1-infected individuals. *Journal of Immunology*, *186*, 584.
- Dekker, S. E., Sillesen, M., Bambakidis, T., Boer, C., Johansson, P. I., Jin, G., ... Alam, H. B. (2014a). Treatment with a histone deacetylase inhibitor, valproic acid, is associated with increased platelet activation in a large animal model of traumatic brain injury and hemorrhagic shock. *Journal of Surgical Research*, *190*, 312–318.
- Dekker, S. E., Bambakidis, T., Sillesen, M., Liu, B., Johnson, C. N., Jin, G., ... Alam, H. B. (2014b). Effect of pharmacologic resuscitation on the brain gene expression profiles in a swine model of traumatic brain injury and hemorrhage. *Journal of Trauma and Acute Care Surgery*, *77*, 906–912.
- Dekker, S. E., Sillesen, M., Bambakidis, T., Jin, G., Liu, B., Boer, C., ... Alam, H. B. (2014c). Normal saline influences coagulation and endothelial function after traumatic brain injury and hemorrhagic shock in pigs. *Surgery*, *156*, 556–563.
- Delcuve, G. P., Khan, D. H., & Davie, J. R. (2012). Roles of histone deacetylases in epigenetic regulation: Emerging paradigms from studies with inhibitors. *Clinical Epigenetics*, *4*, 5.
- Duchesne, J. C., Hunt, J. P., Wahl, G., Marr, A. B., Wang, Y. Z., Weintraub, S. E., ... McSwain, N. E., Jr. (2008). Review of current blood transfusions strategies in a mature level I trauma center: Were we wrong for the last 60 years? *Journal of Trauma*, *65*, 272–278.
- Etemadrezai, H., Baharvahdat, H., Shariati, Z., Lari, S. M., Shakeri, M. T., & Ganjeifar, B. (2007). The effect of fresh frozen plasma in severe closed head injury. *Clinical Neurology and Neurosurgery*, *109*, 166–171.
- Faul, M. X. L., Wald, M. M., & Coronado, V. G. (2010). *Traumatic brain injury in the United States: Emergency department visits, hospitalizations, and deaths*. Atlanta, GA: Centers for Disease Control and Prevention, National Center for Injury Prevention and Control.
- Fu, P., & Myhre, B. (1977). Chemical analysis of a 30-year-old bottle of lyophilized plasma. *Transfusion*, *17*, 32–45.
- Fukudome, E. Y., Li, Y., Kochanek, A. R., Lu, J., Smith, E. J., Liu, B., ... Alam, H. B. (2012). Pharmacologic resuscitation decreases circulating cytokine-induced neutrophil chemoattractant-1 levels and attenuates hemorrhage-induced acute lung injury. *Surgery*, *152*, 254–261.
- Georgoff, P., Halaweish, I., Nikolian, V., Higgins, G., Bonham, T., Tafatia, C., ... Alam, H. B. (2016). Alterations in the human proteome following administration of valproic acid. *Journal of Trauma and Acute Care Surgery*, *81*, 1020–1027.
- Gesundheit, B., Kirby, M., Lau, W., Koren, G., & Abdelhaleem, M. (2002). Thrombocytopenia and megakaryocyte dysplasia: An adverse effect of valproic acid treatment. *Journal of Pediatric Hematology/Oncology*, *24*, 589.
- Gidal, B., Spencer, N., Maly, M., Pitterle, M., Williams, E., Collins, M., & Jones, J. (1994). Valproate-mediated disturbances of hemostasis: Relationship to dose and plasma concentration. *Neurology*, *44*, 1418.
- Glozak, M. A., Sengupta, N., Zhang, X., & Seto, E. 2005. Acetylation and deacetylation of non-histone proteins. *Gene*, *363*, 15–23.
- Goldberg, A. D., Allis, C. D., & Bernstein, E. (2007). Epigenetics: A landscape takes shape. *Cell*, *128*, 635–638.
- Grande, P. O., Asgeirsson, B., & Nordstrom, C. H. (1997). Physiologic principles for volume regulation of a tissue enclosed in a rigid shell with application to the injured brain. *Journal of Trauma*, *42*, S23–S31.
- Halaweish, I., Bambakidis, T., He, W., Linzel, D., Chang, Z., Srinivasan, A., ... Alam, H. B. (2015a). Early resuscitation with fresh frozen plasma for traumatic brain injury combined with hemorrhagic shock improves neurologic recovery. *Journal of the American College of Surgeons*, *220*, 809–819.
- Halaweish, I., Bambakidis, T., Chang, Z., Wei, H., Liu, B., Li, Y., ... Alam, H. B. (2015b). Addition of low-dose valproic acid to saline resuscitation provides neuroprotection and improves long-term outcomes in a large animal model of combined traumatic brain injury and hemorrhagic shock. *Journal of Trauma and Acute Care Surgery*, *79*, 911–919.
- Halaweish, I., Nikolian, V., Georgoff, P., Li, Y., & Alam, H. B. (2015c). Creating a “prosurvival phenotype” through histone deacetylase inhibition: Past, present, and future. *Shock*, *44*, 6–16.

- Halaweish, I., Bambakidis, T., Nikolian, V. C., Georgoff, P., Bruhn, P., Piascik, P., ... Alam, H. B. (2016). Early resuscitation with lyophilized plasma provides equal neuroprotection compared with fresh frozen plasma in a large animal survival model of traumatic brain injury and hemorrhagic shock. *Journal of Trauma and Acute Care Surgery*, *81*, 1080–1087.
- Hammell, C. L., & Henning, J. D. (2009). Prehospital management of severe traumatic brain injury. *BMJ*, *338*, b1683.
- Hwabejire, J. O., Imam, A. M., Jin, G., Liu, B., Li, Y., Sillesen, M., ... Alam, H. B. (2013a). Differential effects of fresh frozen plasma and normal saline on secondary brain damage in a large animal model of polytrauma, hemorrhage and traumatic brain injury. *Journal of Trauma and Acute Care Surgery*, *75*, 968–974.
- Hwabejire, J. O., Jin, G., Imam, A. M., Duggan, M., Sillesen, M., Deperalta, D., ... Alam, H. B. (2013b). Pharmacologic modulation of cerebral metabolic derangement and excitotoxicity in a porcine model of traumatic brain injury and hemorrhagic shock. *Surgery*, *154*, 234–243.
- Ichiyama, T., Okada, K., Lipton, J. M., Matsubara, T., Hayashi, T., & Furukawa, S. (2000). Sodium valproate inhibits production of TNF-alpha and IL-6 and activation of NF-kappaB. *Brain Research*, *857*, 246–251.
- Imam, A. M., Jin, G., Sillesen, M., Duggan, M., Jepsen, C. H., Hwabejire, J. O., ... Alam, H. B. (2013a). Early treatment with lyophilized plasma protects the brain in a large animal model of combined traumatic brain injury and hemorrhagic shock. *Journal of Trauma and Acute Care Surgery*, *75*, 976–983.
- Imam, A. M., Jin, G., Duggan, M., Sillesen, M., Hwabejire, J. O., Jepsen, C. H., ... Alam, H. B. (2013b). Synergistic effects of fresh frozen plasma and valproic acid treatment in a combined model of traumatic brain injury and hemorrhagic shock. *Surgery*, *154*, 388–396.
- Imam, A., Jin, G., Sillesen, M., Dekker, S. E., Bambakidis, T., Hwabejire, J. O., ... Alam, H. B. (2015). Fresh frozen plasma resuscitation provides neuroprotection compared to normal saline in a large animal model of traumatic brain injury and polytrauma. *Journal of Neurotrauma*, *32*, 307–313.
- Jepsen, C. H., deMoya, M. A., Perner, A., Sillesen, M., Ostrowski, S. R., Alam, H. B., & Johansson, P. I. (2014). Effect of valproic acid and injury on lesion size and endothelial glycocalyx shedding in a rodent model of isolated traumatic brain injury. *Journal of Trauma and Acute Care Surgery*, *77*, 292–297.
- Jin, G., Duggan, M., Imam, A., Demoya, M. A., Sillesen, M., Hwabejire, J., ... Alam, H. B. (2012). Pharmacologic resuscitation for hemorrhagic shock combined with traumatic brain injury. *Journal of Trauma and Acute Care Surgery*, *73*, 1461–1470.
- Jin, G., DeMoya, M. A., Duggan, M., Knightly, T., Mejaddam, A. Y., Hwabejire, J., ... Alam, H. B. (2012). Traumatic brain injury and hemorrhagic shock: Evaluation of different resuscitation strategies in a large animal model of combined insults. *Shock*, *38*, 49–56.
- Jin, G., Liu, B., You, Z., Bambakidis, T., Dekker, S. E., Maxwell, J., ... Alam, H. B. (2014). Development of a novel neuroprotective strategy: Combined treatment with hypothermia and valproic acid improves survival in hypoxic hippocampal cells. *Surgery*, *156*, 221–228.
- Ker, K., Roberts, I., Shakur, H., & Coats, T. J. (2015, May 9). Antifibrinolytic drugs for acute traumatic injury. *Cochrane Database of Systematic Reviews*, CD004896.
- Kim, H., & Bae, S. (2011). Histone deacetylase inhibitors: Molecular mechanisms of action and clinical trials as anti-cancer drugs. *American Journal of Translational Research*, *3*, 166–179.
- Kis, B., Szupera, Z., Mezei, Z., Gecse, A., Telegdy, G., & Vecsei, L. (1999). Valproate treatment and platelet function: The role of arachidonate metabolites. *Epilepsia*, *40*, 307.
- Kouzarides, T. (2000). Acetylation: A regulatory modification to rival phosphorylation? *EMBO Journal*, *19*, 1176–1179.
- Kutcher, M. E., Redick, B. J., McCreery, R. C., Crane, I. M., Greenberg, M. D., Cachola, L. M., ... Cohen, M. J. (2012). Characterization of platelet dysfunction after trauma. *Journal of Trauma and Acute Care Surgery*, *73*, 13–19.
- Kwan, I., Bunn, F., & Roberts, I. (2003). WHO Pre-Hospital Trauma Care Steering Committee: Timing and volume of fluid administration for patients with bleeding following trauma. *Cochrane Database of Systematic Reviews*, CD002245.
- Lee, J. Y., Kim, H. S., Choi, H. Y., Oh, T. H., Ju, B. G., & Yune, T. Y. (2012). Valproic acid attenuates blood-spinal cord barrier disruption by inhibiting matrix metalloproteinase-9 activity and improves functional recovery after spinal cord injury. *Journal of Neurochemistry*, *121*, 818–829.
- Li, Y., Yuan, Z., Liu, B., Sailhamer, E. A., Shults, C., Velmahos, G. C., ... Alam, H. B. (2008). Prevention of hypoxia-induced neuronal apoptosis through histone deacetylase inhibition. *Journal of Trauma*, *64*, 863–870.
- Li, Y., Zhao, T., Liu, B., Halaweish, I., Mazitschek, R., Duan, X., & Alam, H. B. (2015). Inhibition of histone deacetylase 6 improves long-term survival in a lethal septic model. *Journal of Trauma and Acute Care Surgery*, *78*, 378–385.
- Lipponen, A., Paananen, J., Puhakka, N., & Pitkänen, A. (2016). Analysis of post-traumatic brain injury gene expression signature reveals tubulins, Nfe2l2, Nfkb, Cd44, and S100a4 as treatment targets. *Scientific Reports*, *6*, 31570.
- Lin, T., Alam, H. B., Chen, H., Britten-Webb, J., Rhee, P., Kirkpatrick, J., & Koustova, E. (2016). Cardiac histones are substrates of histone deacetylase activity in hemorrhagic shock and resuscitation. *Surgery*, *139*, 365–376.
- Liu, Z., Li, Y., Chong, W., Deperalta, D. K., Duan, X., Liu, B., ... Alam, H. B. (2014). Creating a prosurvival phenotype through a histone deacetylase inhibitor in a lethal two-hit model. *Shock*, *41*, 104–108.
- Maegele, M., Lefering, R., Paffrath, T., Tjardes, T., Simanski, C., & Bouillon, B. (2008). Redblood-cell to plasma ratios transfused during massive transfusion are associated with mortality in severe multiple injury: A retrospective analysis from the Trauma Registry of the Deutsche Gesellschaft für Unfallchirurgie. *Vox Sanguinis*, *95*, 112–119.
- Manoguerra, A. S., Erdman, A. R., Woolf, A. D., Chyka, P. A., Caravati, E. M., Scharman, E. J., ... Troutman, W. G. (2008). American Association of Poison Control Centers. Valproic acid poisoning: An evidence-based consensus guideline for out-of-hospital management. *Clinical Toxicology*, *46*, 661–676.
- McMahon, C. G., Yates, D. W., Campbell, F. M., Hollis, S., & Woodford, M. (1999). Unexpected contribution of moderate traumatic brain injury to death after major trauma. *Journal of Trauma*, *47*, 891–895.
- Myburgh, J., Cooper, D. J., Finfer, S., Bellomo, R., Norton, R., Bishop, N., ... Vallance, S. (2007). Saline or albumin for fluid resuscitation in patients with traumatic brain injury. *New England Journal of Medicine*, *357*, 874–884.
- Nascimento, B., Callum, J., Rubenfeld, G., Neto, J. B., Lin, Y., & Rizoli, S. (2010). Clinical review: Fresh frozen plasma in massive bleedings—more questions than answers. *Critical Care*, *14*, 202.
- Nikolian, V. C., Bruhn, P. J., Georgoff, P. E., Halaweish, I., Pan, B., Liu, B., ... Alam, H. B. (2016). Histone deacetylase 6 inhibition attenuates neuronal cell death after oxygen-glucose deprivation and reoxygenation. *Journal of the American College of Surgeons*, *223*, S82.
- Nikolian, V., Dekker, S., Bambakidis, T., Bruhn, P. J., Georgoff, P. E., Andjelkovic, A. V., ... Alam, H. B. (2016). Pharmacologic resuscitation

- decreases blood-brain barrier permeability in a porcine model of traumatic brain injury and hemorrhagic shock. *Journal of the American College of Surgeons*, 223, S83.
- Norris, K. L., Lee, J. Y., & Yao, T. P. (2009). Acetylation goes global: The emergence of acetylation biology. *Science Signaling*, 2, pe76.
- Pareti, F. I., Capitanio, A., Mannucci, L., Ponticelli, C., & Mannucci, P. M. (1980). Acquired dysfunction due to the circulation of "exhausted" platelets. *American Journal of Medicine*, 69, 235–240.
- Peiniger, S., Nienaber, U., Lefering, R., Braun, M., Wafaisade, A., Wutzler, S., ... Maegele, M. (2011). Trauma registry of the Deutsche Gesellschaft für Unfallchirurgie. Balanced massive transfusion ratios in multiple injury patients with traumatic brain injury. *Critical Care*, 15, R68.
- Perel, P., & Roberts, I. (2012). Colloids versus crystalloids for fluid resuscitation in critically ill patients. *Cochrane Database of Systematic Reviews*, 6, CD000567.
- Posateri, A. E., Given, M. B., Schreiber, M. A., Spinella, P. C., Pati, S., Kozar, R. A., ... Cap, A. P. (2016). Dried plasma: State of the science and recent developments. *Transfusion*, 56(Suppl. 2), S128–S139.
- Rossaint, R., Bouillon, B., Cerny, V., Coats, T. J., Duranteau, J., Fernández-Mondéjar, E., ... Spahn, D. R. (2016). The European guideline on management of major bleeding and coagulopathy following trauma: Fourth edition. *Critical Care*, 20, 100.
- Santry, H. P., & Alam, H. B. (2010). Fluid resuscitation: Past, present, and the future. *Shock*, 33, 229–241.
- Scalea, T. M., Bochicchio, K. M., Lumpkins, K., Hess, J. R., Dutton, R., Pyle, A., & Bochicchio, G. V. (2008). Early aggressive use of fresh frozen plasma does not improve outcome in critically injured trauma patients. *Annals of Surgery*, 248, 578–584.
- Selby, J. B., Mathis, J. E., Berry, C. F., Hagedorn, F. N., Illner, H. P., & Shires, G. T. (1996). Effects of isotonic saline solution resuscitation on blood coagulation in uncontrolled hemorrhage. *Surgery*, 119, 528–533.
- Serdaroglu, G., Tutuncuoglu, S., Kavakli, K., & Tekgul, H. (2002). Coagulation abnormalities and acquired von Willebrand's disease type 1 in children receiving valproic acid. *Journal of Child Neurology*, 17, 41.
- Shang, Y., Jiang, Y. X., Ding, Z. J., Shen, A. L., Xu, S. P., Yuan, S. Y., & Yao, S. L. (2010). Valproic acid attenuates the multiple-organ dysfunction in a rat model of septic shock. *Chinese Medical Journal*, 123, 2682–2687.
- Sillesen, M., Johansson, P. I., Rasmussen, L. S., Jin, G., Jepsen, C. H., Imam, A., ... Alam, H. B. (2013). Valproic acid attenuates platelet dysfunction, endothelial glycocalyx shedding and protein C activation in a porcine model of traumatic brain injury and shock. *Journal of the American College of Surgeons*, 217(3 Suppl.), S51.
- Sillesen, M., Jin, G., Oklu, R., Albadawi, H., Imam, A. M., Jepsen, C. H., ... Alam, H. B. (2013). Fresh-frozen plasma resuscitation after traumatic brain injury and shock attenuates extracellular nucleosome levels and deoxyribonuclease 1 depletion. *Surgery*, 154, 197–205.
- Sillesen, M., Johansson, P. I., Rasmussen, L. S., Jin, G., Jepsen, C. H., Imam, A., ... Alam, H. B. (2014). Fresh frozen plasma resuscitation attenuates platelet dysfunction compared with normal saline in a large animal model of multisystem trauma. *Journal of Trauma and Acute Care Surgery*, 76, 998–1007.
- Sillesen, M., Bambakidis, T., Dekker, S. E., Fabricius, R., Svenningsen, P., Bruhn, P. J., ... Alam, H. B. (2016). Histone deacetylase gene expression profiles are associated with outcomes in blunt trauma patients. *Journal of Trauma and Acute Care Surgery*, 80, 26–32.
- Sillesen, M., Bambakidis, T., Dekker, S. E., Li, Y., & Alam, H. B. (2017). Fresh frozen plasma modulates brain gene expression in a swine model of traumatic brain injury and shock: A network analysis. *Journal of the American College of Surgeons*, 224, 49–58.
- Sondeen, J. L., Coppes, V. G., & Holcomb, J. B. (2013). Blood pressure at which rebleeding occurs after resuscitation in swine with aortic injury. *Journal of Trauma*, 54(Suppl.), S100–S107.
- Tai, Y. T., Lee, W. Y., Lee, F. P., Lin, T. J., Shih, C. L., Wang, J. Y., ... Hung, K. S. (2014). Low dose of valproate improves motor function after traumatic brain injury. *BioMed Research International*, 2014, 980657.
- Verrotti, A., Greco, R., Matera, V., Altobelli, E., Morgese, G., & Chiarelli, F. (1999). Platelet count and function in children receiving sodium valproate. *Pediatric Neurology*, 21, 611.
- Wafaisade, A., Lefering, R., Tjardes, T., Wutzler, S., Simanski, C., Paffrath, T., ... Maegele, M.; Trauma Registry of DGU. (2010). Acute coagulopathy in isolated blunt traumatic brain injury. *Neurocritical Care*, 12, 211–219.
- Wald, S. L., Shackford, S. R., & Fenwick, J. (1993). The effect of secondary insults on mortality and long-term disability after severe head injury in a rural region without a trauma system. *Journal of Trauma*, 34, 377–381.
- Wang, Z., Leng, Y., Tsai, L. K., Leeds, P., & Chuang, D. M. (2011). Valproic acid attenuates blood-brain barrier disruption in a rat model of transient focal cerebral ischemia: The roles of HDAC and MMP-9 inhibition. *Journal of Cerebral Blood Flow and Metabolism*, 31, 52–57.
- Wong, H., Curry, N., & Stanworth, S. J. (2016). Blood products and pro-coagulants in traumatic bleeding: Use and evidence. *Current Opinion in Critical Care*, 22, 598–606.
- Wong, V. S., & Langley, B. (2016). Epigenetic changes following traumatic brain injury and their implications for outcome, recovery and therapy. *Neuroscience Letters*, 625, 26–33.
- Xiao, W., Mindrinos, M. N., Seok, J., Cuschieri, J., Cuenca, A. G., Gao, H., ... Rosenberg, G. A. (2007). Matrix metalloproteinase-mediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *Journal of Cerebral Blood Flow and Metabolism*, 27, 697–709.
- Xiao, W., Mindrinos, M. N., Seok, J., Cuschieri, J., Cuenca, A. G., Gao, H., ... Tompkins, R. G. (2011). A genomic storm in critically injured humans. *J Exp Med*, 208, 2581–2590.
- Yu, F., Wang, Z., Tanaka, M., Chiu, C. T., Leeds, P., Zhang, Y., & Chuang, D. M. (2013). Posttrauma cotreatment with lithium and valproate: Reduction of lesion volume, attenuation of blood-brain barrier disruption, and improvement in motor coordination in mice with traumatic brain injury. *Journal of Neurosurgery*, 119, 766–773.
- Zehtabchi, S., & Nishijima, D. K. (2009). Impact of transfusion of fresh-frozen plasma and packed red blood cells in a 1:1 ratio on survival of emergency department patients with severe trauma. *Academic Emergency Medicine*, 16, 371–378.
- Zehtabchi, S., Abdel Baki, S. G., Falzon, L., & Nishijima, D. K. (2014). Tranexamic acid for traumatic brain injury: A systematic review and meta-analysis. *American Journal of Emergency Medicine*, 32, 1503–1509.

How to cite this article: Dekker SE, Nikolian VC, Sillesen M, Bambakidis T, Schober P, Alam HB. Different resuscitation strategies and novel pharmacologic treatment with valproic acid in traumatic brain injury. *J Neuro Res*. 2018;96:711–719. <https://doi.org/10.1002/jnr.24125>