

“Applications of Sensory and Physiological Measurement in Oral-Facial Dental Pain”

Darya Dabiri, DMD, MS (Pediatric Dentistry), MS (Endodontics), MSCR, Post-doc Fellow in Pain, Chronic Pain & Fatigue Research Center, University of Michigan Clinical Research, 24 Frank Lloyd Wright Drive, Lobby M, Suite 3100, Ann Arbor, MI, USA 48109

Daniel Harper, PhD, , Chronic Pain & Fatigue Research Center, University of Michigan Clinical Research, 24 Frank Lloyd Wright Drive, Lobby M, Suite 3100, Ann Arbor, MI, USA 48109

Yvonne Kapila, DDS, PhD, Professor, Vice-Chair Division of Periodontology, Dept. of Orofacial Sciences, UCSF, University of California San Francisco, 513 Parnassus Avenue, San Francisco CA 94117

Grant Kruger, MS, PhD, Assistant Research Scientist, Mechanical Engineering, 1043C Dow (Herbert H. Dow Building) 2300 Haywar, Ann Arbor, MI 48109-2136

Daniel Clauw, MD, Professor, Department of Anesthesiology, Director, Translational Research, Director, Center for Chronic Pain and Fatigue Research, University of Michigan Clinical Research, Chronic Pain & Fatigue Research Center, 24 Frank Lloyd Wright Drive, Lobby M, Suite 3100, Ann Arbor, MI, USA 48109

Steven Harte, PhD, Assistant Research Scientist, Anesthesiology, Assistant Research Scientist, Internal Medicine (Rheumatology), Research Associate, Medical Research Service, Veterans Affairs Ann Arbor, Chronic Pain & Fatigue Research Center, University of Michigan Clinical Research, 24 Frank Lloyd Wright Drive, Lobby M, Suite 3100, Ann Arbor, MI, USA 48109

Corresponding author:

Darya Dabiri, DMD, MS, MS, MSCR

Diplomat of American Board of Pediatric Dentistry
Board Eligible of American Board of Endodontics,
Chronic Pain & Fatigue Research Center
University of Michigan Clinical Research
24 Frank Lloyd Wright Drive, Lobby M, Suite 3100
Ann Arbor, MI, USA 48109
E-mail: daryad@umich.edu
734-998-6939
FAX: 734-998-6900

Key Words: Quantitative Sensory Testing, Dental Pain, Orofacial Pain, Cognitive Impairment

Acknowledgements: This research was supported in part by National Institute of Dental and Craniofacial Research (grant number: K12 DE023574). Authors thank Dr. Joe Camp, Dr. Daniel

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/scd.12323](https://doi.org/10.1111/scd.12323).

Chiego, Dr. Neville J. McDonald and Dr. Michael Apicella for their insightful comments that greatly improved the manuscript.

Conflict of Interest:

Each author completed a separate conflict of interest form from Wiley's website and included in the uploaded files on this submission.

INTRODUCTION

Pain motivates individuals to seek dental care. [1] The perception of pain is a complex process that involves bidirectional communication between the central and peripheral nervous systems. It is now known that individuals vary widely in their pain sensitivity, and there often is a very poor relationship between the degree of peripheral damage/inflammation within an individual and how much pain he/she is experiencing. This is especially true in sub-acute or chronic pain conditions, where frequently there is little evidence of ongoing damage or inflammation in the periphery. [2] In these cases, intervening in the periphery – as dentists are trained to do – will not relieve pain, and could actually worsen a person's clinical condition. Therefore, it is critically important that the field of dentistry progresses toward a mechanistic understanding of pain to better identify individuals who are at risk of failing to respond to our interventions and to limit procedures to only those who are likely to benefit from them.

Traditionally, dentists and other oral health clinicians have used relatively crude 'chairside' tests to infer whether there is underlying pathology in oral structures. Typical intraoral examinations include palpation, percussion, thermal, electric and periodontal examinations. While valuable, these methods often lack quantifiable and objective outcomes, and there remains considerable inter- and intra-clinician variation in their implementation. In addition, determination of pain and pathology in young children, adults with a language barrier, or individuals of any age with cognitive impairment (CI) is very challenging. In contrast to these subjective measures, standardized tests of perceptual and physiological reactions to externally applied and quantifiable stimuli (i.e., quantitative sensory testing - QST) mitigates these limitations and provides a more accurate approach to identify pain mechanisms and track changes in sensory function over time. In this review, we provide an overview of QST methods and discuss how they can be implemented in standard dental practice. Section 1 reviews

pain mechanisms that contribute to acute and chronic dental pain. Section 2 discusses various physiological signals that can be monitored to determine tooth health and pulpal vitality. Section 3 describes the use of QST procedures in the study of orofacial pain. Section 4 describes challenges in the diagnosis of dental pain in non-communicative individuals. Finally, Section 5 provides recommendations on how these sensory and physiological measurement technologies can be combined to improve clinical dental care.

1. Mechanisms of orofacial pain

1.1. Nociceptive pain

Nociceptive pain is the most common form of pain in the orofacial region and normally occurs following acute stimulation of the nociceptors embedded in the skin, intraoral cavity, and dental pulp. Inflammatory pain is also categorized under this umbrella term, since inflammation in the periphery is known to sensitize nociceptors and increase their spontaneous firing rate and their excitability to stimulation. Clinical pain is a reflection of the nociceptive circuits' overall excitability, and not just a pain system being "turned on" by a pathology. The sensitivity of those nociceptive circuits can be shifted and changed by innocuous stimuli, more like a dial than a switch, and the state of excitability can dictate the level of pain experienced. [3, 4]

The majority of tooth pain is thought to be nociceptive and of odontogenic origin. [5] Odontogenic pain encompasses pain that could originate from either pulpal or periodontal tissue (mucosa, gingiva, or periodontal ligament). [6] Orofacial nociceptive pain often originates from insult of the dental pulp. The rigid compartment that pulp resides in provides a support structure and protects it from the microbes present in the mouth. When that protective chamber is damaged or corroded, the pulp it encapsulates becomes susceptible to the hostile elements present in the oral cavity. The microcirculation within the healthy pulp plays a crucial role in orchestrating inflammatory response in response to pulpal damages. [7, 8] Inflammation of the pulp, or pulpitis, produces increasingly intense and prolonged painful responses to thermal or osmotic stimulation, as distinct from the less painful and phasic responses seen with normal dentine sensitivity. Early stage

inflammatory responses may be reversible, but as the pathology advances, the process becomes irreversible, and may result in the development of spontaneous pain that occurs without provocation, likely because of both peripheral and central sensitization. Chronic inflammation can eventually lead to pulpal necrosis and periapical pathology. [9]

Pulpitis can be either reversible or irreversible. Reversible pulpitis is often characterized by a brief, sharp non-spontaneous pain upon provocation. [10] This transient pulpal inflammation can be reverted once the source of irritation is removed (e.g., caries or occlusal trauma) In contrast, irreversible pulpitis is characterized by pain that lingers following stimulation and spontaneous pain that occurs without provocation. Treatment for irreversible pulpitis involves either excavation of the diseased pulp or tooth extraction. Irreversible pulpitis, left untreated, leads to necrosis -- a necrotic pulp will usually not respond to a thermal or electrical stimulation. Teeth with irreversible pulpitis can be completely asymptomatic or extremely painful on percussion, [10, 11] which limits the usefulness of this technique in diagnosis. Symptoms of odontogenic pain can vary greatly, in some cases pulpal necrosis occurs without any prior symptomatic pulpitis. [12] Studies have shown that clinical pain symptoms do not necessarily correlate with histological findings in pulp. [13-15]

Orofacial nociceptive pain could also originate from temporomandibular joint (TMJ) structures. There are at least three potential etiologies for TMD pain including degradation of temporomandibular joint (TMJ) structures, inflammation in the joint (i.e. degenerative joint disease-arthritis) and myofascial pain. There is limited evidence that peripheral inflammation in local musculature (e.g., temporalis, medial pterygoid and masseter) contributes to myalgic pain in TMD, [16] and instead this type of TMD pain is thought to occur mainly via central nervous system (CNS) mechanisms. [17-19]

In the otherwise healthy individual, acute dental pain is often sudden and debilitating. Despite its unexpected and concerning nature, it has a protective effect as its intensity mirrors the amount of injury to the tissue and its presence motivates behaviors that aid recuperation and healing. [9, 20] However, in patients with chronic or sub-acute pain conditions, pain signals can be augmented or

amplified by the CNS, and pain often occurs either disproportionate to or in the absence of ongoing nociceptive input. Both neuropathic and centralized pain mechanisms, discussed below, are hallmark features of chronic pain.

1.2. Neuropathic pain

Neuropathic pain occurs following destruction to or compression of the peripheral nerves that transmit somatosensory signals to the CNS. It has typical clinical features regardless of where in the body it occurs that aid in its diagnosis. For instance, neuropathic pain is often characterized as waxing and waning, and lancinating, and it may be accompanied by paresthesias, numbness, tingling, and shooting sensations. [21-26] Neuropathic pain also typically follows the distribution of one or more sensory nerves that are damaged or inflamed.

There are specific oral conditions with prominent neuropathic components. Neuropathic pain can be classified according to its location and frequency: unilateral continuous, unilateral episodic, and bilateral continuous. [19, 26-28] Episodic pain typically is of short duration, and produces very sharp or electric-like sensations. Episodic neuralgias include of trigeminal and glossopharyngeal neuralgia - named after the nerve affected. Neuropathic pain in these conditions is typically episodic and unilateral, but it can be bilateral. If bilateral, multiple sclerosis is often suspected as an etiological factor, especially in younger age groups.

The continuous neuropathic pain disorders can be spontaneous or have a trigger zone and are characterized more by a burning-type sensation. The continuous neuralgias are considered a form of deafferentation pain and can be due to trauma including surgery or metabolic disorders such as diabetic neuropathy. Examples of unilateral continuous neuropathic pain in the orofacial region include atypical odontalgia and burning mouth syndrome. Each of these is described below.

Trigeminal neuralgia (TN) and glossopharyngeal neuralgia. TN and glossopharyngeal are defined by unilateral episodic pain that follows the distribution of one or more defined nerve pathways. TN has typical clinical features but is frequently of unknown etiology. TN is classified into

Classical and Symptomatic subtypes. [19, 26] Classical TN typically manifests as sudden, sharp, shooting, shock-like pain, elicited by slight touching of “trigger points,” that can radiate. Symptomatic TN manifests with paroxysmal painful attacks of short duration affecting one or more branches of the trigeminal nerves. Etiology of the pain source varies in classical versus symptomatic TN. Unlike symptomatic TN, vascular compression is the primary cause of classical TN pain. Trigeminal post-herpetic neuralgia (PHN) is an example of symptomatic TN. PHN is defined as the persistence of pain following disappearance of the rash that can last between 1 and 6 months in a herpes zoster virus infection. Sensory changes are sometimes also observed during clinical testing of PHN, including hyperalgesia and/or allodynia. Although some cases present with a clear history of nerve damage (e.g., due to dental procedure or other insult), the actual cause of the neuropathy often remains unknown and secondary causes such as autoimmune, malignancy or infection could be typically considered. [22, 27-34]

Atypical odontalgia (Post traumatic trigeminal). Atypical odontalgia or Persistent Dentoalveolar Pain Disorder (PDAP) is increasingly recognized as being caused not only by trauma to the facial skeleton, but also by various dental procedures including root canal therapy, extractions and/or dental implants. (Baad-Hansen et al., 2015) Pain in atypical odontalgia is very clearly localized to the dentoalveolar region with or without the presence of dentition. [35, 36] Pain can present as throbbing and continuous, and at times sharp. It is often provoked by light touch. History of dental treatment does not affect the onset of the disease. The source of pain is not easily recognized, and the pain can seemingly occur without any reason. This often leads to more and more unnecessary dental procedures that fail to relieve the pain. [35-37]

Burning mouth syndrome. Burning mouth syndrome is an example of bilateral continuous neuropathic pain. [38] It presents as a burning sensation of the intraoral soft tissue with no apparent etiology. There have been several studies suggesting various precipitating factors, but these studies show no consensus and the quality of prospective studies and case reports is lacking. [39] The symptoms can be continuous but the intensity does vary throughout the day. Several local and systemic causes need to be excluded in diagnosing burning mouth syndrome. Local causes include

candidiasis, lichen planus, herpetic infection, and xerostomia, and systemic causes include use of specific medications (e.g., Angiotensin-converting-enzyme (ACE)-inhibitor for hypertension therapy), hematological causes, nutritional deficiencies, and Sjogren's syndrome.

1.3. Centralized pain

“Central pain,” as originally described, referred to chronic pain that occurred as a result of damage to the CNS, such as thalamic pain syndrome following cerebral ischemia [2] Later, new terminology was introduced to describe the amplification and/or maintenance of pain by CNS mechanisms, irrespective of peripheral nociceptive input or structural damage. Terms that have been used include, “central sensitization,” “centralized pain,” “central hyper-excitability,” and others. [40-43] Consensus within the pain field on this new terminology has yet to be reached. In this review, we will use the terms “central sensitization” for all molecular, structural and functional CNS (brain and spinal) mechanisms related to pain and sensory amplification, and “centralized pain” to refer broadly to clinical phenotypes preferentially characterized by underlying mechanisms of central sensitization. [4, 42, 44, 45]

The clinical features of centralized pain differ from those of nociceptive or neuropathic pain. Using a pain diagram can be very helpful in diagnosing any pain patient, but it can be especially helpful in identifying neuropathic pain that follows the distribution of a peripheral nerve, or the more widespread or diffuse pain distribution that occurs with centralized pain states. The “widespreadness” of an individual's pain often reflects the degree to which their pain has been centralized. Centralized pain can manifest anywhere in the body and in any type of tissue, and is generally more diffuse than nociceptive or neuropathic pain since many of the central gain controls for incoming nociceptive signals that are recognized to be dysregulated in centralized pain patients can affect pain signals from throughout the body, e.g., diffuse noxious inhibitory controls (DNIC). [46] The degree to which pain can be well localized by the patient (centralized pain cannot be as well localized) is another distinguishing factor between nociceptive and centralized pain. [2] Other clinical features of centralized pain include hypersensitivity to a variety of painful (e.g., heat, cold, electrical,

pressure) and innocuous sensory stimuli(e.g., bright lights, noises, odors), and a myriad of co-occurring CNS-organized symptoms (e.g., fatigue, sleep difficulties, mood and memory problems).

Referred odontogenic pain is a type of centralized pain condition. In odontogenic pain, central sensitization begins from peripheral tissue injury at the site of tooth and supporting periodontium. At this site, inflammation modulates activation of afferent nociceptive nerve endings by lowering their firing threshold. [47] Prolonged activation of nociceptive input to second-order neurons facilitates augmentation of nociceptive impulses to higher brain centers. This central sensitization can manifest as secondary hyperalgesia and referred pain. Secondary hyperalgesia denotes a change to CNS that causes an augmented reaction to painful stimuli in surrounding tissue, such as the gingiva or skin. [26] Referred pain occurs when pain manifests at a locations remote from its source. [48]

2. Investigating Intraoral Pathology using Physiological Measures

2.1 Clinical Assessment of Dental Pain

Since the complaint of “tooth pain” could have both odontogenic and non-odontogenic etiologies, it is crucial to conduct a comprehensive dental evaluation, including a detailed history of the present illness, and appropriate radiographic imaging. The most precise technique to assess pulp status is by histological examination of pulpal inflammation. Unfortunately, this method cannot be practiced in clinical care as it requires surgical removal of the enamel and dentin to access the pulp; hence clinicians need to use other non-invasive metrics, such as stimulus testing, to provide additional diagnostic information for pulpal diagnosis. In these tests, teeth and surrounding structures are then evaluated via various sensation-evoking methods including thermal (hot and cold sensitivity), electrical, percussion, palpation testing (see Section 3). Interpretation of these tests requires experience, training and knowledge of various test limitations and patient responses. Ideally, a diagnostic test will always be positive when pathology is present, and negative when pathology is absent – however sensitivity and specificity analyses have revealed that the most common pulp tests (i.e., cold and Electrical Pain Threshold [EPT] testing) are imperfect diagnostic tools. Most of the

time, these diagnostic tests show high negative predictive values (80-90%) and lower positive predicted values (30-70%). [49] This wide range could be due to the difficulty in predicting histological states from diagnostic tests. An earlier study suggested that there was “no reliable” association between pulp status and histological assessment. [50]

Obtaining a detailed characterization of a patient’s pain is one of the most useful diagnostic strategies for dental pain, since pulpal pathology follows a sequence of changes and symptoms that vary over time. [51] In 2010, Pho and his group proposed a validated dental pain screening questionnaire to assess patients with odontogenic pain called the Dental Pain Questionnaire (DePaQ). It consists of 14 items that assess location, frequency, and intensity of pain in the orofacial region. It also asks questions regarding tooth specific signs and symptoms including temperature sensitivity, biting sensitivity, and the combination of both. The DePaQ was originally designed to differentiate between three groups of odontogenic pain: a) irreversible pulpitis/acute apical periodontitis, b) reversible pulpitis/dentin hypersensitivity, and c) periocoronitis. The DePaQ showed acceptable sensitivity of 0.85, however its specificity varies across studies. [51, 52] In 2017, Nixdorf stated that the results of DePaQ showed an unacceptable specificity of 0.11, which was substantially smaller than that identified in the original validation studies (i.e., 0.83).[17]

2.2 Measurement of Somatosensory Responses

In an effort to obtain more objective measures of pulpal status, many investigators have examined physiological responses to stimuli applied to teeth and surrounding tissue. These tests have a long history in the study of dental pain and yet they remain poorly understood. Somatosensory responses in orofacial tissue comes from various low-threshold somatosensory receptors originating from structures such as teeth, temporomandibular joint (TMJ), skin, and muscles. These structures are mainly innervated by branches of the trigeminal (V) nerve. Oral sensorimotor dysfunction could potentially cause sensory alteration and orofacial pain. [53]

Early studies examined somatosensory responses in teeth. In 1965, Scott and Tempel developed a technique to conduct electrophysiological studies in animal teeth by placing two metal

electrodes (within few millimeters) in direct contact with exposed dentin through cavities prepared in cats' teeth. [54] In 1972, Scott provided evidence that dentin is innervated [55] Later, the anatomical relationships between the odontoblasts, their processes, and the sensory nerve endings were described in human teeth by studying sensory responses to thermal stimulation. [56] Others demonstrated that dental pain can originate from the activation of nociceptive receptors at the pulp-dentin junction. [57-61]

Olgart (1985) showed that local application of stimuli such as cold and heat could produce nerve impulses that were associated with perceived pain sensations verbally expressed by the subject. We now know that alterations in pulpal blood flow as a result of aging or disease can affect these somatosensory findings. [8] Edwall and Scott (1971) were one of the first groups to show that reductions of pulpal blood flow can increase excitability and evoke action potentials in the nerves innervating teeth. [62]

2.3 Measurements of Circulating Blood Flow and Temperature

Understanding the complex pathophysiology affecting circulating blood flow is also important for accurate pulpal diagnosis. Previous studies have investigated methods such as pulse oximetry, Laser Doppler Flowmetry (LDF), and crown surface temperature change as ways of measuring pulpal blood flow, and each are described below in detail. However, none of these methods have been established as a standard of care in clinical dentistry. [63, 64]

Pulse Oximetry. Earlier studies suggested that blood oxygenation within teeth could be measured [10] Modern pulse oximeters now provide an inexpensive and efficient method to measure blood oxygenation. Pulse oximetry can detect oxygen saturation levels of the pulp tissue based on spectrophotometry using a light source (wavelengths 760 and 850 nm) [65-68] In one of the first studies demonstrating the utility of pulse oximetry in dentistry, Schnettler and Wallace (1991) showed that a pulse oximeter could detect pulse rate and oxygen saturation for vital teeth and not for root canal treated teeth. This work has now been replicated by several groups. [20, 65-67, 69] Interestingly, Gopikishna showed that normal oxygen saturation in human permanent teeth was lower

than systemic blood measured in fingers, 75-85% vs 98% respectively. [70] The sensitivity of the pulse oximeter to detect pulp vitality was 100% in comparison to sensitivity of cold (81%) and EPT (71%) methods. Unlike sensitivity, the specificity for pulse oximetry testing was very similar to cold and less compared to EPT testing for detecting pulp vitality in both mature and immature permanent central incisors. [71] One of the limitations of using a pulse oximeter in dentistry is the differences in the optical properties of the teeth as the infrared light deflects through the teeth to the photodetector. The infrared light could also scatter due to its close proximity to surrounding gingival tissue. [72, 73]

Laser Doppler Flowmetry. LDF is another method to measure blood flow using light (Helium Neon 632.8nm). LDF works on Doppler principle – light undergoes a frequency shift and scatters as it passes through moving red blood cells. [74] The light is scattered back onto photodetectors to measure pulpal blood flow. [75, 76] LDF was originally designed for the measurement of organs that have abundant blood flow, such as the brain and skin, not for the dental pulp due to its low blood volume and/or low blood flow velocity. [77] Previous studies noted many challenges in using LDF, especially when it is used to measure blood flow in the soft tissue of the oral cavity. LDF is spatially limited (1mm³), and the unknown morphology of the vasculature affects probe placement and subsequently the accuracy of LDF output. LDF outputs are also negatively impacted by head movement of the patient and movement of the hand-held probe by the operator. [78] Contamination of signals from backscattered light on surrounding tissue and/or interference of ambient light together lead to non-linear output in LDF measures that are difficult to interpret. [20]

Temperature. Tooth temperature has also been proposed as measure of determine pulp vitality. For instance, Fanibunda et al. reported that vital teeth are warmer at rest and unlike non-vital teeth they can rewarm more quickly when cooled down. [79-82] Other studies showed that despite having the same temperature at rest for vital and non-vital teeth, there is a delay in regaining heat in non-vital teeth versus vital teeth. [83] One of the limiting factors in incorporating temperature measurements in clinical practice is that there is still no study to date that suggests a relationship between temperature and the degree of pulpal inflammation. [84]

Despite these limited data, non-invasive physiological monitoring techniques such as pulse oximetry, LDF, and crown surface temperature change may provide objective measures of pulp status that could be beneficial for clinical pulp vitality diagnosis.

3. Quantitative Sensory Testing in Orofacial Pain and Dentistry

Quantitative sensory testing (QST) refers to a set of non-invasive procedures for assessing sensory function. QST procedures are psychophysical in nature, and involve application of objective, quantifiable physical stimuli that evoke behavioral (e.g., verbal) responses from the individual being tested. QST has been used for decades in clinical research for cutaneous and mucosal assessment of pain sensitivity. [32, 85-90] It can also be used for patient subgroup classification and prognosis. [91] Lastly, QST can help make inferences about the underlying mechanisms of pain and improve diagnostic accuracy in orofacial pain patients. For example, trigeminal neuropathic pain can be more accurately and reliably diagnosed using QST of innocuous mechanical and thermal stimulation [36] In many cases, patients with peripheral nerve damage present with hypoesthesia to warm and sometimes cool stimuli. [92] TMD patients often experience hyperalgesia on palpation and/or thermal stimulation. This hyperalgesia is present in the orofacial region and also at remote, asymptomatic sites, like the forearm. [9, 17, 18, 87, 93-96]. Evidence also supports perceptual amplification of non-painful auditory tones in TMD patients. [97]. On the other hand, QST has revealed a reduced sensitivity to innocuous vibrations applied to the cheek (and to a lesser extent to the arm), compared to healthy participants, suggesting that ongoing pain signals may be “gating” (or masking) innocuous somatosensory inputs. [97] The hyperalgesia observed in TMD may result from increased pain-excitatory processes in the CNS (e.g., measured as increased temporal summation of pain) and/or impaired endogenous inhibition (e.g., measured as reduced conditioned pain modulation). [98]

Dentists have long used externally applied stimuli to determine a tooth’s vitality as a form of “bedside” QST. [49] The most commonly used tests stimulate the pulp by means thermal stimuli (e.g., cold testing) or electrical current (e.g., Electrical Pulp Test –EPT). These tests are used routinely

in clinical practice to evaluate pulpal status, including the presence of healthy, inflamed or necrotic pulp. [49, 99, 100] However, a significant limitation in the current clinical implementation of these techniques in the assessment of dental pain is that perceptual responses are measured only in the form of qualitative and dichotomous “yes / no” subjective reports of the patient’s evoked sensory experience.

Thermal testing includes assessment of tooth sensitivity to temperature changes via application of cold and heat stimuli. In this test, patients report the sensation they perceive during stimulus application. Overall, reports of pain sensations without lingering pain indicate healthy pulp, with lingering pain suggest inflamed pulp, and finally reports of no sensation may indicate necrotic pulp. According to the Brännström Hydrodynamic Theory, temperature changes cause dentinal fluids to rapidly move within the dentinal tubules which in turn induces activation of nociceptive A-delta fibers within the pulp-dentin complex. [70] Application of heat, however, must be used with caution as it produces lingering pain through activation of C-fibers pain and may increase pulp inflammation. [101, 102] Cold application is safer as it does not produce detrimental effects on the pulp tissue. [103, 104] Several methods of cold delivery are routinely used, such as ice stick (0°C), ethyl chloride (-5°C), and dichlorodifluoromethane (DDM). In clinical practice, it is more common to use DDM as it evokes a quicker response from the pulp in comparison to the other methods. [104-106]

Electrical pulp testing (EPT) involves indirect electrical stimulation of the pulpal nerves through the tooth surface. In a series of early studies, Mumford used EPT for the diagnosis of pulp status by measuring the electrical pain threshold – the lowest physical intensity of electrical stimulation that a person perceives to be just barely painful. [50, 107] In theory, pulpal disease and its degenerative changes can lead to changes in pain threshold such that there are lower pain thresholds (increased pain sensitivity) in acute pulpitis and higher pain thresholds (decreased pain sensitivity) in chronic pulpitis. However, in clinical practice, Mumford’s study sensations produced during EPT varied substantially among patients and this measure was not useful for diagnostic purposes. According to Mumford, EPT is a reliable test for identification of vital pulps, but not necessarily a reliable test for classifying pulp pathology using measures of pain sensitivity. [50, 107]

4. Non-Communicative Patients and Challenges in Dental Pain Assessments

Diagnosis and treatment of dental pain in young children, as well as in children or adults with cognitive impairment (CI), is very challenging given the current use of subjective measures for clinical pain assessments. Individuals with cognitive impairment vary greatly in their experience of pain, and in their response to painful interventions based on their cognitive and emotional maturity. [108]. Dental care providers need to acknowledge these variations and be willing to understand the unique presentation of pain in these unique populations whether it is conveyed by verbal self-report or only in behavioral cues such as alternations in their feeding, sleeping and routine activities of daily living.

Self-report of pain is often not reliable and sometimes not available in non-verbal/non-communicative individuals. In these cases, the history of pain relies heavily on remarks reported from a third party e.g. parents, caregivers, teachers etc. It is important to acknowledge observations provided by these caregivers as they know the patients very well and in different environments outside the clinic. They can also differentiate any subtle changes in behavior and notice cues outside the individuals' normative behavior that may suggest the presence of pain.

When treating pediatric patients, it appears that cognitively impaired children experience pain more significantly in comparison to cognitively intact children. [109, 110] Unfortunately, because of their limited verbal communication, pain in children with CI may be undertreated [111]. Therefore, assessment and localization of dental pain is a complicated process when treating these children. Inadequate diagnosis of pain can lead to poor pain management; and unrelieved pain can have a negative physical and cognitive impact on the overall wellbeing of these children. Given these challenges, the assessment of pain and pulp status in these populations may benefit most from the use of objective diagnostic methods.

5. Bridging the Gap between QST and Pulp Vitality Testing to Improve Clinical Care

Dentists frequently struggle to answer questions such as “Is the pain coming from the tooth or another structure?” and “Is the tooth alive or dead?” These are critical questions in a wide variety of

individuals presenting with acute and chronic orofacial pain. We contend that in order to better diagnose pulp vitality and standardize diagnoses across clinical providers, research should focus on improving our diagnostic tools to become more objective and quantifiable. For example, we propose that combining “research-grade” QST methods that use well-controlled, quantifiable stimuli with objective physiological readouts (e.g., pulse oximetry, LDF, temperature change) will significantly improve the likelihood of early diagnosis and treatment of dental pulp pathology, before irreversible damage occurs. Technology such as this would be of particular benefit in non-communicative individuals and those with CI. However, as described earlier, each of these methods, as currently practiced, has limitations and challenges that need to be addressed prior to widespread clinical adoption.

CONCLUSION

The assessment of orofacial pain and pulp vitality is challenging, especially in non-verbal and impaired populations. Dentists regularly employ a variety of self-report and sensory techniques in the clinic to aid in the diagnosis and treatment of tooth-related disease. This review discusses nociceptive, neuropathic, and centralized (CNS) mechanisms that underlie acute and chronic dental pain. It details the measurement of somatosensory responses and pulpal blood flow as objective measures of tooth health and pain. It also introduces the measurement stimulus-evoked sensations (i.e., QST) as practiced in research settings and compares it with existing qualitative and dichotomous “yes / no” aspects of sensory testing currently practiced at the point of care. Finally, it proposes that bridging these varied methodologies will significantly improve diagnosis and treatment of orofacial pain and pathology. It is critical the field of dentistry progresses toward a mechanistic understanding of pain to better identify individuals who are at risk of failing to respond to our interventions and to limit procedures to only those who will benefit from them.

References:

1. Keiser, K., *Strategies for managing the endodontic pain patient*. Tex Dent J, 2003. **120**(3): p. 250-7.

2. Harper, D.E., A. Schrepf, and D.J. Clauw, *Pain Mechanisms and Centralized Pain in Temporomandibular Disorders*. J Dent Res, 2016. **95**(10): p. 1102-8.
3. Latremoliere, A. and C.J. Woolf, *Central sensitization: a generator of pain hypersensitivity by central neural plasticity*. J Pain, 2009. **10**(9): p. 895-926.
4. Woolf, C.J., *Central sensitization: implications for the diagnosis and treatment of pain*. Pain, 2011. **152**(3 Suppl): p. S2-15.
5. Byers, M.R. and M.V. Narhi, *Dental injury models: experimental tools for understanding neuroinflammatory interactions and polymodal nociceptor functions*. Crit Rev Oral Biol Med, 1999. **10**(1): p. 4-39.
6. Renton, T., *Dental (Odontogenic) Pain*. Rev Pain, 2011. **5**(1): p. 2-7.
7. Arwill, T., et al., *Ultrastructure of nerves in the dentinal-pulp border zone after sensory and autonomic nerve transection in the cat*. Acta Odontol Scand, 1973. **31**(5): p. 273-81.
8. Olgart, L.M., *The role of local factors in dentin and pulp in intradental pain mechanisms*. J Dent Res, 1985. **64 Spec No**: p. 572-8.
9. Yu, C. and P.V. Abbott, *An overview of the dental pulp: its functions and responses to injury*. Aust Dent J, 2007. **52**(1 Suppl): p. S4-16.
10. Dummer, P.M., R. Hicks, and D. Huws, *Clinical signs and symptoms in pulp disease*. Int Endod J, 1980. **13**(1): p. 27-35.
11. Seltzer, S., I.B. Bender, and J. Ehrenreich, *Incidence and duration of pain following endodontic therapy. Relationship to treatment with sulfonamides and to other factors*. Oral Surg Oral Med Oral Pathol, 1961. **14**: p. 74-82.
12. Michaelson, P.L. and G.R. Holland, *Is pulpitis painful?* Int Endod J, 2002. **35**(10): p. 829-32.
13. Seltzer, S., *Pain in endodontics. 1986*. J Endod, 2004. **30**(7): p. 501-3; discussion 500.
14. Seltzer, S., *Pain in endodontics*. J Endod, 1986. **12**(10): p. 505-8.
15. Seltzer, S., I.B. Bender, and M. Ziontz, *THE DYNAMICS OF PULP INFLAMMATION: CORRELATIONS BETWEEN DIAGNOSTIC DATA AND ACTUAL HISTOLOGIC FINDINGS IN THE PULP*. Oral Surg Oral Med Oral Pathol, 1963. **16**: p. 969-77.
16. Slade, G.D., et al., *Cytokine biomarkers and chronic pain: association of genes, transcription, and circulating proteins with temporomandibular disorders and widespread palpation tenderness*. Pain, 2011. **152**(12): p. 2802-12.
17. Fonseca Alonso, B., et al., *Examining the Sensitivity and Specificity of 2 Screening Instruments: Odontogenic or Temporomandibular Disorder Pain?* J Endod, 2017. **43**(1): p. 36-45.
18. Maixner, W., et al., *Relationship between pain sensitivity and resting arterial blood pressure in patients with painful temporomandibular disorders*. Psychosom Med, 1997. **59**(5): p. 503-11.
19. Zakrzewska, J.M., *Differential diagnosis of facial pain and guidelines for management*. Br J Anaesth, 2013. **111**(1): p. 95-104.
20. Chen, E. and P.V. Abbott, *Dental pulp testing: a review*. Int J Dent, 2009. **2009**: p. 365785.
21. Dimitroulas, T., et al., *Neuropathic pain in osteoarthritis: a review of pathophysiological mechanisms and implications for treatment*. Semin Arthritis Rheum, 2014. **44**(2): p. 145-54.
22. Jaaskelainen, S.K., T. Teerijoki-Oksa, and H. Forssell, *Neurophysiologic and quantitative sensory testing in the diagnosis of trigeminal neuropathy and neuropathic pain*. Pain, 2005. **117**(3): p. 349-57.

23. Kumar, S., et al., *Pain in trigeminal neuralgia: neurophysiology and measurement: a comprehensive review*. J Med Life, 2013. **6**(4): p. 383-8.
24. Nijs, J., et al., *Brain-derived neurotrophic factor as a driving force behind neuroplasticity in neuropathic and central sensitization pain: a new therapeutic target?* Expert Opin Ther Targets, 2015. **19**(4): p. 565-76.
25. Vickers, E.R. and M.J. Cousins, *Neuropathic orofacial pain. Part 2-Diagnostic procedures, treatment guidelines and case reports*. Aust Endod J, 2000. **26**(2): p. 53-63.
26. Zakrzewska, J.M., *Facial pain: an update*. Curr Opin Support Palliat Care, 2009. **3**(2): p. 125-30.
27. Zakrzewska, J.M., *Assessment and treatment of trigeminal neuralgia*. Br J Hosp Med (Lond), 2010. **71**(9): p. 490-4.
28. Zakrzewska, J.M. and R. McMillan, *Trigeminal neuralgia: the diagnosis and management of this excruciating and poorly understood facial pain*. Postgrad Med J, 2011. **87**(1028): p. 410-6.
29. Flor, H., et al., *Subtle Sensory Abnormalities Detected by Quantitative Sensory Testing in Patients with Trigeminal Neuralgia*. Pain Physician, 2016. **19**(7): p. 507-18.
30. Ichida, M.C., et al., *Functional and sensory evaluation of patients with idiopathic trigeminal neuralgia: comparison with controls*. Clin Neurol Neurosurg, 2015. **130**: p. 114-21.
31. Kalladka, M., et al., *Trigeminal nerve injury following accidental airbag deployment and assessment with quantitative sensory testing*. Cranio, 2007. **25**(2): p. 138-43.
32. Matos, R., et al., *Quantitative sensory testing in the trigeminal region: site and gender differences*. J Orofac Pain, 2011. **25**(2): p. 161-9.
33. Oono, Y., et al., *Effect of conditioned pain modulation on trigeminal somatosensory function evaluated by quantitative sensory testing*. Pain, 2013. **154**(12): p. 2684-90.
34. Sinay, V.J., L.H. Bonamico, and A. Dubrovsky, *Subclinical sensory abnormalities in trigeminal neuralgia*. Cephalalgia, 2003. **23**(7): p. 541-4.
35. Baad-Hansen, L., et al., *Reliability of intra-oral quantitative sensory testing (QST) in patients with atypical odontalgia and healthy controls - a multicentre study*. J Oral Rehabil, 2015. **42**(2): p. 127-35.
36. Jaaskelainen, S.K., *Clinical neurophysiology and quantitative sensory testing in the investigation of orofacial pain and sensory function*. J Orofac Pain, 2004. **18**(2): p. 85-107.
37. Porporatti, A.L., et al., *Diagnostic Accuracy of Quantitative Sensory Testing to Discriminate Inflammatory Toothache and Intraoral Neuropathic Pain*. J Endod, 2015. **41**(10): p. 1606-13.
38. Moisset, X., et al., *Co-occurrence of Pain Symptoms and Somatosensory Sensitivity in Burning Mouth Syndrome: A Systematic Review*. PLoS One, 2016. **11**(9): p. e0163449.
39. Renton, T., *Burning Mouth Syndrome*. Rev Pain, 2011. **5**(4): p. 12-7.
40. Kosek, E., et al., *Do we need a third mechanistic descriptor for chronic pain states?* Pain, 2016. **157**(7): p. 1382-6.
41. Woolf, C.J., *What to call the amplification of nociceptive signals in the central nervous system that contribute to widespread pain?* Pain, 2014. **155**(10): p. 1911-2.
42. Woolf, C.J., *Pain amplification-A perspective on the how, why, when, and where of central sensitization*. Journal of Applied Biobehavioral Research, 2018: p. e12124.
43. Yunus, M.B., *Central sensitivity syndromes: a new paradigm and group nosology for fibromyalgia and overlapping conditions, and the related issue of disease versus illness*. Semin Arthritis Rheum, 2008. **37**(6): p. 339-52.

44. Brummett, C., et al., *We agree with the need for a new term but disagree with the proposed terms*. Pain, 2016. **157**(12): p. 2876.
45. Clauw, D.J., *Fibromyalgia and related conditions*. Mayo Clin Proc, 2015. **90**(5): p. 680-92.
46. Le Bars, D., A.H. Dickenson, and J.M. Besson, *Diffuse noxious inhibitory controls (DNIC). I. Effects on dorsal horn convergent neurones in the rat*. Pain, 1979. **6**(3): p. 283-304.
47. Sandkuhler, J., *Models and mechanisms of hyperalgesia and allodynia*. Physiol Rev, 2009. **89**(2): p. 707-58.
48. Ross, X.L.a.T., *Neuroplasticity, Central Sensitization and Odontogenic Referred Orofacial Pain*. J pain Relief, 2015. **4**(6).
49. Petersson, K., et al., *Evaluation of the ability of thermal and electrical tests to register pulp vitality*. Endod Dent Traumatol, 1999. **15**(3): p. 127-31.
50. Mumford, J.M., *Pain perception threshold on stimulating human teeth and the histological condition of the pulp*. Br Dent J, 1967. **123**(9): p. 427-33.
51. Pau, A., et al., *Development and validation of a dental pain-screening questionnaire*. Pain, 2005. **119**(1-3): p. 75-81.
52. Pau, A., K.P. Viswanath, and R. Croucher, *Validation of a dental pain screening questionnaire in a semi-urban hospital setting in South India*. Int Dent J, 2010. **60**(2): p. 113-21.
53. Sessle, B.J., *Mechanisms of oral somatosensory and motor functions and their clinical correlates*. J Oral Rehabil, 2006. **33**(4): p. 243-61.
54. Scott, D., Jr. and T.R. Tempel, *NEUROPHYSIOLOGICAL RESPONSE OF SINGLE RECEPTOR UNITS IN THE TOOTH OF THE CAT*. J Dent Res, 1965. **44**: p. 20-7.
55. Scott, D., Jr., *The arousal and suppression of pain in the tooth*. Int Dent J, 1972. **22**(1): p. 20-32.
56. Trowbridge, H.O., et al., *Sensory response to thermal stimulation in human teeth*. J Endod, 1980. **6**(1): p. 405-12.
57. Narhi, M. and G. Haegerstam, *Intradental nerve activity induced by reduced pressure applied to exposed dentine in the cat*. Acta Physiol Scand, 1983. **119**(4): p. 381-6.
58. Narhi, M. and T. Hirvonen, *Functional changes in cat pulp nerve activity after thermal and mechanical injury of the pulp*. Proc Finn Dent Soc, 1983. **79**(4): p. 162-7.
59. Narhi, M., T. Hirvonen, and T. Huopaniemi, *Sensitivity of dentine*. Acupunct Electrother Res, 1983. **8**(2): p. 143-8.
60. Narhi, M., et al., *Comparison of electrical thresholds of intradental nerves and jaw-opening reflex in the cat*. Acta Physiol Scand, 1983. **119**(4): p. 399-403.
61. Virtanen, A., et al., *Thresholds of intradental A- and C-nerve fibres in the cat to electrical current pulses of different duration*. Acta Physiol Scand, 1983. **119**(4): p. 393-8.
62. Edwall, L. and D. Scott, Jr., *Influence of changes in microcirculation on the excitability of the sensory unit in the tooth of the cat*. Acta Physiol Scand, 1971. **82**(4): p. 555-66.
63. Ritter, A.L., et al., *Pulp revascularization of replanted immature dog teeth after treatment with minocycline and doxycycline assessed by laser Doppler flowmetry, radiography, and histology*. Dent Traumatol, 2004. **20**(2): p. 75-84.
64. Yanpiset, K., et al., *Efficacy of laser Doppler flowmetry for the diagnosis of revascularization of reimplanted immature dog teeth*. Dent Traumatol, 2001. **17**(2): p. 63-70.
65. Alghaithy, R.A. and A.J. Qualtrough, *Pulp sensibility and vitality tests for diagnosing pulpal health in permanent teeth: a critical review*. Int Endod J, 2017. **50**(2): p. 135-142.

66. Radhakrishnan, S., A.K. Munshi, and A.M. Hegde, *Pulse oximetry: a diagnostic instrument in pulpal vitality testing*. J Clin Pediatr Dent, 2002. **26**(2): p. 141-5.
67. Schnettler, J.M. and J.A. Wallace, *Pulse oximetry as a diagnostic tool of pulpal vitality*. J Endod, 1991. **17**(10): p. 488-90.
68. Nissan, R., et al., *Dual wavelength spectrophotometry as a diagnostic test of the pulp chamber contents*. Oral Surg Oral Med Oral Pathol, 1992. **74**(4): p. 508-14.
69. Gopikrishna, V., K. Tinagupta, and D. Kandaswamy, *Comparison of electrical, thermal, and pulse oximetry methods for assessing pulp vitality in recently traumatized teeth*. J Endod, 2007. **33**(5): p. 531-5.
70. Gopikrishna, V., G. Pradeep, and N. Venkateshbabu, *Assessment of pulp vitality: a review*. Int J Paediatr Dent, 2009. **19**(1): p. 3-15.
71. Samuel, S., A. Thomas, and N. Singh, *A comparative study of pulse oximetry with the conventional pulp testing methods to assess vitality in immature and mature permanent maxillary incisors*. CHRISMED Journal of Health and Research, 2014. **1**(4): p. 235-240.
72. Fein, M.E., et al., *Evaluation of optical methods of detecting dental pulp vitality*. J Biomed Opt, 1997. **2**(1): p. 58-73.
73. Schmitt, J.M., R.L. Webber, and E.C. Walker, *Optical determination of dental pulp vitality*. IEEE Trans Biomed Eng, 1991. **38**(4): p. 346-52.
74. Fratkin, R.D., D.J. Kenny, and D.H. Johnston, *Evaluation of a laser Doppler flowmeter to assess blood flow in human primary incisor teeth*. Pediatr Dent, 1999. **21**(1): p. 53-6.
75. Sasano, T., et al., *Possible application of transmitted laser light for the assessment of human pulp vitality. Part 2. Increased laser power for enhanced detection of pulpal blood flow*. Dent Traumatol, 2005. **21**(1): p. 37-41.
76. Jafarzadeh, H., *Laser Doppler flowmetry in endodontics: a review*. Int Endod J, 2009. **42**(6): p. 476-90.
77. Qu, X., M. Ikawa, and H. Shimauchi, *Improvement of the detection of human pulpal blood flow using a laser Doppler flowmeter modified for low flow velocity*. Arch Oral Biol, 2014. **59**(2): p. 199-206.
78. Kouadio, A.A., et al., *The use of laser Doppler flowmetry to evaluate oral soft tissue blood flow in humans: A review*. Arch Oral Biol, 2018. **86**: p. 58-71.
79. Fanibunda, K.B., *Diagnosis of tooth vitality by crown surface temperature measurement: a clinical evaluation*. J Dent, 1986. **14**(4): p. 160-4.
80. Fanibunda, K.B., *A method of measuring the volume of human dental pulp cavities*. Int Endod J, 1986. **19**(4): p. 194-7.
81. Fanibunda, K.B., *The feasibility of temperature measurement as a diagnostic procedure in human teeth*. J Dent, 1986. **14**(3): p. 126-9.
82. Fanibunda, K.B., *A laboratory study to investigate the differentiation of pulp vitality in human teeth by temperature measurement*. J Dent, 1985. **13**(4): p. 295-303.
83. Pogrel, M.A., C.K. Yen, and R.C. Taylor, *Studies in tooth crown temperature gradients with the use of infrared thermography*. Oral Surg Oral Med Oral Pathol, 1989. **67**(5): p. 583-7.
84. Jafarzadeh, H., C.I. Udoeye, and J. Kinoshita, *The application of tooth temperature measurement in endodontic diagnosis: a review*. J Endod, 2008. **34**(12): p. 1435-40.
85. da Silva, L.A., et al., *Quantitative sensory testing in fibromyalgia and hemisensory syndrome: comparison with controls*. Rheumatol Int, 2013. **33**(8): p. 2009-17.

86. Eliav, E., et al., *Quantitative sensory testing in trigeminal nerve damage assessment*. J Orofac Pain, 2004. **18**(4): p. 339-44.
87. Giesecke, J., et al., *Quantitative sensory testing in vulvodynia patients and increased peripheral pressure pain sensitivity*. Obstet Gynecol, 2004. **104**(1): p. 126-33.
88. Maier, C., et al., *Quantitative sensory testing in the German Research Network on Neuropathic Pain (DFNS): somatosensory abnormalities in 1236 patients with different neuropathic pain syndromes*. Pain, 2010. **150**(3): p. 439-50.
89. Pavlakovic, G. and F. Petzke, *The role of quantitative sensory testing in the evaluation of musculoskeletal pain conditions*. Curr Rheumatol Rep, 2010. **12**(6): p. 455-61.
90. Siao, P. and D.P. Cros, *Quantitative sensory testing*. Phys Med Rehabil Clin N Am, 2003. **14**(2): p. 261-86.
91. Uddin, Z. and J.C. MacDermid, *Quantitative Sensory Testing in Chronic Musculoskeletal Pain*. Pain Med, 2016. **17**(9): p. 1694-703.
92. Kim, H.K., K.S. Kim, and M.E. Kim, *Thermal Perception as a Key Factor for Assessing Effects of Trigeminal Nerve Injury*. J Oral Facial Pain Headache, 2017. **31**(2): p. 129-138.
93. Giesecke, T., et al., *Evidence of augmented central pain processing in idiopathic chronic low back pain*. Arthritis Rheum, 2004. **50**(2): p. 613-23.
94. O'Keefe, E.M., *Pain in endodontic therapy: preliminary study*. J Endod, 1976. **2**(10): p. 315-9.
95. Leffler, A.S., P. Hansson, and E. Kosek, *Somatosensory perception in a remote pain-free area and function of diffuse noxious inhibitory controls (DNIC) in patients suffering from long-term trapezius myalgia*. Eur J Pain, 2002. **6**(2): p. 149-59.
96. Maixner, W., et al., *Sensitivity of patients with painful temporomandibular disorders to experimentally evoked pain*. Pain, 1995. **63**(3): p. 341-51.
97. Hollins, M., et al., *Perceived intensity and unpleasantness of cutaneous and auditory stimuli: an evaluation of the generalized hypervigilance hypothesis*. Pain, 2009. **141**(3): p. 215-21.
98. King, C.D., et al., *Deficiency in endogenous modulation of prolonged heat pain in patients with Irritable Bowel Syndrome and Temporomandibular Disorder*. Pain, 2009. **143**(3): p. 172-8.
99. Salgar, A.R., et al., *Determining predictability and accuracy of thermal and electrical dental pulp tests: An in vivo study*. J Conserv Dent, 2017. **20**(1): p. 46-49.
100. Baumgardner, K.R., et al., *Induced hypoxia in rat pulp and periapex demonstrated by 3H-misonidazole retention*. J Dent Res, 1996. **75**(10): p. 1753-60.
101. Zach, L. and G. Cohen, *PULP RESPONSE TO EXTERNALLY APPLIED HEAT*. Oral Surg Oral Med Oral Pathol, 1965. **19**: p. 515-30.
102. Baldissara, P., S. Catapano, and R. Scotti, *Clinical and histological evaluation of thermal injury thresholds in human teeth: a preliminary study*. J Oral Rehabil, 1997. **24**(11): p. 791-801.
103. Fuss, Z., et al., *Assessment of reliability of electrical and thermal pulp testing agents*. J Endod, 1986. **12**(7): p. 301-5.
104. Peters, D.D., et al., *Evaluation of the effects of carbon dioxide used as a pulpal test. 1. In vitro effect on human enamel*. J Endod, 1983. **9**(6): p. 219-27.
105. Augsburger, R.A. and D.D. Peters, *In vitro effects of ice, skin refrigerant, and CO2 snow on intrapulpal temperature*. J Endod, 1981. **7**(3): p. 110-6.
106. Peters, D.D. and R.A. Augsburger, *In vitro model system to evaluate intrapulpal temperature changes*. J Endod, 1981. **7**(7): p. 320-4.

107. Mumford, J.M., *Thermal and electrical stimulation of teeth in the diagnosis of pulpal and periapical disease*. Proc R Soc Med, 1967. **60**(2): p. 197-200.
108. Perquin, C.W., et al., *Pain in children and adolescents: a common experience*. Pain, 2000. **87**(1): p. 51-8.
109. Massaro, M., et al., *A comparison of three scales for measuring pain in children with cognitive impairment*. Acta Paediatr, 2014. **103**(11): p. e495-500.
110. Breau, L.M., et al., *Preliminary validation of an observational pain checklist for persons with cognitive impairments and inability to communicate verbally*. Dev Med Child Neurol, 2000. **42**(9): p. 609-16.
111. Stallard, P., et al., *Pain in cognitively impaired, non-communicating children*. Arch Dis Child, 2001. **85**(6): p. 460-2.