

INDUCTION OF BOTH CYTOCHROMES P-450 AND P-448
BY 2,3',4,4',5-PENTABROMOBIPHENYL,
A COMPONENT OF FIREMASTER

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Received November 26, 1979

SUMMARY

The synthesis and purification of a component of fireMaster BP-6 and fireMaster FF-1, 2,3',4,4',5-pentabromobiphenyl, is described. The compound was found to be a potent inducer of liver microsomal drug-metabolizing enzymes in the rat, enhancing those enzymic activities induced by both phenobarbitone and 3-methylcholanthrene (i.e. cytochromes P-450 and P-448). The pentabromobiphenyl enhanced the activities of benzo[a]pyrene hydroxylase, dimethylaminoantipyrene N-demethylase and NADPH-cytochrome c reductase. The hepatic cytochromes b₅ and P-450 were increased and the Soret peak maximum of the latter was shifted to 448.5 nm. The relative peak intensities and spectral shifts for the ethylisocyanide-binding difference spectra confirmed the mixed induction characteristics of 2,3',4,4',5-pentabromobiphenyl.

INTRODUCTION

In 1973 and 1974 large quantities of an industrial flame retardant (fireMaster), containing among other chemicals a complex mixture of polybrominated biphenyls (PBBs) were inadvertently introduced into Michigan's food supply. Exposure of farm animals, farm families and the consuming public to fireMaster followed (1,2). The need to identify the composition, the biologic effects and the persistence of fireMaster became apparent.

Induction of the hepatic microsomal drug-metabolizing enzymes has been widely used to compare the biologic effects of various chemicals. Inducers

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Abbreviations: PBEs - polybrominated biphenyls, PB-phenobarbitone, MC-3-methylcholanthrene, NMR-nuclear magnetic resonance, PBBp-2,3',4,4',5-pentabromobiphenyl, GC-gas chromatographic, TBBp-2,3',4',5-tetrabromobiphenyl, B[a]P-benzo[a]pyrene, EIC-ethylisocyanide, DMAP-4-dimethylaminoantipyrene, CO-carbon monoxide

of these enzymes are often categorized as phenobarbitone(PB)-type inducers of cytochrome P-450- or 3-methylcholanthrene(MC)-type inducers of cytochrome P-448-dependent monooxygenase activities. In rats and mice fireMaster is a potent inducer of hepatic microsomal enzymes, producing a pattern of induction consistent with the simultaneous induction of cytochromes P-450 and P-448 (3,4).

For several classes of halogenated aromatic hydrocarbons [chlorinated biphenyls (5,6), dibenzo-p-dioxins and dibenzofurans (7), azobenzenes and azoxybenzenes (8)] there appears to be a good correlation between the toxicity of the chemical and its ability to induce cytochrome P-448. It is of particular interest, therefore, to identify those brominated biphenyls present in fireMaster which are capable of inducing cytochrome P-448.

Only one substance in fireMaster, namely the 2,3',4,4',5,5'-hexabromo-biphenyl, has been found capable of inducing cytochrome P-448 (9).

This study describes the synthesis and purification of a component of fireMaster which is a potent inducer of cytochrome P-448.

METHODS

(syntheses)

2,4,5-Tribromoaniline was synthesized from 2,5-dibromoaniline (Aldrich) by bromination with liquid bromine in carbon tetrachloride in the presence of iron metal filings. Various batches yielded products between 90 and 99.5% pure after methanol/water recrystallization. The major impurity in each batch was the starting material. The proton nuclear magnetic resonance (NMR) spectrum of the product showed 3 singlets at 4.13, 6.98 and 7.57 ppm.

2,3',4,4',5-Pentabromobiphenyl(PBBp) was synthesized by the diazo coupling (10) of 2,4,5-tribromoaniline and *o*-dibromobenzene (Aldrich). The products were purified by alumina/Florisil column chromatography to remove possible brominated dibenzofuran impurities followed by repeated thin-layer chromatography on silica gel HF₂₅₄ to separate the two major products, PBBp and 2,2',3',4,5-pentabromobiphenyl. PBBp was further purified by methanol recrystallization. The NMR proton spectrum confirmed the identification of PBBp as 2,3',4,4',5-pentabromobiphenyl with singlets at 7.92 and 7.54 ppm, doublets at 7.63 [J = 2.0 Hz] and 7.68 [J = 8.2 Hz] ppm and a quartet centered at 7.17 ppm. The proton chemical shifts for PBBp are consistent with published data (11).

By gas chromatographic (GC) response and by comparison with known standards, PBBp was found to be 96.14% pure and was contaminated with the following: 2.83% 2,3',4',5-tetrabromobiphenyl (TBBp), 0.82% 2,2',3',5-tetrabromobiphenyl with 0.21% unidentified.

A sample of the major impurity, TBBp, was kindly supplied by Drs. G. Sundström and O. Hutzinger. The synthetic TBBp was found by GC response to be greater than 99% pure and was used without further cleanup. The synthesis, purification and identification of TBBp have been described (12).

(Biochemicals)

Cytochrome c (horse heart, type III), NADP⁺, NADPH, α -D-glucose-6-phosphate, α -D-glucose-6-phosphate dehydrogenase (Baker's yeast), MC, benzo[a]pyrene (B[a]P) and ethylisocyanide (EIC) were purchased from Sigma Chemical Co.; 4-dimethylaminoantipyrine (DMAP) from Aldrich Chemical Co.; carbon monoxide (CO) (research purity) from Matheson, and PB from the Ontario Veterinary College, Guelph. [³H]-B[a]P was obtained from New England Nuclear Co. and purified by thin-layer chromatography using hexane as eluant.

(Animal treatment and isolation of microsomes)

One month old male Wistar rats, average weight 100 g, were housed in wire cages and allowed free access to Purina Certified Rodent Chow #5002 and water. PBBp was dissolved in corn oil and doses of 30 $\mu\text{mol Kg}^{-1}$ (low dose) and 150 $\mu\text{mol Kg}^{-1}$ (high dose) were administered to four animals at each dose level by intraperitoneal injection (ca. 0.5 ml) on days 1 and 3. The animals were killed by cervical dislocation on day 6. TBBp was administered in a similar manner only at the 150 $\mu\text{mol Kg}^{-1}$ dose level. PB (400 $\mu\text{mol Kg}^{-1}$) dissolved in isotonic saline and MC (100 $\mu\text{mol Kg}^{-1}$) dissolved in corn oil were administered (ca. 0.5 ml) individually as well as coadministered to animals on days 1 and 2 and the animals killed on day 3. Animals injected with corn oil (ca. 0.5 ml) served as controls. All animals were fasted over the last 24 hours to lower liver glycogen levels.

(Assays)

In all assays the final concentration of microsomal protein was 1.0 mg ml^{-1} as determined by the method of Lowry *et al.* (13). The cytochrome P-450 content was determined by the method of Omura and Sato (14), from the CO-difference spectrum of dithionite-reduced microsomes using an extinction coefficient of 91 $\text{cm}^{-1} \text{mM}^{-1}$ between A_{max} and A_{490} . The EIC-difference spectrum was determined in a similar manner to the CO-difference spectrum except that EIC was added to the sample cuvette (final concentration 4.5 mM) instead of CO. The concentration of cytochrome b_5 was determined from the difference spectrum between NADH-reduced microsomes and oxidised microsomes (15) using the corrected extinction coefficient of 185 $\text{cm}^{-1} \text{mM}^{-1}$ (14). All spectra were recorded on a Cary 118C spectrophotometer with a repetitive scan accessory. Holmium oxide was used to calibrate all spectra.

The rate of oxidative N-demethylation of DMAP was measured by quantifying the production of formaldehyde as described (16). The formaldehyde, trapped as the semicarbazone, was developed in double strength Nash reagent (17). The rate of B[a]P hydroxylation was measured by the radiometric assay of DePierre *et al.* (18) as improved by Nesnow *et al.* (19), by quantifying the base-soluble metabolites following hexane-extraction of the unreacted B[a]P. Because an NADPH-regenerating system was employed in both metabolic assays, all tubes were preincubated for 15 minutes. The activity of NADPH-cytochrome P-450 reductase was measured by the rate of reduction of cytochrome c (20). To prolong the linearity of the cytochrome c reduction, samples with high NADPH-cytochrome P-450 reductase activity were diluted to a final microsomal protein concentration of 0.2 mg ml^{-1} .

RESULTS

The effects of pretreatment with corn oil, PB, MC, PB plus MC, PBBp and TBBp on the hepatic drug-metabolizing enzymes are shown in Table 1. The activity of DMAP N-demethylase and of NADPH-cytochrome c reductase was increased by pretreatment with PB, PBBp and TBBp but not by MC. PBBp induced these enzymic activities more than TBBp. Since TBBp was administered at more than 30-times the concentration injected as a contaminant in PBBp, the results suggest that PBBp is itself a PB-type inducer of cytochrome P-450-dependent enzymic activity.

Pretreatment with PB and TBBp increased the activity of B[a]P hydroxylase by 3.2- and 2.8-fold, respectively. In contrast, the activity was stimulated 15.2-fold by MC-pretreatment and 17.1-fold by pretreatment with PBBp at the high dose level. At the lower dose level of $30 \mu\text{mol Kg}^{-1}$, PBBp-pretreatment enhanced the activity of B[a]P hydroxylase by 14.7-fold indicating that PBBp was a potent inducer of cytochrome P-448-dependent monooxygenase activity.

The concentrations of cytochrome b_5 and cytochrome P-450 were increased by PBBp-pretreatment. The increase in cytochrome P-450 was accompanied by a hypsochromic shift in the CO-difference spectrum from 450.0 nm to 448.5 nm. Both Soret peaks of the EIC-difference spectrum were shifted: from 428.0 to 428.6 nm and from 455.0 to 453.0 nm. The ratio of the two Soret peak heights (455:428) increased from 0.49 (controls) to 1.1 and 1.5 for the low and high dose, respectively. The qualitative spectral characteristics displayed by PBBp-induced microsomes were intermediate between PB- and MC-induced microsomes.

The pattern of enzymic activities and the spectral characteristics of PBBp-induced microsomes were simulated by the coadministration of PB and MC. This indicates that PBBp-pretreatment resulted in a mixed-type induction of both cytochromes P-450 and P-448.

TABLE I
THE EFFECTS OF BROMOBIPHENYL ISOMERS AS HEPATIC MICROSOMAL ENZYME INDUCERS IN MALE WISTAR RATS.

Treatment	% Liver Wt. of Body Wt.	mg Protein/g Liver-1	Benzo[a]pyrene Hydroxylase nmol B[a]P me- tabolized/mg protein-1min-1	Dimethylamino- antipyrene N-demethylase nmol HCHO formed mg protein-1min-1	NADPH- Cytochrome c reductase nmol mg pro- tein-1min-1	Cytochrome b ₅ pmol mg pro- tein-1	Cytochrome "P-450" nmol mg pro- tein-1 (peak maximum)	Ethylisocyanide Difference Spectra peak maxima (nm)	455nm to 428nm peak heights
Corn Oil (Control)	4.10 ± 0.31	17.6 ± 1.4	155 ± 17	3.45 ± 0.27	58.4 ± 7.2	233 ± 18	0.640 ± 0.032 (450.0)	428.0, 455.0	0.49 ± 0.04
Phenobarbitone (PB)	5.21 ± 0.46	27.4 ± 3.7	490 ± 43	9.89 ± 0.76	165 ± 13	350 ± 30	1.63 ± 0.09 (450.0)	428.0, 455.0	0.60 ± 0.07
3-Methylcholanthrene (MC)	4.55 ± 0.28	19.3 ± 2.1	2350 ± 180	4.00 ± 0.31	61.4 ± 9.1	322 ± 26	1.28 ± 0.10 (448.0)	429.7, 452.0	1.9 ± 0.2
PB + MC	5.46 ± 0.37	25.0 ± 2.9	2450 ± 210	10.1 ± 0.9	172 ± 18	387 ± 31	2.18 ± 0.17 (448.5)	428.5, 452.0	1.2 ± 0.2
2,3',4',4',5-Penta- bromobiphenyl (PB5p)	4.59 ± 0.20†	19.7 ± 3.3	2280 ± 60	5.40 ± 0.44	80.1 ± 6.4	406 ± 49	1.46 ± 0.12 (448.6)	428.6, 453.0	1.1 ± 0.1
	5.73 ± 0.34*	20.6 ± 1.1	2650 ± 180	6.33 ± 1.10	131 ± 12	423 ± 39	2.27 ± 0.51 (448.5)	428.6, 453.0	1.5 ± 0.1
2,3',4',5-Tetra- bromobiphenyl (TB5p)	4.44 ± 0.25*	23.0 ± 0.9	437 ± 130	4.05 ± 0.65	76.8 ± 6.5	290 ± 36	0.890 ± 0.076 (450.0)	428.0, 455.0	0.57 ± 0.11

† Low dose

* High dose

DISCUSSION

FireMaster BP-6 and FF-1 are commercial mixtures of brominated biphenyls possessing both PB and MC inducing characteristics. Two components, comprising more than 80% of fireMaster, namely the 2,2',4,4',5,5'-hexabromobiphenyl and 2,2',3,4,4',5,5'-heptabromobiphenyl, were PB-type inducers (21, 22).

Aust and coworkers reported that 2,3',4,4',5,5'-hexabromobiphenyl, isolated from fireMaster, was a mixed-type inducer and thereby identified the first component of fireMaster which causes MC-type effects (9). In this report PBBp has also been shown to be a mixed-type inducer. Using the activity of B[a]P hydroxylase as an index of cytochrome P-448-dependent monooxygenase activity, it can be concluded that PBBp is a potent inducer of MC-type characteristics with almost maximum effects observed at the relatively low dose of 30 $\mu\text{mole Kg}^{-1}$. Recently the chloro analog of PBBp, the 2,3',4,4',5-pentachlorobiphenyl, was shown in our laboratory to be a mixed-type inducer. The properties of PBBp, which together with the other pentabromobiphenyl, 2,2',4,5,5'-pentabromobiphenyl, comprises up to 8% of fireMaster mixtures (23), strongly suggests that PBBp contributes significantly to the MC-type characteristics of the commercial PBB mixtures.

The identification of PBBp as a mixed-type inducer may be significant not only in terms of explaining the inducing properties of fireMaster but also in terms of toxicity. It has been suggested for several classes of halogenated aromatic hydrocarbons, including polychlorinated biphenyls, that a correlation exists between various toxic responses to a chemical and its ability to induce cytochrome P-448. It has yet to be shown whether this generalization can be extended to include the brominated biphenyls.

Pharmacokinetic studies *in vivo* have revealed that the individual PBBs in fireMaster are eliminated from the rat at different rates (24). Due to this differential rate of elimination, those components more slowly removed from the body appear to become more concentrated relative to the other PBB isomers and congeners. PBBp is among those components which apparently

concentrate in mammals. Metabolic studies *in vitro* produced similar results in that, due to differential rates of metabolism, PBBp, as well as other poorly metabolized components of fireMaster, appeared to concentrate within the PBB mixture (25). These results show that PBBp is a persistent chemical. This correlates with reports which have demonstrated that 2,3',4,4',5-pentachlorobiphenyl, a component of commercial PCBs, preferentially bioconcentrates in human adipose tissue and breast milk (26).

In conclusion, PBBp has been identified as mixed-type inducer of both cytochromes P-450 and P-448. The present data concerning the toxicity of several classes of halogenated aromatic hydrocarbons suggest that PBBp is potentially toxic. It is interesting to speculate that the persistence of PBBp in mammals together with its potential toxicity may explain in part why PBBs derived from contaminated beef and poultry were more toxic to mink than the original fireMaster (27).

ACKNOWLEDGEMENTS

The authors would like to thank Dr. Albert Woon-Fat for his help in obtaining the NMR spectral data and the Research Programs Directorate, Health and Welfare Canada and the National Cancer Institute (U.S.A.), DHEW, Grant No. 1 B01 CA21814-01 for financial support.

REFERENCES

1. Robertson, L. W. and Chynoweth, D. P. (1975). *Environment* 17, 25-27.
2. Carter, L. J. (1976). *Science* 192, 240-243.
3. Moore, R. W., Dannan, G. A. and Aust, S. D. (1978). *Environ. Health Perspect.* 23, 159-165.
4. Ahotupa, N. and Aitio, A. (1978). *Toxicol.* 11, 309-314.
5. McKinney, J. D., Chae, K., Gupta, N., Moore, J. A. and Goldstein, J. A. (1976). *Toxicol. Appl. Pharmacol.* 36, 65-80.
6. Poland, A. and Glover, E. (1977). *Mol. Pharmacol.* 13, 924-938.
7. Poland, A. and Glover, E. (1973). *Mol. Pharmacol.* 9, 736-747.
8. Poland, A., Glover, E., Kende, A. S., DeCamp, M. and Giandomenico, C. M. (1976). *Science* 194, 627-630.
9. Dannan, G. A., Moore, R. W., Besaw, L. C. and Aust, S. D. (1978). *Biochem. Biophys. Res. Comm.* 85, 450-458.
10. Cadogan, J. I. G. (1962). *J. Chem. Soc. (London)*, 4257-4258.
11. Moore, R. W. and Aust, S. D. (1978). *Biochem. Biophys. Res. Comm.* 84, 936-942.

12. Sundström, G., Hutzinger, O., Safe, S. and Zitko, V. (1976). *Sci. Total Environ.* 6, 15-29.
13. Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. (1951). *J. Biol. Chem.* 193, 265-275.
14. Omura, T. and Sato, R. (1964). *J. Biol. Chem.* 239, 2370-2378.
15. Raw, I. and Mahler, H. R. (1959). *J. Biol. Chem.* 234, 1867-1873.
16. Parkinson, A. and Safe, S. (1979). *J. Pharmac. Pharmacol.* 31, 444-447.
17. Nash, T. (1953). *Biochem. J.* 55, 416-421.
18. DePierre, J. W., Moron, M. S., Johannesen, K. A. M. and Ernster, L. (1975). *Anal. Biochem.* 63, 470-484.
19. Nesnow, S., Fahl, W. E. and Jefcoate, C. R. (1977). *Anal. Biochem.* 80, 258-266.
20. Williams, C. H. and Kamin, H. (1962). *J. Biol. Chem.* 237, 587-595.
21. Moore, R. W., Sleight, S. D. and Aust, S. D. (1978). *Toxicol. Appl. Pharmacol.* 44, 309-321.
22. Moore, R. W., Sleight, S. D. and Aust, S. D. (1979). *Toxicol. Appl. Pharmacol.* 48, 73-86.
23. DeKok, J. J., KeKok, A., Brinkman, U. A. Th. and Kok, R. M. (1977). *J. Chromatogr.* 142, 367-383.
24. Wolff, M. S. and Selikoff, I. J. (1979). *Bull. Environ. Contam. Toxicol.* 21, 771-774.
25. Dannan, G. A., Moore, R. W. and Aust, S. D. (1978). *Environ. Health Perspect.* 23, 51-61.
26. Yakushiji, T., Watanabe, I., Kuwabara, K., Yoshida, S., Koyama, K. and Kunita, N. (1979). *Int. Arch. Occup. Environ. Health* 43, 1-15.
27. Aulerich, R. J. and Ringer, R. K. (1979). *Arch. Environ. Contam. Toxicol.* 8, 487-498.