

Evolution in alternating environments with tunable inter-landscape correlations

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Natural populations are often exposed to temporally varying environments. Evolutionary dynamics in varying environments have been extensively studied, though understanding the effects of varying selection pressures remains challenging. Here we investigate how cycling between a pair of statistically related fitness landscapes affects the evolved fitness of an asexually reproducing population. We construct pairs of fitness landscapes that share global fitness features but are correlated with one another in a tunable way, resulting in landscape pairs with specific correlations. We find that switching between these landscape pairs, depending on the ruggedness of the landscape and the inter-landscape correlation, can either increase or decrease steady-state fitness relative to evolution in single environments. In addition, we show that switching between rugged landscapes often selects for increased fitness in both landscapes, even in situations where the landscapes themselves are anti-correlated. We demonstrate that positively correlated landscapes often possess a shared maximum in both landscapes that allows the population to step through sub-optimal local fitness maxima that often trap single landscape evolution trajectories. Finally, we demonstrate that switching between anti-correlated paired landscapes leads to ergodic-like dynamics where each genotype is populated with nonzero probability, dramatically lowering the steady-state fitness in comparison to single landscape evolution.

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Abstract

Natural populations are often exposed to temporally varying environments. Evolutionary dynamics in varying environments have been extensively studied, though understanding the effects of varying selection pressures remains challenging. Here we investigate how cycling between a pair of statistically related fitness landscapes affects the evolved fitness of an asexually reproducing population. We construct pairs of fitness landscapes that share global fitness features but are correlated with one another in a tunable way, resulting in landscape pairs with specific correlations. We find that switching between these landscape pairs, depending on the ruggedness of the landscape and the inter-landscape correlation, can either increase or decrease steady-state fitness relative to evolution in single environments. In addition, we show that switching between rugged landscapes often selects for increased fitness in both landscapes, even in situations where the landscapes themselves are anti-correlated. We demonstrate that positively correlated landscapes often possess a shared maximum in both landscapes that allows the population to step through sub-optimal local fitness maxima that often trap single landscape evolution trajectories. Finally, we demonstrate that switching between anti-correlated paired landscapes leads to ergodic-like dynamics where each genotype is populated with nonzero probability, dramatically lowering the steady-state fitness in comparison to single landscape evolution.

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I. INTRODUCTION

1 Natural populations experience tremendous environmental diversity, and understand-
2 ing how this spatiotemporal diversity influences evolutionary dynamics is a long-standing
3 challenge. A great deal of work, both theoretical and experimental, has shown that spa-
4 tial (Agarwala and Fisher 2019; Constable and McKane 2014a,b; Farhang-Sardroodi et al.
5 2017; Habets et al. 2006; Hermsen and Hwa 2010; Korona et al. 1994; Lin et al. 2015;
6 Waddell et al. 2010; Whitlock and Gomulkiewicz 2005) and temporal (Acar et al. 2008;
7 Canino-Koning et al. 2019; Cook and Hartl 1974; Cooper and Lenski 2010; Cvijović et al.
8 2015; Gaál et al. 2010; Gillespie and Gues 1978; Gupta et al. 2011; Hartl and Cook 1974;
9 Kashtan et al. 2007; Kussell and Leibler 2005; Lewontin and Cohen 1969; Mustonen and
10 Lässig 2008; Mustonen and Lässig 2009; Patra and Klumpp 2015; Shahrezaei et al. 2008;
11 Skanata and Kussell 2016; Steinberg and Ostermeier 2016; Tan and Gore 2012; Tan et al.
12 2011; de Vos et al. 2015) heterogeneity play an important role in adaptation of asexual
13 communities. For example, temporal or spatial fluctuations may lead to increased fixa-
14 tion probability and adaptation rates (Cvijović et al. 2015; Farhang-Sardroodi et al. 2017;
15 Hermsen and Hwa 2010; Kashtan et al. 2007; Lewontin and Cohen 1969; Mustonen and
16 Lässig 2008; Whitlock and Gomulkiewicz 2005), a phenomenon that is also exploited in ge-
17 netic programming algorithms (O'Neill et al. 2010). In addition, environments that change
18 in systematic ways may promote facilitated variation (Gerhart and Kirschner 2007; Parter
19 et al. 2008), allowing organisms to preferentially harness the beneficial effects of random
20 genetic changes and rapidly adapt to future perturbations. And when phenotypes them-
21 selves fluctuate over time, the frequency of inter-phenotype switching can evolve to match
22 the timescale of environmental fluctuations (Acar et al. 2008; Gupta et al. 2011; Kussell and
23 Leibler 2005; Shahrezaei et al. 2008).

24 It is increasingly clear that these evolutionary dynamics have practical consequences for
25 human health. The rise of drug resistance, which threatens the efficacy of treatments for
26 bacterial infections, cancer, and viruses, is driven—at least in part—by evolutionary adaption
27 occurring in complex, heterogeneous environments. Spatial heterogeneity in drug concen-
28 tration has been shown to accelerate the evolution of resistance (Baym et al. 2016; Fu
29 et al. 2015; Greulich et al. 2012; Hermsen et al. 2012; Moreno-Gamez et al. 2015; Zhang
30 et al. 2011), though adaptation may also be slowed when fitness landscapes (Greulich et al.

31 2012) or drug profiles (De Jong and Wood 2018) are judiciously tuned. Similarly, tempo-
32 ral variations in drug exposure—for example, drug cycling—can slow resistance under some
33 conditions, though hospital-level strategies such as mixing may be more effective at generat-
34 ing the requisite environmental heterogeneity (Bergstrom et al. 2004; Brown and Nathwani
35 2005). Recent studies have also shown the potential of new control strategies that harness
36 so-called *collateral effects* (Barbosa et al. 2018; Dhawan et al. 2017; de Evgrafov et al. 2015;
37 Fuentes-Hernandez et al. 2015; Imamovic et al. 2018; Imamovic and Sommer 2013; Kim
38 et al. 2014; Lazar et al. 2018, 2014, 2013; Maltas et al. 2019; Maltas and Wood 2019; Munck
39 et al. 2014; Nichol et al. 2019; Roemhild et al. 2015, 2018; Yoshida et al. 2017), which occur
40 when resistance to a target drug is accompanied by an increase or decrease in resistance to
41 an unseen stressor. In essence, these strategies force populations to simultaneously adapt to
42 incompatible evolutionary tasks (Hart et al. 2015; Shoval et al. 2012).

43 Evolutionary adaptation is often modeled as a biased random walk on a high-dimensional
44 landscape that links each specific genotype with a particular fitness (Gillespie 1983a,b, 1984).
45 In the simplest scenario, these landscapes represent evolution in the strong selection weak
46 mutation (SSWM) limit, where isogenic populations evolve step-wise as the current geno-
47 type is replaced by that of a fitter descendant. While these idealized models are strictly
48 valid only under certain conditions—for example, SSWM typically holds when mutation rate
49 and effective population size are small—simple models have contributed significantly to our
50 understanding of evolution (Cook and Hartl 1974; Desai and Fisher 2007; Desai et al. 2007;
51 Gerrish and Lenski 1998; Gillespie 1983a, 1984; Hartl and Cook 1974). In the context of fit-
52 ness landscape models, control strategies that exploit collateral effects force the population
53 to adapt to sequences of distinct, but statistically related, landscapes. For example, alter-
54 nating between two drugs that induce mutual collateral sensitivity (adaptation to drug A
55 leads to sensitivity to drug B, and vice versa) corresponds to landscapes with anti-correlated
56 fitness peaks. When environments change in systematic ways—for example, by forcing the
57 population to adapt to modular tasks comprised of related sub-goals—adaptation may select
58 for generalists, genotypes that are fit in a wide range of environments at the cost of subop-
59 timal specialization for any particular task (Parter et al. 2008; Sachdeva et al. 2020; Wang
60 and Dai 2019). Relatively recent theoretical work also shows that conditional effects of
61 evolutionary history can be captured by slowly changing landscapes—*seascapes*—which allow
62 for the incorporation of time-dependent correlations (Agarwala and Fisher 2019; Mustonen

63 and Lässig 2009). In general, however, understanding evolution in correlated landscapes–
64 and in particular, how the choice of that correlation impacts fitness adaptation–remains
65 challenging.

66 In this work, we investigate evolutionary dynamics of asexual populations in rapidly al-
67 ternating environments described by pairs of (potentially rugged) fitness landscapes with
68 tunable inter-landscape correlations (Fig 1). This problem is loosely inspired by adaptation
69 of microbial communities to 2-drug cycles in which each drug induces collateral resistance or
70 sensitivity to the other, though the scenario in question may arise in many different contexts,
71 including evolution in antibodies (Burton et al. 2012) and viruses (Rhee et al. 2010). Our
72 goal is to understand how the interplay between intra-landscape disorder (ruggedness) and
73 inter-landscape fitness correlations impact fitness. By formulating the evolutionary dynam-
74 ics as a simple Markov chain (Durrett and Durrett 1999; Nichol et al. 2015), we are able
75 to efficiently calculate time-dependent genotype distributions and investigate adaptation to
76 ensembles of landscape pairs with various levels of epistasis and fitness correlations–results
77 that would be more difficult to achieve from stochastic simulations alone. We find that rapid
78 switching can either increase or decrease the steady state fitness of the population, depend-
79 ing on both the correlation between landscapes and level of intra-landscape ruggedness (i.e.
80 epistasis). On short timescales, mean fitness is generally highest in static landscapes, but
81 rapid switching between correlated environments can produce fitness gains for sufficiently
82 rugged landscapes on longer timescales. Surprisingly, longer periods of rapid switching can
83 also produce a genotype distribution whose fitness is, on average, larger than that of the
84 ancestor population in both environments, even when the landscapes themselves are anti-
85 correlated. To intuitively understand these results, we visualized genotype distributions
86 and inter-genotype transitions as network diagrams, revealing that rapid switching in highly
87 correlated environments frequently shepherds the population to genotypes that are locally
88 optimal in both landscapes and, in doing so, fosters escape from the locally optimal but
89 globally suboptimal fitness peaks that limit adaptation in static environments. The dynam-
90 ics arise, in part, from the fact that rugged landscape pairs are increasingly likely to exhibit
91 shared maxima as they become more positively correlated, and in turn, for landscapes with
92 positive correlations, the mean fitness of these shared peaks is higher than that of non-
93 shared peaks. By contrast, evolution in anti-correlated landscape pairs sample large regions
94 of genotype space, exhibiting ergodic-like steady-state behavior that results in decreased

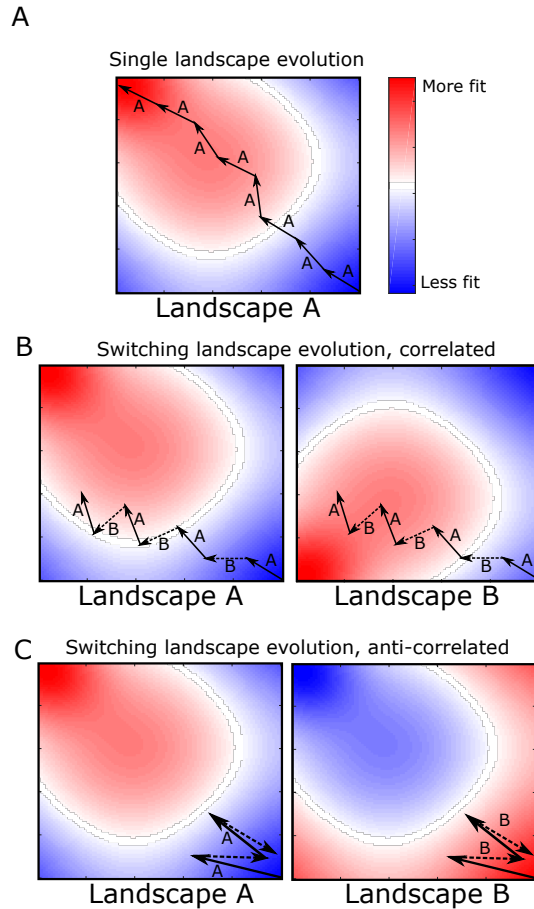


FIG. 1: Adaptation to alternating landscapes may depend on inter-landscape correlations A. Schematic fitness landscape, with fitness varying from less fit (blue) to more fit (red) over the two dimensional genotype space. Starting from a single genotype (lower right hand corner), adaptation follows a biased random walk (arrows) toward local fitness maxima (in this case, in the upper left side of the landscape). B and C. Fitness landscapes A and B are positively (B) or negatively (C) correlated and do not share a global fitness maximum. Adaptation under rapid alternation of landscapes A and B leads to an altered evolutionary trajectory (represented as arrows, with solid arrows indicating steps in A and dashed arrows steps in B). In this example, the final fitness achieved in both correlated (panel B) and anti-correlated (panel C) landscapes is lower than that of static landscape evolution (panel A). Adaptation to anti-correlated landscapes leads to a particularly significant decrease in final fitness, as each step in B effectively reverses the progress made the previous step in A.

95 average fitness.

96 II. RESULTS

97 A. Markov chain model of evolution in alternating landscape pairs with tunable 98 correlations

99 We consider evolution of an asexual haploid genome with N mutational sites. Each
100 mutational site can have one of two alleles (labeled 0 or 1), and a single genotype can
101 therefore be represented by one of the 2^N possible binary sequences of length N . The fitness

102 of each genotype depends on the specific environment in which evolution takes place. We
103 consider two different environments (“A” and “B”), and in each environment, every genotype
104 is assigned a fixed fitness value, which defines the corresponding fitness landscapes (landscape
105 A and landscape B) in each environment. Each fitness landscape is therefore defined on an
106 N -dimensional hypercubic graph, with the nodes corresponding to specific genotypes.

107 To construct the landscape for a given environment, we use a many-peaked “rough Mt.
108 Fuji” landscape (Aita and Husimi 1998; Neidhart et al. 2014; Tan and Gore 2012). Specif-
109 ically, we assume that the fitness of the ancestor genotype (0,0,0...0) is zero and that the
110 fitness f_i associated with a single mutation at mutational site i is drawn from a uniform
111 distribution on the interval $[-1,1]$. Single mutations can therefore lead to increases ($f_i > 0$)
112 or decreases ($f_i < 0$) in fitness. To fully specify the base landscape (i.e. the smooth land-
113 scape in the absence of epistasis), we then assume fitness associated with multiple mutations
114 is additive. Finally, landscape ruggedness is incorporated by adding to the fitness of each
115 genotype j a fixed, random variable ξ_j drawn from a zero-mean normal distribution with
116 variance σ^2 . The variable σ —the amplitude of the noise—determines the level of ruggedness
117 of the landscape, which simulates epistasis (Anderson et al. 2015; Phillips 2008; Ritchie
118 et al. 2001; da Silva et al. 2010; Tsai et al. 2007; Weinreich et al. 2006; Xu et al. 2005). In
119 what follows, we focus on landscapes of size $N = 7$ (128 total genotypes) for computational
120 convenience and limit ourselves primarily to $\sigma = 0$ (smooth landscapes) or $\sigma=1$ (rugged
121 landscapes).

122 Our goal is to investigate evolution in rapidly changing environments that correspond to
123 landscape pairs with correlated fitness peaks. To do so, we generate for each landscape A a
124 “paired” landscape B with similar statistical properties (identical fitness mean and variance)
125 but fitness peaks that are, on average, correlated with those of landscape A in a tunable
126 way. To do so, we represent each landscape A as a vector \bar{A} of length 2^N and use simple
127 matrix algebra to generate a random vector \bar{A}_\perp orthogonal to \bar{A} ; by construction, then, this
128 vector corresponds to a landscape whose fitness values are, on average, uncorrelated with
129 those of landscape A. It is then straightforward to generate a vector \bar{B} , a linear combination
130 of \bar{A} and \bar{A}_\perp , such that the fitness values of landscapes A and B are correlated to a tunable
131 degree $-1 \leq \rho \leq 1$, where ρ is the Pearson correlation coefficient between the two vectors
132 \bar{A} and \bar{B} (see Methods).

133 With the landscapes specified, we then model adaptation in the well-characterized Strong

134 Selection Weak Mutation (SSWM) limit (Gillespie 1983a,b, 1984), which can be formally
135 described by a Markov chain (Durrett and Durrett 1999; Nichol et al. 2015). During each
136 time step, the population transitions with uniform probability to one of the neighboring
137 genotypes with a higher fitness in the current environment. We compare adaptation on a
138 single landscape (single landscape evolution, SLE) with adaptation to rapid alternation of
139 the two correlated landscapes A and B, which we refer to as paired landscape evolution
140 (PLE). We focus here on the limit of rapid environmental switching, where the fitness
141 landscape changes (A-B-A-B...) at each time step. This corresponds loosely to the rapid
142 environmental switching seen in many laboratory experiments (Burch and Chao 1999; Crill
143 et al. 2000; Kim et al. 2014; Lenski 1988).

144 We are primarily interested in comparing the (average) steady-state fitness of populations
145 undergoing SLE to that of populations undergoing PLE. The average fitness, $\bar{F}_X(\bar{p})$, in
146 environment X can be calculated at any time step t using $\bar{F}_X(\bar{p}) = \bar{X} \cdot \bar{p}(t)$, where $\bar{p}(t)$
147 is the vector whose i^{th} component is the probability to be in genotype i at time t and
148 \bar{X} is the landscape vector for environment X . Because the process can be described by a
149 Markov chain, the vector $\bar{p}(t)$ is given by $\bar{p}(t) = T_M \bar{p}(0)$, where the matrix T_M describes
150 the sequence of environments over time (e.g. $T_M = T_A^M$ for M steps in environment A , or
151 $T_M = (T_B T_A)^{M/2}$ for M consecutive A-B cycles, with T_A and T_B the transition matrices
152 corresponding to single steps in environment A and B, respectively). In what follows, we
153 focus primarily on the mean fitness difference between the SLE and PLE adaptation, which
154 is given by $\bar{F}_\Delta^A \equiv \bar{F}_A(\bar{p}_A) - \bar{F}_A(\bar{p}_{AB})$, where \bar{p}_A is the steady state genotype distribution
155 following adaptation to environment A, and \bar{p}_{AB} is the steady state genotype distribution
156 following adaptation to alternating A-B environments. Note that we define this fitness
157 difference, \bar{F}_Δ^A , with respect to landscape \bar{A} (noted by superscript), which allows us to
158 compare adaptation in environment A with adaptation in the alternating A-B environments.
159 In the drug cycling analogy, we are measuring the average fitness in the drug A environment—
160 essentially a measure of resistance to that drug. In all calculations, we consider an ensemble
161 of 1000 landscapes pairs—with each pair sharing the same mean and variance in fitness and
162 the same inter-landscape correlations—and we average the results over this ensemble.

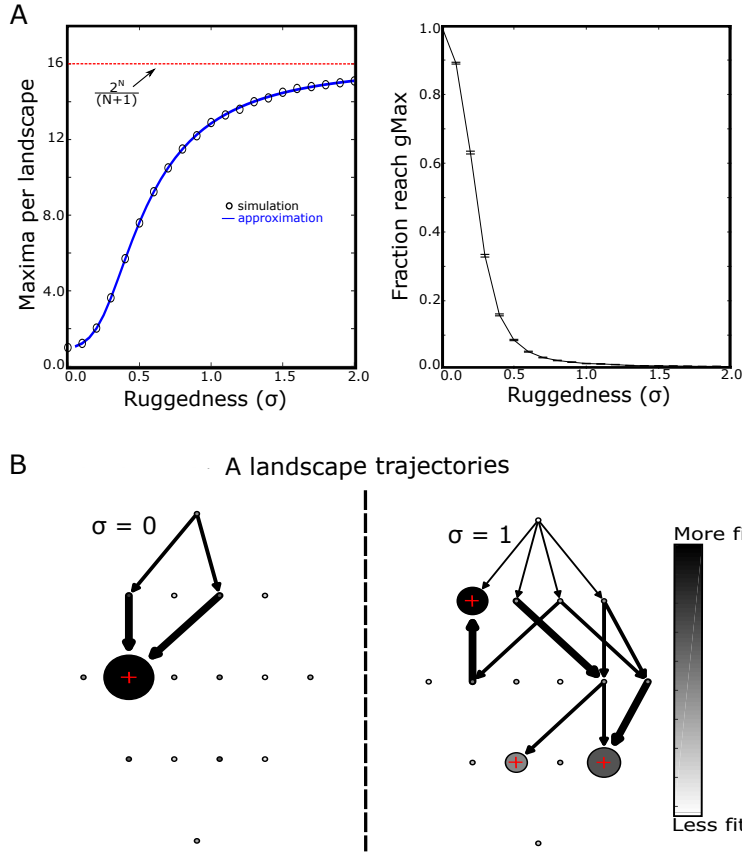


FIG. 2: **Rugged landscapes trap populations in non-optimal fitness maxima** A. Left panel: average number of local fitness maxima per landscape as a function of increasing ruggedness (epistasis, σ). Circles are estimates from simulations, solid curve is semi-analytical approximation (see SI), and dotted red line is the theoretical maximum ($2^N/(N+1) = 16$). Right panel: fraction of adapted populations that reach the global fitness maximum value as a function of ruggedness. Error bars are \pm standard error of the mean in the ensemble of landscapes. B. Sample adaptive trajectories for small landscapes ($N = 4$) and $\sigma = 0$ (left) or $\sigma = 1$ (right). Each circle represents a genotype, with the ancestral genotype at the top. The shading of the circle represents the relative fitness of that genotype (ranging from less fit, white, to more fit, black) and the size of the circle indicates occupation probability in the steady state. Red + symbols mark genotypes corresponding to local fitness maxima. Arrows represent transitions between genotypes that occur with nonzero probability given that adaptation begins in the ancestral genotype. The width of the arrow represents the magnitude of the transition probability.

B. Adaptation in rugged landscapes frequently ends in local, sub-optimal fitness maxima

While adaptation to static, rugged landscapes is well-understood, we first briefly discuss the effects of landscape ruggedness in the context of the current model. In static landscapes, steady state is reached when the genotype corresponds to a local fitness maximum. In the case of smooth, purely additive landscapes ($\sigma = 0$), there is a single fitness peak that corresponds to the global maximum, which we call gMax. However, as the landscape becomes more rugged ($\sigma > 0$), the average number of local maxima increases. For small $\sigma \ll 1$, the average number of local maxima is $N_{max} \approx 1 + 1/2N(N+1)\sigma^2$, while for large σ it

172 approaches the theoretical maximum of $2^N/(N+1)$ (Fig 2A); in the SI, we provide a semi-
173 analytical approximation for intermediate values of σ . In turn, the fraction of adaptation
174 trajectories that reach the global maximum decreases, reflecting the propensity of rugged
175 landscapes to trap evolution in globally sub-optimal genotypes. To visualize these results,
176 we represented the steady state genotype distributions and inter-genotype transitions as a
177 network diagram (Fig 2B), with each node (circle) representing a genotype. The shading
178 of each circle represents the relative fitness of that genotype (ranging from less fit, white,
179 to more fit, black) and the size of the circle indicates occupation probability in the steady
180 state. Arrows connecting different genotypes indicate nonzero transition probabilities, with
181 the thickness of the arrow corresponding to its magnitude. We show only those transitions
182 that can occur when adaptation starts in the ancestor genotype (top of diagram). In the case
183 of evolution on a smooth landscape ($\sigma = 0$, Fig 2B, left panel), all trajectories lead to the
184 single global maximum (indicated by red “+”). However, in the rugged landscape ($\sigma = 1$,
185 Fig 2B, right panel), there is a nonzero probability of settling in each of three local maxima,
186 and the population frequently ends in a non-optimal genotype. Increasing ruggedness would
187 therefore be expected to lower the average fitness achieved in an ensemble of landscapes.

188 C. Switching between positively correlated landscapes can produce higher average 189 fitness than adaptation to a static environment

190 Next, we set out to compare adaptation to landscape A with adaptation to alternating
191 landscapes (A, B) with a tunable level of correlation, ρ , in the absence of epistasis ($\sigma = 0$,
192 Fig 3A, blue). On these smooth landscapes, the fitness is single-peaked (Tan and Gore
193 2012), and in the absence of switching, the population always reaches this global maximum.
194 In alternating environments, adaptation approaches the same average fitness as in static
195 environments (i.e. $\bar{F}_\Delta^A \approx 0$)—implying that it finds the global fitness maximum—for all but
196 the most negatively correlated landscapes ($\rho < -0.85$), where switching leads to steep
197 decreases in fitness. By contrast, when landscapes are rugged ($\sigma = 1$), we find a range of
198 correlations for which switching (PLE) increases the mean fitness ($\bar{F}_\Delta^A < 0$, Fig 3A, orange).
199 Furthermore, as ruggedness increases, the range of correlations leading to increased fitness
200 grows (Fig 3B).

201 **D. Fitness can be maximally increased in either static or alternating environments**
202 **depending on the timescale**

203 We find that adaptation to static environments typically occurs on a faster timescale
204 than adaptation to alternating environments (Fig S3). As a result, the protocol yielding
205 the highest average fitness may differ depending on the timescale over which the compar-
206 ison is made. For example, on short timescales (5 total evolutionary steps; Fig 3C, blue),
207 adaptation to static environments always leads to greater fitness gain, regardless of the cor-
208 relation between landscapes. On moderate (11 total evolutionary steps; Fig 3C, red) to long
209 (Fig 3C, black) timescales, however, we again see a range of positive correlations for which
210 switching improves fitness—first only for highly correlated landscapes, and then eventually
211 for a wider range of positively correlated landscapes. This result indicates that the optimal
212 protocol for increasing fitness may depend on the chosen timescale; notably, recent results
213 indicate that these timescales can also be tuned to maintain generalists successful in different
214 environments (Sachdeva et al. 2020).

215 **E. Adaptation to alternating landscapes can lead to increased mean fitness even**
216 **in anti-correlated landscapes**

217 While we have so far focused on mean fitness defined in landscape A, either due to static
218 ($\bar{F}_A(\bar{p}_A)$) or alternating ($\bar{F}_A(\bar{p}_A)$) environments, we also asked how fitness in landscape B was
219 modulated during adaptation. If adaptation occurs to a static landscape (A), the results are
220 simple: the genotype adapted to A will on average exhibit increased (decreased) fitness in B
221 when landscape B is positively (negatively) correlated with A. This scenario is reminiscent
222 of collateral effects between different drugs, where increased resistance to one drug may
223 be associated with either increased (cross resistance) or decreased (collateral sensitivity)
224 resistance to a different (unseen) drug. In the case of alternating environments, however,
225 the outcome is less clear *a priori*.

226 For smooth landscapes ($\sigma = 0$), we find that adaptation to the alternating landscapes
227 leads to increased fitness in B ($\bar{F}_B(\bar{p}_{AB}) > 0$) when the landscapes are positively corre-
228 lated and decreased fitness when they are negatively correlated (Fig 3D). Nonzero epista-
229 sis shifts the boundary separating increased and decreased fitness toward negative corre-

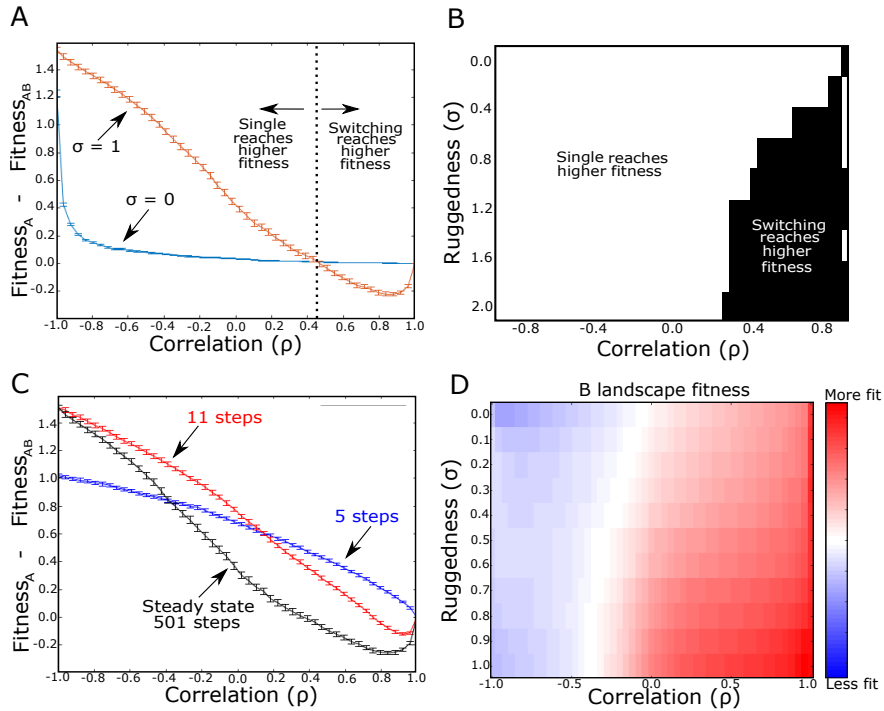


FIG. 3: Modulated fitness in alternating landscapes depends on intra-landscape ruggedness and inter-landscape correlations. A. Difference in average fitness (at steady state) between populations adapted to a single static landscape (A) or rapidly alternating landscape pairs (A-B) as a function of correlation between landscapes A and B. Average fitness is defined as the mean fitness of the steady state genotype distribution (which arises following adaptation to either static or switching protocols) measured in landscape A. Blue curve: $\sigma = 0$ (no epistasis; smooth); Orange curve: $\sigma = 1$ (orange; rugged). Dotted vertical line (corresponding to zero fitness difference) indicates critical value of correlation; above this critical value, switching between rugged landscape pairs ($\sigma = 1$) leads to larger fitness gains than evolution in a static landscape. B. Heatmap showing regions of parameter space (ruggedness σ , inter-landscape correlation) where switching leads to higher (black) or lower (white) fitness than evolution in a static landscape. C. Identical to panel A, but curves are shown for 5 (blue), 11 (red) and 501 (black) total evolutionary steps. $\sigma=1$ for all curves. D. Collateral fitness change, ranging from blue (less fit) to red (more fit), for populations adapted to alternating environments A and B as a function of ruggedness (σ) and inter-landscape correlation. Collateral fitness change is defined as the increase in average fitness in landscape B (relative to ancestor) associated with the steady state genotype distribution arising from adaptation to alternating A-B landscapes. $N = 7$ in all panels, but see also Figure S1. Error bars in panels A and C are \pm standard error of the mean in the ensemble of landscapes.

230 lations. As a result, switching leads to increased fitness in both landscapes for a wider
 231 range of correlations—even, counterintuitively, in cases where the landscapes are (weakly)
 232 anti-correlated. In the context of drug cycling, this result suggests that cross resistance is
 233 likely to arise following repeated cycling of two drugs, even when their fitness landscapes
 234 are anti-correlated (i.e. drugs induce mutual collateral sensitivity).

235 **F. Alternating between highly-correlated landscapes promotes escape from local**
236 **fitness optima**

237 To understand why switching between highly correlated landscapes can increase fitness
238 relative to single landscape adaptation, we again represented adaptation on a simple ($N =$
239 4) network representing a particular pair of fitness landscapes (Fig S7). The choice of
240 $N=4$ allows for a simpler visual interpretation of the results, and the relevant dynamics are
241 qualitatively similar for a broad range of landscape sizes and sigma values (Fig S1, Fig S2).
242 The landscape for environment A is characterized by multiple local maxima (Fig S7A, left
243 panel), and in this example, the adaptation dynamics starting from the ancestral genotype
244 are relatively simple, with only two paths possible (Fig S7A, right panel). With equal
245 probability, the trajectory ends in one of two possible states, one of which is the global
246 maximum.

247 If we now introduce rapid alternation with a second, positively correlated landscape
248 ($\rho = 0.8$), the dynamics are much richer (Fig S7B). In this example, there is a single
249 shared (local) maximum between the two landscapes (marked with red “+”), and adaptation
250 to alternating environments eventually shepherds all trajectories to this shared maximum,
251 which also happens to be the global maximum. As a result, alternating between landscapes
252 leads to (on average) greater fitness increases than that achieved in static landscapes, where
253 trajectories are split between local and global maxima. Intuitively, this example suggests
254 that one advantage of rapid switching is that it dislodges trajectories from suboptimal local
255 maxima—that is, switching between highly (but not perfectly) correlated landscapes provides
256 a source of fluctuations that maximize the likelihood of finding globally optimal genotypes.
257 This result is reminiscent of the observed “ratchet-like” mechanism of the *lac* operon in
258 *Escherichia coli* (de Vos et al. 2015).

259 **G. Evolution in highly anti-correlated paired landscapes broadly samples geno-**
260 **type space resulting in reduced average fitness**

261 We now return to dynamics in strongly anti-correlated landscapes, where shared max-
262 ima may be less likely to occur. To intuitively understand dynamics in this regime, we
263 visualized the fitness landscape and evolutionary trajectories for a pair of simple ($N = 4$)

264 anticorrelated landscapes (Fig S8). In this example, adaptation to the static landscape leads
265 to considerably higher fitness than adaptation to alternating landscapes. Interestingly, we
266 see that the genotype distribution remains broad, even for long times. In fact, the only
267 genotypes that remain unoccupied ($p_i = 0$) are those five that correspond to local minima
268 in the A landscape. Including an additional step in landscape B leads to a similarly broad
269 distribution, now with unoccupied genotypes corresponding to local minima of landscape B
270 (Figure S5). In contrast to adaptation to single landscapes or alternating, positively corre-
271 lated landscapes, the steady state distribution is not dominated by local fitness maxima but
272 instead corresponds to broad genotype distribution and an associated decrease in average
273 fitness.

274 **H. Adaptation to alternating landscapes is frequently dominated by presence or** 275 **absence of shared fitness maxima**

276 We hypothesized that the increased fitness in alternating landscapes is closely linked
277 to the expected number of shared maxima between paired landscapes. To probe this hy-
278 pothesis, we first estimated two quantities: 1) the fraction of local maxima that are shared
279 between landscapes (specifically, the fraction of A-landscape maxima that also correspond
280 to maxima in the paired landscape B) and 2) the fraction of landscape pairs that share at
281 least one maxima. We estimated these quantities by simulating ensembles of landscapes
282 and also developed semi-analytical approximations that reduce to simple evaluations of the
283 cumulative distribution function (CDF) of a multivariate normal variable (SI). As intuition
284 suggests, the fraction of shared maxima increases with correlation, both for smooth and
285 rugged landscapes (Fig 4A). In addition, we estimated the fraction of landscape pairs in the
286 entire ensemble that share at least one shared maximum (Fig 4B). Again we find that this
287 quantity increases with correlation, but it does so much more rapidly for rugged landscapes.
288 For smooth landscapes, the latter fraction increases gradually—and the curve is identical to
289 that in (Fig 4A), a result of the fact that smooth landscapes have only a single (global)
290 maximum.

291 To link these architectural properties of the landscapes with dynamics, we calculated
292 adaptation trajectories under rapid switching of all paired landscapes in these ensembles
293 (Fig 4C). For both smooth landscapes and negatively correlated rugged landscapes, the

294 fraction of trajectories ending in a shared maximum closely mirrors the fraction of landscapes
295 pairs that share a maximum. This correspondence suggests that under these conditions,
296 when landscapes share a local maximum, the adapting system is likely to settle there. On
297 the other hand, for positively correlated rugged landscapes, the likelihood of finding a shared
298 maximum is relatively insensitive to correlation until ρ becomes quite large ($> .80$), when
299 it rapidly increases (Fig 4C).

300 To further clarify the connection between fitness and shared maxima, we divided the local
301 fitness maxima from landscape A into one of two categories: those that also correspond to
302 a local maximum in landscape B, and those that do not. We found, somewhat counter-
303 intuitively, that the mean fitness differs for the two categories (Fig 4D). For negatively
304 correlated landscape pairs, the fitness of shared maxima is less than that of non-shared
305 maxima. By contrast, shared maxima in highly (positively) correlated landscapes have a
306 higher mean fitness than non-shared maxima. In addition, there is a range of positive ρ where
307 the fitness of shared maxima is also greater than the average fitness of maxima in a single A
308 landscape (which corresponds to the $\rho \rightarrow 1$ limit of the curve), offering an explanation for the
309 fitness increase induced by alternating between highly correlated landscapes. Specifically,
310 evolutionary trajectories typically settle into a single local maxima for adaptation to both
311 static and positively correlated, alternating environments; however, for a range of highly
312 (but not perfectly) correlated landscape pairs, the mean fitness of those shared maxima is
313 greater than the mean fitness of local maxima in a single A landscape.

314 **I. Steady-state genotype distributions transition from narrow to broad as corre-** 315 **lation is decreased**

316 To further characterize steady state dynamics, we calculated the entropy of the steady
317 state genotype distribution, defined as $S(p)/S_{max} \equiv -(\sum_i p_i \ln p_i)/S_{max}$, where p_i is the
318 steady state probability of being in genotype i and the expression is normalized by $S_{max} =$
319 $N \ln(2)$, the entropy of a uniform distribution (Fig 4E)—that is, a state where every genotype
320 is equally probable. To capture dynamics associated with potential non-fixed point behavior,
321 for this analysis we slightly modify the definition of steady state to be $p_i = (p_A + p_B)/2$, where
322 p_A is the steady state fitness following a step in landscape A (the previously used definition)
323 and p_B the fitness in the same steady state regime but following a step in landscape B (in

words, we average over a full A-B cycle in the steady state). We find that as correlation (ρ) increases, the entropy of the system decreases, indicating that the dynamics are confined to an ever smaller set of genotypes—presumably those corresponding to shared maxima. Indeed, if we restrict the ensemble to only those landscape pairs that share a maximum, the entropy of the distribution is unchanged for highly correlated landscapes, suggesting that shared maxima dominate the steady state dynamics. By contrast, when landscape pairs are anticorrelated, restricting the ensemble to pairs *without* shared maxima closely approximates the results of the full ensemble, suggesting that dynamics in this regime are dominated by qualitatively different behavior. Consistent with changes in the entropy of the genotype distribution, we also find that correlation dramatically changes the fraction of genotype space occupied (with nonzero probability) in the steady state (Fig 4F). For highly correlated landscapes, only a small fraction of the total genotype space is occupied. By contrast, highly anti-correlated landscapes produce steady state distributions wherein all states are occupied with non-zero probability, suggesting ergodic-like behavior, consistent with the example in Fig S8. The fact that relative entropy remains less than 1 in this regime does indicate, however, that the distribution is not fully uniform.

Finally, in Fig 4G, we plot the difference in steady state fitness achieved in static vs alternating environments for both the full landscape pair ensemble (black) and for a reduced ensemble consisting only of landscapes with shared maxima (red). We find that the curves are nearly identical over a wide range of correlations $\sigma > -0.4$. Similarly, when correlation is strongly anticorrelated, fitness differences are similar between the full ensemble and the reduced ensemble with no shared maxima (Fig 4H). Taken together, these results provide evidence that adaptation in this model is frequently dominated by the presence or absence of shared fitness maxima, which in turn depends on the correlation between landscapes and landscape ruggedness.

J. Clonal interference and slow switching reduce the impact of alternating between anticorrelated landscapes

Our idealized model neglects clonal interference, which could potentially impact the evolutionary dynamics (Gerrish and Lenski 1998). To investigate its potential impacts, we implement a phenomenological model previously used to estimate the effects of clonal inter-

354 ference (Tan and Gore 2012). Briefly, in the absence of clonal interference, the population
355 can be treated as a *random walker* that steps to any nearby more fit genotype with equal
356 probability. In order to simulate clonal interference, the population can be treated as a
357 *greedy walker*, where the fixation probability of advantageous mutations is assumed to be
358 proportional to s^x , where s is the selective advantage and x is the phenomenological pa-
359 rameter. As x increases, the probability of stepping to more fit mutants continues to grow,
360 simulating larger population sizes.

361 We find that small and moderate levels of clonal interference ($x \sim 5$) reduce the observed
362 fitness differences between static and alternating protocols but lead to similar qualitative
363 dynamics (Fig 5). However, as the population size gets large ($x > 5$) the fitness difference,
364 genetic diversity and collateral effects due to switching become quite small; the impact of
365 clonal interference is particularly large when landscape pairs are strongly anticorrelated.

366 We next asked how the period of switching impacts the evolutionary dynamics. To do
367 so, we varied the period of the switching (specifically, the number of consecutive steps taken
368 in one landscape before switching) over approximately an order of magnitude (Fig 6). We
369 find that small changes in the period—for example, doubling it from 1 step to 2—reduces
370 the observed fitness differences and the normalized entropy, particularly for anticorrelated
371 landscapes, but does not dramatically impact the likelihood of ending in a shared maximum
372 or the collateral fitness changes (Fig 6).

373 III. DISCUSSION

374 Our results indicate that both intra-landscape disorder (ruggedness) and inter-landscape
375 fitness correlations impact fitness in rapidly alternating fitness landscapes. Compared with
376 static adaptation, rapid switching can lead to increased or decreased fitness, depending on
377 both the correlation between landscapes and level of intra-landscape ruggedness (i.e. epista-
378 sis). Perhaps most strikingly, switching between highly, but not perfectly, correlated rugged
379 landscapes can increase fitness by promoting escape from local fitness maxima, increasing
380 the likelihood of finding global fitness optima. Furthermore, rapid switching can also pro-
381 duce a genotype distribution whose fitness is, on average, larger than that of the ancestor
382 population in both environments, even when the landscapes themselves are anti-correlated.
383 Adaptation dynamics are often dominated by the presence or absence of shared maxima

384 between landscapes. Rugged landscape pairs are increasingly likely to exhibit shared max-
385 ima as they become more positively correlated, and in turn, for landscapes with positive
386 correlations, the mean fitness of these shared peaks is higher than that of non-shared peaks.
387 By contrast, evolution in anti-correlated landscape pairs samples large regions of genotype
388 space, exhibiting ergodic-like steady-state behavior that results in decreased average fitness.
389 A simple phenomenological model suggests these results are robust to competition due to
390 small and moderate clonal interference, however they disappear as population sizes grow
391 excessively large. In addition, while prolonging the period of switching can alter the dy-
392 namics in anti-correlated landscape evolution, the fitness advantage conferred by alternating
393 evolution in correlated landscape pairs is robust to the period of switching.

394 While our results are loosely inspired by antibiotic cycling, the model is highly idealized
395 and certainly cannot make predictions that apply directly to clinical scenarios. At the same
396 time, the simplicity and relative generality of the model means that it may be relevant for
397 understanding the qualitative behavior of a wide range of systems, including evolution in
398 antibodies (Burton et al. 2012), viruses (Rhee et al. 2010), and bacteria, where ratchet-like
399 mechanisms for rapid adaptation have been observed experimentally (de Vos et al. 2015).
400 Our model relies on the Strong Selection Weak Mutation (SSWM) limit and neglects po-
401 tentially relevant dynamics that could arise due to horizontal gene transfer or population
402 heterogeneity, which could potentially accelerate adaptation, particularly when switching
403 between anticorrelated landscapes. While we also investigated an adapted model that ac-
404 counts for clonal interference (Tan and Gore 2012), the model still assumes a homogeneous
405 population, thus ignoring the genetic diversity necessary of clonal interference, and it ne-
406 glects the possibility for deleterious or multiple simultaneous mutations to fix. In addition,
407 we focus on small (typically $N = 7$) landscapes for tractability, and dynamics could differ
408 for much larger landscapes.

409 It is important to note that we focus on paired landscapes characterized by a prescribed
410 "global" correlation coefficient, but we do not investigate heterogeneity in the correlations
411 at the single node level. In addition, the paired landscapes in our ensembles are constructed
412 to share certain global features—like mean fitness—and are related by a prescribed inter-
413 landscape correlation, but they are not statistically identical. For example, the average
414 number of local maxima can differ between landscape A and B, leading to different levels
415 of evolved fitness for each landscape individually (Figure S6). This indicates that land-

416 scapes A and B have effectively different levels of epistasis, depending on the desired value
417 of ρ , though these differences are most pronounced when A landscapes are very smooth
418 ($\sigma \approx 0$). These differences do not seem to be appreciably impacting fitness dynamics, as
419 removing them by choosing a reduced ensemble (keeping only the B landscapes the exhibit
420 similar fitness gains as A under static adaptation) does not appreciably modify the results
421 (Figure S6). Nevertheless, it is possible that some of these results are specific to the exact
422 manner in which correlated landscapes were produced; for example, in Figure 4D, the mean
423 fitness for shared maxima equals that for unshared maxima at a small but nonzero value of ρ
424 (rather than at $\rho = 0$), a counter intuitive result that may not hold when paired landscapes
425 are generated by different algorithms. Indeed, it may be interesting to investigate switching
426 dynamics using landscapes with different types of statistical similarities—for example, those
427 that differ only in higher-order moments, or those that fully decouple landscape ruggedness
428 and correlation (Wang and Dai 2019). In fact, the results presented here are complementary
429 to recent findings showing that environmental switching can enhance the basin of attraction
430 for generalists, which are genotypes that are fit in multiple environments (Sachdeva et al.
431 2020; Wang and Dai 2019). While the focus of the work is different—and the timescale of
432 environmental switching and the statistical relationships between landscape pairs differ in
433 their model—our results similarly highlight the importance of shared landscape maxima in
434 determining adaptation dynamics. Future work may aim to further elucidate the evolution-
435 ary impacts of varying timescale, ordering, and temporal correlations in landscape dynamics.
436 In the long run, we hope results from idealized models like these offer increased conceptual
437 clarity to complement the rapidly evolving experimental approaches for mapping landscape
438 dynamics in living organisms.

439 IV. METHODS

440 A. Construction of the landscapes

441 We use the “rough Mt. Fiji” landscape model (Aita and Husimi 1998; Neidhart et al.
442 2014; Tan and Gore 2012) where each genotype is represented by a binary sequences of length
443 N . The fitness of the ancestor genotype (0,0,0...0) is set to zero and the fitness f_i associated
444 with a single mutation at mutational site i is drawn from a uniform distribution on the

445 interval $[-1,1]$. The fitness associated with multiple mutations is additive, and landscape
 446 ruggedness is incorporated by adding to the fitness of each genotype j a fixed, random
 447 variable ξ_j drawn from a zero-mean normal distribution with variance σ^2 .

448 To create paired fitness landscapes, we represent each landscape A as a vector \bar{A} of
 449 length 2^N , which we center and rescale to achieve a zero mean, unit variance vector. Then,
 450 we generate a Gaussian random vector \bar{A}_\perp (also with zero mean and unit variance) and
 451 subtract from \bar{A}_\perp its projection onto \bar{A} , making \bar{A}_\perp orthogonal to \bar{A} ; by construction, this
 452 vector corresponds to a landscape whose fitness values are, on average, uncorrelated with
 453 those of landscape A. It is then straightforward to generate a vector \bar{B} , a linear combination
 454 of \bar{A} and \bar{A}_\perp , such that the fitness values of landscapes A and B are correlated to a tunable
 455 degree $-1 \leq \rho \leq 1$, where ρ is the Pearson correlation coefficient between the two vectors
 456 \bar{A} and \bar{B} . At the end of this procedure, we rescale \bar{A} and \bar{B} so that both have mean and
 457 variance equal to that of the original A landscape.

458 B. Evolution on the landscapes

The SSWM assumption allows the evolutionary trajectories to be modeled as a Markov
 chain (Durrett and Durrett 1999; Nichol et al. 2015). We follow the “random move SSWM
 model”, which says that the probability of transitioning between adjacent genotypes $i \rightarrow j$
 is given by $T_{ij} = 1/m$, with m the total number of i -adjacent genotypes with fitness greater
 than that of i in the given environment. Each environment (A or B) has its own transition
 matrix, which we designate as T_A and T_B . Evolution in environment A is then given by

$$\bar{p}(t) = (T_A)^t \bar{p}(0) \quad (1)$$

with $\bar{p}(t)$ the vector whose i^{th} component is the probability to be in genotype i at time
 step t . We refer to the steady state ($t \rightarrow \infty$) limit of this process as \bar{p}_A . Similarly, we can
 describe rapidly alternating landscapes (A-B-A-B...) with

$$\bar{p}(t') = (T_B T_A)^{t'/2} \bar{p}(0) \quad (2)$$

459 with $t' \equiv 2t$ an even time step. We refer to the steady state ($t \rightarrow \infty$) limit of this process
 460 as \bar{p}_{AB} . In practice, we define steady state using the condition $\|(\bar{p}(2t+1) - \bar{p}(2t-1))\| <$

461 $\epsilon = 0.001$. In words, we require the change in \bar{p} between consecutive steps in environment
462 A to be sufficiently small. To facilitate comparison with static evolution in landscape A , we
463 always end the process after a step in landscape A , meaning there are always an odd number
464 of steps. Ending instead in landscape B results in qualitatively similar behavior, though the
465 fitness is often shifted, indicating that a single step in A or B —even in steady state—can lead
466 to significant changes in fitness [S4](#).

467 Acar, M., J. T. Mettetal, and A. van Oudenaarden, 2008. Stochastic switching as a survival
468 strategy in fluctuating environments. *Nature Genetics* 40:471 EP –. URL [https://doi.org/10.](https://doi.org/10.1038/ng.110)
469 [1038/ng.110](https://doi.org/10.1038/ng.110).

470 Agarwala, A. and D. S. Fisher, 2019. Adaptive walks on high-dimensional fitness landscapes and
471 seascapes with distance-dependent statistics. *Theoretical Population Biology* 130:13–49.

472 Aita, T. and Y. Husimi, 1998. Adaptive walks by the fittest among finite random mutants on
473 a mt. fuji-type fitness landscape. *Journal of Theoretical Biology* 193:383 – 405. URL [http:](http://www.sciencedirect.com/science/article/pii/S0022519398907093)
474 [//www.sciencedirect.com/science/article/pii/S0022519398907093](http://www.sciencedirect.com/science/article/pii/S0022519398907093).

475 Anderson, D. W., A. N. McKeown, and J. W. Thornton, 2015. Intermolecular epistasis shaped the
476 function and evolution of an ancient transcription factor and its dna binding sites. *eLife* 4:e07864.
477 URL <https://doi.org/10.7554/eLife.07864>.

478 Barbosa, C., R. Beardmore, H. Schulenburg, and G. Jansen, 2018. Antibiotic combination efficacy
479 (ace) networks for a pseudomonas aeruginosa model. *PLoS biology* 16:e2004356.

480 Baym, M., T. D. Lieberman, E. D. Kelsic, R. Chait, R. Gross, I. Yelin, and R. Kishony, 2016.
481 Spatiotemporal microbial evolution on antibiotic landscapes. *Science* 353:1147–1151.

482 Bergstrom, C. T., M. Lo, and M. Lipsitch, 2004. Ecological theory suggests that antimicrobial
483 cycling will not reduce antimicrobial resistance in hospitals. *Proc. Natl. Acad. Sci. USA* 101:13285–
484 13290.

485 Brown, E. M. and D. Nathwani, 2005. Antibiotic cycling or rotation: a systemic review of the
486 evidence of efficacy. *Journal of Antimicrobial Chemotherapy* 55:6–9.

487 Burch, C. L. and L. Chao, 1999. Evolution by small steps and rugged landscapes in the rna virus
488 phi-6. *Genetics* 151:921–927. URL <https://www.genetics.org/content/151/3/921>.

489 Burton, D. R., P. Poignard, R. L. Stanfield, and I. A. Wilson, 2012. Broadly neutralizing antibodies
490 present new prospects to counter highly antigenically diverse viruses. *Science* 337:183–186.

491 Canino-Koning, R., M. J. Wisner, and C. Ofria, 2019. Fluctuating environments select for short-
492 term phenotypic variation leading to long-term exploration. *PLOS Computational Biology* 15:1–32.
493 URL <https://doi.org/10.1371/journal.pcbi.1006445>.

494 Constable, G. W. and A. J. McKane, 2014a. Fast-mode elimination in stochastic metapopulation
495 models. *Physical Review E* 89:032141.

496 ———, 2014b. Population genetics on islands connected by an arbitrary network: An analytic
497 approach. *Journal of theoretical biology* 358:149–165.

498 Cook, R. D. and D. L. Hartl, 1974. Uncorrelated random environments and their effects on gene
499 frequency. *Evolution* 28:265–274. URL <http://www.jstor.org/stable/2407328>.

500 Cooper, T. F. and R. E. Lenski, 2010. Experimental evolution with *e. coli* in diverse resource
501 environments. i. fluctuating environments promote divergence of replicate populations. *BMC evo-*
502 *lutionary biology* 10:11.

503 Crill, W. D., H. A. Wichman, and J. J. Bull, 2000. Evolutionary reversals during viral adaptation
504 to alternating hosts. *Genetics* 154:27–37. URL <https://www.genetics.org/content/154/1/27>.

505 Cvijović, I., B. H. Good, E. R. Jerison, and M. M. Desai, 2015. Fate of a mutation in a fluctuating
506 environment. *Proceedings of the National Academy of Sciences* 112:E5021–E5028. URL <https://www.pnas.org/content/112/36/E5021>.

507 <https://www.pnas.org/content/112/36/E5021>.

508 David, H. A. and H. N. Nagaraja, 2004. Order statistics. *Encyclopedia of Statistical Sciences* .

509 De Jong, M. G. and K. B. Wood, 2018. Tuning spatial profiles of selection pressure to modulate
510 the evolution of drug resistance. *Physical review letters* 120:238102.

511 Desai, M. M. and D. S. Fisher, 2007. Beneficial mutation–selection balance and the effect of linkage
512 on positive selection. *Genetics* 176:1759–1798. URL [https://www.genetics.org/content/176/](https://www.genetics.org/content/176/3/1759)
513 [3/1759](https://www.genetics.org/content/176/3/1759).

514 Desai, M. M., D. S. Fisher, and A. W. Murray, 2007. The speed of evolution and maintenance of
515 variation in asexual populations. *Current Biology* 17:385–394. URL [https://doi.org/10.1016/](https://doi.org/10.1016/j.cub.2007.01.072)
516 [j.cub.2007.01.072](https://doi.org/10.1016/j.cub.2007.01.072).

517 Dhawan, A., D. Nichol, F. Kinose, M. E. Abazeed, A. Marusyk, E. B. Haura, and J. G. Scott,
518 2017. Collateral sensitivity networks reveal evolutionary instability and novel treatment strategies
519 in alk mutated non-small cell lung cancer. *Scientific Reports* 7.

- 520 Durrett, R. and R. Durrett, 1999. Essentials of stochastic processes, vol. 1. Springer.
- 521 de Evgrafov, M. R., H. Gumpert, C. Munck, T. T. Thomsen, and M. O. A. Sommer, 2015. Collateral
522 resistance and sensitivity modulate evolution in high-level resistance to drug combination treatment
523 in *staphylococcus aureus*. Mol. Biol. Evol. 32:1175–1185.
- 524 Farhang-Sardroodi, S., A. Darooneh, M. Nikbakht, N. Komarova, and M. Kohandel, 2017. The
525 effect of spatial randomness on the average fixation time of mutants. PLoS computational biology
526 13:e1005864.
- 527 Fu, F., M. A. Nowak, and S. Bonhoeffer, 2015. Spatial heterogeneity in drug concentrations can
528 facilitate the emergence of resistance to cancer therapy. PLoS Comput Biol 11:e1004142.
- 529 Fuentes-Hernandez, A., J. Plucain, F. Gori, R. Pena-Miller, C. Reding, G. Jansen, H. Schulenburg,
530 I. Gudelj, and R. Beardmore, 2015. Using a sequential regimen to eliminate bacteria at sublethal
531 antibiotic dosages. PLoS biology 13:e1002104.
- 532 Gaál, B., J. W. Pitchford, and A. J. Wood, 2010. Exact results for the evolution of stochastic
533 switching in variable asymmetric environments. Genetics 184:1113–1119.
- 534 Gerhart, J. and M. Kirschner, 2007. The theory of facilitated variation. Proceedings of the National
535 Academy of Sciences 104:8582–8589.
- 536 Gerrish, P. J. and R. E. Lenski, 1998. The fate of competing beneficial mutations in an asexual
537 population. Genetica 102:127. URL <https://doi.org/10.1023/A:1017067816551>.
- 538 Gillespie, J. H., 1983a. A simple stochastic gene substitution model. Theoretical Population Biology
539 23:202 – 215. URL <http://www.sciencedirect.com/science/article/pii/004058098390014X>.
- 540 ———, 1983b. Some properties of finite populations experiencing strong selection and weak mu-
541 tation. The American Naturalist 121:691–708.
- 542 ———, 1984. Molecular evolution over the mutational landscape. Evolution 38:1116–1129.
- 543 Gillespie, J. H. and H. A. Guess, 1978. The effects of environmental autocorrelations on the
544 progress of selection in a random environment. The American Naturalist 112:897–909. URL
545 <https://doi.org/10.1086/283330>.
- 546 Greulich, P., B. Waclaw, and R. J. Allen, 2012. Mutational pathway determines whether drug
547 gradients accelerate evolution of drug-resistant cells. Physical Review Letters 109:088101.
- 548 Gupta, P. B., C. M. Fillmore, G. Kiang, S. D. Shapira, K. Tao, C. Kuperwasser, and E. S. Lander,
549 2011. Stochastic state transitions give rise to phenotypic equilibrium in populations of cancer cells.
550 Cell 146:633–644.

- 551 Habets, M. G., D. E. Rozen, R. F. Hoekstra, and J. A. G. de Visser, 2006. The effect of popula-
552 tion structure on the adaptive radiation of microbial populations evolving in spatially structured
553 environments. *Ecology letters* 9:1041–1048.
- 554 Hart, Y., H. Sheftel, J. Hausser, P. Szekely, N. B. Ben-Moshe, Y. Korem, A. Tendler, A. E. Mayo,
555 and U. Alon, 2015. Inferring biological tasks using pareto analysis of high-dimensional data. *Nature*
556 *methods* 12:233–235.
- 557 Hartl, D. L. and R. D. Cook, 1974. Autocorrelated random environments and their effects on gene
558 frequency. *Evolution* 28:275–280. URL <http://www.jstor.org/stable/2407329>.
- 559 Hermesen, R., J. B. Deris, and T. Hwa, 2012. On the rapidity of antibiotic resistance evolu-
560 tion facilitated by a concentration gradient. *Proceedings of the National Academy of Sciences*
561 109:10775–10780.
- 562 Hermesen, R. and T. Hwa, 2010. Sources and sinks: a stochastic model of evolution in heterogeneous
563 environments. *Physical review letters* 105:248104.
- 564 Imamovic, L., M. Ellabaan, A. Machado, S. Molin, H. Johansen, and M. Sommer, 2018. Drug-
565 driven phenotypic convergence supports rational treatment strategies of chronic infections. *Cell*
566 172:P121–134.
- 567 Imamovic, L. and M. O. A. Sommer, 2013. Use of collateral sensitivity networks to design drug
568 cycling protocols that avoid resistance development. *Sci. Transl. Med* 5:204ra132.
- 569 Kashtan, N., E. Noor, and U. Alon, 2007. Varying environments can speed up evolution. *Pro-*
570 *ceedings of the National Academy of Sciences* 104:13711–13716. URL [https://www.pnas.org/
571 content/104/34/13711](https://www.pnas.org/content/104/34/13711).
- 572 Kim, S., T. D. Lieberman, and R. Kishony, 2014. Alternating antibiotic treatments constrain
573 evolutionary paths to multidrug resistance. *Proceedings of the National Academy of Sciences*
574 111:14494–14499.
- 575 Korona, R., C. H. Nakatsu, L. J. Forney, and R. E. Lenski, 1994. Evidence for multiple adaptive
576 peaks from populations of bacteria evolving in a structured habitat. *Proceedings of the National*
577 *Academy of Sciences* 91:9037–9041.
- 578 Kussell, E. and S. Leibler, 2005. Phenotypic diversity, population growth, and information in fluctu-
579 ating environments. *Science* 309:2075–2078. URL [https://science.sciencemag.org/content/
580 309/5743/2075](https://science.sciencemag.org/content/309/5743/2075).

- 581 Lazar, V., A. Martins, R. Spohn, L. Daruka, G. Grezal, G. Fekete, M. Szamel, P. Jangir, B. Kintses,
582 B. Csorgo, A. Nyerges, A. Gyorkei, A. Kincses, A. Der, F. Walter, M. Deli, E. Urban, Z. Hegedus,
583 G. Olajos, O. Mehi, B. Balint, I. Nagy, T. Martinek, B. Papp, and C. Pal, 2018. Antibiotic-resistant
584 bacteria show widespread collateral sensitivity to antimicrobial peptides. *Nature Microbiology*
585 3:718–731.
- 586 Lazar, V., I. Nagy, R. Spohn, B. Csorgo, A. Gyorkei, A. Nyerges, B. Horvath, A. Voros, R. Busa-
587 Fekete, M. Hrtyan, B. Bogos, O. Mehi, G. Fekete, B. Szappanos, B. Kegl, B. Papp, and C. Pal,
588 2014. Genome-wide analysis captures the determinants of the antibiotic cross-resistance interaction
589 network. *Nat. Commun.* 5.
- 590 Lazar, V., G. P. Singh, R. Spohn, I. Nagy, B. Horvath, M. Hrtyan, R. Busa-Fekete, B. Bogos,
591 O. Mehi, B. Csorgo, G. Posfai, G. Fekete, B. Szappanos, B. Kegl, B. Papp, and C. Pal, 2013.
592 Bacterial evolution and antibiotic hypersensitivity. *Mol. Syst. Biol.* 9.
- 593 Lenski, R. E., 1988. Experimental studies of pleiotropy and epistasis in *Escherichia coli*. ii. com-
594 pensation for maladaptive effects associated with resistance to virus T4. *Evolution* 42:433–440. URL
595 <http://www.jstor.org/stable/2409029>.
- 596 Lewontin, R. C. and D. Cohen, 1969. On population growth in a randomly varying environment.
597 *Proceedings of the National Academy of Sciences* 62:1056–1060. URL [https://www.pnas.org/
598 content/62/4/1056](https://www.pnas.org/content/62/4/1056).
- 599 Lin, Y. T., H. Kim, and C. R. Doering, 2015. Demographic stochasticity and evolution of dispersion
600 ii: Spatially inhomogeneous environments. *Journal of Mathematical Biology* 70:679–707. URL
601 <https://doi.org/10.1007/s00285-014-0756-0>.
- 602 Maltas, J., B. Krasnick, and K. B. Wood, 2019. Using Selection by Nonantibiotic Stressors to
603 Sensitize Bacteria to Antibiotics. *Molecular Biology and Evolution* URL [https://doi.org/10.
604 1093/molbev/msz303](https://doi.org/10.1093/molbev/msz303). Msz303.
- 605 Maltas, J. and K. B. Wood, 2019. Pervasive and diverse collateral sensitivity profiles inform optimal
606 strategies to limit antibiotic resistance. *PLoS biology* 17.
- 607 Moreno-Gamez, S., A. L. Hill, D. I. Rosenbloom, D. A. Petrov, M. A. Nowak, and P. S. Pennings,
608 2015. Imperfect drug penetration leads to spatial monotherapy and rapid evolution of multidrug
609 resistance. *Proceedings of the National Academy of Sciences* 112:E2874–E2883.
- 610 Munck, C., H. K. Gumpert, A. I. N. Wallin, H. H. Wang, and M. O. A. Sommer, 2014. Prediction
611 of resistance development against drug components by collateral responses to component drugs.

612 Sci. Transl. Med 6:262ra156.

613 Mustonen, V. and M. Lässig, 2008. Molecular evolution under fitness fluctuations. Phys. Rev.
614 Lett. 100:108101. URL <https://link.aps.org/doi/10.1103/PhysRevLett.100.108101>.

615 Mustonen, V. and M. Lässig, 2009. From fitness landscapes to seascares: non-equilibrium dynamics
616 of selection and adaptation. Trends in genetics 25:111–119.

617 Neidhart, J., I. G. Szendro, and J. Krug, 2014. Adaptation in tunably rugged fitness landscapes: the
618 rough mount fuji model. Genetics 198:699–721. URL [https://www.ncbi.nlm.nih.gov/pubmed/
619 25123507](https://www.ncbi.nlm.nih.gov/pubmed/25123507). 25123507[pmid].

620 Nichol, D., P. Jeavons, A. G. Fletcher, R. A. Bonomo, P. K. Maini, J. L. Paul, R. A. Gatenby,
621 A. R. Anderson, and J. G. Scott, 2015. Steering evolution with sequential therapy to prevent
622 the emergence of bacterial antibiotic resistance. PLOS Computational Biology 11:1–19. URL
623 <https://doi.org/10.1371/journal.pcbi.1004493>.

624 Nichol, D., J. Rutter, C. Bryant, A. M. Hujer, S. Lek, M. D. Adams, P. Jeavons, A. R. Ander-
625 son, R. A. Bonomo, and J. G. Scott, 2019. Antibiotic collateral sensitivity is contingent on the
626 repeatability of evolution. Nature communications 10:334.

627 O'Neill, M., L. Vanneschi, S. Gustafson, and W. Banzhaf, 2010. Open issues in genetic programming.
628 Genetic Programming and Evolvable Machines 11:339–363.

629 Parter, M., N. Kashtan, and U. Alon, 2008. Facilitated variation: how evolution learns from past
630 environments to generalize to new environments. PLoS computational biology 4:e1000206.

631 Patra, P. and S. Klumpp, 2015. Emergence of phenotype switching through continuous and dis-
632 continuous evolutionary transitions. Physical biology 12:046004.

633 Phillips, P. C., 2008. Epistasis: the essential role of gene interactions in the structure and evolution
634 of genetic systems. Nature Reviews Genetics 9:855.

635 Rhee, S.-Y., J. Taylor, W. J. Fessel, D. Kaufman, W. Towner, P. Troia, P. Ruane, J. Hellinger,
636 V. Shirvani, A. Zolopa, et al., 2010. Hiv-1 protease mutations and protease inhibitor cross-
637 resistance. Antimicrobial agents and chemotherapy 54:4253–4261.

638 Ritchie, M. D., L. W. Hahn, N. Roodi, L. R. Bailey, W. D. Dupont, F. F. Parl, and J. H.
639 Moore, 2001. Multifactor-dimensionality reduction reveals high-order interactions among estrogen-
640 metabolism genes in sporadic breast cancer. The American Journal of Human Genetics 69:138–147.
641 URL <https://doi.org/10.1086/321276>.

- 642 Roemhild, R., C. Barbosa, R. E. Beardmore, G. Jansen, and H. Schulenburg, 2015. Temporal
643 variation in antibiotic environments slows down resistance evolution in pathogenic *Pseudomonas*
644 *aeruginosa*. *Evolutionary applications* 8:945–955.
- 645 Roemhild, R., C. S. Gokhale, P. Dirksen, C. Blake, P. Rosenstiel, A. Traulsen, D. I. Andersson,
646 and H. Schulenburg, 2018. Cellular hysteresis as a principle to maximize the efficacy of antibiotic
647 therapy. *Proceedings of the National Academy of Sciences* 115:9767–9772.
- 648 Royston, J., 1982. Algorithm as 177: Expected normal order statistics (exact and approximate).
649 *Journal of the royal statistical society. Series C (Applied statistics)* 31:161–165.
- 650 Sachdeva, V., K. Husain, J. Sheng, S. Wang, and A. Murugan, 2020. Tuning environmental
651 timescales to evolve and maintain generalists. *Proceedings of the National Academy of Sciences*
652 117:12693–12699.
- 653 Shahrezaei, V., J. F. Ollivier, and P. S. Swain, 2008. Colored extrinsic fluctuations and stochastic
654 gene expression. *Molecular Systems Biology* 4:196. URL [https://www.embopress.org/doi/abs/
655 10.1038/msb.2008.31](https://www.embopress.org/doi/abs/10.1038/msb.2008.31).
- 656 Shoval, O., H. Sheftel, G. Shinar, Y. Hart, O. Ramote, A. Mayo, E. Dekel, K. Kavanagh, and
657 U. Alon, 2012. Evolutionary trade-offs, pareto optimality, and the geometry of phenotype space.
658 *Science* 336:1157–1160.
- 659 da Silva, J., M. Coetzer, R. Nedellec, C. Pastore, and D. E. Mosier, 2010. Fitness epistasis and
660 constraints on adaptation in a human immunodeficiency virus type 1 protein region. *Genetics*
661 185:293–303. URL <https://www.genetics.org/content/185/1/293>.
- 662 Skanata, A. and E. Kussell, 2016. Evolutionary phase transitions in random environments. *Physical*
663 *review letters* 117:038104.
- 664 Steinberg, B. and M. Ostermeier, 2016. Environmental changes bridge evolutionary valleys. *Science*
665 *advances* 2:e1500921.
- 666 Tan, L. and J. Gore, 2012. Slowly switching between environments facilitates reverse evolution in
667 small populations. *Evolution* 66:3144–3154. URL [https://onlinelibrary.wiley.com/doi/abs/
668 10.1111/j.1558-5646.2012.01680.x](https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1558-5646.2012.01680.x).
- 669 Tan, L., S. Serene, H. X. Chao, and J. Gore, 2011. Hidden randomness between fitness landscapes
670 limits reverse evolution. *Phys. Rev. Lett.* 106:198102. URL [https://link.aps.org/doi/10.
671 1103/PhysRevLett.106.198102](https://link.aps.org/doi/10.1103/PhysRevLett.106.198102).

- 672 Tsai, C.-T., J.-J. Hwang, M. D. Ritchie, J. H. Moore, F.-T. Chiang, L.-P. Lai, K.-L. Hsu, C.-
673 D. Tseng, J.-L. Lin, and Y.-Z. Tseng, 2007. Renin–angiotensin system gene polymor-
674 phisms and coronary artery disease in a large angiographic cohort: Detection of high order
675 gene–gene interaction. *Atherosclerosis* 195:172–180. URL [https://doi.org/10.1016/
676 j.atherosclerosis.2006.09.014](https://doi.org/10.1016/j.atherosclerosis.2006.09.014).
- 677 de Vos, M. G. J., A. Dawid, V. Sunderlikova, and S. J. Tans, 2015. Breaking evolutionary constraint
678 with a tradeoff ratchet. *Proceedings of the National Academy of Sciences* 112:14906–14911. URL
679 <https://www.pnas.org/content/112/48/14906>.
- 680 Waddell, J. N., L. M. Sander, and C. R. Doering, 2010. Demographic stochasticity versus spatial
681 variation in the competition between fast and slow dispersers. *Theoretical population biology*
682 77:279–286.
- 683 Wang, S. and L. Dai, 2019. Evolving generalists in switching rugged landscapes. *PLOS Computa-*
684 *tional Biology* 15:e1007320.
- 685 Weinreich, D. M., N. F. Delaney, M. A. DePristo, and D. L. Hartl, 2006. Darwinian evolution
686 can follow only very few mutational paths to fitter proteins. *Science* 312:111–114. URL [https:
687 //science.sciencemag.org/content/312/5770/111](https://science.sciencemag.org/content/312/5770/111).
- 688 Whitlock, M. C. and R. Gomulkiewicz, 2005. Probability of fixation in a heterogeneous environ-
689 ment. *Genetics* 171:1407–1417.
- 690 Xu, J., J. Lowey, F. Wiklund, J. Sun, F. Lindmark, F.-C. Hsu, L. Dimitrov, B. Chang, A. R.
691 Turner, W. Liu, H.-O. Adami, E. Suh, J. H. Moore, S. L. Zheng, W. B. Isaacs, J. M. Trent,
692 and H. Grönberg, 2005. The interaction of four genes in the inflammation pathway significantly
693 predicts prostate cancer risk. *Cancer Epidemiology and Prevention Biomarkers* 14:2563–2568. URL
694 <https://cebp.aacrjournals.org/content/14/11/2563>.
- 695 Yoshida, M., S. G. Reyes, S. Tsudo, T. Horinouchi, C. Furusawa, and L. Cronin, 2017. Time-
696 programmable dosing allows the manipulation, suppression and reversal of antibiotic drug resis-
697 tance *in vitro*. *Nat. Commun.* 8.
- 698 Zhang, Q., G. Lambert, D. Liao, H. Kim, K. Robin, C.-k. Tung, N. Pourmand, and R. H. Austin,
699 2011. Acceleration of emergence of bacterial antibiotic resistance in connected microenvironments.
700 *Science* 333:1764–1767.

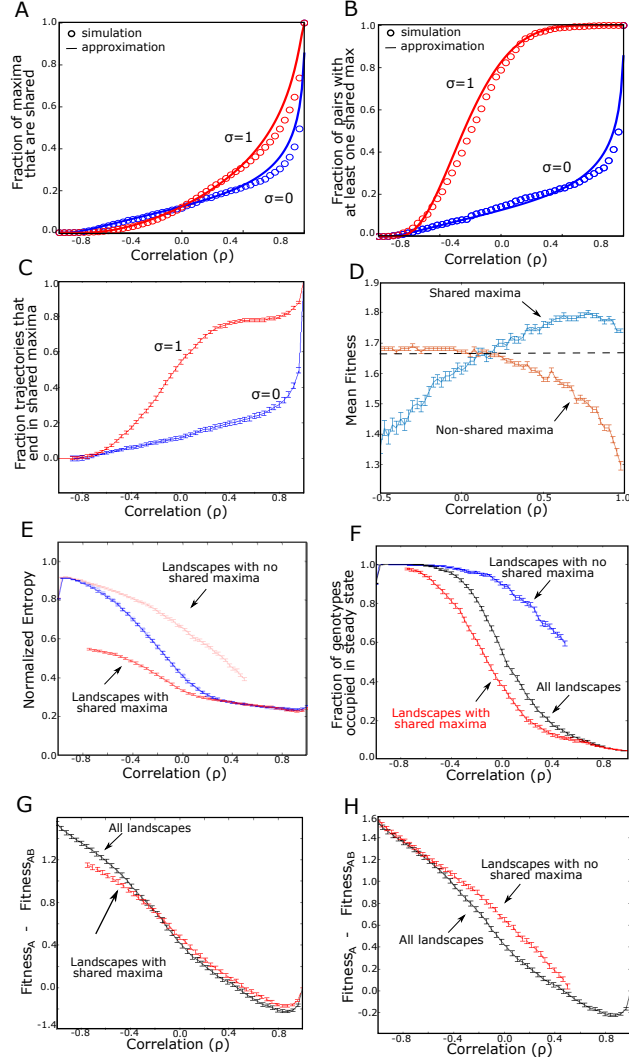


FIG. 4: Evolution in alternating landscapes is frequently dominated by presence or absence of shared fitness maxima. A. Fraction of local maxima in landscape A that also correspond to a shared maxima in landscape B ($\sigma = 0$, blue; and $\sigma = 1$, red). B. Fraction of landscape pairs that share at least one maximum. In panels A and B, circles correspond to simulated landscapes and solid lines are semi-analytic approximations (see SI). C. Fraction of trajectories ending in a shared maximum as a function of correlation. D. Average fitness of shared maxima (blue) and average fitness of non-shared maxima (orange). Dashed line is average fitness of all local maxima in landscape A. E. Normalized entropy of the steady state genotype distribution following adaptation to alternating landscapes. Curves correspond to the full landscape pair ensemble (blue) and a reduced ensemble consisting only of landscapes that contain a shared maximum (red), bottom, and a reduced ensemble consisting only of landscapes with no shared maxima (red, top). The relative entropy is defined as $S(p)/S_{max} \equiv -(\sum_i p_i \ln p_i)/S_{max}$, where p_i is the steady state probability of being in genotype i and S_{max} is the entropy of a uniform distribution. F. Fraction of genotypes that have a nonzero probability of occupation in either the last A step or last B step at steady-state. Curves represent the paired landscape ensemble with no shared maxima (blue), the ensemble where every pair has at least one shared maximum (red), and the full ensemble (black). G. Difference in average fitness achieved in static and switching landscapes. Curves correspond to the full ensemble of paired landscapes (black) or a restricted ensemble that includes only those pairs that share a fitness maximum (red). H. Similar to panel F, with curves corresponding to the full ensemble (black) or a restricted ensemble that includes only those pairs with no shared fitness maxima (red). Error bars are \pm standard error of the mean in the ensemble of landscapes. Error bars are \pm standard error of the mean in the ensemble of landscapes. $N = 7$ for all curves, and $\sigma = 1$ for all curves in panels D-H.

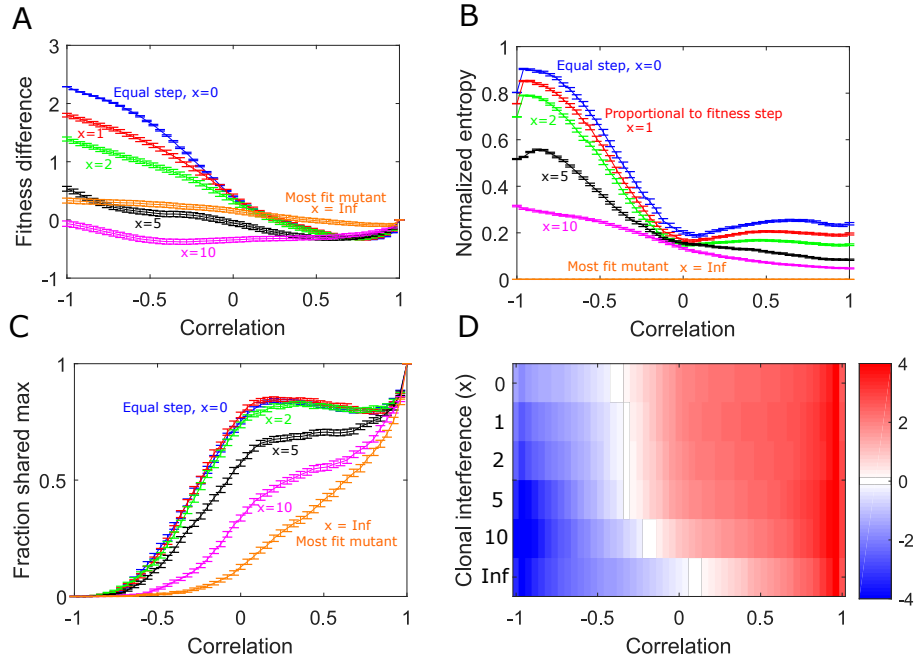


FIG. 5: Clonal interference reduces the effects of alternating landscape evolution. A. Difference in average fitness achieved in static and switching landscapes. Curves correspond to different strengths of clonal interference (blue: random walker, $x = 0$, red: proportional walker, $x = 1$, green: $x = 2$, black: $x = 5$, magenta: $x = 10$, orange: x infinite, always steps to largest fitness neighbor). B. Normalized entropy of the steady state genotype distribution following adaption to alternating landscapes with different clonal interference. C. Fraction of trajectories ending in a shared maximum as a function of correlation with different clonal interference. D. Collateral fitness change, ranging from blue (less fit) to red (more fit), for populations adapted to alternating environments A and B as a function of clonal interference (x).

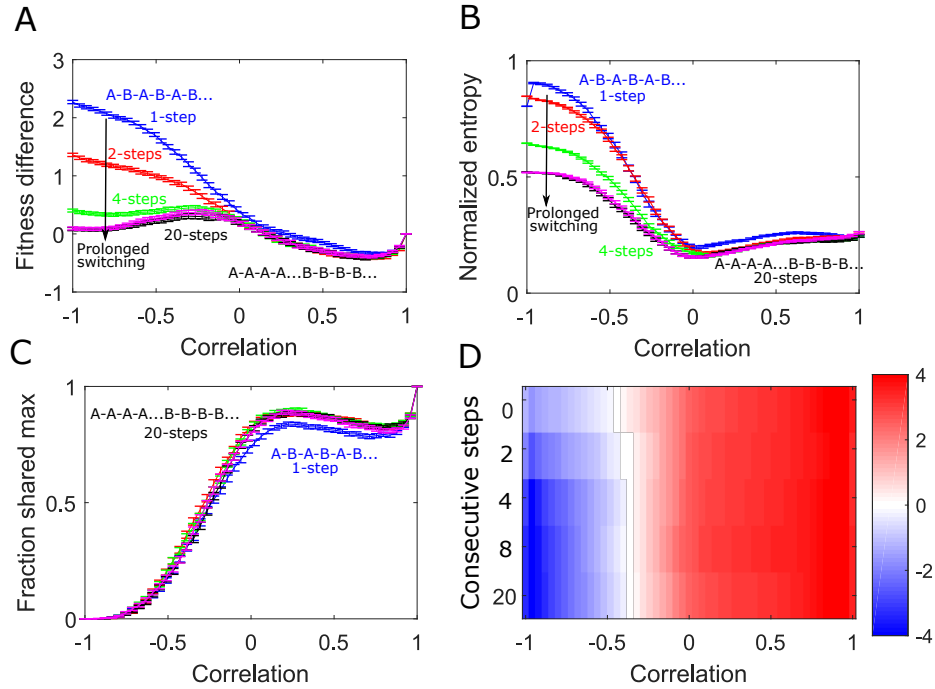


FIG. 6: Consecutive steps in the same landscape before switching lessens the effects of alternating landscape evolution. A. Difference in average fitness achieved in static and switching landscapes. Curves correspond to different evolutionary steps taken in a landscape before switching (blue: 1 step, red: 2 steps, green: 4 steps, magenta: 8 steps, black: 20 steps). B. Normalized entropy of the steady state genotype distribution following adaption to alternating in landscapes with different switching periods. C. Fraction of trajectories ending in a shared maximum as a function of correlation with different switching periods. D. Collateral fitness change, ranging from blue (less fit) to red (more fit), for populations adapted to alternating environments A and B as a function of switching period.

701 **SUPPLEMENTAL MATERIAL**

702 The Supplemental Material contains semi-analytic approximations for the number of local
703 maxima in a single landscape and the probability of shared maxima in paired landscapes.
704 It also includes seven supplemental figures (S1-S7).

705 **Semi-analytical approximations to describe local maxima**

706 Dynamics in alternating environments are often impacted by the presence of shared local
707 maxima. Here we derive semi-analytical approximate expressions for several key quantities.
708 While exact expressions are difficult to obtain, even for the simple model used here, we
709 derive below several approximations that involve cumulative distribution functions (CDFs)
710 for common distributions (e.g. multivariate normal) and/or Gaussian-like integrals that can
711 be easily calculated numerically.

712 **Number of local maxima in a single landscape**

713 Rugged landscapes ($\sigma > 0$) potentially have multiple local maxima. To approximate the
714 expected number of local maxima, consider a landscape with N loci so that each genotype has
715 a total of N nearest neighbor genotypes. In a purely additive ($\sigma = 0$) landscape, a mutation
716 in gene i changes the fitness by an amount ϵ_i drawn from a uniform distribution $[0,1]$. In a
717 rugged landscape, the fitness of each genotype also contains an additive contribution from
718 epistasis—in this case, a zero-mean, normally distributed random variable with variance σ^2 .
719 The fitness of each genotype is therefore a sum of (up to N) uniform variables and a single
720 normally distributed variable.

To estimate the expected number of maxima in a landscape, consider a particular geno-
type with a fixed fitness $f = f_\epsilon + f_\sigma$, where f_ϵ is the total fitness contribution from any
mutations and f_σ is the contribution from epistasis. The genotype will have N neigh-
bors, each differing by a single mutation; the fitness of neighbor i has a fitness of the form
 $f_i = f_\epsilon + \epsilon_i + \sigma_i$, where ϵ_i is a uniform random variable accounting for adding or subtracting
one mutation, and σ_i is a normal random variable accounting for epistasis. The f_ϵ term is a
fixed value—the same as for the focal genotype. In a statistical ensemble of such neighbors,
the probability that $f > f_i$, that is, the probability that the genotype in question has a

higher fitness than one particular neighbor is given by

$$p_{max}^i(f_\sigma) = \int_{-\infty}^{f_\sigma} dx p_+(x), \quad (\text{S1})$$

where $p_+(x)$ is the probability density function (pdf) for the sum $\sigma_i + \epsilon_i$. Since the pdf for a sum of random variables is given by the convolution of their individual pdfs, we have

$$p_{max}^i(f_\sigma) = \frac{1}{2} \int_{-\infty}^{f_\sigma} dx \int_{-1}^1 du \phi_{\sigma_i}(x - u), \quad (\text{S2})$$

where $\phi_\sigma(x)$ is the pdf of a zero-mean normal variable with variance σ^2 ,

$$\phi_\sigma(x) \equiv \frac{1}{\sqrt{2\pi\sigma^2}} \exp\left(-\frac{x^2}{2\sigma^2}\right). \quad (\text{S3})$$

Equation S2 can also be written as

$$p_{max}^i(f_\sigma) = \frac{1}{2} \int_{-1}^1 du F_{\sigma_i}(f_\sigma + u), \quad (\text{S4})$$

721 where $F_{\sigma_i}(x)$ is the cumulative distribution function for the variable σ_i (and in this case,
 722 each σ_i is a zero-mean normal variable with variance σ^2). The integral above can be written
 723 as a linear sum of error functions, though it is somewhat cumbersome and we do not write
 724 it out here.

The probability that the genotype in question is a local max—that is, has a fitness larger than each of its N nearest neighbors—is approximately

$$p_{max}(f_\sigma) \approx (p_{max}^i(f_\sigma))^N \quad (\text{S5})$$

where we have assumed that each neighbor can be treated as independent from the others. The average probability that a genotype is a local maximum is then given by integrating over the distribution for f_σ ,

$$P_{max} = \int_{-\infty}^{\infty} df_\sigma \phi_\sigma(f_\sigma) p_{max}(f_\sigma) = \langle p_{max}(x) \rangle \quad (\text{S6})$$

where brackets $\langle \cdot \rangle$ represent an average over a normal distribution with variance σ^2 . If we

assume that the 2^N different genotypes in a landscape are approximately independent, the expected number of maxima is then given by

$$N_{max} = 2^N P_{max}. \quad (\text{S7})$$

Equation S7 is difficult to evaluate analytically but easy to solve numerically, and the approximation closely matches results from randomly generated landscapes (Figure 3). For small epistasis ($\sigma \ll 1$), we can expand $p_{max}(f_\sigma)$ about the average $\langle f_\sigma \rangle$ to arrive at the approximation

$$N_{max} \approx 1 + \frac{1}{2}N(N-1)\sigma^2. \quad (\text{S8})$$

Similarly, for large epistasis ($\sigma \rightarrow \infty$), we have $P_{max} \approx (N+1)^{-1}$; intuitively, all genotypes in a local neighborhood (the focal genotype and its N nearest neighbors) are equally likely to be the maximum, and the expected number of maxima therefore approaches $N_{max} = 2^N/(N+1)$.

Shared maxima between correlated landscapes

Given that a particular genotype corresponds to a local maximum in landscape A, we would like to estimate the probability that it is also a maximum in the paired landscape B. To do so, consider a genotype that is a local maximum in landscape A. Let the fitness of that genotype be a_1 and the fitness of its N nearest neighbors be $a_2 > a_3 \dots > a_{N+1}$, where we have labeled the neighbors according to their ranked fitness. We would like to calculate the conditional probability $p(b_1, b_2 \dots b_{N+1} | a_1, a_2 \dots a_{N+1})$ that describes the fitness values $\{b_i\}$ of the corresponding genotypes in landscape B, which is correlated with landscape A with correlation ρ .

In the limit of large epistasis ($\sigma \rightarrow \infty$), the fitness variables $\{a_i\}$ and $\{b_i\}$ are jointly distributed normal variables with mean $\bar{\mu} = (\mu_{a_1}, \mu_{a_2}, \dots, \mu_{a_{N+1}}, \mu_{b_1}, \mu_{b_2} \dots \mu_{b_{N+1}}) = (0, 0, \dots, 0)$ and covariance matrix

$$\Sigma = \begin{pmatrix} \Sigma_{aa} & \Sigma_{ab} \\ \Sigma_{ab} & \Sigma_{bb} \end{pmatrix} \quad (\text{S9})$$

made of $(N + 1) \times (N + 1)$ sub-matrices

$$\Sigma_{aa} = \Sigma_{bb} = \sigma^2 \begin{pmatrix} 1 & 0 & \dots & 0 \\ 0 & 1 & \dots & 0 \\ \vdots & \vdots & \ddots & \vdots \\ 0 & 0 & \dots & 1 \end{pmatrix} \quad (\text{S10})$$

and

$$\Sigma_{ab} = \sigma^2 \begin{pmatrix} \rho & 0 & \dots & 0 \\ 0 & \rho & \dots & 0 \\ \vdots & \vdots & \ddots & \vdots \\ 0 & 0 & \dots & \rho \end{pmatrix} \quad (\text{S11})$$

The matrix Σ_{aa} (Σ_{bb}) describes the covariance relationships between the focal genotype in landscape A (B) and each of its N nearest neighbors. The matrix Σ_{ab} describes the covariance between fitness values for the local neighborhood of $N + 1$ genotypes in landscapes A and B. If we treat the fitness values $\{a_i\}$ as fixed and the fitness values $\{b_i\}$ as random variables, the conditional probability $p(b_1, b_2, \dots, b_{N+1} | a_1, a_2, \dots, a_{N+1})$ is also normally distributed, with mean vector $\bar{\mu}_{cond} = \rho(a_1, a_2, \dots, a_{N+1})$ and covariance

$$\Sigma_{cond} = \sigma^2 \begin{pmatrix} 1 - \rho^2 & 0 & \dots & 0 \\ 0 & 1 - \rho^2 & \dots & 0 \\ \vdots & \vdots & \ddots & \vdots \\ 0 & 0 & \dots & 1 - \rho^2 \end{pmatrix} \quad (\text{S12})$$

We would like to know the probability of b_1 corresponding to a local maximum in landscape B given that a_1 corresponds to a local maximum in landscape A. To do so, we consider the N variables $\delta_i \equiv b_i - b_{i-1}$, whose distribution (conditioned on a specific set of values $\{a_i\}$) is a multivariate normal with mean $\bar{\mu}_\delta = \rho(a_1 - a_2, a_1 - a_2, \dots, a_1 - a_{N+1})$ and covariance

$$\Sigma_\delta = \sigma^2 \begin{pmatrix} 2 - 2\rho^2 & 1 - \rho^2 & \dots & 1 - \rho^2 \\ 1 - \rho^2 & 2 - 2\rho^2 & \dots & 1 - \rho^2 \\ \vdots & \vdots & \ddots & \vdots \\ 1 - \rho^2 & 1 - \rho^2 & \dots & 2 - 2\rho^2 \end{pmatrix} \quad (\text{S13})$$

Hence, if we are given a specific set of fitness values $\{a_i\}$, with a_1 the maximum of the local fitness neighborhood in landscape A, the probability that the fitness is also a maximum in the B landscape is given by

$$p_{\text{shared}}(\sigma, \rho) = 1 - \bar{F}_{\delta, \bar{a}}(\bar{0}) \quad (\text{S14})$$

where $\bar{F}_{\delta, \bar{a}}(\bar{x})$ is the cumulative distribution function (CDF) for the multivariate normal with mean $\bar{\mu}_\delta$ and covariance Σ_δ (conditioned on a set of values $\bar{a} = (a_1, a_2, \dots, a_{N+1})$) and $\bar{0}$ is the zero vector. Specifically, we have

$$\bar{F}_{\delta, \bar{a}}(\bar{x}) = \int_{-\infty}^{x_1} dz_1 \int_{-\infty}^{x_2} dz_2 \dots \int_{-\infty}^{x_N} dz_N \frac{1}{(2\pi)^{N/2} |\Sigma_\delta|^{1/2}} \exp\left(-(\bar{z} - \bar{\mu}_\delta)^T \Sigma_\delta^{-1} (\bar{z} - \bar{\mu}_\delta)\right) \quad (\text{S15})$$

where x_i is component i of \bar{x} . While there is no closed expression for the CDF of a multivariate Gaussian, there are many algorithms to rapidly calculate it numerically, and many scientific computing platforms even have built-in functions for this purpose.

To complete our approximation, we must choose specific values of the fixed variables $\{a_i\}$ on which the approximation is conditioned. In what follows, we consider two choices that lead to approximate expressions in the limits of large and small epistasis.

In the limit of large epistasis, the fitness values in the local neighborhood $\{a_i\}$ are uncorrelated, Gaussian variables with variance σ^2 . We therefore choose a_i to be the expected value of the i -th largest value in a sample of Gaussian variables (i.e. an order statistic (David and Nagaraja 2004)); without loss of generality, we assume the variables have mean zero. While there is no analytical expression for the expected value of the order statistics for normal variables, multiple approximations have been proposed. Here, we use the approximation in Royston (1982), which gives for our $N + 1$ fitness variables

$$a_i \approx \Phi^{-1}\left(\frac{N + 1 - i - \alpha}{N - 2\alpha + 2}\right) \quad (\text{S16})$$

where Φ^{-1} is the inverse CDF for the unit Gaussian and $\alpha = 0.375$ (we note that the order statistics can be calculated numerically to high precision, which slightly improves the approximation). Therefore, in the large σ limit, we have

$$p_{\text{shared}}(\sigma, \rho) = 1 - \bar{F}_{\delta, \bar{a}_g}(\bar{0}) \quad (\text{S17})$$

744 where the i -th component of \bar{a}_g is given in Equation S16.

In the limit $\sigma \rightarrow 0$, the fitness values in the local neighborhood $\{a_i\}$ are uncorrelated variables drawn from the uniform distribution $[-1,1]$ (where again we choose a zero mean distribution without loss of generality). In this case, the expected value of the order statistics for uniform variables leads to

$$a_i = \frac{2(N+2-i)}{(N+2)} - 1, \quad (\text{S18})$$

and our approximate expression is therefore

$$p_{\text{shared}}(\sigma, \rho) = 1 - \bar{F}_{\delta, \bar{a}_u}(\bar{0}), \quad (\text{S19})$$

745 where the i -th component of \bar{a}_u is given in Equation S18.

Finally, the fraction of landscape pairs that share at least one maximum is given by

$$f_{\text{shared}} = 1 - (1 - p_{\text{shared}}(\sigma, \rho))^{N_{\text{max}}} \quad (\text{S20})$$

746 The approximations in Equations S17-S20 are not exact, but we find that they agree quite
747 well with results from simulated landscapes in the $\sigma = 0$ (low epistasis) and $\sigma = 1$ (high
748 epistasis) cases (Figure 4A and 4B).

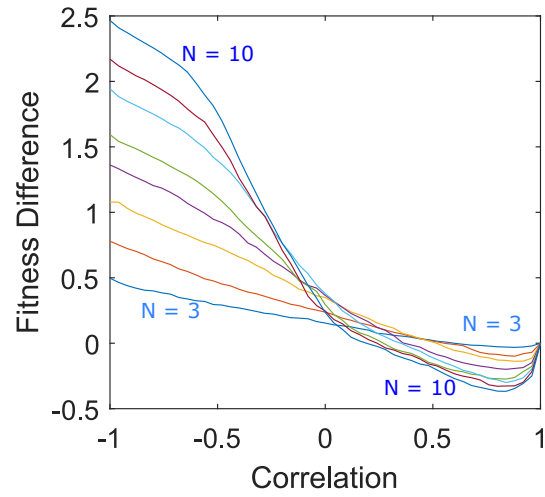


FIG. S1: **Rugged landscapes of different sizes show qualitatively similar changes in fitness as a function of correlation.** Difference in average fitness (at steady state) between populations adapted to a single static landscape (landscape A) or rapidly alternating landscape pairs (A-B cycles) as a function of correlation between landscapes A and B. Average fitness is defined as the mean fitness of the steady state genotype distribution (which arises following adaptation to either static or switching protocols) measured in landscape A. Different curves range from $N = 3$ to $N = 10$, and $\sigma = N/12$ for each landscape to achieve relatively similar magnitudes of epistasis as N varies.

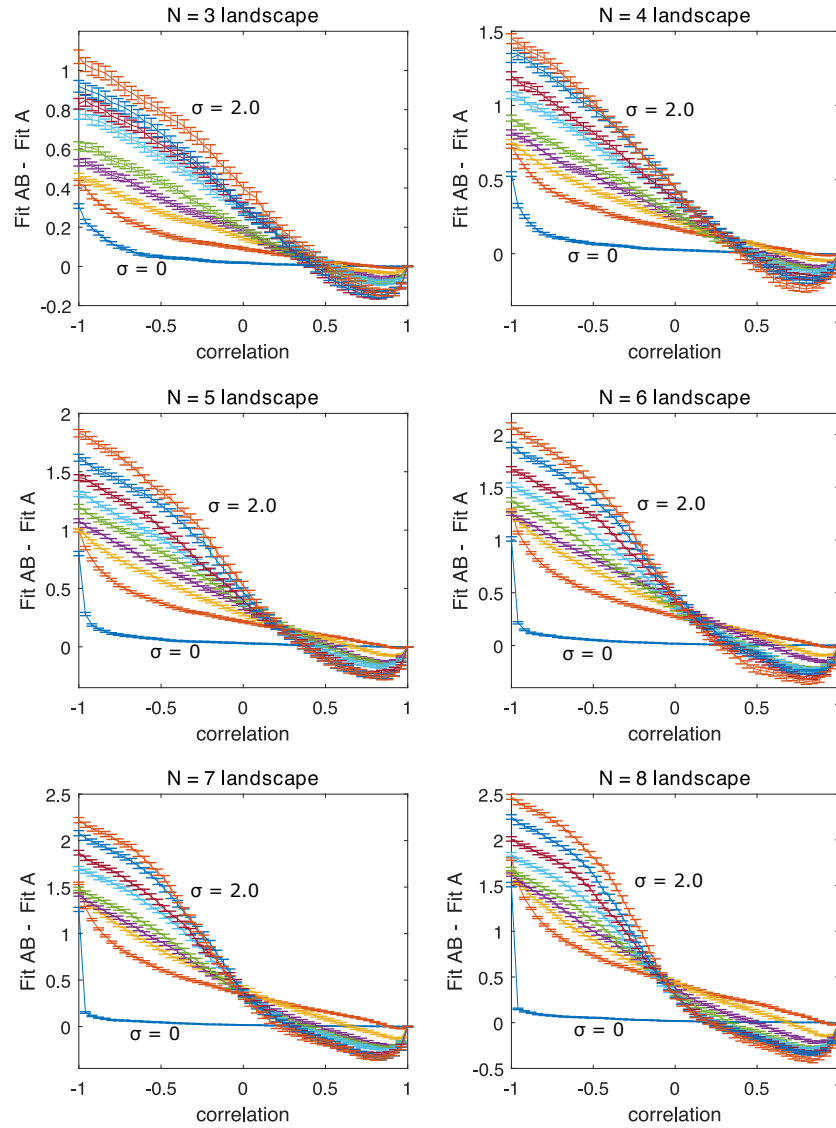


FIG. S2: **Landscapes of different sizes and sigmas show qualitatively similar results.** Difference in average fitness (at steady state) between populations adapted to a single static landscape (landscape A) or rapidly alternating landscape pairs (A-B cycles) as a function of correlation between landscapes A and B. Average fitness is defined as the mean fitness of the steady state genotype distribution (which arises following adaptation to either static or switching protocols) measured in landscape A. Different curves range from $\sigma = 0.0$ (blue, labeled) to $\sigma = 2.0$ (orange, labeled) in increments of 0.25 for each landscape. Error bars represent the standard error of the mean for each simulation.

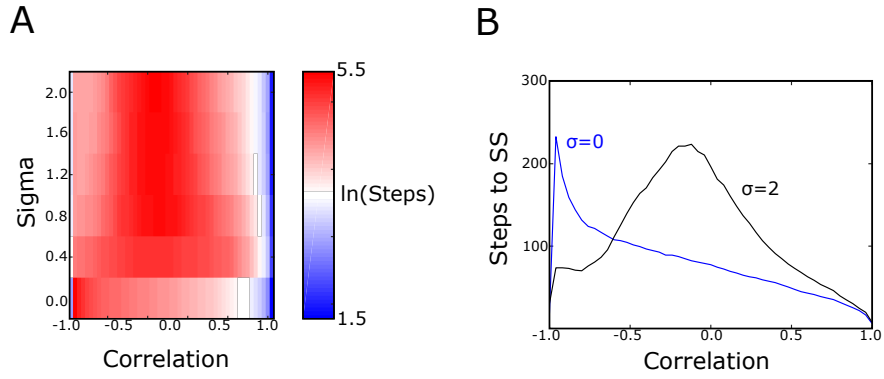


FIG. S3: **Adaptation to static and alternating environments approach steady state at different timescales.** A. Number of time steps (log scale) until steady state for alternating landscapes of a given ruggedness (σ) and correlation (ρ). Full correlated landscapes ($\rho = 1$) correspond to static evolution in a single landscape. B. Example slices through panel A corresponding to $\sigma = 0$ and $\sigma = 2$.

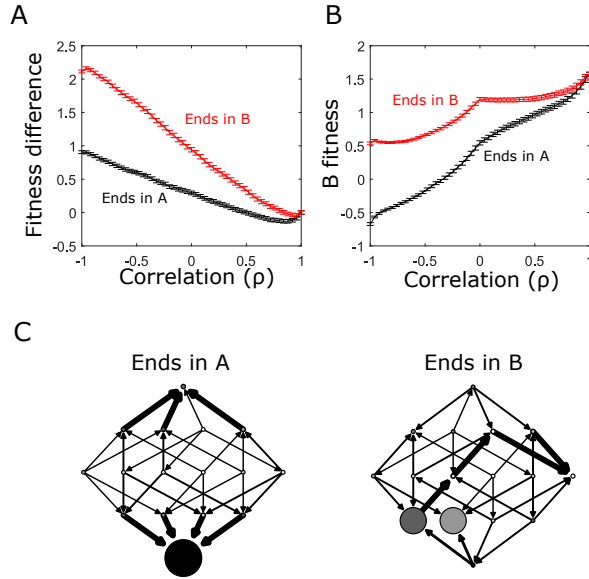


FIG. S4: **Adapted fitness depends on whether final step is taken in landscape A or B when landscapes are anticorrelated.** A. Difference in average fitness (at steady state) between populations adapted to a single static landscape (landscape A) or rapidly alternating landscape pairs (A-B cycles) as a function of correlation between landscapes A and B. Average fitness is defined as the mean fitness of the steady state genotype distribution (which arises following adaptation to either static or switching protocols) measured in landscape A. Curves correspond to steady state with a final step in landscape A (black) or a final step in landscape B (red). B. Collateral fitness change for populations adapted to alternating environments A and B as a function of inter-landscape correlation. Collateral fitness change is defined as the increase in average fitness in landscape B (relative to ancestor) associated with the steady state genotype distribution arising from adaptation to alternating A-B landscapes. C. Network representation of example fitness landscapes and transition probabilities following long-term adaptation to uncorrelated ($\rho = 0$) landscapes; adaptation ends either in landscape A (left) or B (right). $N = 4$ in all panels.

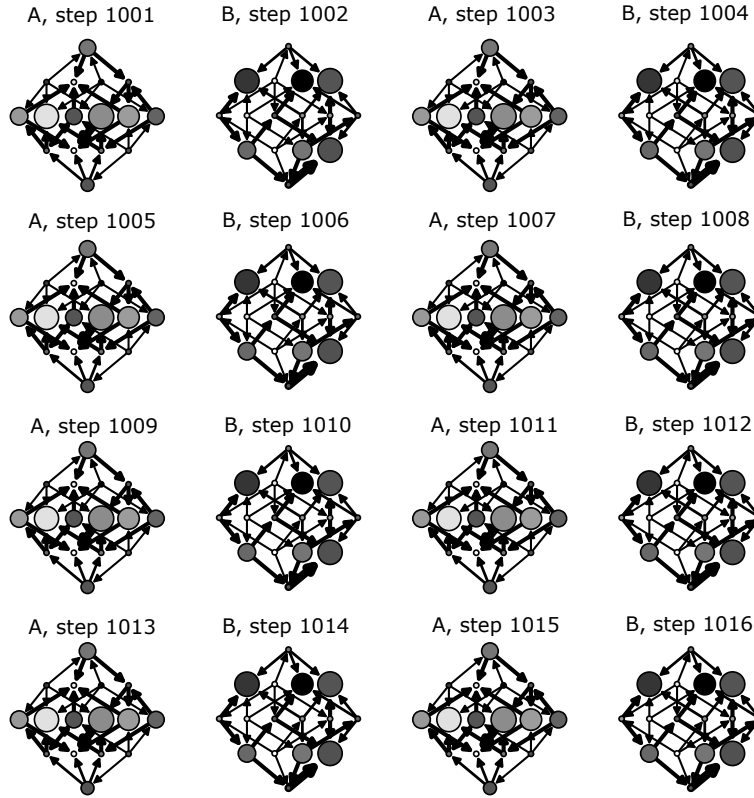


FIG. S5: **Adaptation to anti-correlated landscapes can produce cycles that sample large fractions of genotype space.** Network representations of 16 consecutive steps in the steady state for paired landscape evolution with $\rho = -0.88$. Each circle represents a genotype (ancestral genotype at the top), with shading indicating the relative fitness of that genotype and size representing the occupation probability at that time step. Arrows represent transitions between genotypes that occur with nonzero probability and are accessible starting from the ancestor genotype. The width of the arrow represents the magnitude of the transition probability.

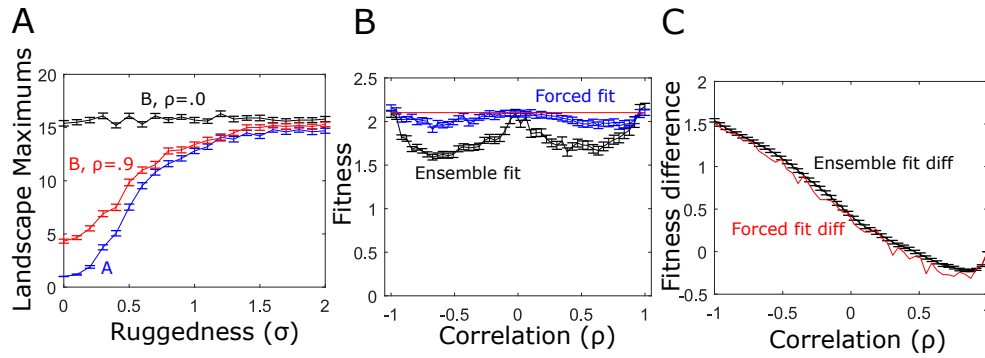


FIG. S6: **Statistical properties of landscape B differ from those of A but do not appreciably impact fitness differences between static and alternating landscapes.** A. Average number of local maxima in landscape A (blue) and two different B landscapes correlated with A to different degrees ($\rho = 0$, black; $\rho = 0.9$, red). B. Evolved fitness following static adaptation to landscape A (red) or B (black). Blue curve is fitness in a reduced “forced fit” ensemble of B landscapes, which includes only those B landscapes that lead to similar levels of fitness as in landscape A. C. Fitness difference between static and switching environments for the full paired landscape ensemble (black) and for the reduced “forced fit” ensemble (black).

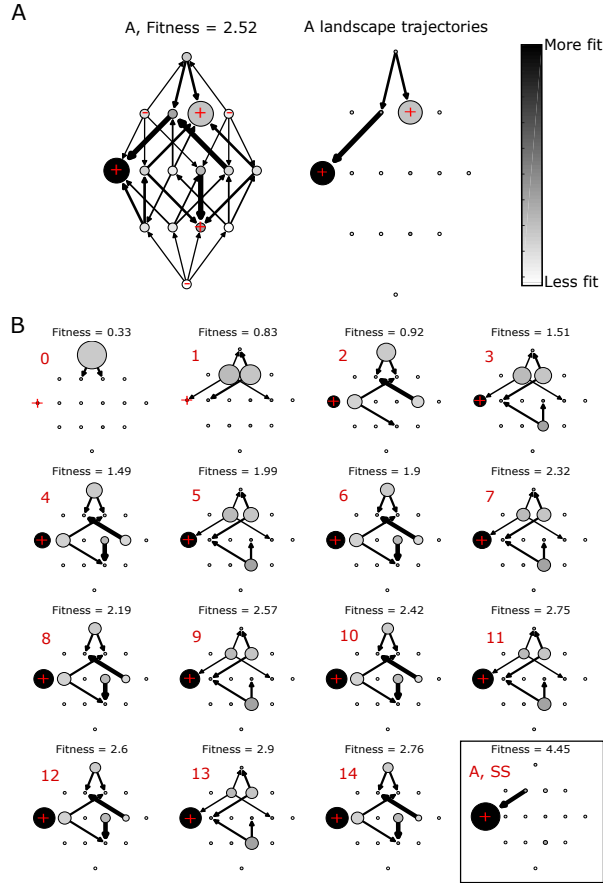


FIG. S7: Evolutionary dynamics in alternating landscapes with positively correlated fitness peaks. A. Left panel: network representation of adaptation on a static landscape (environment A) of size $N = 4$. Each circle represents a genotype (ancestral genotype at the top), with shading indicating the relative fitness of that genotype and size representing the occupation probability in the steady state. Red + symbols mark genotypes corresponding to local fitness maxima. Arrows represent transitions between genotypes that occur with nonzero probability—that is, the entries of the transition matrix. The width of the arrow represents the magnitude of the transition probability. Right panel: same as left panel, but showing only transitions that occur during adaptation starting from the ancestral genotype (top circle). B. Network representations of adaptation (at different time points) in alternating landscapes with positively correlated fitness peaks. Red number above each landscape represents the current evolutionary time point (ranging from 0 to SS, indicating steady state of approximately 200 steps). Directed arrows represent possible transitions between genotypes based on the current genotype distribution (indicated by the circle sizes) and the current landscape (A or B). Average fitness at each time point (calculated over the current genotype distribution) are listed above each plot. Even numbered steps correspond to landscape A, odd to landscape B.

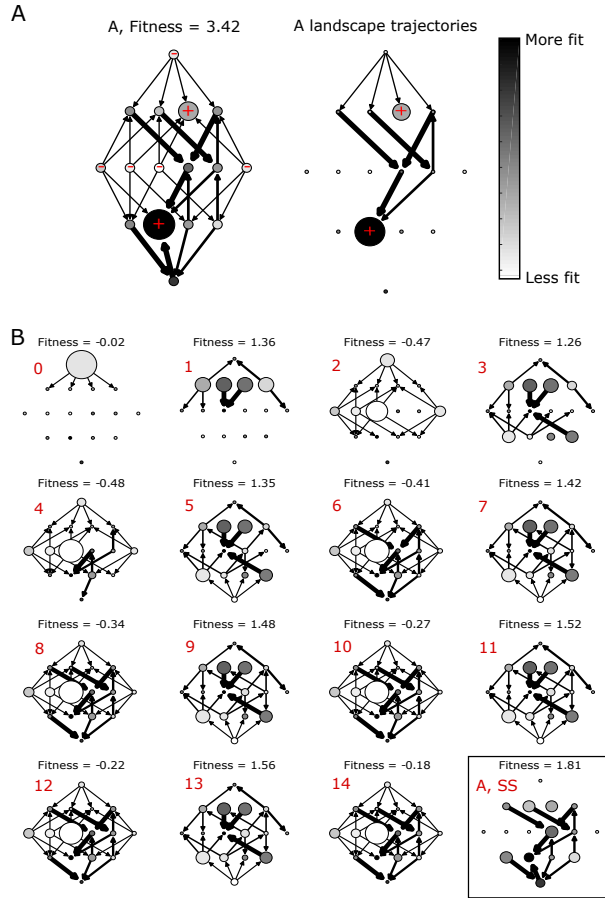


FIG. S8: Evolutionary dynamics in alternating landscapes with negatively correlated fitness peaks. A. Left panel: network representation of adaptation on a static landscape (environment A) of size $N = 4$. Each circle represents a genotype (ancestral genotype at the top), with shading indicating the relative fitness of that genotype and size representing the occupation probability in the steady state. Red + symbols mark genotypes corresponding to local fitness maxima. Arrows represent transitions between genotypes that occur with nonzero probability. The width of the arrow represents the magnitude of the transition probability. Right panel: same as left panel, but showing only transitions that occur during adaptation starting from the ancestral genotype (top circle). B. Network representations of adaptation (at different time points) in alternating landscapes with negatively correlated fitness peaks. Red number above each landscape represents the current evolutionary time point (ranging from 0 to SS, indicating steady state of approximately 200 steps). Directed arrows represent possible transitions between genotypes based on the current genotype distribution (indicated by the circle sizes) and the current landscape (A or B). Average fitness at each time point (calculated over the current genotype distribution) are listed above each plot. Even numbered steps correspond to landscape A, odd to landscape B.