THE ROLE OF HEME SYNTHESIS DURING THE INDUCTION OF HEPATIC MICROSOMAL CYTOCHROME P-450 AND DRUG METABOLISM PRODUCED BY BENZPYRENE

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Summary

 δ -Aminolevulinic acid synthetase, the initial and rate limiting step in hepatic heme synthesis, is induced by both benzpyrene and phenobarbital. Induction of this enzyme by benzpyrene results in the stimulation of glycine-2- ^{14}C incorporation into hepatic microsomal heme in vivo and in the induction of cytochrome P-450 and N-demethylase activity. 3-Amino-1,2,4-triazole, an inhibitor of the second step in hepatic heme synthesis, prevents the stimulation of hepatic heme synthesis and the induction of P-450 and N-demethylase activity. It is suggested that induction of δ -aminolevulinic acid synthetase leading to increases in hepatic heme synthesis may be the mechanism by which benzpyrene induces cytochrome P-450 and certain hepatic microsomal oxidations.

Pretreatment of animals with 3,4-benzpyrene (BP) and other polycyclic aromatic hydrocarbons such as 3-methylcholanthrene results in increased hepatic microsomal levels of cytochrome P-450 and in the induction of the hepatic microsomal oxidations of certain drugs and carcinogens (1-5). Although the exact mechanism by which induction occurs remains to be elucidated, stimulation of both heme synthesis and protein synthesis are undoubtedly essential to the induction process since, at least during the initial stages of induction, there appears to be a good correlation between increases in P-450 and certain microsomal oxidations (10).

Hepatic mitochondrial δ -aminolevulinic acid synthetase (ALA synthetase) is the initial and rate limiting step in hepatic heme synthesis (6,7). Phenobarbital (PB) has been demonstrated to induce ALA synthetase in chick embryo liver cells in culture and in the livers of experimental animals (6,8,9). Previous studies in this laboratory (10) have suggested that PB might exert its stimulatory effect on drug metabolism by increasing the rate of hepatic heme synthesis which directs the increased synthesis of cytochrome P-450. Since the

administration of BP also results in increases in the level of cytochrome P-450 in hepatic microsomes, stimulation of hepatic heme synthesis via induction of ALA synthetase is also a possible mechanism for the induction of the hepatic microsomal drug-metabolizing system by BP.

The herbicide 3-amino-1,2,4-triazole (AT), an inhibitor of the second step in hepatic heme synthesis, has been employed to inhibit hepatic heme synthesis and to prevent PB from stimulating hepatic heme synthesis (10). AT has also been demonstrated to inhibit the induction of certain hepatic microsomal oxidations produced by either PB (10,11,12) or by methylcholanthrene (11).

The present report describes the stimulatory effect of BP on rat hepatic ALA synthetase, hepatic heme synthesis, cytochrome P-450 synthesis and on the N-demethylation of 3-methyl-4-monomethylaminoazobenzene (3-CH $_3$ -MAB). The inhibitory effect of AT on these inductions, and the effects of both BP and AT on the microsomal levels of cytochrome b $_5$ and NADPH-cytochrome c reductase activity are also described.

Methods

Male Holtzman rats weighing 130 to 170 g were used throughout these studies. Drugs were administered intraperitoneally in the following doses: BP, 20 mg/kg in corn oil; AT, 3 g/kg in saline; and PB, 40 mg/kg in saline. Control animals received an equal volume of saline and/or corn oil. Rats were fasted for 24 hr prior to sacrifice but were given water ad libitum. In all experiments, the livers from two rats which received identical treatments were pooled upon homogenization. ALA synthetase activity was determined in hepatic homogenates by a modification of the method of Marver et al. (13). Methods for the preparation of microsomes and measurements of the N-demethylation of 3-CH₃-MAB, the activity of NADPH-cytochrome c reductase, the content of cytochromes P-450 and b₃ and the incorporation of isotopic precursor into hepatic microsomal heme have been described previously (10).

Results

BP administration results in the induction of microsomal cytochrome

Table 1. Effects of benzpyrene (BP) and aminotriazole (AT) on the incorporation of glycine-2-14C into hepatic microsomal heme in vivo. 1

Pretreatment	Percent of control 2
вР	131.8 \pm 1.7 ³
AT	86.1 ± 2.8 ³
BP + AT	109.0 \pm 2.1

Rats received intraperitoneally a 45-min. pulse-dose of 30 μC of glycine-2-14C 16 hr. after the administration of drugs. The control value was 196.9 cpm/mμmole of protoheme.

P-450 and the induction of certain microsomal oxidations (1,4,5). If this phenomenon is related to an increase in the synthesis of cytochrome P-450, a stimulation of hepatic microsomal heme synthesis should be observed after the administration of BP to rats. Table 1 shows that there is a 32% stimulation of the in vivo incorporation of glycine-2-14°C into hepatic microsomal heme 16 hr after BP administration. This stimulation is prevented by AT, an inhibitor of ALA dehydratase (10,14), the second step in hepatic heme synthesis. AT has previously been observed to inhibit the PB-induced increase in hepatic heme synthesis (10).

The mechanism for the induction of heme synthesis and cytochrome P-450 may be related to an increase in ALA synthetase activity. Table 2 shows that both BP and PB produce a marked induction of hepatic ALA synthetase activity 16 hr after administration to rats. The induction is not affected by AT, nor does AT alone alter the level of activity of this enzyme. This indicates that both BP and PB induce cytochrome P-450 by stimulating hepatic microsomal heme synthesis via induction of ALA synthetase. PB has also been shown to stimulate

 $^{^{2}}$ Values represent the mean \pm S.E.M. of at least three experiments.

 $^{^{3}}$ p < 0.05.

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Pretreatment	mµmoles ALA formed/ g of liver/hr	Percent of control
Control	15.3 ± 0.4 ²	
AT	16.0 ± 0.5	104.9 <u>+</u> 1.7
ВР	79.8 <u>+</u> 9.7	522.4 <u>+</u> 15.8 ³
BP + AT	74.3 ± 12.1	519.2 ± 23.6^{3}
PB	225.2 <u>+</u> 28.7	1449.0 ± 87.1^{3}
PB + AT	193.8 <u>+</u> 15.8	1281.5 ± 52.6^{3}

Table 2. Effects of benzpyrene (BP), phenobarbital (PB), and aminotriazole (AT) pretreatments on hepatic ALA synthetase activity. 1

synthesis of microsomal heme (10) and AT inhibits this stimulation. Treatment of rats with actinomycin D (3 mg/kg) at the same time as either BP or PB completely prevented the induction of ALA synthetase.

Table 3 shows that BP induces cytochrome P-450 and the N-demethylation of 3-CH₃-MAB. That these effects are due to an enhancement of hepatic microsomal heme synthesis is indicated from the observation that AT significantly antagonizes the increases of both P-450 content and 3-CH₃-MAB N-demethylase activity. The effects of AT do not result from either interference with the assay or from a direct inhibition (10). However AT begins to lose its effect after the first 24 hr. This is likely due to increased levels of endogenous ALA which would reverse the inhibition.

Although BP pretreatment results in an enhancement of hepatic heme synthesis and an increase in cytochrome P-450 levels, there was no observable effect by either BP or AT on the hepatic microsomal level of cytochrome b_5 during the first 48 hr of treatment (Table 3). This is similar to the effect

Drugs were administered 16 hr. prior to sacrifice.

²Values represent the mean <u>+</u> S.E.M. of at least three experiments.

 $^{^{3}}p < 0.05.$

Table 3.	Effects of	benzpyrene (BP) and a	minotriazole	(AT) p:	retreatments	on
	microsomal	hemoproteins	, 3-CH ₂ -M	AB N-demethy:	lase an	d NADPH-cyto	chrome
	c reductase	≥. ¹	3				

Measurement	Duration of Treatment	Percent of control 2			
		ВР	AT	BP + AT	
P-450 3	24	164 + 13*	70 + 5*	114 + 8	
	48	229 <u>+</u> 19*	70 <u>+</u> 5* 78 <u>+</u> 3*	114 ± 8 162 ± 15	
Cytochrome b ₅ 4	24	123 + 23	101 + 15	126 + 19	
	48	$ \begin{array}{r} 123 \pm 23 \\ 127 \pm 22 \end{array} $	117 ± 31	124 ± 16	
3-CH ₂ -MAB	24	151 ± 17*	76 + 5 *	115 + 18	
3-CH ₃ -MAB N-demethylase ⁵	48	$212 \pm 21*$	76 ± 10*	153 ± 12	
NADPH-cytochrome	24	113 ± 10	116 ± 10	119 + 9	
c reductase 6	48	121 ± 13	113 🛨 13	119 ± 9 138 ± 17	

¹ Drugs were administered every 24 hr.

of PB (10,15). Unlike PB, BP does not induce NADPH-cytochrome c reductase activity (Table 3), the microsomal flavoprotein which may be responsible for the reduction of cytochrome P-450. This may indicate that in those oxidations induced by BP, NADPH-cytochrome c reductase is not a rate limiting component. On the other hand, NADPH-cytochrome c reductase activity may not be representative of NADPH-cytochrome P-450 reductase activity (16).

Discussion

Within 24 hr after the administration of BP to rats there is an increase in the level of microsomal cytochrome P-450 and an induction of certain hepatic microsomal oxidations (1,4,5). The proximal mechanism by which BP and related compounds produce this effect remains to be elucidated. However, the present

 $^{^2}$ Values represent the mean \pm S.E.M. of at least three experiments.

 $^{^3}$ Control value was 0.023 \triangle OD 450-490 m μ /mg protein.

⁴Control value was 0.096 mµmoles cytochrome b_s/mg protein.

⁵Control value was 0.69 mµmoles HCHO formed/mg protein/min.

 $^{^6}$ Control value was 42 mµmoles cytochrome c reduced/mg protein/min.

^{*}p < 0.05.

studies suggest that, like PB (10), BP produces a stimulation of hepatic microsomal heme synthesis through the enhancement of ALA synthetase activity. This leads to an increased synthesis of cytochrome P-450 and the increase in certain microsomal oxidations. However, the species of cytochrome P-450 induced by BP and related compounds may be different from that induced by PB (2,3, 17) which may reflect the fact that certain protein moieties confer specificity to P-450. BP and PB may also influence the synthesis of these apocytochromes.

PB has been demonstrated to induce the rate controlling step in hepatic heme synthesis, ALA synthetase (6,8,9), and has also been demonstrated to stimulate hepatic microsomal heme synthesis (9,10). The present studies have demonstrated that BP similarly produces an induction of ALA synthetase and a stimulation of hepatic microsomal heme synthesis. The latter event is abolished by AT, although AT does not prevent the induction of ALA synthetase. Thus, it appears that both PB and BP stimulate hepatic heme synthesis through the induction of ALA synthetase.

Further support for the postulate that induction of the drug-metabolizing system is dependent upon heme synthesis is seen from the data on the induction of cytochrome P-450 and 3-CH₃-MAB N-demethylation by BP. When AT is administered together with BP, the stimulatory effects of BP are significantly reduced or prevented. Since AT exerts its effect on heme synthesis exclusive of an effect on protein synthesis and since any direct inhibitory effect of AT on microsomal drug metabolism is minimal (10), these results suggest that the stimulation of hepatic heme synthesis may be the mechanism by which BP induces the hepatic microsomal drug-metabolizing system.

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References

1. Silverman, D.A. and Talalay, P. Mol. Pharmacol., 3, 90 (1967).

- Sladek, N.E. and Mannering, G.J. Biochem. Biophys. Res. Commun., 24, 668 (1966).
- Alvares, A.P., Schilling, G., Levin, W., and Kuntzman, R. Biochem. Biophys. Res. Commun., 29, 521 (1967).
- Arcos, J.C., Conney, A.H., and Buu-Hoi, N.P. J. Biol. Chem., 236, 1291 (1961).
- Conney, A.H., Davison, C., Gastel, R., and Burns, J.J. J. Pharmacol. Exptl. Therap., <u>130</u>, 1 (1960). 6. Granick, S. J. Biol. Chem., <u>241</u>, 1359 (1966).

- 7. Granick, S. and Urata, G. J. Biol. Chem., 238, 821 (1963).

 8. Marver, H.S., Schmid, R., and Schutzel, H. Biochem. Biophy Commun., 33, 969 (1968). Biochem. Biophys. Res.
- Wada, O., Yano, Y., Urata, G., and Nakao, K. Biochem. Pharmacol., 17, 595 (1968).
- Baron, J. and Tephly, T.R. Mol. Pharmacol., 5, 10 (1969). 10.
- Kato, R. Jap. J. Pharmacol., 17, 56 (1967). 11.
- 12. Baron, J. and Tephly, T. Federation Proc., $\underline{27}$, 301 (1968).
- Marver, H.S., Tschudy, D.P., Perlroth, M.G., and Collins, A. J. Biol. Chem., 241, 2803 (1966).
- Tschudy, D.P. and Collins, A. Science, 126, 168 (1957).
- Ernster, L. and Orrenius, S. Federation Proc., 24, 1190 (1965). 15.
- 16. Gigon, P.L., Gram, T.E., and Gillette, J.R. Biochem. Biophys. Res. Commun., 31, 558 (1968).
- 17. Hildebrandt, A., Remmer, H., and Estabrook, R.W. Biochem. Biophys. Res. Commun., 30, 607 (1968).