Zinc Inhibition of t-[3 H]Butylbicycloorthobenzoate Binding to the GABA_A Receptor Complex

Akito Kume, Sharin Y. Sakurai, and Roger L. Albin

Department of Neurology, University of Michigan, Ann Arbor, Michigan, U.S.A.

Abstract: The effect of Zn²⁺ on t-[³H]butylbicycloorthobenzoate ([3H]TBOB) binding to the GABA, receptor complex was studied autoradiographically in rat brain. Zn2+ inhibited [3H]TBOB binding in a dose-dependent manner at physiological concentrations. Saturation analysis revealed noncompetitive inhibition in various brain regions. The inhibitory effect of Zn2+ had regional heterogeneity; regions showing the greatest inhibition of [3H]TBOB binding were cortical laminae I-III, most areas of hippocampus, striatum, septum, and cerebellar cortex. Regions with relatively less inhibition of [3H]TBOB binding included cortical laminae V-VI, thalamus, superior colliculus, inferior colliculus, and central gray matter. The effect of Zn2+ and those of other GABA, ligands, such as benzodiazepines, bicuculline, isoguvacine, and picrotoxin, on [3H]TBOB binding seemed to be additive. Ni2+, Cd2+, and Cu2+ also inhibited [3H]TBOB binding with a regional heterogeneity similar to that produced by Zn2+. These results are consistent with Zn2+ acting at the previously detected recognition site on the GABA, receptor complex, distinct from the picrotoxin, GABA, and benzodiazepine sites. The regional heterogeneity of the Zn2+ effect may reflect differential regional distribution of GABA_A receptor subtypes among brain regions. Other divalent cations probably act at the Zn2+ binding site. Key Words: Zinc-t-Butylbicycloorthobenzoate GABA—GABA
receptor complex—Autoradiography. J. Neurochem. 62, 602-607 (1994).

The GABA_A receptor is the major mediator of synaptic inhibition in the CNS and structurally is a heterooligomeric protein comprised of five (or fewer) subunits per active complex (Burt and Kamatchi, 1991). Molecular cloning studies have provided evidence for the existence of a number of different subunits forming the GABA_A receptor, and each of these subunits has a distinct regional distribution in the brain (Olsen and Tobin, 1990; Burt and Kamatchi, 1991; Lüddens and Wisden, 1991; Doble and Martin, 1992; Wisden et al., 1992). Functionally, the GABA_A receptor incorporates a chloride ionophore, as well as binding sites for several classes of ligands that interact allosterically either to potentiate or to inhibit receptor function (Burt and Kamatchi, 1991; Sieghart, 1992).

Divalent cations have been shown to modulate GABA_A receptors in biochemical and electrophysio-

logical studies. Zn²⁺ and other metal cations, including Cd²⁺, Ni²⁺, Mn²⁺, and Co²⁺, inhibited the GABA response of neurons in a variety of preparations (Smart and Constanti, 1982; Kaneko and Tachibana, 1986; Celentano et al., 1991; Smart, 1992; Kilić et al., 1993). Binding studies have provided evidence for Zn²⁺ modulation of GABA ligand binding. Zn²⁺ enhanced the specific binding of [3H]diazepam in the CNS (MacKerer and Kochman, 1978). Although Zn²⁺ seemed to increase [³H]GABA binding to synaptic membranes at physiological concentrations, it inhibited the binding at pharmacotoxicological doses (Baraldi et al., 1984). An autoradiographic study showed that Zn²⁺ inhibited [³H]GABA binding to GABA_B receptors, but not significantly to GABA_A receptors in any brain region (Turgeon and Albin, 1992). For the picrotoxin binding site, a binding site on the GABA_A receptor complex whose binding is modulated by the activation of other ligand binding sites of the GABA_A receptor complex, there are no published data on the effect of Zn²⁺ and other divalent cations.

The effects of drugs on picrotoxin ligand binding have been used frequently to assess ligand interactions with the GABA_A receptor complex. The purposes of this study were to elucidate Zn²⁺ effects on picrotoxin ligand binding, to examine the regional heterogeneity of the Zn²⁺ effect on GABA_A receptors, and to study the possible interaction of Zn²⁺ effects with the effects of other classes of GABA_A receptor ligands. The picrotoxin binding site of the GABA_A receptor complex can be assayed with *t*-[³H]-butylbicycloorthobenzoate ([³H]TBOB) (Lawrence et al., 1985; O'Connor and McEwen, 1986; Sakurai et al., 1992). We used in vitro quantitative autoradiography to determine the effects of Zn²⁺ on [³H]TBOB binding in rat brain.

Received May 20, 1993; revised manuscript received June 30, 1993; accepted July 8, 1993.

Address correspondence to Dr. R. L. Albin at Department of Neurology, University of Michigan, Neuroscience Laboratory Building, 1103 E. Huron, Ann Arbor, MI 48104, U.S.A.

Abbreviations used: TBOB, t-butylbicycloorthobenzoate; TBPS, t-butylbicyclophosphorothionate.

EXPERIMENTAL PROCEDURES

Male Sprague-Dawley rats (175-199 g; Harlan Labs, Indianapolis, IN, U.S.A.), were decapitated, and their brains rapidly removed and frozen in Lipshaw embedding matrix surrounded by powdered dry ice. Twenty-micrometer-thick sections were cut horizontally on a Lipshaw cryostat. Sections were obtained at the level including the caudoputamen, thaw-mounted onto 2 × subbed gelatin-coated slides on a warming plate, and stored at -20°C until the time of assay. All assays were performed 24 h after decapitation. Sections were run in triplicate. Slide-mounted tissue sections were warmed to room temperature and prewashed for 3×10 min in buffer (50 mM Tris-HCl plus 1 mM EDTA, pH 7.4 at 4°C) at 4°C and dried under a stream of cool air. Binding of [3 H]TBOB (20 nM) was carried out in buffer (50 mM Tris-HCl plus 120 mM NaCl, pH 7.4, at room temperature) for 90 min at room temperature. The incubation was terminated by two 30-min rinses in buffer (50 mM Tris-HCl, pH 7.4, at 4°C) at 4°C, followed by a brief dip in distilled water. Each slide was then dried under a stream of hot air. Nonspecific binding was assessed in the presence of $20 \mu M$ picrotoxin. For analysis of the Zn²⁺ effect on [³H]-TBOB binding, tissue sections were incubated with nine concentrations of ZnCl₂ ranging from 1 nM to 1 mM with 20 nM [3 H]TBOB. Saturation studies were performed with nine concentrations of [3 H]TBOB ranging from 1 pM to 100 nM. Specific [3H]TBOB binding and nonspecific binding were determined at all concentrations of [3H]TBOB in both the presence and the absence of $100 \mu M \text{ ZnCl}_2$. To determine the interaction of Zn^{2+} effects with the effects of other GABA_A ligands that modulate [3H]TBOB binding, other GABA_A ligands were included in the incubation mixture with 20 nM [3H]TBOB in both the presence and absence of $100 \,\mu M \, \text{ZnCl}_2$. The effects of other divalent cations on [3H]TBOB binding were analyzed in the incubation mixture with 20 nM [3H]TBOB. Autoradiograms were generated by apposing the slides to ³H-sensitive film (Hyperfilm, Amersham Inc., Arlington Heights, IL, U.S.A.) in lighttight cassettes along with standards containing known amounts of radioactivity. After 3 weeks, films were developed in Kodak D-19. Ligand binding was quantitated with computer-assisted densitometry using the MCID system (Imaging Research Inc., St. Catharines, Ontario, Canada). To quantify ligand binding density, the optical density of coexposed standards was determined and a standard curve generated by fitting standard values with a fourth-degree polynomial regression equation. Standards were commercial ¹⁴C plastic standards (ARC, St. Louis, MO, U.S.A.). Use of the standards and derived standard curve allow conversion of areal optical density to picomoles per milligram of protein. Areas read included the cortical laminae I-III, lamina IV, and laminae V-VI, hippocampal CA1 stratum oriens, CA1 stratum radiatum, CA3 stratum oriens, CA3 stratum radiatum, and dentate gyrus, striatum, thalamus (laterodorsal thalamic nucleus and lateral posterior thalamic nucleus), septum, superior colliculus, inferior colliculus, periaqueductal gray matter, and cerebellar molecular and granule cell layers. Ten to 20 readings per area from triplicate sections were averaged. IC_{50} values for Zn^{2+} were calculated by log-logit analysis. In the saturation experiment, values of specific [3H]TBOB bound were quantified to construct Scatchard plots.

Materials

[3H]TBOB (sp. act. 19.4 Ci/mmol) was purchased from Amersham. Clonazepam was the gift of Dr. Peter Sorter

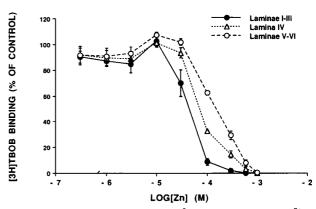


FIG. 1. Curves for the inhibition of [3 H]TBOB binding by Zn 2 + in laminae I–III, lamina IV, and laminae V–VI of the cerebral cortex. Zn 2 + inhibited TBOB binding in a dose-dependent manner. Data are means \pm SEM (bars) from experiments carried out with three animals.

(Hoffmann-La Roche). Zolpidem was the gift of Synthelabo Recherche. Isoguvacine was purchased from Cambridge Research Biochemicals Ltd. (Cambridge, U.K.). Bicuculline and picrotoxin were purchased from Research Biochemicals Inc. (Natick, MA, U.S.A.). The remaining reagents were purchased from Sigma Chemical (St. Louis, MO, U.S.A.) and were of the highest possible purity.

RESULTS

Zn²⁺ modulation of [³H]TBOB binding

 Zn^{2+} produced dose-dependent inhibition of [³H]-TBOB binding in rat brain sections (Fig. 1). Inhibition of [³H]TBOB binding was observed in all areas examined (Fig. 2 and Table 1). Saturation analysis of [³H]TBOB in the presence of 100 μ M Zn²⁺ revealed a significant decrease in B_{max} with little change in K_{D} in most regions (Table 2).

Regional difference of Zn²⁺ modulation of [³H]TBOB binding

The observed regional distribution of [3H]TBOB binding sites was similar to that reported previously by Olsen et al. (1990). Picrotoxin displaced to the same extent all the [3H]TBOB bound in all brain regions (data not shown). On the other hand, there were regional differences in Zn²⁺ inhibition of [³H]TBOB binding (Fig. 1). IC₅₀ values for Zn²⁺ were significantly different among brain regions (p < 0.05 on ANOVA; Table 1), with IC₅₀ values lowest in cortical laminae I-III, the cerebellar molecular layer, and septum. Relatively low IC₅₀ values were observed also in most subfields of the hippocampus, the striatum, and cerebellar granule cell layer. Relatively high IC₅₀ values were found in cortical laminae V-VI, thalamus, superior colliculus, inferior colliculus, and periaqueductal gray matter.

Pharmacological profile of [3H]TBOB binding

Compounds that modulate the activity of the GA-BA_A receptor complex allosterically modulate bind-

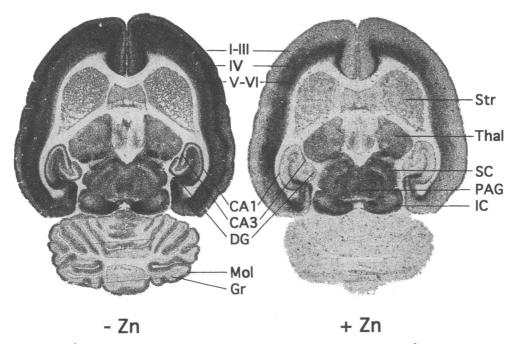


FIG. 2. Autoradiographs of [³H]TBOB binding in the absence (-Zn) and presence (+Zn) of 100 μM Zn²+ in horizontal sections of rat brain. Zn²+ inhibited [³H]TBOB binding in all areas with regional heterogeneity. Regions showing the greatest inhibition of [³H]TBOB binding were cortical laminae I-III, most areas of hippocampus, striatum, septum, and cerebellar cortex. Regions with relatively less inhibition of [³H]TBOB binding included cortical laminae V-VI, thalamus, superior colliculus, inferior colliculus, and periaqueductal gray matter. I-III, cortical laminae I-III; IV, cortical lamina IV; V-VI, cortical laminae V-VI; CA1, CA1 stratum oriens; CA3, CA3 stratum oriens; DG, dentate gyrus; Gr, cerebellar granule cell layer; IC, inferior colliculus; MoI, cerebellar molecular layer; PAG, periaqueductal gray matter; SC, superior colliculus; Str, striatum; Thal, thalamus.

ing of the picrotoxin ligands, t-[35S]butylbicyclophosphorothionate ([35S]TBPS) and [3H]TBOB. The benzodiazepine agonists, clonazepam and zolpidem, enhance TBPS and TBOB binding at concentrations between 100 nM and 1 µM (Lloyd et al., 1990; Sakurai et al., 1992; Kume et al., unpublished observations). GABA agonists, such as muscimol and isoguvacine, inhibit TBPS and TBOB binding (Squires et al., 1983; Lawrence et al., 1985; Edgar and Schwartz, 1990; Sakurai et al., 1992), whereas the GABA_A competitive antagonist bicuculline enhances their binding in the cerebellar granule cell layer (Squires et al., 1983; Korpi et al., 1992; Sakurai et al., 1992). In this experiment, Zn²⁺ inhibited [³H]TBOB binding in the presence of either clonazepam or zolpidem (Fig. 3). In the presence of either isoguvacine or bicuculline, Zn²⁺ produced approximately the same percent reduction of TBOB binding as that produced in the absence of these compounds. The effects of Zn²⁺ and benzodiazepines, isoguvacine, and bicuculline seemed to be essentially additive. Picrotoxin is considered to be a competitive inhibitor of [3H]TBOB (Lawrence et al., 1985). The inhibition by picrotoxin and Zn²⁺ also seemed to be additive (Fig. 3).

Modulation of [3H]TBOB binding by other divalent cations

 Ni^{2+} , Cd^{2+} , and Cu^{2+} at $300 \,\mu M$ also inhibited [³H]-TBOB binding (Fig. 4). There were regional differ-

ences in the inhibition of [³H]TBOB binding similar to that produced by Zn²⁺.

DISCUSSION

The primary finding of this study is that Zn²⁺ produced dose-dependent inhibition of [3H]TBOB binding in rat brain sections. Zn²⁺ inhibition of [³H]TBOB binding was observed in all areas examined, and there were regional differences in the Zn²⁺ effect on [³H]-TBOB binding. Saturation analysis indicated that Zn²⁺ inhibits [³H]TBOB binding in a noncompetitive manner. Divalent cations are known to form soluble complexes with some ligands (Crawford and McBurney, 1977). If Zn²⁺ forms a soluble complex with [³H]-TBOB, reducing the concentration of free [3H]TBOB, Zn²⁺ would decrease the amount of free [³H]TBOB interacting with the GABAA receptor. In this case, however, an autoradiogram obtained from the incubation mixture of [3H]TBOB with Zn2+ would show uniform decrease in regional binding and not the regional differences we documented. Moreover, soluble complex formation should not alter the B_{max} of saturation curves. Our results indicate a noncompetitive interaction, and these observations cannot be explained by the hypothesis of complex formation. We infer that Zn²⁺ is acting as an inhibitor of [3H]TBOB binding by acting at a site on the GABA_A receptor.

TABLE 1. Inhibition of $\lceil ^3H \rceil TBOB$ binding in various brain regions by Zn^{2+}

Brain region	$IC_{50}(\mu M)$			
With high effect of Zn ²⁺ :				
Neocortex				
Laminae I–III	42.2 ± 4.1			
Hippocampus				
CA1 stratum oriens	74.8 ± 5.3			
CA1 stratum radiatum	57.0 ± 16.3			
CA3 stratum oriens	46.9 ± 2.8			
Dentate gyrus	59.4 ± 4.5			
Striatum	83.5 ± 15.7			
Septum	48.3 ± 8.1			
Cerebellum				
Molecular layer	47.7 ± 8.8			
Granule cell layer	70.8 ± 27.6			
With moderate effect of Zn ²⁺ :				
Neocortex				
Lamina IV	102.5 ± 16.8			
Hippocampus				
CA3 stratum radiatum	113.3 ± 10.8			
Pre-/parasubiculum	123.6 ± 17.5			
With low effect of Zn ²⁺ :				
Neocortex				
Laminae V–VI	150.6 ± 9.4			
Thalamus	167.2 ± 14.8			
Superior colliculus	214.2 ± 4.7			
Inferior colliculus	247.9 ± 43.3			
Central gray mater	193.6 ± 12.8			

 IC_{50} values were derived from inhibition curves with $ZnCl_2$. Data are means \pm SEM from experiments with three animals. A significant difference of the IC_{50} values among regions was noted using one-way ANOVA (p < 0.05).

Our results are consistent with prior electrophysiologic data and indicate that our binding data address the same phenomena described in electrophysiologic studies. Our observed IC₅₀ values approximate 100 μM , which is consistent with results obtained in several functional studies (Smart and Constanti, 1982; Westbrook and Mayer, 1987; Celentano et al., 1991; Legendre and Westbrook, 1991; Smart et al., 1991; Smart, 1992; Kilić et al., 1993), and the effects of Zn²⁺ are mimicked by other divalent metal cations, a phenomenon noted also in electrophysiologic studies

(Kaneko and Tachibana, 1986; Celentano et al., 1991).

Zn2+ is present in many synaptic vesicles in the vertebrate CNS (Pérez-Clausell and Danscher, 1985) and is released into the extracellular space during neuronal activity (Assaf and Chung, 1984; Howell et al., 1984). Assaf and Chung (1984) demonstrated extracellular concentrations of Zn^{2+} as high as 300 μM following depolarization. The effective concentration range for Zn²⁺ effects in our study and prior electrophysiologic studies (Smart and Constanti, 1982; Westbrook and Mayer, 1987; Celentano et al., 1991; Legendre and Westbrook, 1991; Smart et al., 1991; Smart, 1992; Kilić et al., 1993) is within the probable physiological concentration range of Zn²⁺ within the synaptic cleft. Zn2+ may function as an endogenous neuromodulator of GABA receptor complex function within the CNS.

Our binding data provide evidence for regional differences in the Zn²⁺ effect on the GABA_A receptor complex. Regions showing the greatest inhibition of [³H]TBOB binding were cortical laminae I-III, most areas of hippocampus, striatum, septum, and cerebellar cortex. Regions with relatively less inhibition of [³H]TBOB binding included cortical laminae V-VI, thalamus, superior colliculus, inferior colliculus, and periaqueductal gray matter.

The pharmacological and molecular biological evidence for regional heterogeneity of the GABA, receptor complex has been widely reported, and regional/ cellular differences in the expression of GABAA receptor subtypes probably underlie, at least in part, regional/cellular differences in receptor function and pharmacology (Olsen and Tobin, 1990; Burt and Kamatchi, 1991). Recent recombinant receptor studies have shown that GABA_A receptors expressing the γ 2 subunit are relatively insensitive to Zn²⁺ (Draguhn et al., 1990; Smart et al., 1991), suggesting that the presence of γ^2 subunits is an important determinant of the relative Zn²⁺ sensitivity of native GABA_A receptors. It is likely that each brain region expresses a variety of GABAA receptors of varying isoform composition with substantial regional differences in the expressed amount of different types of receptor heterooligomers. The γ 2 subunit is expressed in vir-

TABLE 2. Saturation binding parameters of [³H]TBOB binding in several brain regions

Brain region	$K_{\mathbf{D}}\left(-\mathbf{Z}\mathbf{n}\right)$	$K_{\mathbf{D}}(+\mathbf{Z}\mathbf{n})$	B_{max} ($-Z$ n)	$B_{\text{max}} (+Zn)$
Cerebral cortex				
Laminae I-III	24.0 ± 1.5	37.0 ± 2.4^a	2.25 ± 0.21	0.61 ± 0.07^a
Lamina IV	22.7 ± 2.2	28.7 ± 0.6	2.84 ± 0.30	1.46 ± 0.16^a
Laminae V-VI	30.3 ± 3.6	34.2 ± 5.3	2.60 ± 0.27	1.92 ± 0.11
Striatum	55.7 ± 11.1	35.7 ± 5.6	1.80 ± 0.26	0.74 ± 0.13^a

 $K_{\rm D}$ (nM) and $B_{\rm max}$ (pmol/mg of protein) values were determined in the presence and absence of 100 μ M ZnCl₂. Data are means \pm SEM from experiments with three animals.

^a A significant difference between -Zn and +Zn was noted using two-tailed t test (p < 0.05).

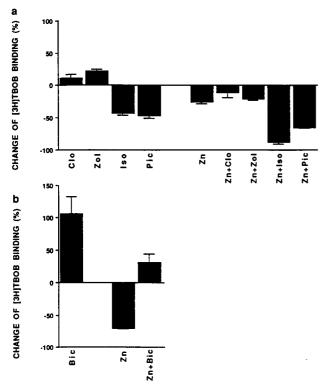


FIG. 3. Zn²+ effects were essentially additive to those of other GABA_A ligands. The graphs show the results in the cortical laminae V–VI (a) and in the cerebellar granule cell layer (b). The experiment was performed in the presence of 500 nM clonazepam (Clo), 500 nM zolpidem (Zol), 10 μ M isoguvacine (Iso), 600 nM picrotoxin (Pic), or 100 μ M bicuculline (Bic), each done in the absence and presence of 100 μ M Zn²+. Zn²+ produced inhibition in the presence of either clonazepam or zolpidem (a). Zn²+ added another inhibitory effect to the inhibition produced by either isoguvacine or picrotoxin. Zn²+ decreased the enhancement of [³H]-TBOB binding by bicuculline in the cerebellar granule cell layer (b). Data are means \pm SEM (bars) from experiments with three animals

tually all brain regions examined (Wisden et al., 1992), albeit at low levels in many regions, and it is likely that most, if not all, brain regions possess γ 2-containing GABA_A receptors. Recombinant receptors without γ 2 subunits [α 1 β 1 (Draguhn et al., 1990), α 1 β 2 (Smart et al., 1991)] have IC₅₀ values for Zn²⁺ approximating 1 μ M. These IC₅₀ values are much lower than those of either recombinant receptors containing γ 2 subunits or native GABA_A receptors, and are significantly lower than the IC₅₀ values of the Zn²⁺ effect on [3 H]TBOB binding. It is likely that the presence of GABA_A receptors containing γ 2 subunits accounts for the relative insensitivity of native receptors compared with recombinant receptors lacking γ 2 receptors.

Our autoradiographic binding data, however, suggest that the relative abundance of $\gamma 2$ subunits is not the sole determinant of regional differences in the Zn^{2+} sensitivity of $GABA_A$ receptors. The neocortex, hippocampus, and cerebellar cortex are brain regions

in which abundant γ^2 mRNA has been detected, and the γ^2 subunit is considered to be a primary component of native GABA_A receptor complexes (Laurie et al., 1992; Wisden et al., 1992). Therefore, those regions are predicted to be less sensitive to Zn²⁺. In our autoradiograms, however, those brain regions showed relatively high sensitivity to the effects of Zn²⁺. Moreover, our data show that thalamus, which contains a relatively low level of γ 2 subunit mRNA (Wisden et al., 1992), was relatively insensitive to Zn²⁺. These results are consistent with the suggestion of Smart et al. (1991) that some combinations of GABA_A receptor subunits that lack the γ 2 subunit are also likely to be insensitive to Zn²⁺. Comparison of our data with the reported distribution of other GABA_A receptor subunit mRNAs (Laurie et al., 1992; Wisden et al., 1992) reveals that the distribution of no single subunit correlates with the distribution of the Zn^{2+} effect on [3H]TBOB binding. The degree of susceptibility to the effects of Zn²⁺ may be regulated by complex combinations of subunits and/or by posttranslational modification of GABA_A receptor complexes.

Electrophysiologic studies (Smart and Constanti, 1982; Celentano et al., 1991; Smart, 1992) indicate that modulation of GABA_A receptor function by benzodiazepines, barbiturates, neurosteroids, bicuculline, and picrotoxin does not interfere with the antagonism exerted by Zn²⁺. These studies suggest the existence of a Zn²⁺ site distinct from other modulatory GABA_A ligand binding sites. Our binding data are consistent with this inference. The effect of Zn²⁺ and those of other GABA_A ligands on [³H]TBOB binding seemed to be additive, a result consistent with the existence of a separate Zn²⁺ site. There is a possible discrepancy between our results and prior electrophysiologic data (Celentano et al., 1991; Smart, 1992). Our results indicate that Zn^{2+} inhibits picrotoxin ligand binding, and this suggests that Zn^{2+} might decrease the effectiveness of picrotoxin, a result not apparently

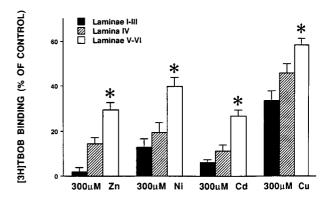


FIG. 4. Divalent metal cations inhibit [3 H]TBOB binding with similar regional differences in effect. [3 H]TBOB binding was more inhibited in laminae I–III than in laminae IV and V–VI of the cerebral cortex. Data are means \pm SEM (bars) from experiments done with three animals. *A significant difference from laminae I–III and IV was noted using ANOVA (p < 0.05).

seen in prior studies. Zn^{2+} , however, is a noncompetitive inhibitor of [${}^{3}H$]TBOB binding with a predominant effect on B_{max} and little effect on binding site affinity. Prior studies of Zn^{2+} -picrotoxin interactions were done with single, subsaturating concentrations of picrotoxin, and full dose-response curves achieving saturating concentrations of picrotoxin might be necessary to document a Zn^{2+} -picrotoxin interaction.

In addition to Zn^{2+} , other divalent metal cations, such as Co^{2+} , Ni^{2+} , Mn^{2+} , and Cd^{2+} , also have been reported to inhibit GABA-induced current noncompetitively (Kaneko and Tachibana, 1986; Celentano et al., 1991). These cations probably act at a common site, but possess different intrinsic efficacies as allosteric inhibitors, such that Zn^{2+} has greater efficacy than Cd^{2+} (Celentano et al., 1991). In the present study, $300~\mu M~Ni^{2+}$, Cd^{2+} , and Cu^{2+} inhibited [³H]-TBOB binding as well as $300~\mu M~Zn^{2+}$. Moreover, there were regional differences in the inhibition of [³H]TBOB binding, similar to that produced by Zn^{2+} . These results suggest that Ni^{2+} , Cd^{2+} , and Cu^{2+} act at the Zn^{2+} binding site.

Further studies will be necessary to establish the subunit composition, location, and functional properties of cation-sensitive GABA_A receptor subtypes.

Acknowledgment: We thank Mr. K. Kaatz, Dr. S. Tallaksen-Greene, and Miss S. M. Turgeon for technical assistance and their helpful advice, and Mr. C. Campbell for assistance with computer programs. This work was supported by grants from Japan Foundation for Aging and Health, and NS19613 from the National Institutes of Health.

REFERENCES

- Assaf S. Y. and Chung S.-H. (1984) Release of endogenous Zn²⁺ from brain tissue during activity. *Nature* **308**, 734–736.
- Baraldi M., Caselgrandi E., and Santi M. (1984) Effect of zinc on specific binding of GABA to rat brain membranes, in *The Neu-robiology of Zinc (Part A)* (Frederickson C. J., Howell G. A., and Kasarskis E. J., eds), pp. 59–71. Alan R. Liss, New York.
- Burt D. R. and Kamatchi G. L. (1991) GABA_A receptor subtypes: from pharmacology to molecular biology. *FASEB J.* **5**, 2916–2923
- Celentano J. J., Gyenes M., Gibbs T. T., and Farb D. H. (1991) Negative modulation of the γ-aminobutyric acid response by extracellular zinc. *Mol. Pharmacol.* **40**, 766-773.
- Crawford A. C. and McBurney R. N. (1977) The synergistic action of L-glutamate and L-aspartate at crustacean excitatory neuro-muscular junctions. *J. Physiol. (Lond.)* **268**, 697–709.
- Doble A. and Martin I. L. (1992) Multiple benzodiazepine receptors: no reason for anxiety. *Trends Pharmacol. Sci.* **13**, 76–81.
- Draguhn A., Verdorn T. A., Ewert M., Seeburg P. H., and Sakmann B. (1990) Functional and molecular distinction between recombinant rat GABA_A receptor subtypes by Zn²⁺. Neuron 5, 781–788.
- Edgar P. P. and Schwartz R. D. (1990) Localization and characterization of ³⁵S-*t*-butylbicyclophosphorothionate binding in rat brain: an autoradiographic study. *J. Neurosci.* **10**, 603–612.
- Howell G. A., Welch M. G., and Frederickson C. J. (1984) Stimulation-induced uptake and release of zinc in hippocampal slices. *Nature* 308, 736–738.

- Kaneko A. and Tachibana M. (1986) Blocking effects of cobalt and related ions on the γ -aminobutyric acid-induced current in turtle retinal cones. *J. Physiol. (Lond.)* **373**, 463–479.
- Kilić G., Moran O., and Cherubini E. (1993) Currents activated by GABA and their modulation by Zn²⁺ in cerebellar granule cells in culture. *Eur. J. Neurosci.* **5,** 65–72.
- Korpi E. R., Lüddens H., and Seeburg P. H. (1992) GABA_A antagonists reveal binding sites for [35S]TBPS in cerebellar granular cell layer. Eur. J. Pharmacol. 211, 427–428.
- Laurie D. J., Seeburg P. H., and Wisden W. (1992) The distribution of 13 GABA_A receptor subunit mRNAs in the rat brain. II. Olfactory bulb and cerebellum. J. Neurosci. 12, 1063-1076.
- Lawrence L. J., Palmer C. J., Gee K. W., Wang X., Yamamura H. I., and Casida J. E. (1985) t-[³H]Butylbicycloorthobenzoate: new radioligand probe for the γ-aminobutyric acid-regulated chloride ionophore. J. Neurochem. 45, 798–804.
- Legendre P. and Westbrook G. L. (1991) Noncompetitive inhibition of γ -aminobutyric acid A channels by Zn. *Mol. Pharmacol.* **39**, 267–274.
- Lloyd G. K., Danielou G., and Thuret F. (1990) The activity of zolpidem and other hypnotics within the γ-aminobutyric acid (GABA_A) receptor supramolecular complex, as determined by ³⁵S-/-butylbicyclophosphorothionate (³⁵S-TBPS) binding to rat cerebral cortex membranes. J. Pharmacol. Exp. Ther. 255, 690-696.
- Lüddens H. and Wisden W. (1991) Function and pharmacology of multiple GABA_A receptor subunits. *Trends Pharmacol. Sci.* 12, 49-51.
- MacKerer C. R. and Kochman R. L. (1978) Effects of cations and anions on the binding of ³H-diazepam to rat brain. *Proc. Soc. Exp. Biol. Med.* **158**, 393–397.
- O'Connor L. H. and McEwen B. S. (1986) Autoradiographic localization of GABA-regulated chloride ionophore binding sites using [³H]*t*-butylbicycloorthobenzoate. *Eur. J. Pharmacol.* **120**, 141–142.
- Olsen R. W. and Tobin A. J. (1990) Molecular biology of GABA_A receptors. *FASEB J.* **4**, 1469–1480.
- Olsen R. W., McCabe R. T., and Wamsley J. K. (1990) GABA_A receptor subtypes: autoradiographic comparison of GABA, benzodiazepine, and convulsant binding sites in the rat central nervous system. *J. Chem. Neuroanat.* 3, 59–76.
- Pérez-Clausell J. and Danscher G. (1985) Intravesicular localization of zinc in rat telencephalic boutons: a histochemical study. *Brain Res.* 337, 91–98.
- Sakurai S. Y., Burdette D. E., and Albin R. L. (1992) Quantitative autoradiography of [³H]TBOB binding to GABA_A receptors. *Soc. Neurosci. Abstr.* **18**, 655.
- Sieghart W. (1992) GABA_A receptors: ligand-gated Cl⁻ ion channels modulated by multiple drug-binding sites. *Trends Pharmacol. Sci.* 13, 446-450.
- Smart T. G. (1992) A novel modulatory binding site for zinc on the GABA_A receptor complex in cultured rat neurons. *J. Physiol.* (Lond.) **447**, 587–625.
- Smart T. G. and Constanti A. (1982) A novel effect of zinc on the lobster muscle GABA receptor. Proc. R. Soc. Lond. [Biol.] 215, 327–341.
- Smart T. G., Moss S. J., Xie X., and Huganir R. L. (1991) GABA_A receptors are differentially sensitive to zinc: dependence on subunit composition. *Br. J. Pharmacol.* 103, 1837–1839.
- Squires R. F., Casida J. E., Richardson M., and Saederup E. (1983) [35S]*t*-Butylbicyclophosphorothionate binds with high affinity to brain-specific sites coupled to γ-aminobutyric acid-A and ion recognition sites. *Mol. Pharmacol.* **23**, 326–336.
- Turgeon S. M. and Albin R. L. (1992) Zinc modulates GABA_B binding in rat brain. *Brain Res.* **30**, 30–34.
- Westbrook G. L. and Mayer M. L. (1987) Micromolar concentrations of Zn²⁺ antagonize NMDA and GABA responses of hippocampal neurons. *Nature* **328**, 640–643.
- Wisden W., Laurie D. J., Monyer H., and Seeburg P. H. (1992) The distribution of 13 GABA_A receptor subunit mRNAs in the rat brain. I. Telencephalon, diencephalon, mesencephalon. J. Neurosci. 12, 1040-1062.