Maintenance of Respiratory Control in Mitochondria after Rate Zonal Centrifugation

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Abstract

The respiratory control of rat liver mitochondria is lost when they are subjected to rate zonal centrifugation in a sucrose gradient (8.0% to 46.6%, w/w) at values for $\omega^2 t$ necessary for resolution. High sucrose concentration and high $\omega^2 t$ are both responsible. Respiratory control can be maintained in iso-osmotic Ficoll + 8.3% sucrose media, and after zonal centrifugation in such media at values of $\omega^2 t$ sufficient for resolution.

Introduction

We have observed that rat liver mitochondria prepared by the usual differential centrifugation in 8.3% (0.25 M) sucrose lose respiratory control during a subsequent rate zonal centrifugation through a sucrose gradient (8% w/w to 46.6\%). Little reference has been made to changes in mitochondrial oxidative phosphorylation after zonal centrifugations, although "damage" has been recognized from electron micrographs and from the solubilizations of enzymes [1, 2]. To extend the usefulness of the zonal centrifugation method, we have studied the effects of suspending medium and of force on oxidative phosphorylation, and find that respiratory control can be preserved in an iso-osmotic Ficoll gradient at low rotor speeds sufficient for resolution.

Materials and Methods

Rat liver was pulped and then homogenized in 8.3% sucrose, pH 7.4. Heavy debris and cells were removed by spinning twice at $600 g_{av}$ for 5 min in a refrigerated centrifuge. A mitochondrial fraction was

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sedimented at 13,500 g_{av} for 20 min in a \cdot 30 rotor in a refrigerated ultracentrifuge (Spinco Division, Beckman Instruments). The mitochondrial pellet was resuspended in 0.25 M sucrose alone, or in sucrose with 5% or 15% (w/v) Ficoll (a synthetic high copolymer of sucrose and epichlorohydrin, obtained from Pharmacia Fine Chemicals, Inc.), pH 7.4.

Rate zonal centrifugation was then performed according to Price [3]. All mitochondrial samples were adjusted to 8.0% sucrose and introduced into a Ti-14 zonal rotor (Beckman). Sucrose gradients ran from 8.0% (w/w) to 46.6%. Ficoll gradients were from 5% (w/v) to 12\%, and the presence of 8.3% sucrose throughout the gradient maintained an iso-osmotic condition. Gradient concentrations were determined with an Abbe refractometer.

Oxygen consumption was determined polarographically (Oxygraph, Gilson Medical Electronics). The reaction mixture of 3 ml volume, pH 7.4, 25°C, contained 0.25 M sucrose, 7.4 mM KCl, 7.4 mM Tris, 3.0 mM potassium mono- and dihydrogen phosphates, 0.16 mM EDTA, 2.7 mM succinate or glutamate; $58 \,\mu\text{M}$ ADP was added to measure respiratory control (the ratio, State 3 : State 4 respiration) and phosphorylation (the ADP : 0 ratio).

Results

Mitochondrial pellets prepared by differential centrifugation have a respiratory control ratio of 2.75 (substrate, succinate) when suspended in 8.3% sucrose (Table I). Suspension in 40% sucrose abolishes respiratory control, but suspension in 7.5% Ficoll + 8.3% sucrose depresses respiratory control only slightly, by -18%. Sucrose obtained from commercial sources usually contains calcium in significant amounts [4], and concentrated sucrose might thus abolish respiratory control via massive Ca⁺⁺-loading of the mitochondria [5]. However, this does not seem to be the case, because mitochondrial pellets suspended in 40% sucrose containing 1-10 mM EDTA, to lower free [Ca⁺⁺], have no respiratory control. Nor do our 40% sucrose solutions contain enough calcium (0.7 mM) to produce massive loading. The mitochondria in such solutions are exposed to to 0.2 μ moles of Ca⁺⁺ per mg protein; approximately 0.1100 μ moles of Ca⁺⁺ per mg are required to produce even limited loading [6], which does not affect respiratory control. The concentrated sucrose solutions thus probably lower respiratory control through their high osmotic pressures, whereas the Ficoll-sucrose medium is iso-osmotic.

When mitochondria, suspended in 8.0% sucrose, are separated by zonal centrifugation through a sucrose gradient, their capacity for

	Respiratory control					
Mitochondria isolated by:	in 8.3% Sucrose	in 40% Sucrose	in 7.5% Ficoll + 8.3% Sucrose			
Differential centrifugation (13,500 g _{av} × 20 m)	2.75	1.00	2.24			
Rate zonal centrifugation						
$\omega^{2} t (\times 10^{-5})$		1.00				
12.5	1.00	1.00				
1.97	1.00	1.00	1.00			
0.164			2.61			
0.082	2.08					

TABL	E	I.	Effects	of	$\omega^2 t$	and	suspending	medium	on	respiratory	$\operatorname{control}$	in	rat
liver 1	mit	ocl	hondria										

Mitochondrial pellets were prepared by differential centrifugation, suspended in one of three media shown, and assayed for respiratory control (substrate, 2.7 mM succinate). Mitochondrial samples also were prepared, suspended in 8.3% sucrose, and subjected to rate zonal centrifugation through a sucrose gradient (8% to 46.6%) or an iso-osmotic sucrose-Ficoll gradient (5% to 12%). Fractions were obtained at the specified concentrations of medium after sedimentation corresponding to the given values of $\omega^2 t$, and assayed for respiratory control.

respiratory control also depends on the $\omega^2 t$ value (Table I). At $\omega^2 t = 12.5 \times 10^9$ (25,000 rpm × 30 m) or 1.97×10^9 (10,000 rpm × 30 m), which are usual values for resolving mitochondria [3], respiratory control is abolished in the fractions containing 8.3% or 40% sucrose. At low $\omega^2 t$, 0.082 × 10⁹, 75% of the original respiratory control is preserved in the 8.3% sucrose fractions; however, this low force is not sufficient to migrate any mitochondria to the heavy end of the sucrose gradient. In the iso-osmotic sucrose-FicoII gradient fractions at the 7.5% FicoII level have no respiratory control at $\omega^2 t = 1.31 \times 10^9$ (10,000 rpm × 10 m), but show excellent respiratory control at $\omega^2 t = 0.164 \times 10^9$ (5,000 rpm × 5 m). At the latter force-time value, the mitochondria maintain control for at least 1½ h; thereafter respiration in State 4 increases and the ADP : O ratio decreases.

Suspending mitochondria in 5% or 15% Ficoll + 8.3% sucrose has little effect (ca. -20%) on respiratory control or phosphorylation, with either succinate or glutamate as substrate (Table II).

Discussion

High concentrations of sucrose depress mitochondrial respiratory control, probably due to the incident osmotic stress [7]. Ficoll, with

Resuspending medium:	Substrate used:	Respiratory control	ADP : O	
1. 0.25 M sucrose	succinate glutamate	$2.39 \\ 2.24$	$\begin{array}{c} 1.14 \\ 2.40 \end{array}$	
 2. 5% Ficoll + 0.25 M sucrose 3. 15% Ficoll + 0.25 M sucrose 	succinate glutamate succinate glutamate	1.79 2.90 2.06 2.24	$1.20 \\ 2.10 \\ 0.95 \\ 1.87$	

TABLE II. Effects of Ficoll on oxidative phosphorylation in rat liver mitochondria.

Mitochondria were isolated by differential centrifugation at 13,500 $g_{av} \times 20$ min and resuspended in 0.25 M sucrose alone or with 5% or 15% (w/v) Ficoll. Oxidative phosphorylation was measured as described in Methods.

a molecular weight of about 400,000, is relatively inert osmotically, and has minor effects on respiratory control. Although Ficoll gradients at relatively high $\omega^2 t$ are reported to depress glutamate dehydrogenase activity in rat liver mitochondria [8], under the conditions shown here concentrations of Ficoll up to 15% do not change O₂ consumption during glutamate oxidation, although they slightly decrease phosphorylative efficiency. To maintain respiratory control, it is also necessary to use lower centrifugal forces with either a Ficoll or a sucrose gradient. With sucrose, a gradient sufficiently concentrated for resolution cannot be used without introducing osmotic stress. Iso-osmotic Ficoll gradients at low speeds seem useful in resolving functionally intact rat liver mitochondria by zonal centrifugation.

Acknowledgements

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Note added in proof:

Observations that high sucrose concentrations suppress the transport of inorganic phosphate ions across the mitochondrial membranes (D. E. Green, personal communication) provide an alternative explanation for the depression of phosphorylation.