

# Pathophysiology of pachyonychia congenita-associated palmoplantar keratoderma: new insights into skin epithelial homeostasis and avenues for treatment\*

A.G. Ziemann <sup>1</sup> and P.A. Coulombe<sup>1,2</sup>

Departments of <sup>1</sup>Cell and Developmental Biology and <sup>2</sup>Dermatology; University of Michigan Medical School, 3071 Biomedical Sciences Research Building, 109 Zina Pitcher Place, Ann Arbor, MI 48109, U.S.A.

**Linked Comment:** Leube and Schwarz. *Br J Dermatol* 2020; **182**:525–526.

## Summary

### Correspondence

Pierre A. Coulombe.  
E-mail: coulombe@umich.edu

### Accepted for publication

22 April 2019

### Funding sources

These studies were supported by grant AR044232 issued to P.A.C. from the National Institute of Arthritis, Musculoskeletal and Skin Disease (NIAMS). A.G.Z. received support from grant T32 CA009110 from the National Cancer Institute.

### Conflicts of interest

None declared.

\*Plain language summary available online

DOI 10.1111/bjd.18033

**Background** Pachyonychia congenita (PC), a rare genodermatosis, primarily affects ectoderm-derived epithelial appendages and typically includes oral leukokeratosis, nail dystrophy and very painful palmoplantar keratoderma (PPK). PC dramatically impacts quality of life although it does not affect lifespan. PC can arise from mutations in any of the wound-repair-associated keratin genes *KRT6A*, *KRT6B*, *KRT6C*, *KRT16* or *KRT17*. There is no cure for this condition, and current treatment options for PC symptoms are limited and palliative in nature.

**Objectives** This review focuses on recent progress made towards understanding the pathophysiology of PPK lesions, the most prevalent and debilitating of all PC symptoms.

**Methods** We reviewed the relevant literature with a particular focus on the *Krt16* null mouse, which spontaneously develops footpad lesions that mimic several aspects of PC-associated PPK.

**Results** There are three main stages of progression of PPK-like lesions in *Krt16* null mice. Ahead of lesion onset, keratinocytes in the palmoplantar (footpad) skin exhibit specific defects in terminal differentiation, including loss of *Krt9* expression. At the time of PPK onset, there is elevated oxidative stress and hypoactive Keap1–Nrf2 signalling. During active PPK, there is a profound defect in the ability of the epidermis to maintain or return to normal homeostasis.

**Conclusions** The progress made suggests new avenues to explore for the treatment of PC-based PPK and deepens our understanding of the mechanisms controlling skin tissue homeostasis.

### What's already known about this topic?

- Pachyonychia congenita (PC) is a rare genodermatosis caused by mutations in *KRT6A*, *KRT6B*, *KRT6C*, *KRT16* and *KRT17*, which are normally expressed in skin appendages and induced following injury.
- Individuals with PC present with multiple clinical symptoms that usually include thickened and dystrophic nails, palmoplantar keratoderma (PPK), glandular cysts and oral leukokeratosis.
- The study of PC pathophysiology is made challenging because of its low incidence and high complexity. There is no cure or effective treatment for PC.

### What does this study add?

- This text reviews recent progress made when studying the pathophysiology of PPK associated with PC.
- This recent progress points to new possibilities for devising effective therapeutics that may complement current palliative strategies.

Pachyonychia congenita (PC; OMIM #1672000 and 167210) is a rare genodermatosis with a collection of symptoms primarily affecting ectoderm-derived appendages; it includes oral leukokeratosis, nail dystrophies, sebaceous cysts, natal teeth and palmoplantar keratoderma (PPK). While PC does not impact lifespan, it dramatically impacts quality of life for affected individuals. For instance, individuals with PC experience severe plantar pain from PPK lesions daily, often making everyday tasks difficult. There is currently no known cure or effective therapeutics for the treatment of PC.<sup>1</sup>

PC can arise from autosomal dominant mutations in any of five keratin genes including *KRT6A*, *KRT6B*, *KRT6C*, *KRT16* or *KRT17*.<sup>1–5</sup> These keratins are normally expressed in epithelial appendages and are otherwise robustly inducible, e.g. after injury or exposure to environmental stresses, together accounting in part for the clinical presentation of this disorder. Most PC-causing mutations are missense alleles, with occasional small insertions or deletions in the keratin coding sequence. Until recently, two major types of PC – type 1 (Jadassohn–Lewandowsky<sup>6</sup>) and type 2 (Jackson–Lawler<sup>7</sup>) – were recognized based on their prevalent clinical features. Nowadays, five subtypes of PC are recognized based on genetic aetiology – for example, PC caused by a *KRT6A* mutation corresponds to the PC-K6a subtype. Owing in part to the heterogeneity in the clinical presentation of PC (even among patients with very similar alleles), a definitive diagnosis can be ascertained only through sequencing of these keratin genes.<sup>1,8,9</sup>

### The Pachyonychia Congenita Project

The Pachyonychia Congenita Project is a U.S. public charity that was founded in 2003 and has evolved into a life-changing resource for individuals with PC and for clinicians and researchers interested in this condition. This organization connects individuals with PC and their families to others with this condition, and to clinicians, translational and basic science researchers. The Pachyonychia Congenita Project provides assistance to individuals with PC to attend support meetings and qualify for genetic testing. Further, the Pachyonychia Congenita Project is home to the International Pachyonychia Congenita Research Registry (IPCRR), which gathers extremely valuable data from questionnaires, photos and notes on genetically confirmed PC cases. As of January 2019, the IPCRR includes 864 genetically confirmed cases of PC in 49 countries. This PC registry has evolved into a transformative resource for patients, clinicians and researchers working together towards understanding this disorder and developing

effective therapeutics. Finally, the Pachyonychia Congenita Project plays a lead role in fostering basic and clinical research on PC. More information about the Pachyonychia Congenita Project and how to get involved can be found on the publicly available website: [www.pachyonychia.org](http://www.pachyonychia.org).

### Asserting a focus on palmoplantar keratoderma

While individuals with PC present with many symptoms of significance, PPK is highly penetrant and reportedly the most debilitating (Fig. 1a).<sup>1</sup> Virtually all individuals with PC (> 90%)<sup>1</sup> present with PPK lesions restricted primarily to pressure points in the palmar and/or plantar epidermis and consisting of dramatic epidermal thickening and hyperkeratosis.<sup>3–5,10</sup> PPK lesions are debilitating in part because of the extreme pain associated with them.<sup>11–13</sup> Interestingly, these lesions do not display signs of keratinocyte fragility and/or lysis. The latter represents a predominant element in epidermolysis bullosa simplex (EBS), a genetically determined skin blistering condition caused by mutations in either *KRT14* or *KRT5*.<sup>14–17</sup> Keratinocyte fragility is also a dominant pathophysiological feature in epidermolytic PPK, which is often caused by mutations in *KRT9*,<sup>18</sup> the major differentiation-specific keratin in the volar epidermis.<sup>19,20</sup> The greater complexity of keratin gene expression in the volar epidermis likely contributes to maintain keratinocyte structural integrity in spite of mutations in individual genes such as *KRT6A-C*, *KRT16* and *KRT17*. That said, the pathophysiology of PC-associated PPK is only partially understood at present, reflecting significant limitations related to the low incidence of this orphan disease and the severe pain associated with these lesions.<sup>21,22</sup> Accordingly, there is no effective treatment for PC-based PPK. The current standard of care for PPK consists of routine removal of calluses followed by treatment with moisturizers (see below for details).<sup>23</sup> A deeper understanding of the pathophysiology of PPK might spearhead the development of effective therapeutics for individuals with PC and also inform researchers, clinicians and drug developers on other genetic and clinical subtypes of PPK (Fig. 1b). This text focuses on recent progress made in deciphering the pathophysiology of PC-associated PPK lesions.

### A short primer on the nomenclature of pachyonychia congenita-associated keratin genes

The original catalogue of human keratin proteins devised by Moll et al.<sup>24</sup> already recognized the existence of K6 as a type II

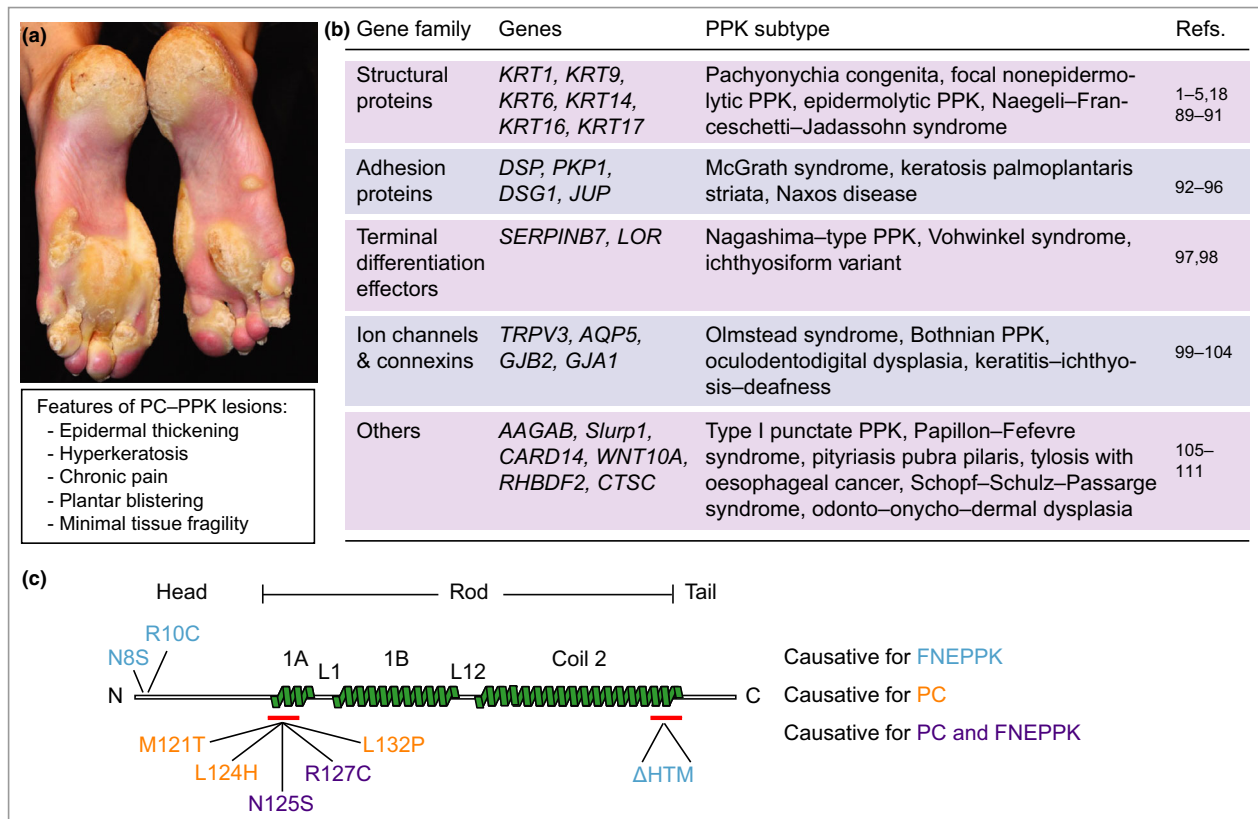


Fig 1. PPK, a genetically heterogeneous disorder. (a) Photograph of PC-based PPK lesions from an individual with a *KRT16* L124R mutation. Source: Pachyonychia Congenita Project ([www.pachyonychia.org](http://www.pachyonychia.org)). (b) Table summarizing the diversity of genes which, when mutated, can elicit a PPK clinical presentation. Various clinical subtypes of PPK are accounted for.<sup>18,89-111</sup> (c) Schematic of select mutations in K16 protein that are causative for PC, FNEPPK, or both PC and FNEPPK. K16 exhibits the tripartite domain structure shared by all IF proteins, with an N-terminal 'head' domain, central  $\alpha$ -helical 'rod' domain and C-terminal 'tail' domain. The central rod domain is comprised of heptad repeat-containing  $\alpha$ -helical coils (1A, 1B, Coil 2) separated by non-heptad repeat linkers (L1 and L12). Many attributes of the central rod domain (red bars) are highly conserved and represent a signature element among IF proteins. Representative mutations that are causative for FNEPPK are in blue text, mutations causative for PC are in gold text, and mutations that are causative for both FNEPPK and PC are in purple text. FNEPPK, focal nonepidermolytic PPK; IF, intermediate filament; K16, keratin 16; PC, pachyonychia congenita; PPK, palmoplantar keratoderma.

keratin, and of K16 and K17 as type I keratins. However, the true diversity of keratin genes and proteins was underestimated until the advent of whole-genome sequencing efforts,<sup>25</sup> which necessitated a revision of the Moll nomenclature.<sup>26</sup> As per the internationally accepted nomenclature, human genes are designated using upper-case lettering (e.g. *KRT16*) and mouse genes are designated using lower-case lettering (e.g. *Krt16*). The multiplicity of K6 sequences was originally uncovered in the human.<sup>27</sup> Currently, we know of two functional genes in the mouse, *Krt6a* and *Krt6b*,<sup>28</sup> and three functional K6 genes in the human, *KRT6A*, *KRT6B* and *KRT6C*.<sup>2,29</sup> In contrast, a single gene codes for each of K16<sup>30,31</sup> and K17 proteins<sup>32,33</sup> in the human and mouse genomes. The high degree of conservation known to apply to orthologous keratin genes in the mouse and human, in terms of sequence features and regulation, applies to the PC-associated keratin genes.<sup>29,31,32</sup> This information is relevant to discussing the utilization of transgenic mouse models to study keratin mutation-based human conditions such as PC.

## Lessons learned from transgenic mouse models

As there are no *in vitro* human cell culture models that can be used to investigate the cellular and molecular mechanisms underlying PPK pathophysiology or screen potential therapeutics, researchers have relied on the use of transgenic mouse models (summarized in Table 1) to study PC and PPK.<sup>34</sup> Among the models available, the *Krt16* null mouse strain is the only one that spontaneously develops footpad skin lesions mimicking PC-associated PPK lesions. Characterization of *Krt16* null mice has revealed three phases in PPK, each with a somewhat unique molecular signature: pre-PPK, PPK onset and active PPK (Fig. 2).

In 2-week-old *Krt16* null mice, corresponding to the 'pre-PPK stage', footpad skin keratinocytes exhibit defects in selective aspects of terminal differentiation. At this early time point there are minimal alterations to the skin tissue histology but, already, a dramatic loss of K9 (*Krt9/K9*) expression has occurred, which

**Table 1** Mouse models with phenotypes that are potentially relevant to pachyonychia congenita (PC)

Year	Mouse model	Genetic modification	Main phenotype(s)	References
1996	<i>Krt6a</i> $\Delta$ 21P	Deletion of 52 amino acids (residues 125–176) between head and 1A helix domain	Intraepidermal blistering	82
1999	<i>Krt6a</i> transgenic	Truncation deleting the 2B region of the central rod domain	Lethal blister or alopecia	83
1999	<i>Krt6a</i> transgenic	Replacement of E2 by HK1-tag	Hyperkeratosis and late-onset alopecia	83
2000	<i>Krt6a/Krt6b</i> <sup>-/-</sup>	Deletion of <i>Krt6a</i> and <i>Krt6b</i> loci	Oral lesions	49,84
2000	<i>Krt6a</i> <sup>-/-</sup>	Deletion of <i>Krt6a</i> loci	Delay of reepithelialization after wounding	85
2002	<i>Krt17</i> <sup>-/-</sup>	Deletion of <i>Krt17</i> locus	Age- and strain-dependent alopecia	48
2005	<i>Krt6a/Krt6b</i> <sup>-/-</sup> ; <i>Krt17</i> <sup>-/-</sup>	Deletion of <i>Krt6a</i> , <i>Krt6b</i> and <i>Krt17</i> loci	Severe cell lysis in nail bed epithelium	86
2008	<i>Krt75</i> knock-in	Point mutation of codon N158 (corresponding to mutation N171 in PC case)	Defects in hair shaft, nail fragility	87
2011	KRT6A N171K humanized skin	Bioengineered skin equivalents derived from individuals with PC with N171K mutation engrafted onto immunodeficient mice	Acanthosis and epidermal blistering	88
2012	<i>Krt16</i> <sup>-/-</sup>	Deletion of <i>Krt16</i> locus	Oral lesions, footpad lesions resembling human PPK	38,47

PPK, palmoplantar keratoderma

then persists throughout lesion progression.<sup>35</sup> *Krt9* occurs exclusively in differentiating keratinocytes of volar skin and represents a predominant marker gene in this setting.<sup>19,20,36</sup> In contrast to *Krt9*, several differentiation markers appear to be upregulated in *Krt16* null footpad skin, potentially as a compensatory mechanism.<sup>35</sup> While this partial defect in terminal differentiation is currently unexplained,<sup>35</sup> it occurs independent and ahead of the oxidative stress phenotype observed at a later stage of progression of PPK-like lesions in this mouse model.<sup>35,37</sup>

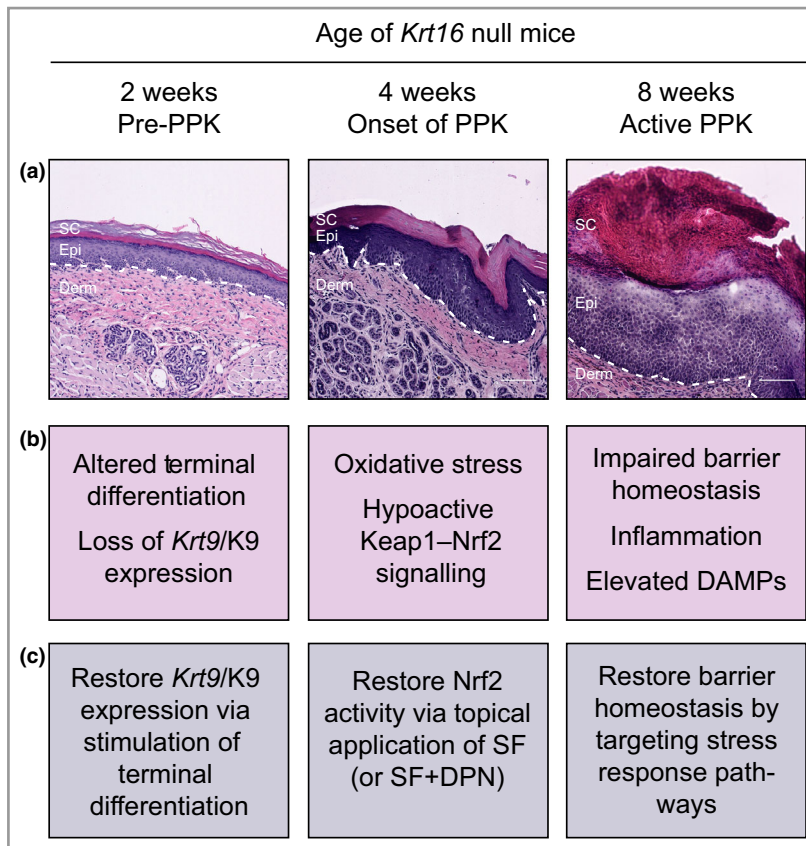
In 1-month-old *Krt16* null mice, corresponding to 'onset stage' of PPK-like lesions, footpad skin epidermis displays several features of oxidative stress, including decreased levels of the master cell antioxidant glutathione and decreased expression of glutathione synthesis genes compared with WT controls. Keap1–Nrf2 signalling, a central regulator of the cellular antioxidant response, is markedly attenuated at that time while Nrf2 itself, a transcription factor, is upregulated though ineffective in *Krt16* null footpad skin (likely reflecting an attempt to restore redox homeostasis). While difficult to ascertain given restricted access to plantar skin biopsies from patients with PC, there is evidence of reduced Nrf2 activity in PC–PPK lesions of individuals with KRT16 mutations.<sup>37</sup>

In 2-month-old *Krt16* null mice, corresponding to an 'active stage' of PPK, there is a profound defect in the ability of footpad skin to maintain or return to normal tissue homeostasis. By this time all *Krt16* null mice have spontaneously developed PPK-like lesions on their paws, and while these lesions preferentially arise in areas exposed to the substratum and thus experience mechanical stress, they are not associated with keratinocyte fragility.<sup>38,39</sup> At a molecular level, the *Krt16* null footpad lesions exhibit a gross misregulation of many danger-associated molecular patterns (DAMPs) and barrier homeostasis genes, which mimics human PC-based PPK lesions.<sup>39</sup>

## Lessons learned from computational endeavours

Along with targeted molecular analyses, computational analysis of genomic datasets has also provided significant insight into the pathophysiology of PC-based PPK. Systems genetics has been used to explore the role of *K16* in regulating the skin's response to stress. Re-analysing a powerful systems genetics dataset that related the risk of developing skin tumours to the regulation of skin inflammation and barrier function<sup>40</sup> revealed a tight link between *Krt16*, skin barrier genes and innate immunity effectors including DAMPs.<sup>39,41</sup> Moreover, in this dataset, *Krt16* expression is significantly correlated with expression of barrier homeostasis and inflammation genes in tail skin, both at baseline and in response to 12-O-tetradecanoylphorbol-13-acetate (TPA), which acts as a chemical irritant.<sup>41</sup> The discovery that *Krt16* belongs to a network of barrier homeostasis genes pointed to a role for *Krt16* in calibrating the skin's response to barrier-compromising stresses,<sup>39</sup> which converged nicely with the phenotype of PPK-like lesions exhibited by *Krt16* null mice. These efforts lent strong support to the notion that a better understanding of how *K16* calibrates the skin's stress response could be applicable to PC as KRT16 expression is often elevated in PC-based PPK lesions.

The availability of genome-wide surveys of gene expression from both *Krt16* null footpad skin lesions<sup>35</sup> and PPK lesions from individuals with PC<sup>22</sup> has provided an excellent opportunity to further test the strengths and limitations of the *Krt16* null mouse as a valid model for PC-based PPK. Merging the human PPK datasets with the murine *Krt16* null footpad lesions dataset, based on human–mouse orthologous gene pairings, enabled multiple computational analyses.<sup>35</sup> Pairwise comparisons of global transcriptional changes in *Krt16* null footpad



**Fig 2.** Development of PPK-like lesions in *Krt16* null footpad skin proceeds in three stages: pre-PPK (at 2 weeks of age), onset of PPK (at 4 weeks) and active PPK (at 8 weeks). (a) Representative histology of *Krt16* null footpad skin at 2, 4 and 8 weeks of age. At 2 weeks, the epidermis shows a normal thickness and overall architecture but, on closer inspection, alterations including the abnormal appearance of the granular layer, crowding of basal keratinocytes, and a decreased nuclear aspect ratio of basal keratinocytes can be seen. At 4 weeks, prior to macroscopic appearance of lesions, mild epidermal thickening is observed. By 8 weeks, there is dramatic thickening of the living epidermis (Epi) and the stratum corneum (SC), infiltration of immune cells, and limited suprabasal cell lysis. The dotted line shows the epidermal–dermal junction. Scale bar = 100  $\mu$ m. Images acquired using a Zeiss microscope with Apotome attachment and processed using Zen 2.3 software. (b) Summary of key molecular changes that occur at 2, 4 and 8 weeks of age in *Krt16* null footpad skin.<sup>35,37–39,47,77</sup> (c) Potential therapeutic interventions for each stage of lesion development.<sup>35,37,39,77</sup> DAMPs, damage-associated molecular pattern molecules; Derm, dermis; DPN, diarylpropionitrile; PPK, palmoplantar keratoderma; SF, sulforaphane.

lesions and individual PC cases (three *KRT6* cases, three *KRT16* cases) generated statistically significant positive correlation values in all cases (Fig. 3a,b). Additionally, pairwise comparisons of global transcriptional changes further highlight the high degree of heterogeneity between individual cases involving different keratin mutations, and between cases with the same mutated keratin allele (Fig. 3c). Altogether these comparisons provided a strong case that lesional *Krt16* null mouse footpad skin mimics PC-associated PPK lesions at a global gene expression level, reinforcing and extending the notion that the *Krt16* null mouse is an appropriate model for the study of pathogenesis of PC-associated PPK lesions.

### A role for keratin imbalances and genetic background in the pathophysiology of palmoplantar keratoderma

Because the presentation of PC symptoms varies greatly between individuals even with similar or the same mutated

keratin allele,<sup>1,42–44</sup> there is likely a role for genetic background and gene modifiers in the pathophysiology of this condition. Remarkably, similar alleles in *KRT16* (N125S and R127C) can elicit a presentation of focal nonepidermolytic PPK vs. full-blown PC (Fig. 1c),<sup>44–46</sup> suggesting that the consequences associated with alterations in *KRT16* are subject to modifier gene(s) effects. Consistent with this notion, despite the immunological differences between mice and humans, several phenotypic aspects of *Krt16* null mice including the PPK-like lesions are modestly impacted by genetic strain background.<sup>47</sup> Interestingly, select phenotypic traits in *Krt17* null mice<sup>48</sup> and *Krt6a/Krt6b* double-null mice<sup>49</sup> also exhibit a dependence on genetic background.

In addition to genetic background, imbalances in keratin expression also appear likely to play a significant role in the pathophysiology of PC-based PPK. For example, the differentiation-specific keratins *KRT2* and *KRT9* are both decreased in *Krt16* null footpad lesions and human PC-based PPK.<sup>22,35,50</sup> Of note, mice that are double-null for the differentiation-specific

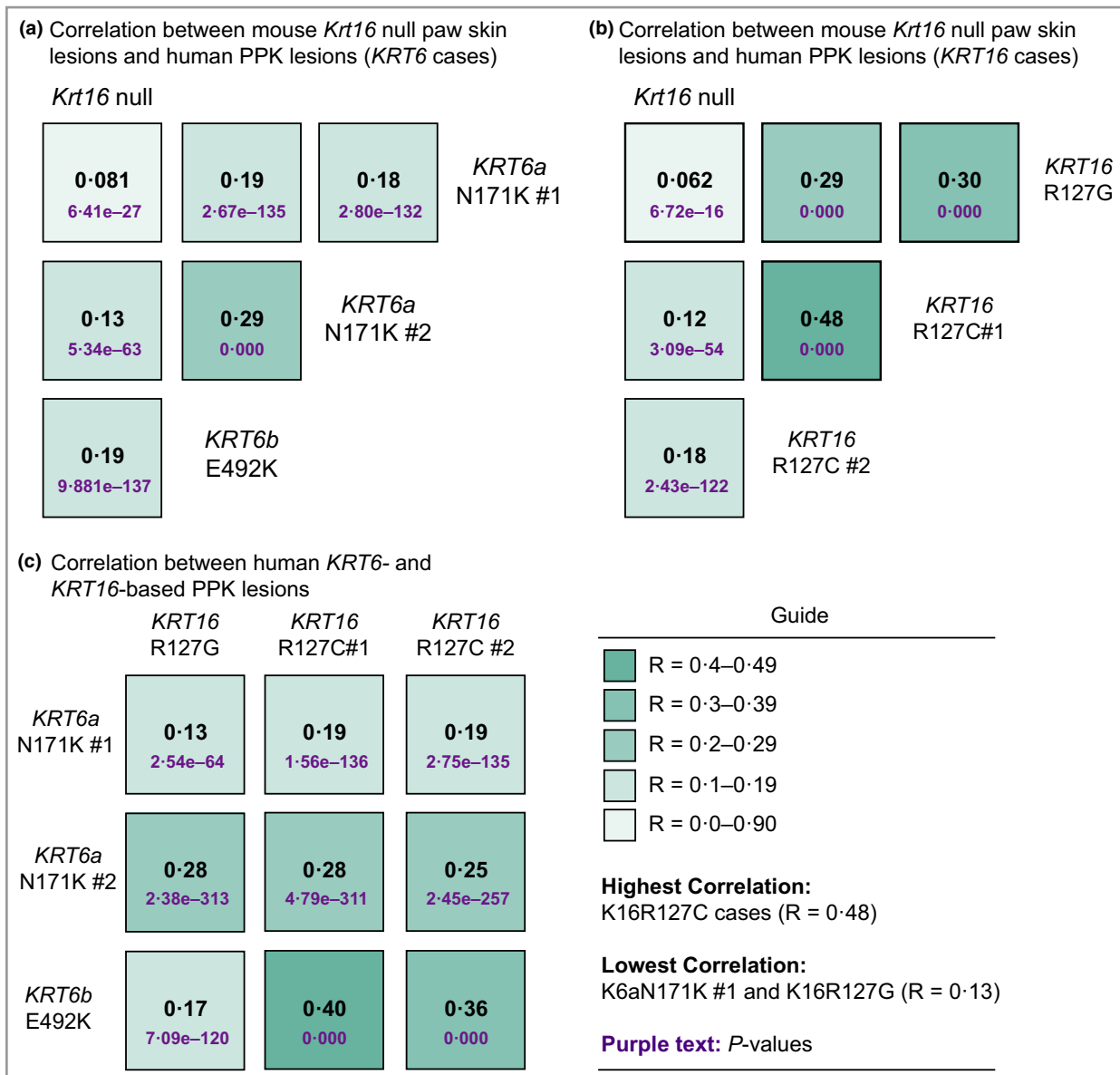


Fig 3. Correlation of transcriptional changes between *Krt16* null footpad lesions and human PC-based PPK lesions. (a,b) Correlation coefficients (R) calculated from pairwise comparisons of microarray data from *Krt16* null footpad lesions<sup>35</sup> and human PC-based PPK lesions<sup>22</sup> resulting from mutations in *KRT6* (a) or *KRT16* (b). (c) Correlation coefficients calculated from pairwise comparisons between individual cases of PC-based PPK with either *KRT6* or *KRT16* mutations. An R-value of -1 would convey perfect negative correlation, whereas an R-value of 1 conveys perfect correlation between samples. P-values for each correlation are denoted in purple text underneath the corresponding R-values. Figure adapted from Ziemann et al.<sup>35</sup> PC, pachyonychia congenita; PPK, palmoplantar keratoderma

*Krt2* and *Krt10* develop a keratoderma-like phenotype on footpad skin<sup>51</sup> while mice null for *Krt9* develop an epidermolytic PPK that closely resembles the corresponding human disorder.<sup>36</sup> Aside from these (and other) alterations,<sup>22,35</sup> the expression of *KRT6A*, *KRT6B*, *KRT16* and/or *KRT17* (including the mutated alleles) is dramatically increased in PC-based PPK, as expected given the stress- and wound-sensitive regulation of these genes.<sup>22</sup> Given the knowledge that K6, K16 and K17 proteins have pleiotropic and context-dependent properties,<sup>35,37,39,52–56</sup> such alterations in keratin protein levels and

balance among them are poised to have a striking impact on the development and evolution of PPK lesions.

### Pathophysiological unknowns in palmoplantar keratoderma and other clinical features of pachyonychia congenita

While all clinical manifestations associated with PC are worth a deep investigation, two stand out as remaining particularly intriguing at a cellular and molecular level. One is

the occurrence of individual or multiple cysts (steatocystoma multiplex; see OMIM entries #184500 and #184510) in patients with PC, which are benign fluid-filled cysts believed to originate from sebaceous glands and which can occur all over the body and arise preferentially in individuals with mutations in *KRT17*.<sup>57–59</sup> These cysts often require surgical drainage or removal as their rupture and/or inflammation pose a risk of infection and can be painful for patients.<sup>1</sup> Another intriguing manifestation is natal teeth, which refers to presence of teeth in newborns and is also preferentially associated with mutations in *KRT17*.<sup>1,59</sup> Natal teeth are soft, friable and prone to caries, and are usually lost within the first few months of life.<sup>1,60–63</sup> Of note, *Krt17* is expressed at a very early stage of the development of ectodermal appendages, including the tooth.<sup>32</sup> Recent studies have shown that genetic variants in the PC-associated keratin genes increase susceptibility to tooth decay.<sup>64</sup> There is currently no model to study the cystic skin lesions and phenomenon of natal teeth associated with PC.

### Limitations of past and current therapeutic strategies for pachyonychia congenita

PC-associated PPK has been treated with a combination of keratolytics, pain medication, orthotics and mechanical removal of calluses.<sup>23,65</sup> While the keratolytics salicylic acid and urea soften calluses, they cannot control the significant overgrowth associated with most cases of PC–PPK. Pain medication and custom orthotics can partially alleviate discomfort, but do not treat the underlying PPK. Routine mechanical removal of calluses by filing, grinding or cutting has been the most satisfying treatment for individuals with PC.<sup>23</sup> Significant efforts are currently under way to develop new and effective therapeutics for the management of these lesions. Two distinct strategies are highlighted here. The first strategy involves the development of short interfering RNAs (siRNAs) that specifically target mutant keratin alleles and reduce their expression. It has shown some promise in a trial of the siRNA TD101, which targets the *KRT6A* N171K allele, albeit in a single patient.<sup>21,66,67</sup> In its current form, this approach suffers from the limitation that delivery of such nucleic acid-based therapeutics requires intradermal injections that cause intense pain to the patient. The generation of self-delivery siRNAs for mutant keratin alleles improves the uptake of siRNAs by keratinocytes<sup>68</sup> but does not improve penetration through the stratum corneum. Accordingly, a method to deliver siRNA-based therapeutics that involves topical application of therapeutic agents is sorely needed.<sup>69</sup>

The second strategy to treat PC-based PPK consists of drug-based interventions aimed at reducing mutant keratin gene expression. The mammalian target of rapamycin (mTOR) inhibitor rapamycin/sirolimus suppresses K6a expression and, when taken orally, improves PC symptoms.<sup>70</sup> However, severe side-effects associated with systemic rapamycin treatment prevent it from being a viable long-term treatment for PC. Recently, topical sirolimus treatment of two patients with

K6a-based disease improved PC–PPK without the toxicity of systemic treatment<sup>71</sup> but requires additional studies to confirm the safety and efficacy of this treatment. Oral retinoids successfully reduced callus thickness in some individuals with PC, but like rapamycin, adverse side-effects including increased pain prevent oral retinoids from being a viable long-term treatment for PC.<sup>72</sup> Statins can also downregulate *KRT6A* expression,<sup>73</sup> but so far only oral rosuvastatin has been shown to be effective in a single case of K6a-based PC.<sup>74</sup> Finally, injections of botulinum toxin (Botox) into plantar calluses improved plantar blistering and pain associated with PC–PPK lesions,<sup>75</sup> but injections are costly and must be performed under anaesthesia. While each of these drug-based interventions provides some relief, none of them in present form provide viable long-term treatment strategies for PC-based PPK.

### Opportunities for novel therapies

A promising opportunity to complement ongoing efforts to develop effective therapeutics for PC-based PPK would be to target stress response pathways and/or pathways capable of promoting the restoration of normal epidermal differentiation. In *Krt16* null mice, topical treatment with the small natural molecule sulforaphane (SF), which activates Nrf2 signalling by modifying Keap1,<sup>76</sup> can prevent PPK-like lesions in male mice.<sup>37</sup> Addition of the ER- $\beta$  agonist diarylpropionitrile to the SF treatment regimen is necessary for successful activation of Nrf2<sup>77</sup> and prevention of PPK-like lesions in female mice. SF is available in pure form or as part of broccoli sprout extract,<sup>78</sup> can be safely delivered topically, and has shown therapeutic promise in the treatment of EBS arising from mutations in either keratins K5 or K14.<sup>79,80</sup> The sexual dimorphism in response to SF treatment in mice is a reminder that sex-based differences are important considerations when developing therapeutics for any disease.<sup>81</sup> Whether there is a sexual dimorphism in the setting of PC remains an open question.

Another strategy worth considering is to normalize terminal differentiation in volar skin. In male *Krt16* null mice treated with SF prior to lesion onset, restoration of Nrf2 activity coincided with induction of *Krt9* expression.<sup>35</sup> Additional efforts should be focused on testing this specific strategy. In the end, the prospect of combining treatment modalities that act to prevent and/or treat active lesions represent an attractive prospect for the treatment of a condition featuring the complexity of PC-associated PPK.

### Future directions

PC is a monogenic skin disease with a complex, polygenic presentation. Despite the plethora of challenges that arise in studying this disease, the use of transgenic mouse models and of computational biology has been invaluable and has provided novel insight into the pathophysiology of PC-based PPK, one of the most debilitating symptoms for individuals with PC. The study of PPK pathophysiology not only paves the way for researchers to devise therapeutics to treat PC, but also

provides an opportunity to better understand the mechanisms that control skin tissue homeostasis.

## Acknowledgments

The authors wish to thank members of the Coulombe laboratory for their support. This work was supported by grant AR044232 from the National Institutes of Health.

## References

- Leachman SA, Kaspar RL, Fleckman P et al. Clinical and pathological features of pachyonychia congenita. *J Invest Dermatol* 2005; **10**:3–17.
- Wilson NJ, Messenger AG, Leachman SA et al. Keratin K6c mutations cause focal palmoplantar keratoderma. *J Invest Dermatol* 2010; **130**:425–9.
- McLean WH, Rugg EL, Lunny DP et al. Keratin 16 and keratin 17 mutations cause pachyonychia congenita. *Nat Genet* 1995; **9**:273–8.
- Smith FJ, Jonkman MF, van Goor H et al. A mutation in human keratin K6b produces a phenocopy of the K17 disorder pachyonychia congenita type 2. *Hum Mol Genet* 1998; **7**:1143–8.
- Bowden PE, Haley JL, Kinsky A et al. Mutation of a type II keratin gene (K6a) in pachyonychia congenita. *Nat Genet* 1995; **10**:363–5.
- Franklin J. Pachyonychia congenita (Jadassohn and Lewandowski). *Proc R Soc Med* 1939; **32**:263–5.
- Jackson AD, Lawler SD. Pachyonychia congenita; a report of six cases in one family, with a note on linkage data. *Ann Eugen* 1951; **16**:142–6.
- Liao H, Sayers JM, Wilson NJ et al. A spectrum of mutations in keratins K6a, K16 and K17 causing pachyonychia congenita. *J Dermatol Sci* 2007; **48**:199–205.
- Fu T, Leachman SA, Wilson NJ et al. Genotype–phenotype correlations among pachyonychia congenita patients with K16 mutations. *J Invest Dermatol* 2011; **131**:1025–8.
- Lin MT, Levy ML, Bowden PE et al. Identification of sporadic mutations in the helix initiation motif of keratin 6 in two pachyonychia congenita patients: further evidence for a mutational hot spot. *Exp Dermatol* 1999; **8**:115–19.
- Krupiczkoj MA, O'Toole EA. Plantar pain in pachyonychia congenita. *Br J Dermatol* 2018; **179**:11–12.
- Brill S, Sprecher E, Smith FJD et al. Chronic pain in pachyonychia congenita: evidence for neuropathic origin. *Br J Dermatol* 2018; **179**:154–62.
- Weinberg RL, Coulombe PA, Polydefkis M, Caterina MJ. Pain mechanisms in hereditary palmoplantar keratoderms. *Br J Dermatol* 2020; **182**:543–51.
- Coulombe PA, Fuchs E. Epidermolysis bullosa simplex. *Semin Dermatol* 1993; **12**:173–90.
- Coulombe PA, Hutton ME, Letai A et al. Point mutations in human keratin 14 genes of epidermolysis bullosa simplex patients: genetic and functional analyses. *Cell* 1991; **66**:1301–11.
- Coulombe PA, Hutton ME, Vassar R et al. A function for keratins and a common thread among different types of epidermolysis bullosa simplex diseases. *J Cell Biol* 1991; **115**:1661–74.
- Coulombe PA, Kerns ML, Fuchs E. Epidermolysis bullosa simplex: a paradigm for disorders of tissue fragility. *J Clin Invest* 2009; **119**:1784–93.
- Reis A, Hennies HC, Langbein L et al. Keratin 9 gene mutations in epidermolytic palmoplantar keratoderma (EPPK). *Nat Genet* 1994; **6**:174–9.
- Langbein L, Heid HW, Moll I, Franke WW. Molecular characterization of the body site-specific human epidermal cytokeratin 9: cDNA cloning, amino acid sequence, and tissue specificity of gene expression. *Differentiation* 1993; **55**:57–71.
- Kim D, Hossain MZ, Nieves A et al. To control site-specific skin gene expression, autocrine mimics paracrine canonical Wnt signaling and is activated ectopically in skin disease. *Am J Pathol* 2016; **186**:1140–50.
- Leachman SA, Hickerson RP, Schwartz ME et al. First-in-human mutation-targeted siRNA phase Ib trial of an inherited skin disorder. *Mol Ther* 2010; **18**:442–6.
- Cao YA, Hickerson RP, Seegmiller BL et al. Gene expression profiling in pachyonychia congenita skin. *J Dermatol Sci* 2015; **77**:156–65.
- Goldberg I, Fruchter D, Meilick A et al. Best treatment practices for pachyonychia congenita. *J Eur Acad Dermatol Venereol* 2014; **28**:279–85.
- Moll R, Franke WW, Schiller DL et al. The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. *Cell* 1982; **31**:11–24.
- Hesse M, Magin TM, Weber K. Genes for intermediate filament proteins and the draft sequence of the human genome: novel keratin genes and a surprisingly high number of pseudogenes related to keratin genes 8 and 18. *J Cell Sci* 2001; **114**:2569–75.
- Schweizer J, Bowden PE, Coulombe PA et al. New consensus nomenclature for mammalian keratins. *J Cell Biol* 2006; **174**:169–74.
- Tyner AL, Fuchs E. Evidence for posttranscriptional regulation of the keratins expressed during hyperproliferation and malignant transformation in human epidermis. *J Cell Biol* 1986; **103**:1945–55.
- Takahashi K, Yan B, Yamanishi K et al. The two functional keratin 6 genes of mouse are differentially regulated and evolved independently from their human orthologs. *Genomics* 1998; **53**:170–83.
- Takahashi K, Paladini RD, Coulombe PA. Cloning and characterization of multiple human genes and cDNAs encoding highly related type II keratin 6 isoforms. *J Biol Chem* 1995; **270**:18581–92.
- Rosenberg M, RayChaudhury A, Shows TB et al. A group of type I keratin genes on human chromosome 17: characterization and expression. *Mol Cell Biol* 1988; **8**:722–36.
- Bernot KM, Coulombe PA, McGowan KM. Keratin 16 expression defines a subset of epithelial cells during skin morphogenesis and the hair cycle. *J Invest Dermatol* 2002; **119**:1137–49.
- McGowan KM, Coulombe PA. Onset of keratin 17 expression coincides with the definition of major epithelial lineages during skin development. *J Cell Biol* 1998; **143**:469–86.
- Troyanovsky SM, Leube RE, Franke WW. Characterization of the human gene encoding cytokeratin 17 and its expression pattern. *Eur J Cell Biol* 1992; **59**:127–37.
- Chen J, Roop DR. Mouse models in preclinical studies for pachyonychia congenita. *J Invest Dermatol Symp Proc* 2005; **10**:37–46.
- Ziemann AG, Poll BG, Ma J, Coulombe PA. Altered keratinocyte differentiation is an early driver of keratin mutation-based palmoplantar keratoderma. *Hum Mol Genet* 2019; **28**:2255–70.
- Fu DJ, Thomson C, Lunny DP et al. Keratin 9 is required for the structural integrity and terminal differentiation of the palmoplantar epidermis. *J Invest Dermatol* 2014; **134**:754–63.
- Kerns ML, Hakim JM, Lu RG et al. Oxidative stress and dysfunctional NRF2 underlie pachyonychia congenita phenotypes. *J Clin Invest* 2016; **126**:2356–66.



- 38 Lessard JC, Coulombe PA. Keratin 16-null mice develop palmo-plantar keratoderma, a hallmark feature of pachyonychia congenita and related disorders. *J Invest Dermatol* 2012; **132**:1384–91.
- 39 Lessard JC, Pina-Paz S, Rotty JD et al. Keratin 16 regulates innate immunity in response to epidermal barrier breach. *Proc Natl Acad Sci U S A* 2013; **110**:19537–42.
- 40 Quigley DA, To MD, Perez-Losada J et al. Genetic architecture of mouse skin inflammation and tumour susceptibility. *Nature* 2009; **458**:505–8.
- 41 Quigley DA, Kandyba E, Huang P et al. Gene expression architecture of mouse dorsal and tail skin reveals functional differences in inflammation and cancer. *Cell Reports* 2016; **16**:1153–65.
- 42 Covello SP, Smith FJ, Sillevs Smitt JH et al. Keratin 17 mutations cause either steatocystoma multiplex or pachyonychia congenita type 2. *Br J Dermatol* 1998; **139**:475–80.
- 43 Smith FJ, Corden LD, Rugg EL et al. Missense mutations in keratin 17 cause either pachyonychia congenita type 2 or a phenotype resembling steatocystoma multiplex. *J Invest Dermatol* 1997; **108**:220–3.
- 44 Smith FJ, Fisher MP, Healy E et al. Novel keratin 16 mutations and protein expression studies in pachyonychia congenita type 1 and focal palmoplantar keratoderma. *Exp Dermatol* 2000; **9**:170–7.
- 45 Shamsheer MK, Navsaria HA, Stevens HP et al. Novel mutations in keratin 16 gene underly focal non-epidermolytic palmoplantar keratoderma (NEPPK) in two families. *Hum Mol Genet* 1995; **4**:1875–81.
- 46 Smith FJ, Liao H, Cassidy AJ et al. The genetic basis of pachyonychia congenita. *J Invest Dermatol Symp Proc* 2005; **10**:21–30.
- 47 Zieman A, Coulombe PA. The keratin 16 null phenotype is modestly impacted by genetic strain background in mice. *Exp Dermatol* 2018; **27**:672–4.
- 48 McGowan KM, Tong X, Colucci-Guyon E et al. Keratin 17 null mice exhibit age- and strain-dependent alopecia. *Genes Dev* 2002; **16**:1412–22.
- 49 Wong P, Colucci-Guyon E, Takahashi K et al. Introducing a null mutation in the mouse K6alpha and K6beta genes reveals their essential structural role in the oral mucosa. *J Cell Biol* 2000; **150**:921–8.
- 50 Rice RH, Durbin-Johnson BP, Salemi M et al. Proteomic profiling of pachyonychia congenita plantar callus. *J Proteomics* 2017; **165**:132–7.
- 51 Fischer H, Langbein L, Reichelt J et al. Keratins K2 and K10 are essential for the epidermal integrity of plantar skin. *J Dermatol Sci* 2016; **81**:10–16.
- 52 Tong X, Coulombe PA. Keratin 17 modulates hair follicle cycling in a TNFalpha-dependent fashion. *Genes Dev* 2006; **20**:1353–64.
- 53 Depianto D, Kerns ML, Dlugosz AA et al. Keratin 17 promotes epithelial proliferation and tumor growth by polarizing the immune response in skin. *Nat Gen* 2010; **42**:910–14.
- 54 Rotty JD, Coulombe PA. A wound-induced keratin inhibits Src activity during keratinocyte migration and tissue repair. *J Cell Biol* 2012; **197**:381–9.
- 55 Chung BM, Arutyunov A, Ilagan E et al. Regulation of C-X-C chemokine gene expression by keratin 17 and hnRNP K in skin tumor keratinocytes. *J Cell Biol* 2015; **208**:613–27.
- 56 Hobbs RP, Depianto DJ, Jacob JT et al. Keratin-dependent regulation of Aire and gene expression in skin tumor keratinocytes. *Nat Gen* 2015; **47**:933–8.
- 57 McLean WH, Hansen CD, Eliason MJ et al. The phenotypic and molecular genetic features of pachyonychia congenita. *J Invest Dermatol* 2011; **131**:1015–17.
- 58 Wilson NJ, Leachman SA, Hansen CD et al. A large mutational study in pachyonychia congenita. *J Invest Dermatol* 2011; **131**:1018–24.
- 59 Eliason MJ, Leachman SA, Feng BJ et al. A review of the clinical phenotype of 254 patients with genetically confirmed pachyonychia congenita. *J Am Acad Dermatol* 2012; **67**:680–6.
- 60 Clementi M, Cardin de Stefani E, Dei Rossi C et al. Pachyonychia congenita Jackson–Lawler type: a distinct malformation syndrome. *Br J Dermatol* 1986; **114**:367–70.
- 61 Feinstein A, Friedman J, Schewach-Millet M. Pachyonychia congenita. *J Am Acad Dermatol* 1988; **19**:705–11.
- 62 Munro CS, Carter S, Bryce S et al. A gene for pachyonychia congenita is closely linked to the keratin gene cluster on 17q12–q21. *J Med Genet* 1994; **31**:675–8.
- 63 Terrinoni A, Smith FJ, Didona B et al. Novel and recurrent mutations in the genes encoding keratins K6a, K16 and K17 in 13 cases of pachyonychia congenita. *J Invest Dermatol* 2001; **117**:1391–6.
- 64 Duverger O, Carlson JC, Karacz CM et al. Genetic variants in pachyonychia congenita-associated keratins increase susceptibility to tooth decay. *PLoS Genet* 2018; **14**:e1007168.
- 65 Porter RM, Bravo AA, Smith FJD. Management of plantar keratodermas: lessons from pachyonychia congenita. *J Am Podiatr Med Assoc* 2017; **107**:428–35.
- 66 Leachman SA, Hickerson RP, Hull PR et al. Therapeutic siRNAs for dominant genetic skin disorders including pachyonychia congenita. *J Dermatol Sci* 2008; **51**:151–7.
- 67 Hickerson RP, Smith FJ, Reeves RE et al. Single-nucleotide-specific siRNA targeting in a dominant-negative skin model. *J Invest Dermatol* 2008; **128**:594–605.
- 68 Hickerson RP, Flores MA, Leake D et al. Use of self-delivery siRNAs to inhibit gene expression in an organotypic pachyonychia congenita model. *J Invest Dermatol* 2011; **131**:1037–44.
- 69 Kaspar RL, Leachman SA, McLean WH et al. Toward a treatment for pachyonychia congenita: report on the 7th Annual International Pachyonychia Congenita Consortium meeting. *J Invest Dermatol* 2011; **131**:1011–14.
- 70 Hickerson RP, Leake D, Pho LN et al. Rapamycin selectively inhibits expression of an inducible keratin (K6a) in human keratinocytes and improves symptoms in pachyonychia congenita patients. *J Dermatol Sci* 2009; **56**:82–8.
- 71 Teng JMC, Bartholomew FB, Patel V et al. Novel treatment of painful plantar keratoderma in pachyonychia congenita using topical sirolimus. *Clin Exp Dermatol* 2018; **43**:968–71.
- 72 Gruber R, Edlinger M, Kaspar RL et al. An appraisal of oral retinoids in the treatment of pachyonychia congenita. *J Am Acad Dermatol* 2012; **66**:e193–9.
- 73 Zhao Y, Gartner U, Smith FJ et al. Statins downregulate K6a promoter activity: a possible therapeutic avenue for pachyonychia congenita. *J Invest Dermatol* 2011; **131**:1045–52.
- 74 Abdollahimajd F, Rajabi F, Shahidi-Dadras M et al. Pachyonychia congenita: a case report of a successful treatment with rosuvastatin in a patient with a KRT6A mutation. *Br J Dermatol* 2019; **181**:584–6.
- 75 Swartling C, Karlqvist M, Hymnelius K et al. Botulinum toxin in the treatment of sweat-worsened foot problems in patients with epidermolysis bullosa simplex and pachyonychia congenita. *Br J Dermatol* 2010; **163**:1072–6.
- 76 Hu C, Eggler AL, Mesecar AD, van Breemen RB. Modification of keap1 cysteine residues by sulforaphane. *Chem Res Toxicol* 2011; **24**:515–21.
- 77 Kerns ML, Hakim JMC, Zieman A et al. Sexual dimorphism in response to an NRF2 inducer in a model for pachyonychia congenita. *J Invest Dermatol* 2018; **138**:1094–100.
- 78 Zhang Y, Talalay P, Cho CG et al. A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure. *Proc Natl Acad Sci U S A* 1992; **89**:2399–403.

- 79 Kerns ML, DePianto D, Dinkova-Kostova AT et al. Reprogramming of keratin biosynthesis by sulforaphane restores skin integrity in epidermolysis bullosa simplex. *Proc Natl Acad Sci U S A* 2007; **104**:14460–5.
- 80 Kerns ML, Guss L, Fahey J et al. Randomized, split-body, single-blinded clinical trial of topical broccoli sprout extract: assessing the feasibility of its use in keratin-based disorders. *J Am Acad Dermatol* 2017; **76**:449–53.e1.
- 81 Leube RE, Schwarz N. Sex matters: interfering with the oxidative stress response in pachyonychia congenita. *J Invest Dermatol* 2018; **138**:1019–22.
- 82 Takahashi K, Coulombe PA. A transgenic mouse model with an inducible skin blistering disease phenotype. *Proc Natl Acad Sci U S A* 1996; **93**:14776–81.
- 83 Wojcik SM, Imakado S, Seki T et al. Expression of MK6a dominant-negative and C-terminal mutant transgenes in mice has distinct phenotypic consequences in the epidermis and hair follicle. *Differentiation* 1999; **65**:97–112.
- 84 Wojcik SM, Longley MA, Roop DR. Discovery of a novel murine keratin 6 (K6) isoform explains the absence of hair and nail defects in mice deficient for K6a and K6b. *J Cell Biol* 2001; **154**:619–30.
- 85 Wojcik SM, Bundman DS, Roop DR. Delayed wound healing in keratin 6a knockout mice. *Mol Cell Biol* 2000; **20**:5248–55.
- 86 Wong P, Domergue R, Coulombe PA. Overcoming functional redundancy to elicit pachyonychia congenita-like nail lesions in transgenic mice. *Mol Cell Biol* 2005; **25**:197–205.
- 87 Chen J, Jaeger K, Den Z et al. Mice expressing a mutant Krt75 (K6hf) allele develop hair and nail defects resembling pachyonychia congenita. *J Invest Dermatol* 2008; **128**:270–9.
- 88 Garcia M, Larcher F, Hickerson RP et al. Development of skin-humanized mouse models of pachyonychia congenita. *J Invest Dermatol* 2011; **131**:1053–60.
- 89 Kimonis V, DiGiovanna JJ, Yang JM et al. A mutation in the V1 end domain of keratin 1 in non-epidermolytic palmar-plantar keratoderma. *J Invest Dermatol* 1994; **103**:764–9.
- 90 Arin MJ, Longley MA, Kuster W et al. An asparagine to threonine substitution in the 1A domain of keratin 1: a novel mutation that causes epidermolytic hyperkeratosis. *Exp Dermatol* 1999; **8**:124–7.
- 91 Lugassy J, Itin P, Ishida-Yamamoto A et al. Naegeli-Franceschetti-Jadassohn syndrome and dermatopathia pigmentosa reticularis: two allelic ectodermal dysplasias caused by dominant mutations in KRT14. *Am J Hum Genet* 2006; **79**:724–30.
- 92 Armstrong DK, McKenna KE, Purkis PE et al. Haploinsufficiency of desmoplakin causes a striate subtype of palmoplantar keratoderma. *Hum Mol Genet* 1999; **8**:143–8.
- 93 Rickman L, Simrak D, Stevens HP et al. N-terminal deletion in a desmosomal cadherin causes the autosomal dominant skin disease striate palmoplantar keratoderma. *Hum Mol Genet* 1999; **8**:971–6.
- 94 McGrath JA, McMillan JR, Shemanko CS et al. Mutations in the plakophilin 1 gene result in ectodermal dysplasia/skin fragility syndrome. *Nat Genet* 1997; **17**:240–4.
- 95 Whittock NV, Smith FJ, Wan H et al. Frameshift mutation in the V2 domain of human keratin 1 results in striate palmoplantar keratoderma. *J Invest Dermatol* 2002; **118**:838–44.
- 96 McKoy G, Protonotarios N, Crosby A et al. Identification of a deletion in plakoglobin in arrhythmogenic right ventricular cardiomyopathy with palmoplantar keratoderma and woolly hair (Naxos disease). *Lancet* 2000; **355**:2119–24.
- 97 Kubo A, Shiohama A, Sasaki T et al. Mutations in SERPINB7, encoding a member of the serine protease inhibitor superfamily, cause Nagashima-type palmoplantar keratosis. *Am J Hum Genet* 2013; **93**:945–56.
- 98 Maestrini E, Monaco AP, McGrath JA et al. A molecular defect in loricrin, the major component of the cornified cell envelope, underlies Vohwinkel's syndrome. *Nat Genet* 1996; **13**:70–7.
- 99 Lin Z, Chen Q, Lee M et al. Exome sequencing reveals mutations in TRPV3 as a cause of Olmsted syndrome. *Am J Hum Genet* 2012; **90**:558–64.
- 100 Lind L, Lundstrom A, Hofer PA et al. The gene for diffuse palmoplantar keratoderma of the type found in northern Sweden is localized to chromosome 12q11–q13. *Hum Mol Genet* 1994; **3**:1789–93.
- 101 Blaydon DC, Lind LK, Plagnol V et al. Mutations in AQP5, encoding a water-channel protein, cause autosomal-dominant diffuse nonepidermolytic palmoplantar keratoderma. *Am J Hum Genet* 2013; **93**:330–5.
- 102 Richard G, White TW, Smith LE et al. Functional defects of Cx26 resulting from a heterozygous missense mutation in a family with dominant deaf-mutism and palmoplantar keratoderma. *Hum Genet* 1998; **103**:393–9.
- 103 van Steensel MA, Spruijt L, van der Burgt I et al. A 2-bp deletion in the GJA1 gene is associated with oculo-dento-digital dysplasia with palmoplantar keratoderma. *Am J Med Genet A* 2005; **132A**:171–4.
- 104 Gong XQ, Shao Q, Lounsbury CS et al. Functional characterization of a GJA1 frameshift mutation causing oculodentodigital dysplasia and palmoplantar keratoderma. *J Biol Chem* 2006; **281**:31801–11.
- 105 Giehl KA, Eckstein GN, Pasternack SM et al. Nonsense mutations in AAGAB cause punctate palmoplantar keratoderma type Buschke-Fischer-Brauer. *Am J Hum Genet* 2012; **91**:754–9.
- 106 Fischer J, Bouadjar B, Heilig R et al. Mutations in the gene encoding SLURP-1 in Mal de Meleda. *Hum Mol Genet* 2001; **10**:875–80.
- 107 Vanderhooft SL, Francis JS, Holbrook KA et al. Familial pityriasis rubra pilaris. *Arch Dermatol* 1995; **131**:448–53.
- 108 Nagy N, Wedgeworth E, Hamada T et al. Schopf-Schulz-Passarge syndrome resulting from a homozygous nonsense mutation in WNT10A. *J Dermatol Sci* 2010; **58**:220–2.
- 109 Adaimy L, Chouery E, Megarbane H et al. Mutation in WNT10A is associated with an autosomal recessive ectodermal dysplasia: the odonto-onycho-dermal dysplasia. *Am J Hum Genet* 2007; **81**:821–8.
- 110 Blaydon DC, Etheridge SL, Risk JM et al. RHBDF2 mutations are associated with tylosis, a familial esophageal cancer syndrome. *Am J Hum Genet* 2012; **90**:340–6.
- 111 Toomes C, James J, Wood AJ et al. Loss-of-function mutations in the cathepsin C gene result in periodontal disease and palmoplantar keratosis. *Nat Genet* 1999; **23**:421–4.

## Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's website:

**Video S1** Author video.