

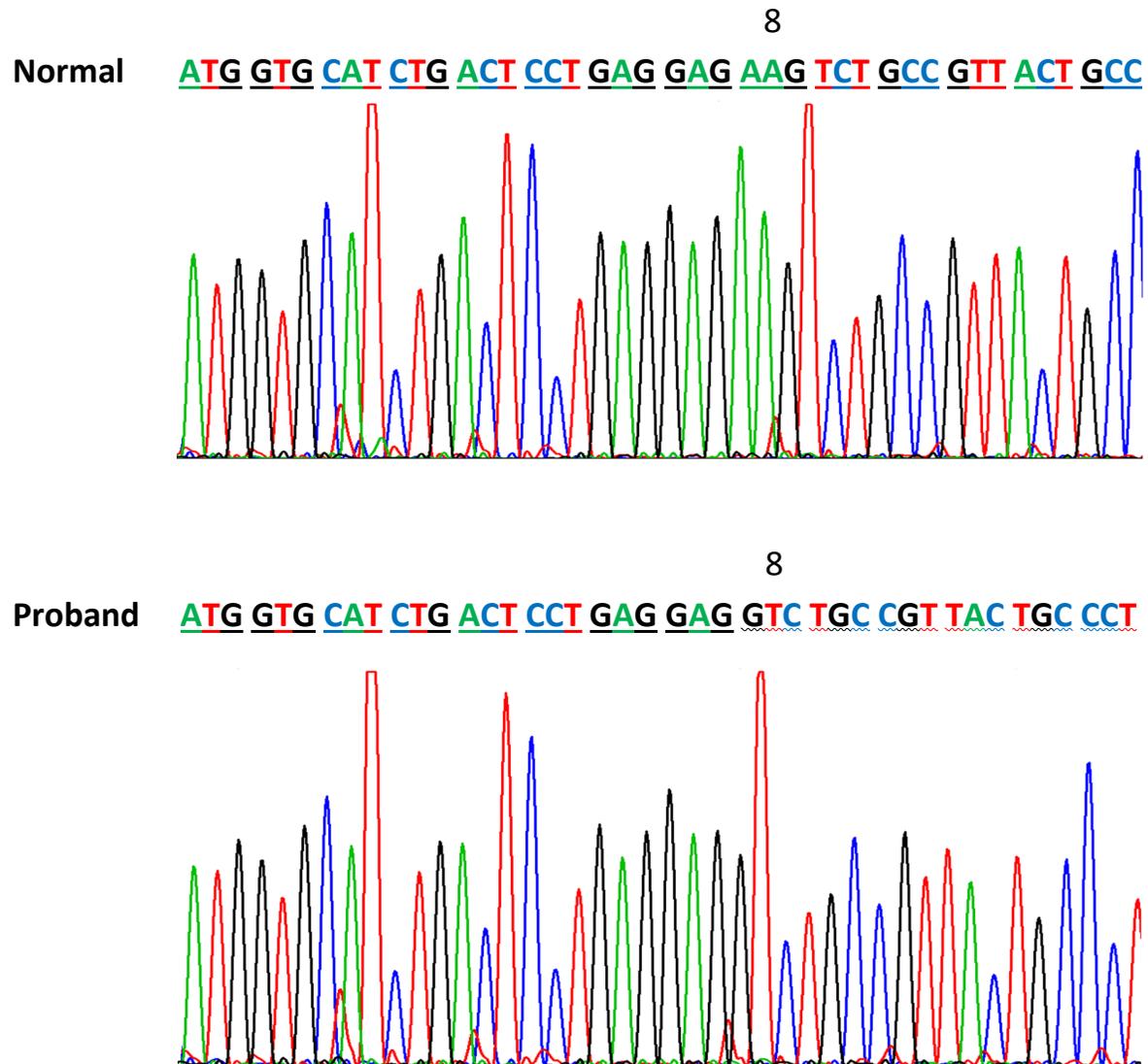
# The Genetic Basis of Asymptomatic Codon 8 Frame-Shift $\beta^0$ -Thalassaemia Homozygotes

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## Supplementary data

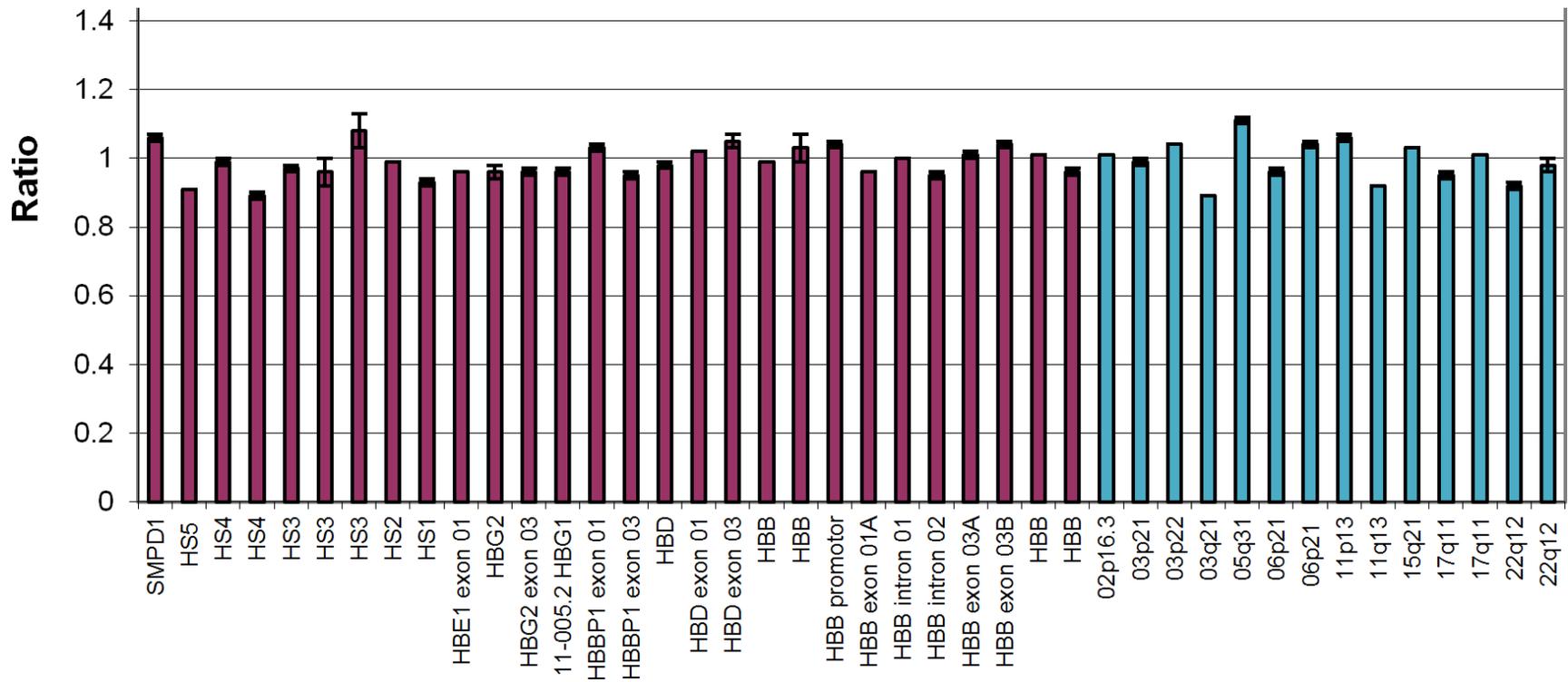
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Figure S1 Nucleotide Sequences of *HBB* from Initiation Codon ATG to Codon 13



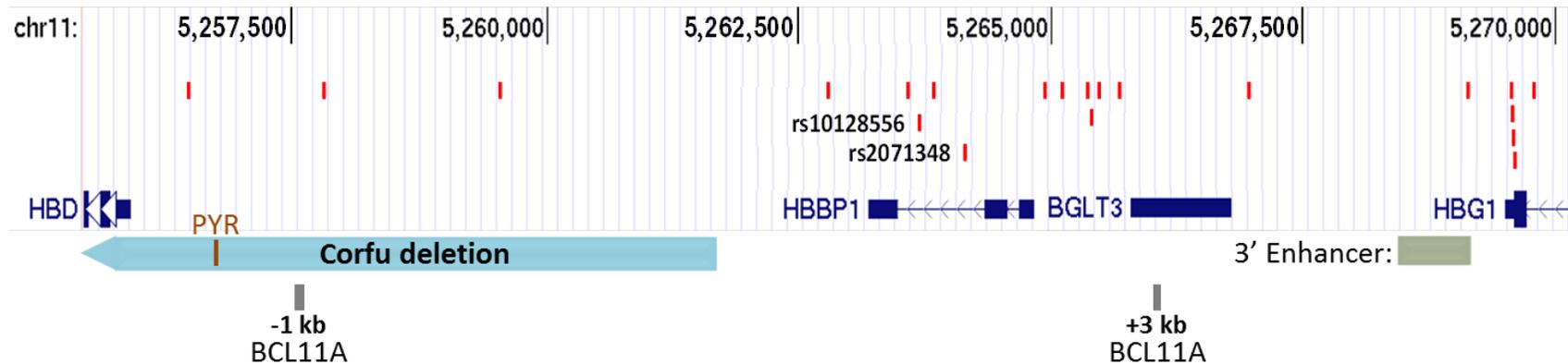
**Figure Legend:** The top figure shows the nucleotide sequences from a normal individual beginning from the *HBB* initiation codon ATG to codon 13. Note codon 8 is AAG. The lower figure shows the nucleotide sequences from the proband. Note codon 8 is GTC, as a result of homozygous deletion of dinucleotides AA from the normal codon 8 sequences.

Figure S2 MLPA on Proband's *HBB* Gene Cluster



**Figure Legend:** MLPA on proband's *HBB* gene cluster showing equal signal strength of probes from LCR to 9 kb downstream from *HBB*. The blue-colored bars represent probes for genes on other chromosomes which serve as control.

Figure S3 SNPs Present in Proband's *HBD* to *HBG1* Intergenic Region



**Figure Legend:** The figure shows ~14.5 kb *HBG1* – *HBD* intergenic region. The numbers on the first row represent hg19 Chr11 positions. Each perpendicular red bar represents a single SNP found in the proband, all homozygous for the alternate alleles (Supplementary Table 2). The light green bar represents the enhancer 3' to *HBG1* (Bodine, Ley, EMBO J 1987; 6:2997-3004). BGLT3 is the template for a long non-coding RNA (Kiefer et al, Blood 2011; 118:6200-6208). The light blue bar represents the Corfu deletion (Kulozik et al, Blood 1988; 71:457-462). The brown strip represents PYR complex DNA-binding site (Bank, Blood 2006; 107:435-443). The grey strips represent two purported BCL11A binding sites, at 3 kb downstream of *HBG1*, and 1 kb upstream of *HBD* (Sankaran et al, N Engl J Med 2011; 365:807-814). SNP rs10128556 is associated with elevated HbF in sickle cell anemia patients (Galarneau et al, Nat Genet 2010; 42:1049-1051). SNP rs2071348 is associated with elevated HbF and milder disease severity in patients with HbE/ $\beta^0$ -thalassemia (Nuinoon et al, Hum Genet 2010; 127:303-314).

Table SI ARMS tests to determine HbF QTL genotypes

HbF QTL	Primer name	Primer sequences 5' to 3'	PCR amplicons	SNP genotype	Final concentrations of primers picomoles/μL	PCR conditions (Temperature / Time)			
						Denaturing	Annealing	Extension	PCR cycles
rs9399137 Chr.6q23	F-9399137	GTTCTGCTTCTACAATAGACATTTTTTCATG			0.1	98°C / 40sec	56°C / 40sec	72°C / 2min	35
	R-9399137	TAAAAACCTGGCAAGATAACCATTTAAAGTA			0.1				
	F-9399137-C	TAATGTAATTAACCTGAACATATGGTTACTC	250 bp	C	0.2				
	R-9399137-T	AGCAGGGTTGCTTGTGAAAAAAGTTA	141 bp	T	0.1				
rs66650371 3-bp deletion Chr.6q23	TAC-1F:	TCACTCTGGACAGCAGATGTTACTAT			0.2	98°C / 20sec	58°C / 40sec	72°C / 40sec	30
	TAC-1R	CTCAGTGATGGTATTTCTGGAGAC	206 bp	no del	0.8				
	TAC-0F	AGCCCGTCCAGACACTCATTGTT			0.1				
	TAC-0R	GCCCTGATAACATTTTGTGGTTTTTCATTTAACAT	276 bp	3-bp del	0.8				
rs766432 Chr.2p15	F-766432	ACAGCGAGACCCTGTCTCAAAAACAA			0.1	98°C / 40sec	56°C / 40sec	72°C / 2min	35
	R-766432	TTCCTTAGTGTAGTGGAGAGGTTTCTA			0.1				
	F-766432-C	GTTTCGCTTAGCTTTATTAAGGTACAC	726 bp	C	0.1				
	R-766432-A	CTTAAAATGAATGACTTTTGTGTATGTACAAT	521 bp	A	0.2				

Genomic DNA was extracted from peripheral blood leukocytes by BioRobot EZ1 (Qiagen). PCR was performed using a thermocycler (ABI). In 10 ul reaction mixture, there were 100 ng of genomic DNA, 5 ul Multiplex mix (x2, Qiagen), 2 ul of Q-solution (x5, Qiagen) and the primer mix as shown in the table. The PCR reaction began with denaturation at 95°C for 15 minutes, followed by amplification cycles as summarized in the table. After PCR, 5 ul of reaction mixture was run on 8% polyacryamide gel at 200V for 40 minutes. Note that genotyping for rs9399137 and rs766432 can be done in a multiplex reaction within a single tube. The alternate alleles are colored red in the table.

Table SII Non-identical SNP Genotypes at 7 Loci Found in Two Dizygotic Twins

Gene locus	SNP ID	hg19 position	Reference allele	Alternate allele	Proband	Twin brother
LFNG	rs61564232	Chr7:2,552,881	–	<b>GATG</b>	<b>GATG / GATG</b>	– / <b>GATG</b>
TFDP1	rs750015004	Chr13:114,290,958	G	<b>A</b>	G / <b>A</b>	G / G
CES3	rs145544825	Chr16:66,998,390	G	<b>A</b>	<b>A / A</b>	G / <b>A</b>
EXO3L1	rs150417999	Chr16:67,222,999	G	<b>A</b>	<b>A / A</b>	G / <b>A</b>
CTRL	rs774504385	Chr16:67,963,944	C	<b>T</b>	<b>T / T</b>	C / <b>T</b>
ILF3	rs200928937	Chr19:10,798,066	G	<b>A</b>	G / <b>A</b>	G / G
ZBTB32	rs202219173	Chr19:36,207,494	C	<b>T</b>	<b>T / T</b>	C / <b>T</b>

Table SIII 21 SNPs Present within Proband's *HBD* to *HBG1* Intergenic Region

Chr11 position (hg19)	SNP ID	Reference allele	Alternate allele	Alternate allele frequency, according to 1,000 Genome Project	Proband's Genotype
5,256,431	rs7948416	A	<b>C</b>	C = 0.84	<b>C / C</b>
5,257,778	rs3759074	G	<b>A</b>	A = 0.19	<b>A / A</b>
5,259,534	rs6578590	T	<b>C</b>	C = 0.86	<b>C / C</b>
5,262,782	rs11036415	A	<b>C</b>	C = 0.84	<b>C / C</b>
5,263,577	rs10128555	C	<b>G</b>	G = 0.89	<b>G / G</b>
5,263,683	<b>rs10128556</b>	C	<b>T</b>	T = 0.20	<b>T / T</b>
5,263,831 - 5,263,828	rs59215941	TGTA	-	N.A.	<b>- / -</b>
5,264,146	<b>rs2071348</b>	T	<b>G</b>	G = 0.21	<b>G / G</b>
5,264,929	rs7934275	A	<b>G</b>	G = 0.89	<b>G / G</b>
5,265,106	rs12417960	C	<b>A</b>	A = 0.24	<b>A / A</b>
5,265,366	rs4910739	G	<b>C</b>	C = 0.83	<b>C / C</b>
5,265,396 - 5,265,394	rs143325232	AAG	-	N.A.	<b>- / -</b>
5,265,475	rs4910545	C	<b>T</b>	T = 0.75	<b>T / T</b>
5,265,680	rs11036431	G	<b>A</b>	A = 0.28	<b>A / A</b>
5,266,961	rs6578591	T	<b>C</b>	C = 0.88	<b>C / C</b>
5,269,140	rs6578592	C	<b>A</b>	A = 0.86	<b>A / A</b>
5,269,583	rs5009539	G	<b>A</b>	A = 0.48	<b>A / A</b>
5,269,584	rs1065686	T	<b>G</b>	G = 0.72	<b>G / G</b>
5,269,585	rs61893083	G	<b>A</b>	A = 0.72	<b>A / A</b>
5,269,586	rs62755960	A	<b>G</b>	G = 0.72	<b>G / G</b>
5,269,799	rs28440105	A	<b>C</b>	C = 0.84	<b>C / C</b>

Table SIV FSC8 Homozygotes, with No or Rare Transfusion

Ethnic background	Patient	Age - Gender	Age at dx	Hb (g/dL)	Splenectomy	Transfusions	$\alpha$ -Globin genotype	Chr11p15	Chr6q23		Chr2p16	HBB cluster haplotype
								rs7482144 ( <i>Xmn I</i> )	rs9399137 (HMIP)	rs66650371 (3-bp deletion)	rs766432 ( <i>BCL11A</i> )	
Turkish	3	32 – F	3 yrs	10 at dx. Now 8.6	At age 7 yrs	Yes before splenectomy; none since	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / C	IV / IV
Turkish	4	30 – F	2 yrs	4.2 at dx. Now 9.3	At age 11 yrs	Yes before splenectomy; none since	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / A	IV / IV
Turkish	5	7 – M	---	7.7	No	Rarely	$\alpha\alpha / \alpha\alpha$	A / A	T / T	WT / WT	A / A	IV / IV
Turkish	6	13 – M	---	9.8	No	No	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / C	IV / IV

Patients #3 and #4 were sisters. They were originally reported in 1989 by Gurgey et al.

Patients #6 and #16 (Table 4) were siblings.

SNP alternate alleles are colored red.

Table SV FSC8 Homozygotes, Transfusion Dependent

Ethnic background	Patient	Age - Gender	Age at dx	Hb (g/dL)	Splenectomy	Transfusions	$\alpha$ -Globin genotype	Chr11p15	Chr6q23		Chr2p16	HBB cluster haplotype
								rs7482144 (Xmn I)	rs9399137 (HMIP)	rs66650371 (3-bp deletion)	rs766432 (BCL11A)	
Iraqi	7	19 – F	7 yrs		At age 18 yrs	Yes, since age 7 yrs	$\alpha\alpha / \alpha\alpha$	A / A	T / T	WT / WT	C / C	IV / IV
Iraqi	8	21 – M	8 yrs	7.1 at dx	At age 14 yrs	Yes, since age 8 yrs	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / C	IV / IV
Iraqi	9	15 - M	3 yrs	7.5	At age 6 yrs	Yes	$\alpha\alpha / \alpha\alpha$	A / A	T / T	WT / WT	A / A	IV / IV
Iraqi	10	14 - F	1 yr	7.9	At age 10 yrs	Yes	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / A	IV / IV
Iraqi	11	14 - M	4 yrs	6.8	At age 5 yrs	Yes	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / C	IV / IV
Iraqi	12	18 - M	1 yr		At age 12 yrs	Yes	$-\alpha^{3.7} / \alpha\alpha$	A / A	T / T	WT / WT	A / A	IV / IV
Turkish	13	16 – M	18-month	6.5 at dx	At age 8 yrs	Yes, since age 18-month	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / A	IV / IV
Turkish	14	21 – F	4 yrs	7.6 at dx	No	Yes, since age 6 yrs	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / C	IV / IV
Turkish	15	20 – M	9-month	8.2 at dx	At age 6 yrs	Yes, since age 1 yr	$\alpha\alpha / \alpha\alpha$	A / A	T / T	WT / WT	A / C	IV / IV
Turkish	16	7 – F		8.0 when transfusion began	No	Yes, since age 4 yrs	$\alpha\alpha / \alpha\alpha$	A / A	T / C	WT / del	A / A	IV / IV
Turkish	17	25 – F			No	Yes	$\alpha\alpha / \alpha\alpha$	A / A	T / T	WT / WT	A / A	IV / IV

Table SV FSC8 Homozygotes, Transfusion Dependent (continued)

Ethnic background	Patient	Age - Gender	Age at dx	Hb (g/dL)	Splenectomy	Transfusions	$\alpha$ -Globin genotype	Chr11p15	Chr6q23		Chr2p16	HBB cluster haplotype
								rs7482144 (Xmn I)	rs9399137 (HMIP)	rs66650371 (3-bp deletion)	rs766432 (BCL11A)	
Moroccan	18	F				Yes	$\alpha\alpha/\alpha\alpha$	A / A	T / T	WT / WT	A / C	IV / IV
Moroccan	19	15 – F				Yes	$\alpha\alpha/\alpha\alpha$	A / A	T / T	WT / WT	A / A	IV / IV
North African	20	DOB 1993 – F			Yes	Yes, since age 3 years	$\alpha\alpha/\alpha\alpha$	A / A	T / T	WT / WT	A / C	IV / IV
North African	21	DOB 1989 – F			At age 20 yrs	Yes, since age 2 years		A / A	T / C	WT / del	A / A	IV / IV
Turkish	22	4 – F	6 month	9.8 at dx	No	Yes, since age 3	$\alpha\alpha/\alpha\alpha$	G / G	T / C	WT / del	C / C	VII / VII
North African	23	DOB 2008 – F			No	Yes, since age 3 months	$\alpha\alpha/\alpha\alpha$	G / G	T / T	WT / WT	A / C	VII / VII
North African	24	DOB 2005 – M			No	Yes, since age 4 months	$\alpha\alpha/\alpha\alpha$	G / G	T / T	WT / WT	A / A	VII / VII

Patients #13 and #14 were siblings.

Patients #16 and #6 (Table 3) were siblings.

Patients #23 and #24 were siblings.

Patient #23 underwent bone marrow transplantation in 2011 at age 3.

SNP alternate alleles are colored red.