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The gene coding for variant hepatic nuclear factor 1 (Tcf-2), maps between the Edp-1 and Erba genes on mouse Chromosome 11

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Variant Hepatic Nuclear Factor 1 (vHNF1), also called TCF2, HNF1\u03b3, or LFB3, is a nuclear protein that binds to proximal promoter sequences of α and β fibrinogen and albumin. The protein was first identified in dedifferentiated rat hepatoma cell lines (Baumhueter et al. 1988; Cereghini et al. 1988). The mRNA is found in significant levels in adult rat and mouse kidney and in low levels in liver, intestine, and lung (Rey-Campos et al. 1991; De Simone et al. 1991; Mendel et al. 1991). The cDNA corresponding to the rat mRNA has been isolated and sequenced from the dedifferentiated hepatoma H5 cell line (Rey-Campos et al. 1991). The human and mouse cDNAs have also been isolated and share approximately 93% sequence homology with the rat cDNA (Bach et al. 1991; De Simone et al. 1991; Mendel et al. 1991). The gene codes for a homeoprotein homologous to Hepatic Nuclear Factor 1 (HNF1) or LFB1 in regions important for DNA binding and dimerization while the sequence is highly divergent in the transactivation domain. Both HNF1 and vHNF1 act as transcriptional activators of the albumin promoter in transient co-transfection experiments, and these proteins form heterodimers (Rey-Campos et al. 1991). Though HNF1 and vHNF1 are two closely related homeoproteins, they are localized on different chromosomes. HNF1 has been previously mapped to mouse Chr 12 and human Chr 5 (Bach et al. 1990).

Transcription factor 2 (Tcf-2) has been assigned to mouse Chr 11 and human Chr 17 by analysis of somatic cell hybrids (Milatovich et al. 1991). In situ hybridization has localized the gene to human Chr 17q11.2-q21.1 and mouse Chr 11B4-D (Bach et al. 1991). We report chromosomal location of mouse Tcf-2 relative to other previously mapped Chr 11 loci using an intersubspecific mouse backcross. This (DF/B-df/df × CASA/Rk)F₁ × DF/B-df/df backcross has been typed for 12 loci on Chr 11 (Buckwalter et al.

1991). Southern blotting and probe labeling was done by standard methods.

An informative CASA/Rk BamHI polymorphism was detected with the Tcf-2 probe (Fig. 1). A DF/B-df/df specific fragment is located at 7.5 kb and a CASA/Rk specific fragment at 6.8 kb. The results of typing 86 animals for endothelial cell-derived protein (Edp-1; Wolf et al. 1992; Buckwalter et al. 1991), Tcf-2, and Avian erythroblastosis virus oncogene A (Erba) (Lazar et al. 1988; Buckwalter et al. 1991) are given in Table 1. Five recombinants were found between Edp-1 and Tcf-2, placing Tcf-2 5.8 \pm 2.5 cM (centimorgans \pm standard deviation) distal to Edp-1 on Chr 11. Fourteen recombinants were found between Tcf-2 and Erba, placing Tcf-2 16.3 \pm 4.0 cM proximal to Erba on this chromosome. This mapping generates the following gene order: Edp-1-5.8 cM-Tcf-2-16.3 cM-Erba.

This localization corrects the preliminary mapping of *Tcf-2*, also known as *D11Pas3*, in an interspecific backcross with *Mus spretus* performed in the laboratory of J.-L. Guénet (Buchberg et al. 1991). Further analysis of those data places *Tcf-2* between *Csfgm* and *Gfap*, confirming our mapping of *Tcf-2*. In 22 progeny, there was one recombinant between *Csfgm* and *Tcf-2*, and there were four between *Tcf-2* and *Gfap*.

Homeobox-2, (Hox-2) a cluster of homeotic genes on Chr 11, has been mapped 0.7 ± 0.7 cM proximal to Erba (Buchberg et al. 1989). The 95% confidence limits for the Erba-Hox-2 and Erba-Tcf-2 distances are not overlapping. Thus, Tcf-2 is localized outside this cluster of other homeotic genes. This was further confirmed by detection of four recombinants out of 22 progeny typed for both Tcf-2 and Hox-2 in the interspecific backcross with M. spretus mentioned above.

This multipoint molecular genetic localization of *Tcf-2* is consistent with the in situ hybridization of *Tcf-2* to mouse 11B4-D. Our localization of *Tcf-2* corresponds to 11B4 (Buchberg et al. 1991). Our mapping is also consistent with the in situ localization of the human gene to 17q11.2-21.1 because the genes that we

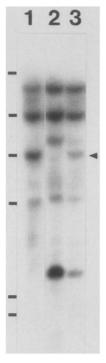


Fig. 1. Southern blot identification of a unique *M. castaneus* (CASA/Rk) RFLP in Tcf-2. An arrowhead signifies the 6.8-kb fragment present in genomic DNA from CASA/Rk (lane 1) and from an animal heterozygous for DF/B-df/df and CASA/Rk (lane 3), but absent in an animal homozygous for DF/B-df/df at this locus (lane 2). Digests were performed with the restriction endonuclease BamHI. Molecular size standard is λ DNA-HindIII fragments.

mapped proximal (*Edp-1*) and distal (*Erba*) to *Tcf-2* also map to human Chr 17. *Edp-1* maps to human 17q22-23 (Wolf et al. 1992), and *Erba* maps to human 17q11.2-21 (Buchberg et al. 1991). The localization of *Tcf-2* near *Edp-1* is consistent with the synteny con-

Table 1. Segregation of three loci on Chr 11 among 86 backcross offspring from the cross (DF/B- $df/df \times CASA/Rk$)F₁ × DF/B-df/df. The symbols D and C represent DF/B-df/df-derived and CASA/Rk-derived alleles, respectively.

| | Loci inherited from F ₁ parent | | | | | Number of |
|-----------------|---|---|-------|---|------|-----------|
| | Edp-1 | | Tcf-2 | | Erba | progeny |
| Nonrecombinants | D | | D | | D | 34 |
| | C | | C | | C | 33 |
| Single | D | | C | | C | 4 |
| recombinants | | × | | | | |
| | C | | D | | D | 1 |
| | D | | D | | C | 5 |
| | | | | × | | |
| | C | | C | | D | 9 |
| | | | _ | | | |

servation and lack of linkage conservation observed between mouse Chr 11 and human Chr 17 (Buchberg et al. 1989).

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