Modeling plant–water interactions: an ecohydrological overview from the cell to the global scale



Simone Fatichi,^{1*} Christoforos Pappas^{1,2} and Valeriy Y. Ivanov³

Vegetation and the water cycles are inherently coupled across a wide range of spatial and temporal scales. Water availability interacts with plant ecophysiology and controls vegetation functioning. Concurrently, vegetation has direct and indirect effects on energy, water, carbon, and nutrient cycles. To better understand and model plant-water interactions, highly interdisciplinary approaches are required. We present an overview of the main processes and relevant interactions between water and plants across a range of spatial scales, from the cell level of leaves, where stomatal controls occur, to drought stress at the level of a single tree, to the integrating scales of a watershed, region, and the globe. A review of process representations in models at different scales is presented. More specifically, three main model families are identified: (1) models of plant hydraulics that mechanistically simulate stomatal controls and/or water transport at the tree level; (2) ecohydrological models that simulate plot- to catchmentscale water, energy, and carbon fluxes; and (3) terrestrial biosphere models that simulate carbon, water, and nutrient dynamics at the regional and global scales and address feedback between Earth's vegetation and the climate system. We identify special features and similarities across the model families. Examples of where plant-water interactions are especially important and have led to key scientific findings are also highlighted. Finally, we discuss the various data sources that are currently available to force and validate existing models, and we present perspectives on the evolution of the field. © 2015 Wiley Periodicals, Inc.

> How to cite this article: WIREs Water 2016, 3:327–368. doi: 10.1002/wat2.1125

INTRODUCTION

Transpiration is the process of water transfer from the soil to the atmosphere through plant tissues; during this process, water changes its phase and exits from the plant. On average, transpiration amounts to roughly 40 % of the land precipitation,¹⁻³ and the corresponding latent heat constitutes about 38 % of the net radiation absorbed by the land surface.⁴ Therefore, it is not surprising that vegetation plays a fundamental role in hydrology and climatology.^{5,6} Transpiration occurs almost entirely through small openings in the leaves called stomata.⁷ This is the same pathway through which plants acquire carbon and, thus, represents a major nexus between hydrology and the carbon cycle. Plants lose about 100-500 molecules of water to fix one molecule of CO_2 ⁸ with water representing a key element determining vegetation function and performance. The coupling of transpiration and photosynthesis creates important feedback between water, carbon, and nutrient cycles. Soil biogeochemistry and nutrient dynamics are in fact directly connected to water availability because soil biota dynamics and rock weathering depend on

^{*}Correspondence to: simone.fatichi@ifu.baug.ethz.ch

¹Institute of Environmental Engineering, ETH Zurich, Zurich, Switzerland

²Département de géographie, Université de Montréal, Montréal, Québec, Canada

³Department of Civil and Environmental Engineering, University of Michigan, Ann Arbor, MI, USA

Conflict of interest: The authors have declared no conflicts of interest for this article.

soil moisture content^{9,10} and, also indirectly, through the effect that soil water content has on vegetation growth and litter production. Despite the strong connection between vegetation and hydrology, historically, in hydrological applications, vegetation has been represented with constant factors embedded in equations for computing bulk evapotranspiration, for example in the Penman–Monteith equation.¹¹ Concurrently, in many ecological applications with detailed characterization of plant processes, hydrology has been represented with a simple bucket approximation.^{12,13}

Given the central role of water-plant interactions in the climate system, representations of vegetation attributes and especially stomatal functioning were included in early Earth System Models (ESMs).^{13–16} Since then, in the last two decades, ecohydrology (or hydroecology) has been an emerging scientific field as testified by the growth of the number of published articles and citation metrics beyond the average increase in scientific productivity (Figure 1). This has also been the result of the emphasis placed on water-vegetation interactions in a series of seminal works.²¹⁻²⁴ Ecohydrology has been less recognized as an emerging field in ecology because many ecologists and ecophysiologists would have regarded themselves as 'ecohydrologists' well before this term was popularized by hydrologists.²⁵ Despite this, the emergence of ecohydrology as a well-distinguished discipline helped a better exchange and an explicit linkage between the two scientific communities. Along with the emergence of ecohydrology, an increasing number of quantitative studies focusing on the carbon, water, and nutrient cycles, on regional and global scales, have appeared within the realm of 'biogeosciences' under the growing pressure to better understand the carbon cycle feedback to climate change.²⁶⁻²⁸ There are recent studies in which the boundaries between the fields of ecohydrology, ecophysiology, and biogeosciences are very subtle and mostly a matter of spatiotemporal scales. Beyond affecting water, carbon, and nutrient cycles, vegetation functioning has important implications in addressing other relevant questions, for example in the assessments of land-use change impacts,¹⁷ evaluation of ecosystem services,²⁹ soil resources, and



FIGURE 1 Plant–water interactions occurr over a broad range of spatial scales from (a) leaf interior ($\approx \mu m$), to (b) individual plant ($\approx m$), to (c) catchments ($\approx km$), up to (d) the entire Earth. A map of annual evapotranspiration (ET) on the Rietholzbach catchment¹⁷ and the annual global estimate of annual evapotranspiration (ET) from the MOD16 product¹⁸ are shown. (e) The increasing attention that is paid by the scientific community to ecohydrology is reflected in the number of published articles and the citations they received during the last 16 years (Source: ISI Web of knowledge, August, 2015). The average increasing rate of publication in scientific literature is also shown as a benchmark.¹⁹ The MOD16 map is reprinted with permission from Ref 18. Copyright 2011 Elsevier; the leaf section is reprinted with permission from Ref 20. Copyright 2013 John Wiley and Sons.

landscape formation³⁰ as well as crop productivity.³¹ These are all topics where water availability interacts with plant ecophysiology to produce complex dynamics that are unlikely to be uncovered with narrowly disciplinary approaches that ignore or trivialize either hydrological or vegetation components. Clearly, the perception of the significance and integration of vegetation processes into various fields of environmental research has recently undergone a considerable transformation.

It is important to underline that ecohydrology should not be regarded as the analysis of how vegetation properties and organization affect streamflow. It is a broader discipline that examines the two-way interactions between the entire hydrological cycle and plant functioning. When the main interest is on relatively short-term (≈decade) analyses of streamflow and water availability or in hydrological engineering design, classic hydrological tools, such as the Penman-Monteith equation, can often suffice for the purpose of representing vegetation and approaches that involve multiple processes and feedbacks are unlikely to considerably improve predictive capabilities or specific design criteria. However, when the focus shifts toward land-surface climate feedback, carbon and nutrients cycles, or the aim is to elucidate ecological mechanisms through which water availability affects vegetation functioning, then the necessity of explicitly accounting for vegetation physiology and anatomy and/or soil biogeochemistry becomes extremely relevant. Furthermore, the feedbacks between vegetation and the water cycle cannot be captured if these are not explicitly represented in models, and hypothesis testing can be severely impaired by simplified model structures. An illustrative example is provided by the potential effects that model complexity and explicit consideration of elevated CO2 feedback can have while evaluating drought trends^{32,33} or aridity projections in a changing climate.^{34,35} A too simple representation of vegetation has been shown to provide misleading assessments. Another example is the study of the effects of land-cover change (e.g., deforestation, grassland management) on evapotranspiration and streamflow. Detailed numerical analyses^{17,36} and actual observations^{37–39} suggest that the impact of land-cover change on the hydrological cycle may typically be less pronounced in comparison with the results obtained with model simulations using simplified approaches (e.g., Ref 40) or from small-scale manipulation experiments.⁴¹ However, if the effects of disturbances or long-term analyses (≈multiple decades) are sought, then the water cycle is more tightly connected to the vegetation cover (e.g., Refs 42,43).

For instance, shifts in forest composition because of species-specific mortality have been shown to produce hydrologically relevant consequences.44 This is the result of processes that are typically unaccounted for in classic ecohydrology such as soilbiogeochemistry, forest demography, and disturbances, which therefore may represent fundamental 'ecohydrological' components. Another example of ecohydrological links among vegetation, energy, and the water cycle is vegetation-snow interaction. Plant canopy that intercepts snow typically favors sublimation, thereby reducing the amount of snowpack on the ground. Vegetation also alters the radiation balance, shading ground snowpack from direct radiation, yet typically increasing the longwave radiation reaching the ground during snow-melting periods.^{45–47} Additionally, the presence of vegetation affects the turbulent exchanges, modifying the wind profile.48,49 All of these processes are highly nonlinear and can contribute to the increase or decrease of total snow melt below vegetation when compared to cleared areas.⁵⁰

In this overview, we provide an essential description of the main processes and relevant interactions between water and plants across a range of spatial scales, from the cell level of leaves, where stomatal controls occur, to drought stress at the level of a single tree to the integrating scales of a watershed, region, and the globe (Figure 1). We treat only terrestrial ecosystems and do not consider aquatic plants in water bodies and oceans.⁵¹ Water controls are also regarded from the plant's perspective and, therefore, issues related with vegetation such as a mechanical roughness element and its effect on modulating river and overland flows⁵²⁻⁵⁴ are not addressed. In combination with the description of the physical and ecological processes, we also refer to models available in literature that simulate specific processes at different scales. We only focus on those models that simulate the temporal and, eventually, spatial dynamics of key vegetation states (e.g., leaf area index, LAI). Because of this constraint, we do not explicitly discuss steady-state models and analytical derivations (e.g., Refs 55-60), which have led to considerable advances in the understanding of plantwater interactions but are less amenable to address real case studies. Models developed in forestry research to simulate long-term (≈century) forest succession and management^{61,62} are also excluded because of the rather minimalistic representation of hydrological processes. Finally, we favor models with a certain degree of generality that can be applied beyond the specific conditions for which they were derived. Furthermore, examples where plant-water interactions are especially important and have already led to key scientific findings are highlighted. We also present various data sources that are currently available to force and validate the current models, concluding with perspectives on the evolution of the field.

CELL TO PLANT SCALE

Plants developed a vascular system consisting of nonliving and living cells in which water, carbohydrates, and nutrients can move without an active 'pump' such as a heart designed to control and coordinate such movements.⁶³ Vascular plants need to solve the problem of water and nutrient transport from soil to different plant organs, especially leaves, which are exposed to continued dehydration because of partial contact with the sub-saturated atmosphere through stomatal apertures. Concurrently, plants have to transport the products of photosynthesis, such as sugars, from the leaves to the other living organs. To overcome the lack of a central pump, plants exploit physical gradients in water potential (the energy state of water⁶⁴) and concentrations of osmotic sub-stances.⁶⁵ The mechanisms for water entering the roots and leaving the leaves are described in the 'Stomatal Controls' and 'Root Controls' section, while in the 'Plant Vascular Transport' section, we provide the key elements of water transport mechanisms within plants. Fluxes of water or mass are expressed using a 'flux-gradient' relation, where the flux is proportional to a gradient in the 'concentration' of the driving quantity times a conductance term. Throughout the text, we reference existing models with different degrees of approximation of the current process understanding. We refer the reader to more specific reviews and books for a complete treatment of plant physiology and plant vascular transport.^{8,66–70}

Stomatal Controls

Leaves are the sites where water transported from the xylem, i.e., the non-living vascular conduits of plants used for water transport from roots to leaves (Box 1), is evaporated (commonly referred to as 'transpired') to the atmosphere. At the same time, carbon from the atmosphere is assimilated through the photosynthetic reaction, which takes place in the chloroplasts, mostly located in the mesophyll cells^{71–73} (Figure 2). Water is transported to the terminal part of the xylem network in the leaf veins, while carbon is loaded and exported in the opposite direction in the phloem, the vascular conduits that

BOX 1

PLANT PHYSIOLOGY: DEFINITIONS

Concise definitions of plant physiological terms are provided to guide readers. Xylem refers to the non-living vascular conduits of plants that transport water from the roots to the leaves (Figure 4). Phloem refers to the vascular conduits that transport sugars produced by photosynthesis to other tissues in the plant (Figure 4). Water potential is the potential energy of water per unit volume relative to pure water in reference conditions. Water potential quantifies the tendency of water to move from one area to another due to osmosis, mechanical pressure, or matric effects such as capillary action. Cavitation is the processes induced by a change of pressure, that leads to the formation of vapor bubbles in a liquid; in plants, it occurs within the xylem. Non-structural carbohydrates (NSC) are the free, low molecular weight sugars (glucose, fructose, and sucrose) plus starch that are readily available for transport and plant metabolism. Abscisic acid (ABA) is a plant hormone involved in many plant processes, including stomatal closure, bud dormancy, and leaf abscission. Aquaporins are integral membrane proteins that form pores in the membrane of biological cells and allow for the selective passage of water. Cytoplasm is the portion of the cell that is enclosed within the cell membrane and is composed by organelles and a gel-like substance. Suberin is a waterproofing, waxy substance present in the cell walls of certain plant tissue.

transport sugars produced by photosynthesis to other tissues in the plant (Figure 2). Stomatal size and density and leaf venation have been recognized to be important for plant functioning and evolution.^{20,74,75} At the same time, the bulk of the resistance to water transport in the leaf occurs outside the venation network.⁷⁶⁻⁷⁸ Transpiration to the external atmosphere takes place through stomatal apertures of few micrometers in size ($\sim 2-40 \ \mu m$), mostly located on the lower side of the leaf.^{7,79} Stomatal apertures are the common pathways for water and carbon fluxes and therefore represent one of the most essential linkages between ecological and hydrological dynamics. There is a large amount of literature on environmental factors influencing stomatal apertures, such as light, temperature, CO₂, vapor pressure deficit,⁸⁰⁻⁸³



FIGURE 2 A leaf is mostly composed of mesophyll and epidermal cells. The mesophyll is subdivided into palisade and spongy mesophyll. The epidermis secretes a waxy substance called the cuticle to separate the leaf interior from the external atmosphere. Among the epidermal cells, there are pairs of guard cells. Each pair of guard cells forms a pore called stoma. Water and CO_2 enter and exit the leaf mostly through the stomata. The vascular network of the plant is composed of xylem (blue) that transports water to the leaf cells and of phloem (red), which transports sugars from the leaf to the rest of the plant. Water that exits the xylem is evaporated in the leaf interior (dashed lines). The terms $\Psi x_v \Psi m$, Ψe , Ψg , Ψi , and Ψa are the water potential in the xylem of the leaf vein, mesophyll cell, epidermal cell, guard cell, leaf interior, and atmosphere, respectively. Stomatal aperture responds positively to guard cell turgor pressure (P_g) and negatively to epidermal cell turgor pressure (P_e) (hydromechanical feedback). The conductance of the stomatal aperture (g_s) decreases with water potential in the leaf because of a combination of hydraulic and chemical factors.

but despite the critical role that stomata play, the details of their regulation are still not fully understood.⁸⁴ Ultimately, stomata are largely regulated biologically, and it is through these tiny apertures (or lack thereof if leaves are shed) that vegetation imprints a unique signature on the water cycle.

Each stoma is surrounded by a pair of guard cells that are, in turn, in contact with multiple epidermal cells (Figure 2). Stomata tend to open when guard cells increase their turgor (the sum of water potential and osmotic pressure, see Eq. (4)), while an increase in epidermal cell turgor results in the opposite reaction, exerting a hydromechanically negative feedback $^{85-87}$ (Figure 2(b)). As the guard cell turgor is the sum of osmotic pressure and water potential, stomatal apertures are controlled by both hydraulic and chemical factors⁸⁸ (Figure 2(c)). Stomata close when water potential in the leaf drops because of a large transpiration flux or low water potential in the upstream xylem conduits.^{89–91} The hydraulic control acts directly in the reduction of guard cell turgor, while chemical signals are less well quantified.92 However, it is well established that chemical factors

are essential for stomata opening in response to light.⁹³⁻⁹⁵ Furthermore, chemical compounds, such as ABA, are typically released in response to water stress from the leaves and roots $^{96-98}$ and contribute to a reduction in the stomatal aperture.⁹⁹ Release of ABA is an important evolutionary trait as in early plants such as lycophyte and ferns, stomatal closure is purely hydraulically controlled.¹⁰⁰ A differential sensitivity of stomata aperture to chemical compounds is a likely explanation why certain plants close stomata considerably in response to dehydratation, keeping a fairly constant leaf water potential (commonly referred to as 'isohydric behavior'), while others tend to keep stomata open to favor carbon assimilation, experiencing larger fluctuations and lower values of the leaf water potential ('anisohydric behavior').

Models have been presented to mechanistically describe stomatal behavior and reproduce the hydraulic dynamics in the leaf^{86,101–108} or simply to reproduce functional relations in agreement with observations.^{80,109,110} Models that represent the exact mechanisms through which stomata respond to

equations^{115,116}:

the external environment and chemical signals^{111–113} still represent an open research frontier. Mechanistic models have been mostly applied in plant physiological studies, while ecohydrological models and ESMs adopted empirical/conceptual solutions, where different environmental factors are treated independently⁸⁰ or the stomatal conductance (g_s) is assumed to be proportional to carbon assimilation (A_n) and inversely related to leaf interior CO₂ concentration (c_i) and a function of relative humidity or vapor pressure deficit (VPD). These are, for instance, the

structures of the Ball-Berry¹¹⁴ and Leuning

$$g_s = g_0 + a \frac{A_n}{C_i - \Gamma} f(\text{VPD}), \qquad (1)$$

where g_0 is the residual (or 'leakage') stomatal conductance, when A_n is equal to zero, *a* is an empirical parameter, and Γ is the CO₂ compensation point, i.e., the concentration of CO₂ at which the rate of CO₂ uptake is exactly balanced by the rate of CO₂ release in respiration. The Leuning equation (Eq (1)) is widely used and requires an estimate of net assimilation A_n , which is carried out using biochemical models of photosynthesis, such as the Farquhar model or its subsequent refinements.^{117–121}

An alternative approach to characterizing stomatal regulation is to assume that stomatal apertures are regulated to maximize carbon gain, while minimizing water loss.^{122,123} These approaches, known as 'optimality arguments', are appealing from a theoretical standpoint because they are based on ecological evolutionary principles and appear to be corroborated by observed stomatal behavior.¹²³ However, they require specific assumptions for the optimality function and temporal and spatial scales of integration, which have all been questioned.⁸⁴ Specific formulations of the optimality principle lead to different analytical expressions for leaf-scale conductance,^{124–126} and the approach has also started to be adopted in ESMs.¹²⁷

Note that most of the models referenced above do not directly account for the effects induced by plant water stress beyond VPD, which need to be introduced empirically by either directly modifying g_s , the parameters of the biochemical photosynthesis model, ^{128,129} or some of the terms in the analytical form of optimal stomatal conductance.¹³⁰

Root Controls

Roots represent an interface between plants and soil, providing entrance and initially distributed pathways

for fluxes of water and nutrients from the external environment to the plant. The soil environment surrounding the root is called 'rhizosphere' and can present strong gradients of water potentials and nutrient concentrations.^{131–134} Roots with diameters smaller than 2 mm are generally defined as 'fine roots', while roots of a larger diameter are named 'coarse roots'. Fine roots are mostly composed of living tissues and are essential for the uptake of water and nutrients, while coarse roots serve as aggregated conduits for transport and structural stability of the plant.^{135,136} Water flows into the roots because the water potential is typically larger in the soil than inside the plants.¹³⁷ Situations of reverse flow from roots to soil may also occur and are typically referred to as 'hydraulic redistribution'.^{138,139} There is still a debate as to whether the phenomenon is a part of a typical plant life process or confined to specific conditions; however, it is generally believed that it is of considerably smaller magnitude than water uptake.¹⁴⁰ Water moves from the soil to the inner part of fine roots overcoming a barrier called the Casparian strip, which is mostly composed of suberin for isolating the plant interior and preventing, for instance, the entrance of pathogens. Afterwards, water movement continues across cell cytoplasm (sympalstic pathway) or through cell walls (apoplaspathway) and reaches the xylem and, tic therefore, the vascular network where it can flow to the main stem.¹⁴¹⁻¹⁴³ Given a certain plant demand, water uptake is a function of the rhizosphere water potential distribution, axial and radial root three-dimensional conductivities. and the architecture of the root system.^{144–146} The axial and especially the radial conductivities to water flow are not constant but vary with water potential and can be partially controlled chemically by the plant through aquaporins^{147,148} even though the exact mechanisms underlying this control are still uncertain. Nutrient uptake can be passive and follow water uptake or can be enhanced through osmotic gradients at the root interface or by the synthesis of various specialized transporters that are dynamically integrated into cell the membrane.^{149–151}

The variability in soil water potential, axial and radial conductivities, and three-dimensional root architecture (Figure 3) is preserved in a number of plant-scale models that generally have a specific emphasis on root processes.^{154–161} The most common approach in ecohydrological models and ESMs is to only consider root depth (zero-dimensional model) in combination with some empirical function of water content or water potential limiting water



FIGURE 3 Representation of plant- and patch-scale root systems suitable for explicit 3-D hydraulic models of plant water and nutrient uptake. (a) Spatial distribution of tree stems and their root systems based on measurements at the University of Michigan Biological Station¹⁵²: the central dot is tree stem (diameter ≥ 10 cm); the solid line shows the maximum lateral root extent, while the dashed line delineates distance corresponding to 95% of vertically and radially integrated root length. (b) Plant-scale properties of root distribution are controlled by using explicit root architecture obtained with the RootBox software.¹⁵³ Roots of different order (color-coded) as well as overlapped areas where competition for soil water and nutrients occurs are shown for three exemplary trees. (c) Patch-scale property of root distribution with depth is inferred from in situ observations of the bulk biomass density¹⁵² converted to the length density from variations of root diameters and root specific density. For each depth, the median density, 25–75%, and 10–90% ranges of the obtained distribution are shown.

uptake or transpiration in conditions considered to be 'water stress'.^{161–163} Several models started also considering the vertical distribution of root biomass (one-dimension^{164,165}), and others include a bulk soil-to-root conductance,^{166,167} which is a function of soil hydraulic conductivity and the amount of roots expressed through some index such as rootlength-index or root-area-index.¹⁶⁸ A hybrid approach has recently been developed that preserves three-dimensional information and hydraulic conductivities in a simplified form¹⁴⁵ and may represent an important way forward in representing the hydraulic relations (and constraints) of describing the process of plant water uptake. However, an important limitation of the studies of root functioning and uptake capacity, which has likely prevented the development of a larger number of models or conceptualization of intermediate complexity, is the immense challenge of instrumenting and monitoring roots in field conditions.

Plant Vascular Transport

It is currently accepted by the scientific community that water movement in the xylem is governed by the cohesion-tension theory,^{63,169,170} while the flow of water in the phloem follows the Münch hypothesis.^{171–173} According to the cohesion-tension theory, water molecules are transported in the plant in a metastable state generated by surface tension at the air–water interfaces in the leaves; the tension is transmitted throughout the entire water column in the plants, and molecules remain cohesive among themselves and adherent to the walls because of the small size of conduits and cells. Under extreme negative potentials or external perturbations, water transport can be interrupted by the formation of air bubbles (the process of cavitation), and, thus, the plant's vascular transport becomes impaired.¹⁷⁰

The vertical upward direction of the water flow in the xylem (J_{xyl}) in a conduit of length (ΔL) can be expressed through a 'flux-gradient' relation as the product of the xylem conductivity (K_{xyl}) and the gradient of water potential $(\Delta \Psi_{xyl})$ minus the gravitational potential $(\rho g \Delta z)$:

$$J_{\rm xyl} = K_{\rm xyl} \left(T, \Psi_{\rm xyl} \right) \frac{\Delta \Psi_{\rm xyl} - \rho g \Delta Z}{\Delta L}, \tag{2}$$

where the xylem conductivity K_{xyl} is a plant/tissuespecific parameter that varies with temperature (T)because temperature affects the water viscosity and water potential itself. The dependence of K_{xyl} on the water potential is typically described with a sigmoidal shape, and it is particularly important because it defines the resistance to cavitation, i.e., the formation of air bubbles (embolism) in the xylem. Cavitation implies a reduction of the conductive capacity of the xylem conduits,^{174–177} typically indicated as percentage loss of conductivity (PLC) (Figure 4). The xylem vulnerability curve (PLC vs Ψ_{xvl}) can be described by the knowledge of the water potential at 50% loss of conductivity $(\Psi_{xyl,50})$ and of another characteristic value (e.g., $\Psi_{xyl,12}$), typically correlated to $\Psi_{xyl,50}$.^{178,179} The term J_{xyl} is called sap flow and can be measured directly in the plants with various methods.180

Similarly, following the Münch hypothesis of a turgor-pressure driven flow, the water flux in the phloem in the vertical downward direction (J_{phl}) in a conduit of length (ΔL) is driven by the gradient of turgor (ΔP_{phl}) plus the gravitational potential $(\rho g \Delta z)$

times the phloem conductivity (K_{phl}) , which depends on temperature and on the concentration of sucrose in the phloem (C_{phl}) because sucrose concentration affects the fluid viscosity¹⁸¹ (Figure 4):

$$J_{\rm phl} = K_{\rm phl} \left(T, C_{\rm phl} \right) \frac{\Delta P_{\rm phl} + \rho g \Delta Z}{\Delta L}.$$
 (3)

The turgor (*P*) is defined as the sum of osmotic pressure (Π), directly related to the concentration of osmotic solutes through the Van't Hoff relation or other empirical functions^{182,183} and water potential^{184,185}:

$$P = \Pi + \Psi. \tag{4}$$

Conversely to sap flow, phloem flow is particularly challenging to measure because of the size of the phloem tissue and the disturbances induced by the measurement itself. It has only been measured in laboratory experiments, ^{68,186} although promising techniques for field measurements are emerging.¹⁸⁷

In order for water transport to occur, there should be a difference larger than the gravitational potential in the water potential between roots and leaves, which is typically the case during daytime in the growing season. Water is not only exchanged vertically in the phloem and xylem, but it can also move radially between the phloem and xylem and between storage reserves (mostly composed by living parenchyma cells residing near xylem conduits) and the xylem. These exchanges make the xylem and phloem hydraulically coupled^{188,189} and confer on the plant a given capacitance for absorbing fluctuations between the water demand of the leaves (transpiration) and the water uptake from the roots.^{$19\bar{0}-194$} Capacitance is typically more significant in stems of larger trees,¹⁹⁵ and it is because of a combination of the elastic shrinkage and swelling of the phloem, living bark, and xylem tissue and because of the release of water from 'stiff' storage in the parenchyma cells near the xylem. Elastic shrinkage and swelling of the tree is the main mechanism used to release stored water in young trees, but it becomes marginal in older trees where larger 'stiff volumes' of stored water are the main contributors to tree capacitance.^{196,197} In both young and old trees, the elastic behavior of the xylem and phloem is large enough to be clearly measured with high-resolution dendrometers^{187,198,199} that can provide important information on plant hydraulic behavior.^{200–202} Time scales through which capacitance can buffer the difference between root water uptake and transpiration demand are confined to less than an hour for small, young



FIGURE 4 Global view of long-distance water and carbohydrate transport pathways in a vascular plant. The xylem mediates the net transfer of water from the soil to the atmosphere down a gradient in water potential, $\nabla \Psi$. The phloem carries a flow of sugars and other metabolites down the plant from the leaves to the tissues. Optical micrographs show cross sections of a leaf, stem, and root with the approximate location of the xylem (blue) and phloem (orange). Root nutrient uptake is also shown (green). Typical curves for changes in stem relative water content (RWC) (black line) and xylem cavitation (red line) expressed as percentage loss of conductivity (PLC) as a function of water potential are shown along with the increase in fluid viscosity as a function of sucrose osmotic concentration (blue line). (Reprinted with permission from Ref 69. Copyright 2014 Annual Reviews)

trees and several hours, or even days during droughts, for large, old trees.^{194,203}

Only a few models have been currently presented to describe plant vascular transport in detail as the prevalent option has been to lump the entire plant system into a single conductance or resistance term²⁰⁴ or to ignore plant hydraulics completely and use the root zone soil water potential to directly simulate the controls at the leaf level (the case of ecohydrological models and ESMs, but see Ref 127). Important exceptions are the Sperry model¹⁴² and the TREES model,^{205,206} which use a series of resistances from the soil to the leaves and xylem vulnerability curves to describe plant hydraulic behavior. The FETCH model,²⁰⁷ the XWF model,²⁰⁸⁻²¹⁰ and the model presented by Chuang et al.,²¹¹ describe water flow through the plants with a porous media analogy using the one-dimensional Richards equation for the water potential along the hydraulic path. Attempts to include numerical descriptions of phloem transport, tissue growth, and diameter variations have been even more rare, although important examples are available with different degrees of complexity^{181,212-218} as a follow-up of the first pioneering attempts.²¹⁹

Plant Physiology During a Progressive Drought

When the soil water potential near the roots decreases (or VPD substantially increases), a plant may be unable to fully satisfy the transpiration demand and thus progressively enters into a phase of water stress. This is characterized by a drop in water potential throughout the various plant compartments, accompanied by a reduction of turgor in living tissues, as the plant is only partially able to compensate for low water potentials through osmotic adjustments.²²⁰⁻²²² The first process to be affected by the low water potentials is growth (Figure 5). Cell growth is mostly the result of cell division, enlargement, and cell wall synthesis.^{223,224} Cell enlargement is a function of turgor pressure.^{185,225-227} A small reduction of water potential can significantly decrease or stop growth well before photosynthesis or stomatal conductance are affected.²²⁸⁻²³⁰ As the growth stops, respiration associated with the growth also ceases.^{231,232} Growth impairment has been neglected by ecohydrological literature and models, but, in fact, it represents the first consequence of the onset of water stress.

Process or variables affected	Reduction in tissue water potential ψ and turgor P 0 -2/-12 MPa
Cell growth	
Growth respiration	
ABA release	
Stomatal conductance /transpiration	
Leaf energy budget	
Photosynthesis	
Xylem cavitation	
Root disconnection from soil	
Maintenance respiration	
NSC transport	
Leaf turgor loss	— — — — — —
Leaf shedding	
Plant mortality	

FIGURE 5 Effects on plant physiology caused by a decrease in water potential and turgor. The length of the horizontal lines represent the range of stress levels within which a process becomes first affected. Two different levels of minimum water potential, Ψ , are given: -2 and -12 MPa. These are indicative, and the former corresponds to a value characteristic for drought intolerant plants/crops and the latter for drought-adapted plants in deserts. Dashed lines signify an incipient or vanishing effect. The figure is inspired by Hsiao et al.²²⁰ and Porporato et al.²²

If water potential continues dropping, plants need to preserve sufficiently high water potentials to avoid catastrophic levels of cavitation and thus start reducing the stomatal conductance (see the 'Stomatal Controls' section). A reduction of stomatal conductance typically leads to a decrease in gas exchanges, e.g., photosynthesis and transpiration. These are the controls that have been typically placed at the foundation of ecohydrology²² and embedded in ESMs.¹⁶ The decrease in turgor and photosynthesis further modifies plant metabolism and carbon allocation strategies.^{233,234} The decrease in transpiration alters the leaf energy budget with lower latent heat and higher sensible heat fluxes and eventually higher emitted longwave radiation because, given constant forcing, leaf temperature tends to increase with lower stomatal conductance. A similar response in the energy budget is also observed at the canopy or larger spatial scales.²³⁵

If the drought persists further, plant vascular transport is also affected because the leaf and xylem conductivities decrease at water potentials low enough to allow cavitation and, thus, the formation of emboli in the xylem conduits ('Plant Vascular Transport' section, Figure 5). At relatively low water potentials, leaves lose turgor, and the photosynthetic machine starts to be structurally damaged.^{222,236,237} At this stage, roots can also become completely disconnected from the soil, a process typically unaccounted for in models (but see Ref 206). Maintenance respiration has also been found to decrease as the drought progresses.^{232,238} This is probably because of the slowing down of metabolic activities, although specific mechanisms are yet not fully understood. It is not rare that plants start to shed leaves when exposed to a long severe drought.^{239,240} For some plants, such as droughtdeciduous trees, this is a routinely adopted strategy.^{241–243}

The decrease or even cessation of photosynthesis poses a challenge for the plant that is called to rely only on NSC reserves for its maintenance. The permanent hydraulic failure of vascular transport and carbon starvation because of exhaustion or impossibility to access NSC or a combination of these mechanisms have been hypothesized as reasons for plant mortality.^{244,245} Recent evidence supports an earlier occurrence of hydraulic failure and a consequent blockage of NSC transport.^{238,246} Regardless of the exact mechanisms, a severe and extended drought unavoidably leads to plant mortality, a phenomenon occurring in many ecosystems worldwide, even in those that are not typically associated with droughts.^{247–249} This is particularly relevant as drought stress conditions can be potentially exacerbated by higher temperatures and VPD in the future.^{250–252} Given the fact that the exact mechanisms that lead to plant mortality are still unknown, its modeling is a daunting task^{253,254} and represents an important challenge to improving models of water-vegetation interactions. Widespread plant mortality can, in fact, lead to a considerable shift in species composition and/or vegetation cover fraction, with potentially long-lasting consequences for surface energy exchanges and the water cycle, even though recent observations suggest that observed changes may be smaller than expectations in several ecosystems.^{38,39,255}

PLOT TO CATCHMENT SCALE

Historically, a representation of vegetation was included in land-surface and hydrological models because it affects transpiration through surface roughness, albedo, and canopy resistance $r_{\rm c} = r_{\rm s}/\text{LAI}$, where $r_s = 1/g_s$ is the reciprocal of stomatal conductance. Through r_c vegetation modifies water and energy exchanges at the land surface. It may offer a larger resistance between the soil and atmosphere with respect to bare ground in well-water conditions or a lower resistance in dry conditions, accessing water storages through relatively deep roots. The equation^{11,256} well-known Penman-Monteith accounts for these vegetation properties, and it has been widely used in hydrological models. In its basic form, it provides a static representation of vegetation functioning in space and time, which has been recognized to be a severe limitation in numerous contexts (e.g., Refs 164,257) but still provides a reasonable approximation for many other studies.²⁵⁸ Consequently, models that explicitly and simultaneously solve hydrological and vegetation dynamics (Box 2) have been presented in the last 15 years (Table 1). In literature, the definition of 'ecohydrological model' has been used in very different contexts, and in this article, we refer to ecohydrological models as those models that, in time and, eventually, space, evolve some of the key vegetation states and concurrently solve the water budget. In the simplest case, they only dynamically simulate canopy resistance or LAI, while in the most complex case, a series of carbon pools and vegetation attributes (e.g., plant height or root density) are prognostically simulated. Note that with the above definition, we do not include steady-state models and analytical derivations (e.g., Refs 55,60). Rather than describing each ecohydrological model separately, a non-exhaustive list of models is

BOX 2

VEGETATION DYNAMICS

The term 'vegetation dynamics', when referred to models, may have different meanings according to the context in which it is used. In a large fraction of terrestrial biosphere model literature, vegetation dynamics is used to refer to models where the vegetation type (e.g., PFT or species) is prognostic in space and time. In other words, vegetation dynamics are related to processes such plant establishment and mortality, forest demography, and disturbances that may modify the amount and type of vegetation occurrence. In ecohydrological model literature, vegetation dynamics typically refer to models that prognostically simulate vegetation attributes such as LAI, root density, carbon allocation, or, more generally, a number of carbon pools. Finally, in some context, vegetation dynamics may simply refer to a model that simulates the temporal evolution of processes connected to vegetation but not directly linked to the plant structure such as photosynthesis, respiration, or stomatal conductance. For the sake of generality, in the article, we use the broadest definition of vegetation dynamics that includes both fast and slow processes through which models simulate the temporal evolution of plant properties.

presented in Table 1. In the following, we summarize the processes that these models represent. Ecohydrological models typically solve water, energy, carbon, and, quite rarely, nutrient cycles at the land surface (Figure 6).

Water Budget

In terms of water budget, models use precipitation as an input that can be partitioned to rainfall and snowfall. Liquid precipitation can fall directly on the ground or be intercepted by the canopy, where it can subsequently evaporate or drip. Depending on the intensity of water flux reaching the ground and on soil water content (or, more appropriately, water potential) near the surface, water can either infiltrate the soil or run off from the surface. Solid precipitation can fall directly to the ground or become intercepted by vegetation where it can sublimate or subsequently fall to the ground. Snow accumulates on the ground in the form of a snowpack, where it melts or sublimates. Water in the soil undergoes vertical and horizontal redistribution following gravitational and soil matric potential gradients, typically modeled with the Richards equation.²⁷⁸⁻²⁸¹ Depending on the vertical profile of soil water potential and plant demand, water is evaporated from the surface as ground evaporation and/or taken up by plants and transpired. As water storage in the plants is typically ignored, root uptake and transpiration are exactly the same flux in most of the models. Water that percolates to deeper layers or into fractured bedrocks remains mostly inaccessible to plants (but see Ref 282) and typically contributes to aquifer recharge first and streamflow at a later stage. In models that do not have an explicit representation of the spatial dimension, this water is subtracted from the water budget as 'deep leakage' or 'recharge'. In models with an explicit spatial representation, after reaching the stream network as surface or sub-surface flow, water is routed through the channel network downstream to a specified outlet.

Energy Budget

Latent heat flux is the energy equivalent of the sum of all water vapor fluxes (transpiration and evaporation); therefore, it makes the water and energy cycles tightly coupled. Models that explicitly solve the energy budget compute the latent heat flux and its partition into different components (evaporation and transpiration) concurrently with the water budget (e.g., Refs 164,165,259,264,275,283). Latent heat flux (λ E) is typically solved assuming conservation of energy in a given domain:

$$R_{\rm n} - H - \lambda E - G - \lambda_{\rm p} A_{\rm n} - A_{\rm H} = \frac{\mathrm{d}S}{\mathrm{d}t},\tag{5}$$

where R_n is the net radiation, H is the sensible heat flux, G is the ground heat flux, λ_p is the specific energy consumed in the process of photosynthesis (A_n) , A_H is the advected energy to the domain either from lateral or vertical fluxes and dS/dt is the change in the energy stored in the system. Most of the models assume negligible storage capacity of energy, i.e., dS/dt = 0, and also neglect the last two terms of the left side of Eq. (5). Even with this assumption, the problem remains difficult to solve because all of the energy fluxes depend on one or, generally, more prognostic surface temperatures (e.g., soil and sunlit and shaded canopy temperatures). Models that do solve the energy budget face the problem of solving one or a system of non-linear equations embedded in Eq. (5) to derive the unknown surface temperature(s). This operation is typically

									Plant
Nodel	Key Reference	Water Budget	Energy Budget	Carbon Pools	Nutrients	Photosynthesis and Stomata	Spatial Representatior	Water Routing	Spatial Dynamics
Tethys-Chloris (T&C)	Fatichi et al. ^{17,165}	Y (Richards -Multilayer)	Y (1 prognostic temp.)	Y (7 Pools)	z	Y (Biochemical + Leuning)	Y (Grid)	~	z
tRIBS-VEGGIE	lvanov et al. ¹⁶⁴	Y (Richards -Multilayer)	Y (2 prognostic temp.)	Y (4 pools)	z	Y (Biochemical + Leuning)	Y (TIN)	≻	z
MLCan	Drewry et al ²⁵⁹	Y (Richards -Multilayer)	Y (Multilayer temp.)	z	z	Y (Biochemical + Ball-Berry)	z	z	z
RHESSys	Tague and Band, Tague et al. ^{260,261}	Y (Two layers)	N (Penman- Monteith)	Y (3 pools)	≻	Y (Empirical)	Y (Semi- distributed)	≻	z
CATHY/ NoahMP	Niu et al. ²⁶²	Y (Richards -Multilayer)	Y (2 prognostic temp.)	z	z	Z	Y (Grid)	≻	z
PAWS+CLM	Shen et al. ²⁶³	Y (Richards -Multilayer)	Y (3 prognostic temp.)	Y (4 Pools)	≻	Y (Biochemical + Leuning)	Y (Grid)	≻	z
Geotop-dv	Della Chiesa et al ²⁶⁴	Y (Richards -Multilayer)	Y (1 prognostic temp.)	Y (3 Pools)	z	Y (Empirical)	Y (Grid)	≻	z
CATGraSS	Zhou et al. ²⁶⁵	Y- Bucket	N (Penman- Monteith)	Y (3 pools)	z	Y (Empirical)	Y (Grid)	z	7
I	Van Wijk and Rodriguez- Iturbe, ²⁶⁶	Y (Bucket)	z	z	z	Z	۲	z	≻
VELMA	Abdelnour et al. ⁴⁰	Y (Four layers)	N (Empirical)	7	۲	Z	7	≻	z
CANOAK	Baldocchi and Wilson, ²⁶⁷	Y (Multilayer)	Y (Multilayer temp.)	z	z	Y (Biochemical + Ball-Berry)	z	z	Z
I	Nouvellon et al. ²⁶⁸	Y (Three layers)	N (Penman- Monteith)	Y (3 Pools)	z	Y (Empirical)	z	z	z
VDM	Montaldo et al. ²⁶⁹	Y (Bucket)	N (Penman- Monteith)	Y (3 Pools)	z	Y (Empirical)	z	z	z

TABLE 1 | A Non-Exhaustive List of Ecohydrological Models

Volume 3, May/June 2016

TABLE 1 C	ontinued								
				Carbon		Photosynthesis and	Spatial	Water	Plant Spatial
Model	Key Reference	Water Budget	Energy Budget	Pools	Nutrients	Stomata	Representatio	n Routing	Dynamics
Topog-IRM	Vertessy et al. ²⁷⁰	Y (Richards -Multilayer)	N (Penman- Monteith)	Y (3 pools)	z	/ (Empirical + Leuning)	Y (Flow-net)	≻	z
I	Cervarolo et al. ²⁷¹	Y (Richards -Multilayer)	Y (1 prognostic temp.)	Y (3 pools)	z	/ (Empirical + Jarvis)	Y (Grid)	≻	z
CABALA	Battaglia et al. ²⁷²	Y (Bucket)	N (Penman- Monteith)	Y (6 pools)	~	/ (Biochemical + Ball-Berry)	z	z	z
CenW 3.1	Kirschbaum et al. ²⁷³	Y (Multilayer)	N (Penman- Monteith)	Y (10 pools)	~	/ (Biochemical + Ball-Berry)	z	z	z
CASTANEA	Dufrene et al. ²⁷⁴	Y (Three layers)	Y (2 prognostic temp.)	Y (5 pools)	z	/ (Biochemical + Ball-Berry)	z	z	z
MuSICA	Ogée et al. ²⁷⁵	Y (Two layers)	Y (1 prognostic temp.)	z	z	/ (Biochemical + Leuning)	z	z	z
WEB-DHM	Wang et al. ²⁷⁶	Y (Richards -Multilayer)	~	z	z	/ (Biochemical + Ball-Berry)	Y (Grid)	≻	z
BEPS- TerrainLab	Govind et al. ²⁷⁷	Y (Two layers)	N (Penman- Monteith)	Y (4 Pools)	~	/ (Biochemical + Empirical)	~	~	z
The presence or a	bsence of specific processes is indicated	with a yes (Y) or no (N) a	nd with additional speci	fications.					



FIGURE 6 | Ecohydrological and terrestrial biosphere models have components and parameterizations to simulate the (1) surface energy exchanges, (2) the water cycle, (3) the carbon cycle, and (4) soil biogeochemistry and nutrient cycles. Many models do not include all the components presented in the figure.

computationally demanding. Therefore, several ecohydrological models still use simplified solutions of the energy budget such as the Penman–Monteith or Priestley-Taylor equations (e.g., Refs 260,265,269,273).

Carbon Budget

The carbon cycle is linked to the water and energy cycles because carbon assimilated through photosynthesis uses the same pathway between the outer atmosphere and leaf interior as transpired water (see the 'Stomatal Controls' section) and because changes in vegetation properties (e.g., plant height and LAI) modify boundary conditions for energy and water exchanges (Figure 6). For instance, a change in LAI modifies interception capacity, energy absorption and emission as well as roughness; a change in photosynthetic rate, A_n (Eq. (1)), may change stomatal conand therefore transpiration. ductance The computation of carbon assimilation can be carried out with various degrees of complexity. Some models use a biochemical model of photosynthesis in which $A_{\rm n}$ and leaf internal CO₂ concentration ($c_{\rm i}$) are computed as prognostic variables in a non-linear equation (e.g., Refs 164,165,259,275,276); others have simpler approaches exploiting the water use efficiency (WUE; i.e., the ratio between net carbon assimilation and transpiration²⁸⁴) or light use efficiency (LUE; i.e., the efficiency through which radiation absorbed by vegetation is converted into

carbon²⁸⁵) concepts that empirically link carbon assimilation to the transpired water or intercepted light (e.g., Refs 264,265,271,286,287). In some ecohydrological models, vegetation dynamics are essentially reduced to the simulation of carbon assimilation only (e.g., Refs 262,276). In others, the assimilated carbon is used to grow plants and to evolve a given number of carbon pools. Carbon pools are the way models account for the size and dynamics of different plant compartments.²⁸⁸ The number of carbon pools varies from model to model, but a typical set is composed of at least of a foliage pool, a fine-root pool, a sapwood or stem pool, and, more recently, a carbon reserve pool (e.g., Refs 165,273). Carbon reserves have been ignored in early ecohydrological models and ESMs with rare exceptions (e.g., Ref 289), but it is currently recognized that plant dynamics cannot be simulated meaningfully without accounting for carbon reserves.²⁹⁰⁻²⁹² Models that use carbon pools can also simulate the dynamics of the biophysical structure of vegetation, e.g., LAI, vegetation height, and root biomass.

Soil Biogeochemistry

Water, energy, and carbon fluxes are additionally connected through soil biogeochemistry and nutrient dynamics (Figure 6). Soil biogeochemistry is typically simulated to account for a given number of carbon and nitrogen pools.²⁹³ Other nutrients, such as phosphorous, sulfur, or potassium, are not typically

included in ecohydrological models and rarely so in ESMs (see the 'Terrestrial Biosphere Models' section). The number of pools varies with model complexity, but a typical approach is to explicitly represent the carbon and nitrogen contents in plant litter material and soil organic matter (SOM), partitioned into various subcomponents, as well as inorganic nutrients in mineral pools, which are directly available for plant uptake (e.g., Refs 294-298). In more recent model formulations, the SOM pool is partitioned among the substrate (available SOM) and the microbial biomass (bacteria, mycorrhiza) that feeds on the substrate,²⁹⁹ while soil fauna is typically neglected. In some models, extracellular enzymes catalyzing the decomposition reactions are also explicitly represented.³⁰⁰⁻³⁰² Exchanges between the different pools are typically simulated as first-order kinetics processes that account for temperature and soil moisture effects or as higher-order kinetics of the type described by the Michaelis-Menten equation.^{293,300,303,304} A fraction of carbon is lost in the decomposition process as CO2 through heterotrophic respiration. This is controlled by the capacity of the soil microbial biomass to assimilate carbon, defined as carbon use efficiency (CUE). The production of mineral nutrients (e.g., NO₃⁻, NH₄⁺) is mostly connected to the dynamics of the carbon pools and to the stoichiometry (e.g., carbon-nitrogen ratio) of litter and SOM.^{305,306} Part of the nutrients can be lost through deep leakage, erosion, or, in the case of nitrogen, through denitrification and volatilization; the remaining can be absorbed by roots. Plant nutrient uptakes represent an essential component of vegetation functioning. Nutrient uptake depends on the availability of mineral nutrients, root density, mycorrhiza symbiosis, and plant requirements, but exact mechanisms are not fully understood, and their simulation is typically quite simplified.^{298,299,307,308} The link with the carbon cycle is evident as soil biogeochemistry regulates the amount of heterotrophically respired CO_2 and controls the nutrients accessible to the plants. Nutrient availability affects plant development because of stoichiometric constraints in building plant tissues³⁰⁹ and because leaf nitrogen is known to affect photosynthetic capacity.³¹⁰⁻³¹³ The linkage with plant development offers important feedback on water and energy exchanges because plant structure (e.g., LAI, vegetation height) modifies boundary conditions at the land surface.²⁶³

Spatial Representation of Processes

The description of how models deal with water, energy, carbon, and nutrient cycles is weakly related

to the spatial dimension of the domain of simulation.

wires.wilev.com/water

In other words, the discussed processes can be used for a domain of $5 \times 5 \text{ m}^2$ as well as $100 \times 100 \text{ km}^2$, changing only the climate forcing and boundary conditions but largely preserving the same model structure. Although, the process' importance and heterogeneity clearly changes with the spatial scale, the conceptualization remains the same, and similar approaches have been used on very different scales. The main issue with an increasing spatial scale is the difficulty of prescribing appropriate boundary conditions and parameter values with a consequent problem related to the 'fallacy of average', i.e., the false assumption that the mean of a non-linear function of several variables equals the function of the means of these variables.^{314,315} Regardless of the scale, a spatial domain has a well-defined topography that needs to be accounted for because it affects climatic inputs such as shortwave and longwave radiation³¹⁶⁻³¹⁹ and rainfall³²⁰ and because lateral exchanges of water and nutrients may become significant. Lateral exchanges of energy are believed to be less important, but they are essentially unstudied. Distributed ecohydrological models are typically designed to account for the lateral exchanges of water^{320,321} and. sometimes, nutrients^{322,323} even when process description is simplified in a comparison with mechanistic solutions.²⁶⁵ The spatial redistribution of these resources in the simulated domain allows for an emergence of topographically controlled patterns in vegetation dynamics (e.g., different species composition in southern and northern exposed hillslopes, higher plant density near the stream network, etc.) that can eventually provide feedbacks about water and energy exchanges.

Another important aspect of the spatial dimension is represented by competition, establishment, and mortality of vegetation. There are a few ecohydrological models that aim to reproduce this specific behavior, and they are especially developed to study vegetation patterns semi-arid in environments.^{265,266,324} These models typically compromise the rigorousness of process description in terms of water, energy, and carbon dynamics, but they explicitly introduce ways, mostly based on stochastic approaches, to simulate seed dispersal, plant establishment and mortality, and their interaction with resource availability (water, radiation), which are absent in more complex models. At an even simpler level, models have been proposed to describe selforganized (or self-emergent) vegetation patterns in semi-arid environments using systems of partial differential equations borrowed from the dynamic systems theory (e.g., Refs 325). While these studies are



quite distant from the representation of the physical processes, they are important as they put forward hypotheses on the controls of vegetation spatial organization and regular patterns observed in semi-arid ecosystems.^{326–332}

GLOBAL SCALE

Terrestrial Biosphere Models

The request for quantifications of global carbon and water cycles and plant biogeography, especially in the context of understanding the consequences of climate change, was conducive to the development of models able to simulate vegetation dynamics (Box 2) on a global scale.^{13,333–337} These models appear with various names: dynamic global vegetation models (DGVMs), terrestrial biosphere models, terrestrial ecosystem models, and more recently, as vegetation components of ESMs (e.g., Refs 14,338-342). Here, the term terrestrial biosphere model is used to refer to this family of models.¹³ Typically, terrestrial biosphere models simulate water, energy, carbon, and nutrient cycles using an approach similar to the ecohydrological models described earlier. As a matter of fact, in most cases, formulations of carbon dynamics and soil biogeochemistry have been first developed for these models (e.g., Refs 15,343,344) and subsequently adopted by ecohydrological models. A detailed description of terrestrial biosphere models can be found in recent reviews by Prentice et al.,³⁴⁵ Levis,³⁴⁶ Quillet et al.,³⁴⁷ Medlyn et al.,³⁴⁸ and Fisher et al.¹³; a list of popular models is presented in Table 2 along with a summary of the specifications of each model. The simulated biophysical and biochemical processes are, to a large extent, similar to the processes described in the 'Plot to Catchment Scale' section. We rather emphasize the main differences in comparison to ecohydrological models related to (1) spatiotemporal scale of application, (2) model initialization and boundary conditions, and (3) biogeography and forest demography. We acknowledge that there is a tendency toward convergence in process descriptions between certain ecohydrological and terrestrial biosphere models, and in the future, the separation may become semantic or mostly a function of the type and scale of application.

A major distinction between ecohydrological and terrestrial biosphere models relies on the different spatiotemporal scales of their applications and, thus, the generality that the latter are meant to achieve. Terrestrial biosphere models are not tailored to any particular place or catchment and have to

simulate vegetation occurring on the entire Earth's surface. In addition, terrestrial biosphere models typically address research questions for longer time scales when compared with the time horizons of ecohydrological studies, even though exceptions exist, and some ecohydrological models have also been developed for long-term analyses.^{260,322} This implies that their structure needs to be flexible enough to include various processes and parameterizations and to simulate vegetation response across a large range of climatic conditions and time scales. However, structural and parameterization issues may emerge because of simplified assumptions introduced for the sake of generality and flexibility (e.g., Ref 371). Furthermore, because the spatial representation in terrestrial biosphere models is in the order of several km², they cannot account explicitly for topography. Each computational element may receive a different climate forcing, but, in essence, terrestrial biosphere models are one-dimensional as they do not exchange mass and nutrient fluxes laterally, and the effect of local topography in controlling meteorological inputs is not accounted for.

The large spatiotemporal scales of the typical applications carried out with terrestrial biosphere models follow a different philosophy for assigning the initial conditions (state variables) for the numerical simulations. For ecohydrological models, an initialization period of a few years is typically able to bring the examined ecosystem to a representative state (as the vegetation component is often assumed to be in a mature state). Longer spin-up periods are required for the initialization of terrestrial biosphere models that simulate short- and long-term soil biogeochemical processes (e.g., soil organic matter and nutrient dynamics). All terrestrial biosphere models account for soil carbon dynamics, many of them for nitrogen (e.g., Refs 297,298,349), and a minor frac-tion for phosphorous cycles.^{363,372,373} It is worth noting that, although often unrealistic, the assumption of vegetation in equilibrium with the observed environmental conditions (climate, nutrient deposition, weathering rates) is commonly and unavoidably applied for the spin-up of terrestrial biosphere models (e.g., Refs 315). In addition, contrary to the confined spatial extent (i.e., plot and catchment scales) and to the relatively short time-scales (i.e., less than 30 years) of the ecohydrological applications, the applications of terrestrial biosphere models address research questions on regional or global scales with time horizons of several decades or centuries. Therefore, land use and vegetation cover are often not pre-assigned, as typically done in many ecohydrological applications, but are prognostic variables

Model	Key Reference	Energy Fluxes	Water Fluxes	Carbon Module	Nitrogen Module	Plant Establ./Mortality/ Disturbances	Carbon Reserves	Root Exudation	Mychorrizal
CN-CLASS	Arain et al. ³⁴⁹	7	7	~	~	Z	7	z	z
JULES- TRIFFID	Clark et al. ³⁵⁰	~	~	7	z	٨	z	z	Z
Noah-MP	Niu et al. ³⁵¹	۲	۲	۲	Z	Z	z	z	z
LPJ	Sitch et al. ³⁵²	z	≻	۲	z	×	z	z	z
ORCHIDEE	Krinner et al. ³⁵³	۲	۲	۲	z	۲	7	Z	z
CLM-DGVM	Levis et al. ³⁵⁴	۲	≻	۲	z	×	z	z	z
CLM4CN	Lawrence et al. ³⁵⁵	۲	۲	٢	~	Z	z	z	z
IBIS	Kucharick et al. ³⁵⁶	۲	۲	٢	~	×	z	z	z
ISAM	Yang et al. ³⁵⁷	۲	۲	۲	~	Z	z	z	z
DLEM	Tian et al. ³⁵⁸	۲	۲	7	~	×	z	z	z
Biome-BGC	Thornton et al. ³⁵⁹	۲	۲	7	7	×	z	z	z
ED2	Medvigy et al. ³⁶⁰	۲	≻	۲	~	×	7	z	z
Ecosys	Grant et al. ³⁶¹	۲	۲	7	~	Z	7	z	۲
MOSES- TRIFFID	Cox ³⁶²	7	7	≻	z	۲	z	z	z
JSBACH	Goll et al. ³⁶³	۲	≻	۲	7	Z	7	۲	z
LPJ-DyN	Xu-Ri and Prentice ²⁹⁷	z	7	7	7	~	z	z	z
0-CN	Zaehle and Friend ²⁹⁸	≻	~	≻	~	7	~	z	z
Hybrid3.0	Friend et al. ²⁸⁹	۲	≻	7	~	7	7	z	z
ANAFORE	Deckmyn et al. ¹⁶⁷	۲	≻	7	7	۲	×	z	z
SEIB-DGVM	Sato et al. ³⁶⁴	z	≻	۲	z	۲	×	z	z
Sheffield- DGVM	Woodward and Lomas ³⁶⁵	z	7	~	7	٨	z	z	z
LPJ-GUESS	Smith et al. ³⁶⁶	z	۲	٢	z	۲	z	z	z
aDGVM2	Scheiter et al. ³⁶⁷	z	≻	۲	z	×	z	z	z
FoBAAR	Keenan et al. ³⁶⁸	z	z	7	z	Z	7	z	z
CASA	Carvalhais et al. ³⁶⁹	z	≻	۲	~	Z	z	z	z
JeDi-DGVM	Pavlick et al. ³⁷⁰	z	7	~	Z	,	~	Z	z

computed by the model. For the same reasons, processes such as forest demography and natural or anthropogenic disturbances such as fire, windthrow, plant diseases, insect attacks, forest management, and plant competition needs to be represented. Feedback on vegetation dynamics caused by nutrient cycles is also much more relevant at those temporal scales than they are for ecohydrological applications, at least in forests.³⁷⁴ Forest demography in terrestrial biosphere models is similar or identical to the one developed for forest-gap models in forestry research.^{61,375,376} Forest demography can be handled by an average tree individual^{352,353,377} evolving in time or by simulating cohorts of individuals with different age classes and size in the stand (e.g., Refs 61,315,360,366,378-380). This allows, for instance, an explicit representation of the early and late successional species and more generally of heterogeneous forests with gaps, uneven stand height, and density.^{381,382} Disturbances are typically very difficult to simulate mechanistically even though a general framework has been recently presented³⁸³. They are mostly simulated by removing a certain amount of biomass in a given domain with a given probability of disturbance occurrence. Plant competition is often simulated by implicitly assigning 'bioclimatic limits' of existence (temperature, soil moisture, or light thresholds for establishment) to specific species or plant functional types or removing plants that are underperforming from a carbon balance perspective (e.g., Refs 352,370). Mortality is far from being mechanistically simulated³⁸⁴ and is typically related to plant-negative carbon balances or prescribed with given constant probabilities or as a function of plant age or size.^{385,386} This background mortality is typically summed with the probability of disturbance occurrence.

Vegetation Feedback to the Climate System

There are several examples of the successful applications of terrestrial biosphere models, which elucidate the feedback between vegetation dynamics and climate or hydrological cycles on global and regional scales. One example is the predictions of the fate of the Amazon forest. Early findings suggested the potential dieback of the Amazon rainforest in response to projected droughts with huge implications for the future of Earth's climate.³⁸⁷ While these extreme projections have been moderated using more refined models, the response of the Amazon forest to a changing climate remains very uncertain (e.g., Refs 388–391). There is a large community effort dedicated to better understand the effect of drought on the Amazon forest (e.g., Refs 392–395) because current observations and future predictions are suggesting a potential lengthening of the dry intervals and an increase of drought risk.^{396,397} On local scales, the Amazon's deforestation has been demonstrated to affect cloud formation and convective precipitation because it changes the partition of net radiation into sensible and latent heat, affecting boundary layer development. This could possibly increase or decrease convective precipitation downwind of the cleared patches as a function of the extent of deforestation.^{398–400}

Another significant example is represented by the tree and shrub encroachment observed at high latitudes. The prognostic nature of vegetation cover in terrestrial biosphere models allows them to simulate shrub encroachment and tree line shifts following, for example, Earth's warming. While the exact causes of tree and shrub encroachment are still uncertain,^{401,402} shrub presence in previously non-vegetated or poorly vegetated areas of the Arctic has the capacity to modify the hydrology and the energy exchanges between the land and the atmosphere. If shrubs grow taller than typical winter snowpack height, the albedo of the surface is substantially decreased, and at the same time, the longwave radiation regime is modified.⁴⁰³ These changes in vegetation have a positive effect on local and global warming with the potential to further push shrub encroachment or northern tree line expansion.43

A final example of important interactions between plants and the water cycle is related to the consequences of increasing atmospheric CO₂ concentration. Elevated CO2 concentration has a wellknown direct physiological effect at the leaf level, increasing photosynthesis and reducing stomatal conductance.^{404,405} The increased plant productivity associated with elevated CO₂ is typically indicated as the CO₂ fertilization effect (e.g., Ref 406). The integration of this effect from the leaf to the global scale is still debated, 407,408 but it has been postulated to have contributed to an increase runoff^{409,410} and be largely responsible for the terrestrial carbon sink with direct feedback on Earth's climate.^{27,411,412} While the magnitude of CO₂ fertilization on the ecosystem scale and the persistence of the carbon sink in time are still open research areas,⁴¹³⁻⁴¹⁵ the expected consequences of elevated CO2 represent one additional example that robust, long-term projections of future water cycles cannot be made without accounting for vegetation physiology.

PROCESS UP-SCALING

Forcing and Boundary Conditions

The models described in the previous sections (plant hydraulic, ecohydrological, and terrestrial biosphere models) are designed to address scientific questions on different spatiotemporal scales. Plant hydraulic models are typically forced with very detailed meteorological inputs and boundary conditions (e.g., soil type, tree height and diameter, root depth, plant hydraulic traits, etc.), but they are used to simulate a single tree or a specific forest stand and for a relatively short period of time (a growing season, few years). It would be unrealistic to simulate each individual tree on the continental scale. At larger spatiotemporal scales, different model types are typically applied. Ecohydrological models are normally employed from the plot to the catchment scale, while terrestrial biosphere models are used from the plot to the global scale. As described previously, terrestrial biosphere models cannot account for fine-scale heterogeneities (e.g., topography, climate) because the spatial resolution at which they usually operate is in the order of several km². This limitation has been shown to be more relevant than previously thought because the mean response obtained with meteorological forcing and boundary conditions mediated over a large area can be different from the mean of the responses obtained with a detailed treatment of input and boundary conditions.³¹⁵ In other words, the non-linear dynamics inherent to hydrological and vegetation processes play a considerable role. In an analogy, the temporal resolution of the model is also very important^{416,417} because coarse temporal resolutions do not allow the solving of non-linear effects produced by short-term variability in the forcing (e.g., shortwave radiation). In this regard, however, differences among models are less marked; most of the ecohydrological and terrestrial biosphere models use an hourly or half-hourly time step for the forcing, at least to solve hydrological and energy dynamics, with a few models still using daily or longer scales (e.g., Ref 366). Plant growth, soil biogeochemistry, and forest demography processes have typically slower dynamics that can be simulated on the daily time scale or even annually for processes such as forest demography.

Vegetation Heterogeneity

A very important 'boundary condition' in any model that deals with vegetation is how vegetation itself is parameterized. Historically, the existing diversity in species and plant traits (e.g., Ref 418) has been

summarized using a given number (~8-25) of plant functional types (PFTs) that aimed to distinguish climatic zones (e.g., temperate, boreal, tropical), phenology (evergreen, deciduous), and leaf morphology (e.g., needle leaves, broadleaves) of the plant.⁴¹⁹ This approach has been utilized in both ecohydrological and terrestrial biosphere models because it allowed a synthesis of important differences in plant properties as well as phenological behavior without entering in the details of exact species or intra-species variability. Recently, the PFT approach has been widely criticized, and new approaches based on the definition of plant traits and their distributions are becoming progressively popular (e.g., Refs 36,420-422). These approaches rely on the knowledge of the observed distributions of plant properties, 'traits', which can correspond or be related to model parameters, such as maximum photosynthetic capacity, leaf nitrogen content, specific leaf area, root depth and turnover rates, leaf dimension, etc. Beyond the distribution of the values of a given plant trait, there are several ecological studies that show a certain degree of coordination between many of these traits.423-425 An illustrative example is the 'leaf economics spectrum' that suggests that leaf traits co-vary in a continuum from leaves with a high photosynthetic capacity, nitrogen content, specific LAI, and a short lifespan typical of fast-growing species to the opposite combination typical of slow-growing species.^{423,426} While cross-correlations have been mostly studied for leaf traits, there is emerging evidence that this holds true for many plant traits, 427 and such information can be transferred to the model parameters. Observed trait distributions and cross-correlations can be used to create 'proxy species', which do not correspond exactly to an observed species but are deemed as realistic within the range of observed natural variation. Models can then use a large number of these 'proxy species' to investigate the importance of variability of plant traits in carbon and water dynamics in a given region or directly use observed distributions of plant traits as input. Note that the latter should be typically weighted for the occurrence of a given species in a specific place or region. In fact, despite the huge amount of biodiversity, forest biomass and productivity are mostly controlled by a limited number of dominant species.⁴²⁸ A great advantage of the probabilistic approximation of plant trait diversity, based on multivariate distributions of whole-plant trait spectra rather than on few PFTs or species, is that it allows for a probabilistic assessment of ecosystem response and thus provides a framework for quantifying uncertainties related to vegetation spatiotemporal heterogeneity.³⁶

DATA FOR MODEL PARAMETERIZATION AND CONFIRMATION

Any model, regardless of the scale, needs climate forcings as well as data that can be used for confirmation of model results. Meteorological forcing requirements are similar in most of the models and include air temperature, precipitation, relative humidity, wind speed, atmospheric pressure, CO₂ concentration, and incoming shortwave and longwave radiation. Boundary conditions consist of soil properties, such as soil depth, soil texture, or soil hydraulic properties (e.g., Refs 429,430), and vegetation properties represented by functional and morphological plant traits (see the 'Vegetation Heterogeneity' section). In the case of distributed simulations, land cover, topographical, and geological information is also required. Modeling of soil biogeochemistry and nutrient cycle theoretically needs additional data

about initial values of nutrient content, nutrient deposition, and primary minerals weathering together with a number of parameters that characterize kinetics and turnover rates of microbial biomass and organic substrate in the soil (e.g., Refs 300,431,432).

There are multiple measurements that can potentially be used in models and that are carried out directly on the plant scale in one or multiple individuals. These are leaf and stem water potential, sap flow, stem diameter changes, leaf turgor, leaf temperature, and gas exchange measurements at the leaf, root, and stem level (Table 3). Additionally, plant hydraulic traits such as stem capacitance and vulnerability curves or biochemical traits such as the ones related to photosynthesis are necessary for the accurate parameterization of stomata and plant hydraulic models (e.g., Ref 253). When combined together, these measurements provide a rather complete characterization of plant behavior with a notable exception represented by short-term carbon allocation

 TABLE 3
 List of Variables That Can Be Potentially Observed on Different Scales

Scale	Variable	Instrument/Product
Plant	Sap flow	Sap flow meter
Plant	Leaf water potential	Pressure chamber
Plant	Stem water potential	Psychrometer
Plant/stand	Tree/branch diameter	Forest inventory
Leaf	Photosynthesis, stomatal conductance	Gas analyzers
Leaf/stand	Leaf/canopy temperature	IR thermometers, thermocouple, thermal cameras
Leaf	Turgor	Turgor probes or estimated from water potential and osmolyte concentration
Plant	Plant respiration	Gas analyzers
Plant	Stem water content	Coring, moisture probe
Stand	Evapotranspiration	Lysimeter, flux towers
Point/stand	Soil respiration	Gas analyzers
Plant/stand/global	Leaf Area Index	Direct sampling, optical sensors, remote sensing
Plant	NSC dynamics (leaves and branches and fine roots)	Sampling and laboratory analysis
Point/stand	Soil water potential (tensiometers) and volumetric water content	Soil moisture probe and tensiometer, remote sensing
Point/stand	Soil temperature	Thermometers, optical sensor cable
Stand	Energy fluxes (λΕ, Η)	Flux towers
Stand	Carbon fluxes (NEE)	Flux towers
Stand/regional	Aboveground carbon stocks	Forest inventories, remote sensing
Stand/regional	Tree height	Forest inventories, remote sensing
Regional/global	Vegetation reflectivity indices (NDVI, EVI, chlorophyll fluorescence)	Remote sensing
Regional/global	Surface temperature	Remote sensing, IR thermometers
Regional/global	CO ₂ anomalies	Remote sensing, ground long-term observatories
Regional/global	Albedo	Remote sensing

wires.wiley.com/water

dynamics (phloem transport, tissue growth, and respiration), which are currently very difficult or impossible to measure in the field and mostly limited to leaf growth and point measurements of NSCs repeated a few times during a season.⁴³³

Moving from the plant to the plot/stand scale, measurements are not typically carried out anymore on the individual plants but express an integrated quantity over a given area. This type of measurement includes: (1) energy and mass exchange between the land surface and the atmosphere obtained with fluxtower monitoring systems that employ the eddy covariance technique $^{434-438}$ and (2) soil water content and temperature and soil respiration at different locations. Measurements of soil water content provide a direct quantification of one of the key variables in the water-vegetation interaction but are difficult to upscale from the sensor level to stand because considerable scale of spatial heterogeneities.^{152,439-441} Flux-tower measurements are instead an up-scaled quantity but have problems of continuity and interpretation.442,443 Despite wellrecognized issues,444,445 flux-tower observations have represented and still represent the major source of validation for ecohydrological and terrestrial biosphere models as they provide values for variables such as latent and sensible heat, net radiation, and mass fluxes of water vapor and carbon. Lysimeters can also be used to directly measure the leaching of water and nutrients and the water content variation in soil monoliths, and, therefore, evapotranspiration.446,447 This measurement is probably the most precise and continuous estimate of evapotranspiration fluxes, but it is very expensive to set up and maintain. Moving from fluxes to stocks, forest inventories represent an important source of information that characterizes standing and dead biomass and plant demography (age, size, and species). When inventories are repeated systematically in time, they also provide information on plant turnover rates, mortality, and growth (e.g., Refs 448-451), and they can be used for model testing (e.g., Refs 315,382). Recently, tree ring dendrochronology has been presented as a possibility to test simulations of carbon accumulation as well as the effect of infrequent disturbances.⁴⁵² Despite the numerous problems that exist in the scaling from single trees to areal estimates, it represents a potential additional source for model validation.

An alternative way to test models is the recourse to observations carried out during ecological manipulation experiments, e.g., rainfall exclusion and addition, warming, CO₂ enrichment (FACE), girdling, and nutrient additions. Such experiments

can be used to test whether models can reproduce the effect of treatment at least in terms of observed sensitivity (e.g., Refs 394,453–456). When models are able to do so without changing their parameterization, this represents an important validation of their predictive skills for conditions different from the current climate. Manipulation experiments have mostly been focused on measuring vegetation productivity, especially in grasslands, but other metrics can also be monitored and used for model validation, especially if new experiments will be designed with this additional scope in mind.^{456–458} Despite their potential importance, manipulation experiments have been seldom used for model testing so far.

Moving to larger spatial scales, direct observations become more challenging, and remote-sensing devices such as Lidar and hyperspectral cameras have been recently employed. Observational devices can be placed over land or aircraft platforms or on satellites.^{459,460} Remote sensing observations can be carried out with optic or radar sensors and are mostly used to compute vegetation reflectivity indices, carbon stocks, and soil moisture. Carbon stocks are typically observed with radar sensors on a regional scale (e.g., Refs 461,462). Optic observations in multiple spectral bands typically map the entire Earth, and they are used to derive vegetation indices as the Enhanced Vegetation Index (EVI), the Normalized Difference Vegetation Index (NDVI), or chlorophyll fluorescence.^{463,464} Signals in multiple bands can also be processed and converted into variables such as LAI, carbon assimilation,^{465,466} or nutrient content of the leaves,⁴⁶⁷ although the reliability of these estimates is still uncertain (e.g., Refs 468,469). A few remote-sensing products are also available for nearsurface soil water content (e.g., Ref 470), and promising results in this direction are expected from campaigns such as Soil Moisture Active Passive (SMAP).⁴⁷¹ Finally, anomalies of atmospheric CO₂ concentration are an important source of information to estimate the net carbon uptake or release over very large areas and on the seasonal and annual time scale,^{412,472,473} and they may become even more so in the future.⁴⁷⁴

CONCLUSION

The examined literature suggests that approaches to simulate the intertwined dynamics of water, energy, carbon, and nutrient cycles are becoming widely available and that a number of models of different complexity has been presented to address problems on various scales. While the combination of hydrology and ecology can be considered a reality, additional efforts need to be devoted to integrate solutions developed for different scales.

The issue of scale is not merely an operational one related to finding better parameterizations but requires innovative approaches to preserve key physical and ecological mechanisms operating locally as well as existing heterogeneities in larger-scale applications. We do not mean that the entire complexity of plant-hydraulic or detailed biogeochemistry models should be retained in ecohydrological or terrestrial biosphere models, but there is a wide margin to better use the insights gained at local scales in watershed, regional, and global applications (see Ref 127). For instance, models simulating plant-hydraulic and carbon dynamics mechanistically are still rare and confined to specialized studies,^{206,212,213,218} but they have the potential to considerably improve the representation of fundamental aspects such as plant growth, response to elevated CO₂, response to severe droughts or pathogen attacks, and ultimately plant mortality. These are subjects where we still have gaps in process understating and where a well-structured combination of modeling and observations is likely to provide insights well beyond model improvements, potentially increasing fundamental knowledge of plant-water interactions. Modeling plant mortality has been regarded as one of the great scientific challenges⁴⁷⁵ and surely it is, but we should not forget that current models still struggle to correctly simulate the response to water limitations⁴⁷⁶ and have a limited flexibility to accommodate diverging responses across species or within a landscape.^{36,315}

Most of the processes underlying plant-water interactions are highly non-linear; this implies that

averaging significant heterogeneities in space and time may lead to considerable biases. While the importance of solving ecohydrological processes at the sub-daily temporal scales is progressively recognized^{416,417,477} and implemented in most of the models, the relevance of being spatially explicit has been less emphasized in literature.³¹⁵ Only a few studies presented process-based simulations accounting explicitly for topographic attributes and lateral water and mass exchanges.^{261,263,320,321} Applications of ecohydrological models at a regional or continental scale or the introduction of topography in terrestrial biosphere models will represent possible solutions. These solutions lead to operational issues related to computational resources and accurate model parameterizations. Large spatial and long temporal scales pose great challenges to model initialization and validation as current available data are typically inadequate when compared to the diversity of model outputs. A better integration of data from multiple platforms and scales and especially of remote-sensing estimates is thus essential for improving distributed and/or continental scale models.^{478,479} A more rigorous treatment of uncertainty with stochastic analyses and probabilistic predictions should also represent an important avenue for the future as many model parameters (e.g., plant and microbial traits) are unlikely to be known with a high accuracy everywhere and vary not only in space but also in time.³⁶ Studies and modeling of plant-water interactions have seen great advancements in the past years, but current shortcomings and the challenges posed by environmental changes and preservation of water resources and ecosystem services suggest that exciting research opportunities are still ahead.

ACKNOWLEDGMENTS

SF thanks the Stavros Niarchos Foundation and the ETH Zurich Foundation (Grant ETH-29 14-2) for their support. CP acknowledges the support of the Stavros Niarchos Foundation and the ETH Zurich Foundation (Grant P2EZP2_162293) through the SNSF Early Postdoc Mobility Fellowship. VI acknowledges the support of NSF Grant EAR 1151443.

REFERENCES

- 1. Oki T, Kanae S. Global hydrological cycles and world water resources. *Science* 2006, 313:1068–1072. doi:10.1126/science.1128845.
- 2. Katul GG, Oren R, Manzoni S, Higgins C, Parlange MB. Evapotranspiration: a process driving mass transport and energy exchange in the soil-plant-atmosphere-climate system. *Rev Geophys* 2012, 50: RG3002.
- Schlesinger WH, Jasechko S. Transpiration in the global water cycle. Agr Forest Meteorol 2014, 189-190:115–117. doi:10.1016/j.agrformet.2014.01.011.
- 4. Wild M, Folini D, Hakuba MZ, Schär C, Seneviratne SI, Kato S, Rutan D, Ammann C, Wood EF, König-Langlo G. The energy balance over land and oceans: an assessment based on direct observations and

CMIP5 climate models. *Climate Dynam* 2015, 44:3393–3429.

- Eagleson PS. Climate, soil, and vegetation 1. Introduction to water balance dynamics. Water Resour Res 1978, 14:705-712.
- 6. Bonan GB. *Ecological Climatology*. Cambridge, UK: Cambridge University Press; 2008.
- 7. Berry JA, Beerling DJ, Franks PJ. Stomata: key players in the Earth system past and present. *Curr Opin Plant Biol* 2010, 13:232–239.
- 8. Nobel PS. *Physicochemical and Environmental Plant Physiology*. Oxford, UK: Elsevier Academic Press; 2009.
- 9. Brown AD. *Microbial Water Stress Physiology, Principles and Perspectives*. Chichester: John Wiley and Sons; 1990.
- 10. Anderson SP, von Blanckenburg F, White AF. Physical and chemical controls on the critical zone. *Elements* 2007, 3:315–319.
- McMahon TA, Peel MC, Lowe L, Srikanthan R, McVicar TR. Estimating actual, potential, reference crop and pan evaporation using standard meteorological data: a pragmatic synthesis. *Hydrol Earth Syst Sci* 2013, 17:1331–1363. doi:10.5194/hess-17-1331-2013.
- Gerten D, Schaphoff S, Haberlandt U, Lucht W, Sitch S. Terrestrial vegetation and water balancehydrological evaluation of a dynamic global vegetation model. J Hydrol 2004, 286:249–270.
- Fisher JB, Huntzinger DN, Schwalm CR, Sitch S. Modeling the Terrestrial Biosphere. Annu Rev Environ Resour 2014, 39:91–123. doi:10.1146/ annurev-environ-012913-093456.
- 14. Sellers PJ, Mintz Y, Sud YC, Dalcher A. A simple biosphere model (SiB) for use within general circulation models. *J Atmos Sci* 1986, 43:505–531.
- 15. Sellers PJ, Randall DA, Collatz GJ, Berry JA, Field CB, Dazlich CA, Zhang C, Collelo D, Bounoua L. A revised land surface parameterization (SiB2) for atmospheric GCMs. 1. Model formulation. *J Climate* 1996, 9:674–705.
- 16. Sellers PJ, Dickinson RE, Randall DA, Betts AK, Hall FG, Berry JA, Collatz GJ, Denning AS, Mooney HA, Nobre CA, et al. Modeling the exchanges of energy, water and carbon between continents and the atmosphere. *Science* 1997, 275:502–509.
- Fatichi S, Zeeman MJ, Fuhrer J, Burlando P. Ecohydrological effects of management on subalpine grasslands: from local to catchment scale. Water Resour Res 2014, 50:148–164. doi:10.1002/ 2013WR014535.
- Mu Q, Zhao M, Running SW. Improvements to a MODIS global terrestrial evapotranspiration algorithm. *Remote Sens Environ* 2011, 115:1781–1800.

- 19. Bornmann L, Mutz R. Growth rates of modern science: a bibliometric analysis based on the number of publications and cited references. J Assoc Inform Sci Technol 2015, 66:2215–2222. doi:10.1002/asi.23329.
- Sack L, Scoffoni C. Leaf venation: structure, function, development, evolution, ecology and applications in the past, present and future. *New Phytol* 2013, 198:983–1000. doi:10.1111/nph.12253.
- 21. Rodriguez-Iturbe I. Ecohydrology: a hydrological perspective of climate-soil-vegetation dynamics. *Water Resour Res* 2000, 36:3–9.
- 22. Porporato A, Laio F, Ridolfi L, Rodriguez-Iturbe I. Plants in water-controlled ecosystems: active role in hydrologic processes and response to water stress III. Vegetation water stress. *Adv Water Resour* 2001, 24:725–744.
- 23. Eagleson PS. Ecohydrology: Darwinian Expression of Vegetation Form and Function. Cambridge: Cambridge University Press; 2002.
- 24. Rodriguez-Iturbe I, Porporato A. *Ecohydrology of Water-Controlled Ecosystems*. Cambridge, UK: Cambridge University Press; 2004.
- 25. Bond B. Hydrology and ecology meet-and the meeting is good. *Hydrol Process* 2003, 2087:2089. doi:10.1002/hyp.5133.
- 26. Friedlingstein P, Cox PM, Betts RA, Bopp L, Von-Bloh W, Brovkin V, Cadule P, Doney S, Eby M, Fung I, et al. Climate-carbon cycle feedback analysis: results from the C4MIP model intercomparison. J Climate 2006, 19:3337–3353.
- 27. Arora VK, Boer GJ, Friedlingstein P, Eby M, Jones CD, Christian JR, Bonan LBG, Brovkin V, Cadule P, Hajima T, et al. Carbon-concentration and carbon-climate feedbacks in CMIP5 Earth System models. *J Climate* 2013, 26:5289–5314. doi:10.1175/JCLI-D-12-00494.1.
- Friedlingstein P, Meinshausen M, Arora VK, Jones CD, Anav A, Liddicoat SK, Knutti R. Uncertainties in CMIP5 climate projections due to carbon cycle feedbacks. J Climate 2014, 27:511–526.
- Mooney H, Larigauderie A, Cesario M, Elmquist T, Hoegh-Guldberg O, Lavorel S, Mace GM, Palmer M, Scholes R, Yahara T. Biodiversity, climate change, and ecosystem services. *Curr Opin Environ Sustain* 2009, 1:46–54. doi:10.1016/j.cosust.2009.07.006.
- Istanbulluoglu E, Bras RL. Vegetation-modulated landscape evolution: effects of vegetation on landscape processes, drainage density, and topography. *J Geophys Res* 2005, 110:F02012. doi:10.1029/ 2004JF000249.
- Lobell DB, Schlenker W, Costa-Roberts J. Climate trends and global crop production since 1980. *Science* 2011, 333:616–620. doi:10.1126/science.1204531.

- 32. Dai A, Trenberth KE, Qian T. A global dataset of Palmer drought severity index for 1870-2002: relationship with soil moisture and effects of surface warming. *J Hydrometeorol* 2004, 5:1117–1130.
- Sheffield J, Wood EF, Roderick ML. Little change in global drought over the past 60 years. *Nature* 2012, 491:435-438. doi:10.1038/nature11575.
- Fu Q, Feng S. Responses of terrestrial aridity to global warming. J Geophys Res Atmos 2014, 119:7863-7875. doi:10.1002/2014JD021608.
- 35. Roderick ML, Greve P, Farquhar GD. On the assessment of aridity with changes in atmospheric CO₂. Water Resour Res 2015, 2:5450–5463. doi:10.1002/2015WR017031.
- Pappas C, Fatichi S, Burlando P. Modeling terrestrial carbon and water dynamics across climatic gradients: does plant diversity matter? *New Phytol* 2015. doi:10.1111/nph.13590.
- Inauen N, Körner C, Hiltbrunner E. Hydrological consequences of declining land use and elevated CO₂ in alpine grassland. *J Ecol* 2013, 101:86–96. doi:10.1111/1365-2745.12029.
- 38. Gough CM, Hardiman BS, Nave LE, Bohrer G, Maurer KD, Vogel CS, Nadelhoffer KJ, Curtis PS. Sustained carbon uptake and storage following moderate disturbance in a great lakes forest. *Ecol Appl* 2013, 23:1202–1215.
- Biederman JA, Harpold AA, Gochis DJ, Ewers BE, Reed DE, Papuga SA, Brooks PD. Increased evaporation following widespread tree mortality limits streamflow response. Water Resour Res 2014, 50:5395–5409. doi:10.1002/2013WR014994.
- 40. Abdelnour A, Stieglitz M, Pan F, McKane R. Catchment hydrological responses to forest harvest amount and spatial pattern. *Water Resour Res* 2011, 47: W09521. doi:10.1029/2010WR010165.
- 41. Brown AE, Zhang L, McMahon TA, Western AW, Vertessy RA. A review of paired catchment studies for determining changes in water yield resulting from alterations in vegetation. *J Hydrol* 2005, 310:28–61. doi:10.1016/j.jhydrol.2004.12.010.
- 42. Guardiola-Claramonte M, Troch PA, Breshears DD, Huxman TE, Switanek MB, Durcik M, Cobb NS. Decreased streamflow in semi-arid basins following drought-induced tree die-off: a counter-intuitive and indirect climate impact on hydrology. *J Hydrol* 2011, 406:225–233.
- 43. Pearson RG, Phillips SJ, Loranty MM, Beck PSA, Damoulas T, Knight SJ, Goetz SJ. Shifts in Arctic vegetation and associated feedbacks under climate change. *Nat Clim Change* 2013, 3:673–677.
- 44. Matheny A, Bohrer G, Vogel CS, Morin TH, He L, Frasson RPM, Mirfenderesgi G, Schäfer KVR, Gough CM, Ivanov VY, et al. Species-specific transpiration responses to intermediate disturbance in a

northern hardwood forest. J Geophys Res 2014, 119:2292-2311.

- 45. Pomeroy JW, Marks DM, Link T, Ellis C, Hardy J, Rowlands A, Granger R. The impact of coniferous forest temperature on incoming longwave radiation to melting snow. *Hydrol Process* 2009, 23:2513–2525.
- Ellis CR, Pomeroy JW, Brown T, MacDonald J. Simulation of snow accumulation and melt in needleleaf forest environments. *Hydrol Earth Syst Sci* 2010, 14:925–940. doi:10.5194/hess-14-925-2010.
- 47. Broxton PD, Harpold AA, Biederman JA, Troch PA, Molotch NP, Brooks PD. Quantifying the effects of vegetation structure on snow accumulation and ablation in mixed-conifer forests. *Ecohydrology* 2015, 8:1073–1094. doi:10.1002/eco.1565.
- Bohrer G, Katul GG, Walko RL, Avissar R. Exploring the effects of microscale structural heterogeneity of forest canopies using large-eddy simulations. *Bound-Lay Meteorol* 2009, 132:351–382. doi:10.1007/s10546-009-9404-4.
- Mahat V, Tarboton DG, Molotch NP. Testing aboveand below-canopy representations of turbulent fluxes in an energy balance snowmelt model. *Water Resour Res* 2013, 49:1107–1122. doi:10.1002/wrcr.20073.
- Lundquist JD, Dickerson-Lange SE, Lutz JA, Cristea NC. Lower forest density enhances snow retention in regions with warmer winters: a global framework developed from plot-scale observations and modelling. Water Resour Res 2013, 49:6356-6370. doi:10.1002/wrcr.20504.
- 51. Duarte CM, Middelburg JJ, Caraco N. Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* 2005, 2:1–8.
- Katul GG, Poggi D, Ridolfi L. A low resistance model for assessing the impact of vegetation on flood routing mechanics. *Water Resour Res* 2011, 47:W08533. doi:10.1029/2010WR010278.
- 53. Kim J, Ivanov VY, Katopodes ND. Hydraulic resistance to overland flow on surfaces with partially submerged vegetation. *Water Resour Res* 2012, 48: W10540. doi:10.1029/2012WR012047.
- 54. Marjoribanks TI, Hardy RJ, Lane SN. The hydraulic description of vegetated river channels: the weaknesses of existing formulations and emerging alternatives. WIREs Water 2014, 1:549–560. doi:10.1002/ wat2.1044.
- 55. Rodriquez-Iturbe I, D'Odorico P, Porporato A, Ridolfi L. On the spatial and temporal links between vegetation, climate, and soil moisture. *Water Resour Res* 1999, 35:3709–3722.
- Ridolfi L, D'Odorico P, Porporato A, Rodriguez-Iturbe I. Duration and frequency of water stress in vegetation: an analytical model. *Water Resour Res* 2000, 36:2297–2307.

- Guswa AJ. The influence of climate on root depth: a carbon cost-benefit analysis. Water Resour Res 2008, 44:W02427. doi:10.1029/2007WR006384.
- Thompson SE, Harman CJ, Troch PA, Brooks PD, Sivapalan M. Spatial scale dependence of ecohydrologically mediated water balance partitioning: a synthesis framework for catchment ecohydrology. *Water Resour Res* 2011, 47:W00J03. doi:10.1029/ 2010WR009998.
- 59. Manzoni S, Vico G, Katul GG, Palmroth S, Jackson RB, Porporato A. Hydraulic limits on maximum plant transpiration and the emergence of the safety-efficiency trade-off. *New Phytol* 2013, 198:169–178. doi:10.1111/nph.12126.
- 60. Manzoni S, Vico G, Katul GG, Palmroth S, Porporato A. Optimal plant water use strategies under stochastic rainfall. *Water Resour Res* 2014, 2014:5379–5394. doi:10.1002/2014WR015375.
- 61. Bugmann H. A review of forest gap models. *Clim Change* 2001, 51:259–305.
- 62. Weiskittel AR, Hann DW, Kershaw JA, Vanclay JK. Forest Growth and Yield Modeling. Oxford: Wiley-Blackwell; 2011.
- 63. Tyree MT. The ascent of water. *Nature* 2003, 423:923.
- 64. Campbell GS, Norman JM. An Introduction to Environmental Biophysics. New York: Springer-Verlag New York, Inc.; 1998.
- 65. Zimmermann MH. Transport in the phloem. Annu Rev Plant Physiol 1960, 11:167–190.
- 66. Pickard WF. The ascent of sap in plants. Prog Biophys Mol Biol 1981, 37:181-229.
- 67. Holbrook NM, Zwieniecki MA. Vascular Transport in Plants. Amsterdam: Elsevier; 2005.
- Jensen KH, Berg-Sørensen K, Friis SMM, Bohr T. Analytic solutions and universal properties of sugar loading models in Münch phloem flow. *J Theor Biol* 2012, 304:286–296.
- 69. Stroock AD, Pagay VV, Zwieniecki MA, Holbrook NM. The physicochemical hydrodynamics of vascular plants. *Annu Rev Fluid Mech* 2014, 46:615–642.
- 70. Taiz L, Zeiger E. *Plant Physiology*. Sunderland, MA: Sinauer Associates Inc; 2006.
- 71. von Caemmerer S, Farquhar GD. Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves. *Planta* 1981, 153:376–387.
- Evans JR, Kaldenho R, Genty B, Terashima I. Resistances along the CO₂ diffusion pathway inside leaves. *J Exp Bot* 2009, 60:2235–2248. doi:10.1093/jxb/erp117.
- 73. Bernacchi CJ, Bagley JE, Serbin SP, Ruiz-Vera UM, Rosenthal DM, Van-Loocke A. Modelling C3 photosynthesis from the chloroplast to the ecosystem. *Plant Cell Environ* 2013, 36:1641–1657. doi:10.1111/ pce.12118.

- 74. Franks PJ, Beerling DJ. Maximum leaf conductance driven by CO₂ effects on stomatal size and density over geologic time. *Proc Natl Acad Sci USA* 2009, 106:10343–10347.
- Assouline S, Or D. Plant water use efficiency over geological time –evolution of leaf stomata configurations affecting plant gas exchange. *PLoS One* 2013, 8:e67757. doi:10.1371/journal.pone.0067757.
- 76. Cochard H, Nardini A, Coll L. Hydraulic architecture of leaf blades: where is the main resistance? *Plant. Cell Environ* 2004, 27:1257–1267. doi:10.1111/ j.1365-3040.2004.01233.x.
- 77. Sack L, Holbrook NM. Leaf hydraulics. Annu Rev Plant Biol 2006, 57:361–381.
- 78. Brodribb TJ, Field TS, Jordan GJ. Leaf maximum photosynthetic rate and venation are linked by hydraulics. *Plant Physiol* 2007, 144:1890–1898.
- Hetherington AM, Woodward FI. The role of stomata in sensing and driving environmental change. *Nature* 2003, 424:901–908.
- Jarvis PG. The interpretation of the variances in leaf water potential and stomatal conductance found in canopies in the field. *Philos Trans R Soc Lond Ser B* 1976, 273:593-610.
- 81. Monteith JL. A reinterpretation of stomatal responses to humidity. *Plant Cell Environ* 1995, 18:357–364.
- 82. Assmann SM. The cellular basis of guard cell sensing to rising CO₂. *Plant Cell Environ* 1999, 22:629–637.
- Merilo E, Joesaar I, Brosché M, Kollist H. To open or to close: species-specific stomatal responses to simultaneously applied opposing environmental factors. *New Phytol* 2014, 202:499–508. doi:10.1111/ nph.12667.
- Buckley TN, Schymanski SJ. Stomatal optimisation in relation to atmospheric CO₂. *New Phytol* 2014, 201:372–377. doi:10.1111/nph.12552.
- 85. Fischer RA. The relationship of stomatal aperture and guard-cell turgor pressure in *Vicia faba*. J Exp Bot 1973, 24:387–399.
- Buckley TN, Mott KA, Farquhar GD. A hydromechanical and biochemical model of stomatal conductance. *Plant Cell Environ* 2003, 26:1767–1785.
- Buckley TN. The control of stomata by water balance. New Phytol 2005, 169:275–292. doi:10.1111/ j.1469-8137.2005.01543.x.
- 88. Comstock JP. Hydraulic and chemical signalling in the control of stomatal conductance and transpiration. *J Exp Bot* 2002, 53:195–200.
- Brodribb TJ, Holbrook NM. Stomatal closure during leaf dehydration, correlation with other leaf physiological traits. *Plant Physiol* 2003, 132:2166–2173.
- Brodribb TJ, Holbrook NM. Stomatal protection against hydraulic failure: a comparison of coexisting ferns and angiosperms. *New Phytol* 2004, 162:663– 670. doi:10.1111/j.1469-8137.2004.01060.x.

- 91. Guyot G, Scoffoni C, Sack L. Combined impacts of irradiance and dehydration on leaf hydraulic conductance: insights into vulnerability and stomatal control. *Plant Cell Environ* 2012, 35:857–871. doi:10.1111/ j.1365-3040.2011.02458.x.
- 92. Pantin F, Monnet F, Jannaud D, Costa JM, Renaud J, Muller B, Simonneau T, Genty B. The dual effect of abscisic acid on stomata. *New Phytol* 2013, 197:65– 72. doi:10.1111/nph.12013.
- 93. Zeiger E. The biology of stomatal guard cells. Annu Rev Plant Physiol 1983, 34:441-475.
- 94. Talbott LD, Zeiger E. The role of sucrose in guard cell osmoregulation. J Exp Bot 1998, 49:329-337.
- 95. Roelfsema MRG, Hedrich R. In the light of stomatal opening: new insights into the watergate. *New Phytol* 2005, 167:665–691. doi:10.1111/ j.1469-8137.2005.01460.x.
- Wilkinson S, Davies WJ. ABA-based chemical signalling: the co-ordination of responses to stress in plants. *Plant Cell Environ* 2002, 25:195–210.
- 97. Brodribb TJ, McAdam SAM. Abscisic acid mediates a divergence in the drought response of two conifers. *Plant Physiol* 2013, 162:1370–1377.
- Dodd IC. Abscisic acid and stomatal closure: a hydraulic conductance conundrum? *New Phytol* 2013, 197:6–8. doi:10.1111/nph.12052.
- 99. Franks PJ, Farquhar GD. The effect of exogenous abscisic acid on stomatal development, stomatal mechanics, and leaf gas exchange in *Tradescantia Virginiana*. *Plant Physiol* 2001, 125:935–942.
- 100. Brodribb TJ, McAdam SAM. Passive origins of stomatal control in vascular plants. *Science* 2011, 331:582–585. doi:10.1126/science.1197985.
- 101. Dewar RC. Interpretation of an empirical model for stomatal conductance in terms of guard cell function. *Plant Cell Environ* 1995, 18:365–372.
- 102. Dewar RC. The Ball-Berry-Leuning and Tardieu-Davies stomatal models: synthesis and extension within a spatially aggregated picture of guard cell function. *Plant Cell Environ* 2002, 25:1383–1398.
- 103. Gao Q, Xhao P, Zeng X, Cai X, Shen W. A model of stomatal conductance to quantify the relationship between leaf transpiration, microclimate, and soil water stress. *Plant Cell Environ* 2002, 25:1373–1381.
- 104. Franks PJ. Stomatal control and hydraulic conductance, with special reference to tall trees. *Tree Physiol* 2004, 24:865–878.
- 105. Franks PJ, Drake PL, Froend RH. Anisohydric but isohydrodynamic: seasonally constant plant water potential gradient explained by a stomatal control mechanism incorporating variable plant hydraulic conductance. *Plant Cell Environ* 2007, 30:19–30. doi:10.1111/j.1365-3040.2006.01600.x.

- 106. Peak D, Mott KA. A new, vapour-phase mechanism for stomatal responses to humidity and temperature. *Plant Cell Environ* 2011, 34:162–178. doi:10.1111/ j.1365-3040.2010.02234.x.
- 107. de Boer HJ, Eppinga MB, Wassen MJ, Dekker SC. A critical transition in leaf evolution facilitated the Cretaceous angiosperm revolution. *Nat Commun* 2012, 3. doi:10.1038/ncomms2217.
- 108. Mott KA, Peak D. Testing a vapour-phase model of stomatal responses to humidity. *Plant Cell Environ* 2013, 36:936–944.
- 109. Tuzet A, Perrier A, Leuning R. A coupled model of stomatal conductance, photosynthesis and transpiration. *Plant Cell Environ* 2003, 26:1097–1116.
- 110. Damour G, Simonneau T, Cochard H, Urban L. An overview of models of stomatal conductance at the leaf level. *Plant Cell Environ* 2010, 33:1419–1438. doi:10.1111/j.1365-3040.2010.02181.x.
- 111. Tardieu F, Davies WJ. Integration of hydraulic and chemical signalling in the control of stomatal conductance and water status of droughted plants. *Plant Cell Environ* 1993, 16:341–349.
- 112. Tardieu F, Simonneau T. Variability among species of stomatal control under fluctuating soil water status and evaporative demand: modelling isohydric and anisohydric behaviours. *J Exp Bot* 1998, 49:419–432.
- 113. Buckley TN, Mott KA. Modelling stomatal conductance in response to environmental factors. *Plant Cell Environ* 2013, 36:1691–1699. doi:10.1111/ pce.12140.
- 114. Ball JT, Woodrow IE, Berry JA. A model predicting stomatal conductance and its contribution to the control of photosynthesis under different environmental conditions. In: Biggins J, ed. *Progress in Photosynthesis Research*. Netherlands: Martinus Nijho; 1987, 221–224.
- 115. Leuning R. Modelling stomatal behaviour and photosynthesis of *Eucalyptus Grandis*. *Aust J Plant Physiol* 1990, 17:159–175.
- 116. Leuning R. A critical appraisal of a combined stomatal- photosynthesis model for C3 plants. *Plant Cell Environ* 1995, 18:357–364.
- 117. Farquhar GD, von Caemmerer S, Berry JA. A biochemical model of photosynthetic CO₂ assimilation in leaves of C3 species. *Planta* 1980, 149:78–90.
- 118. Collatz GJ, Ball JT, Grivet C, Berry JA. Physiological and environmental regulation of stomatal conductance, photosynthesis and transpiration: a model that includes a laminar boundary-layer. *Agr Forest Meteorol* 1991, 54:107–136.
- 119. Collatz GJ, Ribas-Carbo M, Berry JA. Coupled photosynthesis-stomatal conductance model for leaves of C4 plants. *Aust J Plant Physiol* 1992, 19:519–538.

- 120. Wang YP, Leuning R. A two-leaf model for canopy conductance, photosynthesis and partitioning of available energy I: model description and comparison with a multi-layered model. *Agr Forest Meteorol* 1998, 91:89–111.
- 121. Bonan GB, Lawrence PJ, Oleson KW, Levis S, Jung M, Reichstein M, Lawrence DM, Swenson SC. Improving canopy processes in the Community Land Model version 4 (CLM4) using global flux fields empirically inferred from FLUXNET data. *Journal of Geophysical Research* 2011, 116:G02014. doi:10.1029/2010JG001593.
- 122. Medlyn BE, Duursma RA, De Kauwe MG, Prentice IC. The optimal stomatal response to atmospheric CO_2 concentration: alternative solutions, alternative interpretations. *Agr Forest Meteorol* 2013, 182-183:200-203.
- 123. Lin ZS, Medlyn BE, Duursma RA, Prentice IC, Wang H, Baig S, Eamus D, de Dios VR, Mitchell P, Ellsworth DS, et al. Optimal stomatal behaviour around the world. *Nat Clim Change* 2015, 5:459–464.
- 124. Katul G, Manzoni S, Palmroth S, Oren R. A stomatal optimization theory to describe the effects of atmospheric CO_2 on leaf photosynthesis and transpiration. *Ann Bot* 2010, 105:431–442.
- 125. Medlyn BE, Duursma RA, Eamus D, Ellsworth DS, Prentice IC, Barto CVM, Crous KY, De Angelis P, Freeman MC, Wingate L. Reconciling the optimal and empirical approaches to modelling stomatal conductance. *Glob Change Biol* 2011, 17:2134–2144.
- 126. Manzoni S, Vico G, Palmroth S, Porporato A, Katul G. Optimization of stomatal conductance for maximum carbon gain under dynamic soil moisture. Adv Water Resour 2013, 62:90–105.
- 127. Bonan GB, Williams M, Fisher RA, Oleson KW. Modeling stomatal conductance in the earth system: linking leaf water-use efficiency and water transport along the soil-plant–atmosphere continuum. *Geosci Model Dev* 2014, 7:2193–2222. doi:10.5194/gmd-7-2193-2014.
- 128. Egea G, Verhoef A, Vidale PL. Towards an improved and more flexible representation of water stress in coupled photosynthesis-stomatal conductance models. *Agr Forest Meteorol* 2011, 151:1370-1384.
- 129. Zhou S, Duursma RA, Medlyn BE, Kelly JWG, Prentice IC. How should we model plant responses to drought? An analysis of stomatal and non-stomatal responses to water stress. *Agr Forest Meteorol* 2013, 182-183:204–214.
- 130. Manzoni S, Vico G, Katul G, Fay PA, Polley W, Palmroth S, Porporato A. Optimizing stomatal conductance for maximum carbon gain under water stress: a meta-analysis across plant functional types and climates. *Funct Ecol* 2011, 25:456–467. doi:10.1111/j.1365-2435.2010.01822.x.

- 131. Clode PL, Kilburn MR, Jones DL, Stockdale EA, Cliff JB III, Herrmann AM, Murphy DV. In situ mapping of nutrient uptake in the rhizosphere using nanoscale secondary ion mass spectrometry. *Plant Physiol* 2009, 151:1751–1757.
- 132. Jones DL, Nguyen C, Finlay RD. Carbon flow in the rhizosphere: carbon trading at the soil-root interface. *Plant and Soil* 2009, 321:5–33. doi:10.1007/s11104-009-9925-0.
- 133. Hinsinger P, Bengough AG, Vetterlein D, Young IM. Rhizosphere: biophysics, bio-geochemistry and ecological relevance. *Plant and Soil* 2009, 321:117–152. doi:10.1007/s11104-008-9885-9.
- 134. Carminati A, Moradi AB, Vetterlein D, Vontobel P, Lehmann E, Weller U, Vogel HJ, Oswald SE. Dynamics of soil water content in the rhizosphere. *Plant and Soil* 2010, 332:163–176. doi:10.1007/s11104-010-0283-8.
- 135. Jackson RB, Mooney HA, Schulze ED. A global budget for net root biomass, surface area, and nutrient contents. *Proc Natl Acad Sci USA* 1997, 94:7362–7366.
- 136. McCormack ML, Dickie IA, Eissenstat DM, Fahey TJ, Fernandez CW, Guo D, Helmisaari HS, Hobbie EA, Iversen CM, Jackson RB, et al. Redefining fine roots improves understanding of below-ground contributions to terrestrial biosphere processes. *New Phytol* 2015, 207:505–518. doi:10.1111/nph.13363.
- 137. Boyer JS. Water transport. Annu Rev Plant Physiol 1985, 36:473-516.
- 138. Nadezhdina N, David TS, David JS, Ferreira MI, Dohnal M, Tesar M, Gartner K, Leitgeb E, Nadezhdin V, Cermak J, et al. Trees never rest: the multiple facets of hydraulic redistribution. *Ecohydrology* 2010, 3:431–444. doi:10.1002/eco.148.
- 139. Goldsmith GR. Changing directions: the atmosphereplant-soil continuum. *New Phytol* 2013, 2013:4–6. doi:10.1111/nph.12332.
- 140. Neumann RB, Cardon ZG. The magnitude of hydraulic redistribution by plant roots: a review and synthesis of empirical and modeling studies. *New Phytol* 2012, 194:337–352. doi:10.1111/j.1469-8137.2012.04088.x.
- 141. Gardner WR. Relation of root distribution to water uptake and availability. *Agron J* 1964, 56:41–45.
- 142. Sperry JS, Adler FR, Campbell GS, Comstock JB. Limitation of plant water use by rhizosphere and xylem conductance: results from a model. *Plant Cell Environ* 1998, 21:347–359.
- 143. Steudle E. Water uptake by plant roots: an integration of views. *Plant and Soil* 2000, 226:45–56.
- 144. Doussan C, Vercambre G, Pages L. Modelling of the hydraulic architecture of root systems: an integrated approach to water absorption - distribution of axial

and radial conductances in maize. Ann Bot 1998, 81:225-232.

- 145. Couvreur V, Vanderborght J, Javaux M. A simple three-dimensional macroscopic root water uptake model based on the hydraulic architecture approach. *Hydrol Earth Syst Sci* 2012, 16:2957–2971. doi:10.5194/hess-16-2957-2012.
- 146. Javaux M, Vanderborght J, Couvreur V, Vereecken H. Root water uptake: from 3D biophysical processes to macroscopic modeling approaches. *Vadose Zone J* 2013, 12:1–16. doi:10.2136/vzj2013.02.0042.
- 147. Johnson DM, Sherrard ME, Domec JC, Jackson RB. Role of aquaporin activity in regulating deep and shallow root hydraulic conductance during extreme drought. *Trees* 2014, 28:1323–1331. doi:10.1007/ s00468-014-1036-8.
- 148. Moshelion M, Halperin O, Wallach R, Oren R, Way DA. Role of aquaporins in determining transpiration and photosynthesis in water-stressed plants: crop water-use efficiency, growth and yield. *Plant Cell Environ* 2014, 38:1785–1793. doi:10.1111/ pce.12410.
- 149. Jungk AO. Dynamics of nutrient movement at the soil-root interface. In: *Plant Roots: The Hidden Half.* New York: Marcel Dekker Inc.; 2002, 587–616.
- 150. Chapman N, Miller AJ, Lindsey K, Richard Whalley W. Roots, water, and nutrient acquisition: let's get physical. *Trends Plant Sci* 2012, 17:701–710.
- 151. Zelazny E, Vert G. Plant nutrition: root transporters on the move. *Plant Physiol* 2014, 166:500–508. doi:10.1104/pp.114.244475.
- 152. He L, Ivanov VY, Bohrer G, Thomsen JE, Vogel CS, Moghaddam M. Temporal dynamics of soil moisture in a northern temperate mixed successional forest after a prescribed intermediate disturbance. *Agr Forest Meteorol* 2013, 180:22–33.
- 153. Leitner D, Klepsch S, Bodner G, Schnepf A. A dynamic root system growth model based on L-systems. *Plant and Soil* 2010, 332:177–192.
- 154. Vrugt JA, vanWijk MT, Hopmans JW, Simunek J. One-, two-, and three-dimensional root water uptake functions for transient modeling. *Water Resour Res* 2001, 37:2457–2470. doi:10.1029/2000WR000027.
- 155. Doussan C, Pierret A, Garrigues E, Pages L. Water uptake by plant roots: II—modelling of water transfer in the soil root system with explicit account of flow within the root system—comparison with experiments. *Plant and Soil* 2006, 283:99–117.
- 156. Javaux M, Schroder T, Vanderborght J, Vereecken H. Use of a three-dimensional detailed modeling approach for predicting root water uptake. *Vadose Zone J* 2008, 7:1079–1088.
- 157. Schneider CL, Attinger S, Delfs JO, Hildebrandt A. Implementing small scale processes at the soil-plant interface - the role of root architectures for

calculating root water uptake profiles. *Hydrol Earth Syst Sci* 2010, 14:279–289. doi:10.5194/hess-14-279-2010.

- 158. Dunbabin VM, Postma JA, Schnepf A, Pagès L, Javaux M, Wu L, Leitner D, Chen YL, Rengel Z, Diggle AJ. Modelling root-soil interactions using three-dimensional models of root growth, architecture and function. *Plant and Soil* 2013, 372:93–124. doi:10.1007/s11104-013-1769-y.
- 159. Manoli G, Bonetti S, Domec JC, Putti M, Katul G, Marani M. Tree root systems competing for soil moisture in a 3D soil-plant model. *Adv Water Resour* 2014, 66:32-42.
- 160. Postma JA, Schurr U, Fiorani F. Dynamic root growth and architecture responses to limiting nutrient availability: linking physiological models and experimentation. *Biotechnol Adv* 2014, 32:53–65.
- 161. Warren JM, Hanson PJ, Iversen CM, Kumar J, Walker AP, Wullschleger SD. Root structural and functional dynamics in terrestrial biosphere models – evaluation and recommendations. *New Phytol* 2015, 205:59–78. doi:10.1111/nph.1303.
- 162. Feddes RA, Kowalik P, Kolinska-Malinka K, Zaradny H. Simulation of field water uptake by plants using a soil water dependent root extraction function. *J Hydrol* 1976, 31:13–26.
- 163. Feddes RA, Ho H, Bruen M, Dawson T, de Rosnay P, Dirmeyer P, Jackson RB, Kabat P, Kleidon A, Lilly A, et al. Modeling root water uptake in hydrological and climate models. *Bull Am Meteorol Soc* 2001, 82:2797–2809.
- 164. Ivanov VY, Bras RL, Vivoni ER. Vegetationhydrology dynamics in complex terrain of semiarid areas: 1. A mechanistic approach to modeling dynamic feedbacks. *Water Resour Res* 2008, 44: W03429. doi:10.1029/2006WR005588.
- 165. Fatichi S, Ivanov VY, Caporali E. A mechanistic ecohydrological model to investigate complex interactions in cold and warm water-controlled environments. 1. Theoretical framework and plotscale analysis. *J Adv Model Earth Syst* 2012, 4: M05002.
- 166. Daly E, Porporato A, Rodriguez-Iturbe I. Coupled dynamics of photosynthesis, transpiration, and soil water balance. Part I: upscaling from hourly to daily level. *J Hydrometeorol* 2004, *5*:546–558.
- 167. Deckmyn G, Verbeeck H, de Beeck MO, Vansteenkiste D, Steppe K, Ceulemans R. ANAFORE: a stand-scale process-based forest model that includes wood tissue development and labile carbon storage in trees. *Ecol Model* 2008, 215:345–368.
- 168. Newman EI. Resistance to water flow in soil and plant. I. Soil resistance in relation to amounts of root: theoretical estimate. *J Appl Ecol* 1969, 6:1–12.
- 169. Dixon HH, Joly J. On the ascent of sap. *Philos Trans R Soc Lond Ser B* 1894, 186:563–576.

- 170. Tyree MT. The cohesion-tension theory of sap ascent: current controversies. J Exp Bot 1997, 48:1753-1765.
- 171. Münch E. Die Stoffbewegungen in der Pflanze. Jena, Germany: Gustav Fischer; 1930.
- 172. De Schepper V, De Swaef T, Bauweraerts I, Steppe K. Phloem transport: a review of mechanisms and controls. *J Exp Bot* 2013, 64:4839–4850. doi:10.1093/ jxb/ert302.
- 173. Ryan MG, Asao S. Phloem transport in trees. *Tree Physiol* 2014, 34:1–4. doi:10.1093/treephys/tpt123.
- 174. Pockman WT, Sperry JS. Vulnerability to xylem cavitation and the distribution of Sonoran vegetation. *Am J Bot* 2000, 87:1287–1299.
- 175. Martínez-Vilalta J, Prat E, Oliveras I, Piñol J. Xylem hydraulic properties of roots and stems of nine mediterranean woody species. *Oecologia* 2002, 133:19–29.
- 176. Lopez OR, Kursar TA, Cochard H, Tyree MT. Interspecific variation in xylem vulnerability to cavitation among tropical tree and shrub species. *Tree Physiol* 2005, 25:1553–1562.
- 177. Maherali H, Pockman WT, Jackson RB. Adaptive variation in the vulnerability of woody plants to xylem cavitation. *Ecology* 2004, 85:2184–2199.
- 178. Meinzer FC, Johnson DM, Lachenbruch B, McCulloh KM, Woodruff DR. Xylem hydraulic safety margins in woody plants: coordination of stomatal control of xylem tension with hydraulic capacitance. *Funct Ecol* 2009, 23:922–930. doi:10.1111/j.1365-2435.2009.01577.x.
- 179. Domec JC, Gartner BL. Cavitation and water storage capacity in bole xylem segments of mature and young Douglas-fir trees. *Trees* 2001, 15:204–214.
- 180. Steppe K, De Pauw DJW, Doody TM, Teskey RO. Comparison of sap flux density using thermal dissipation, heat pulse velocity and heat field deformation methods. *Agr Forest Meteorol* 2010, 150:1046-1056.
- 181. Hölttä T, Vesala T, Sevanto S, Perämäki M, Nikinmaa E. Modeling xylem and phloem water flows in trees according to cohesion theory and Münch hypothesis. *Trees Struct Funct* 2006, 20:67–78.
- 182. Thompson MV, Holbrook NM. Application of a single-solute non-steady-state phloem model to the study of long-distance assimilate transport. *J Theor Biol* 2003, 220:419–455.
- Cochrane TT, Cochrane TA. Differences in the way potassium chloride and sucrose solutions effect osmotic potential of significance to stomata aperture modulation. *Plant Physiol Biochem* 2009, 47:205–209.
- 184. Woodruff DR, Bond BJ, Meinzer FC. Does turgor limit growth in tall trees? *Plant Cell Environ* 2004, 27:229–236.

- 185. Woodruff DR, Meinzer FC. Size-dependent changes in biophysical control of tree growth: the role of turgor. In: *Size- and Age-Related Changes in Tree Structure and Function*. Dordrecht, Netherlands: Springer; 2011, 363–384.
- 186. Turgeon R. The puzzle of phloem pressure. *Plant Physiol* 2010, 154:578–581.
- 187. Mencuccini M, Hölttä T, Sevanto S, Nikinmaa E. Concurrent measurements of change in the bark and xylem diameters of trees reveal a phloem-generated turgor signal. *New Phytol* 2013, 198:1143–1154.
- 188. Hölttä T, Mencuccini M, Nikinmaa E. Linking phloem function to structure: analysis with a coupled xylem-phloem transport model. *J Theor Biol* 2009, 259:325–337.
- 189. Sevanto S, Hölttä T, Holbrook NM. Effects of the hydraulic coupling between xylem and phloem on diurnal phloem diameter variation. *Plant Cell Envi*ron 2011, 34:690–703. doi:10.1111/j.1365-3040.2011.02275.x.
- 190. Meinzer FC, James SA, Goldstein G, Woodruff D. Whole-tree water transport scales with sapwood capacitance in tropical forest canopy trees. *Plant Cell Environ* 2003, 2003:1147–1155.
- 191. Scholz FG, Bucci SJ, Goldstein G, Meinzer FC, Franco AC, Miralles-Wilhelm F. Biophysical properties and functional significance of stem water storage tissues in neotropical savanna trees. *Plant Cell Environ* 2007, 30:236–248.
- 192. Barnard DM, Meinzer FC, Lachenbruch B, McCulloh KA, Johnson DM, Woodruff DR. Climate-related trends in sapwood biophysical properties in two conifers: avoidance of hydraulic dysfunction through coordinated adjustments in xylem efficiency, safety and capacitance. *Plant Cell Environ* 2011, 34:643–654.
- 193. Verbeeck H, Steppe K, Nadezhdina N, De Beeck MO, Deckmyn GO, Meiresonne L, Lemeur R, Cermák J, Ceulemans R, Janssens IA. Stored water use and transpiration in Scots pine: a modeling analysis with ANAFORE. *Tree Physiol* 2007, 27:1671–1685.
- 194. Cermák J, Kucera J, Bauerle WL, Phillips N, Hinckely TM. Tree water storage and its diurnal dynamics related to sap flow and changes in stem volume in old-growth douglas-fir trees. *Tree Physiol* 2007, 27:181–198.
- 195. Scholz FG, Phillips NG, Bucci SJ, Meinzer FC, Goldstein G. Hydraulic capacitance: biophysics and functional significance of internal water sources in relation to tree size. In: *Size- and Age-Related Changes in Tree Structure and Function*. Dordrecht, Netherlands: Springer; 2011, 341–361.
- 196. Domec JC, Gartner BL. How do water transport and storage differ in coniferous earlywood and latewood? *J Exp Bot* 2002, 53:2369–2379.

- 197. Domec JC, Gartner BL. Relationship between growth rates and xylem hydraulic characteristics in young, mature and old-growth ponderosa pine trees. *Plant Cell Environ* 2003, 26:471–483.
- 198. Génard M, Fishman S, Vercambre G, Huguet JG, Bussi C, Besset J, Habib R. A biophysical analysis of stem and root diameter variations in woody plants. *Plant Physiol* 2001, 126:188–202.
- 199. Zweifel R, Item H, Häsler R. Link between diurnal stem radius changes and tree water relations. *Tree Physiol* 2001, 21:869–887.
- 200. Zweifel R, Zimmermann L, Zeugin F, Newbery DM. Intra-annual radial growth and water relations of trees: implications towards a growth mechanism. *J Exp Bot* 2006, 57:1445–1459. doi:10.1093/jxb/ erj125.
- 201. Fernández JE, Cuevas MV. Irrigation scheduling from stem diameter variations: a review. *Agr Forest Meteorol* 2010, 150:135–151.
- 202. King G, Fonti P, Nievergelt D, Buntgen U, Frank D. Climatic drivers of hourly to yearly tree radius variations along a 6 °C natural warming gradient. Agr Forest Meteorol 2013, 2013:36–46.
- 203. Köcher P. V. Horna, and C. Leuschner. Stem water storage in five coexisting temperate broad-leaved tree species: significance, temporal dynamics and dependence on tree functional traits. *Tree Physiol* 2013, 33:817–832. doi:10.1093/treephys/tpt055.
- 204. Katul GG, Leuning R, Oren R. Relationship between plant hydraulic and biochemical properties derived from a steady-state coupled water and carbon transport model. *Plant Cell Environ* 2003, 26:339–350.
- 205. Loranty MM, Mackay DS, Ewers BE, Traver E, Kruger EL. Competition for light between individual trees lowers reference canopy stomatal conductance: results from a model. *J Geophys Res Biogeosci* 2010, 115:G04019. doi:10.1029/2010JG001377.
- 206. Mackay DS, Roberts DE, Ewers BE, Sperry JS, McDowell NG, Pockman WT. Interdependence of chronic hydraulic dysfunction and canopy processes can improve integrated models of tree response to drought. *Water Resour Res* 2015, 51:6156–6176. doi:10.1002/2015WR017244.
- 207. Bohrer G, Mourad H, Laursen TA, Drewry D, Avissar R, Poggi D, Oren R, Katul GG. Finite element tree crown hydrodynamics model (FETCH) using porous media flow within branching elements: a new representation of tree hydrodynamics. *Water Resour Res* 2005, 41:W11404. doi:10.1029/2005WR004181.
- 208. Janott M, Gayler S, Gessler A, Javaux M, Klier C, Priesack E. A one-dimensional model of water flow in soil-plant systems based on plant architecture. *Plant* and Soil 2011, 341:233–256. doi:10.1007/s11104-010-0639-0.
- 209. Bittner S, Legner N, Beese F, Priesack E. Individual tree branch-level simulation of light attenuation and

water flow of three F. sylvatica L. trees. J Geophys Res 2012, 117:G01037. doi:10.1029/2011JG001780.

- 210. Hentschel R, Bittner S, Janott M, Biernath C, Holst J, Ferrio JP, Gessler A, Priesack E. Simulation of stand transpiration based on a xylem water flow model for individual trees. *Agricultural and Forest Meteorology* 2013, 182–183:31–42.
- 211. Chuang YL, Oren R, Bertozzi AL, Phillips N, Katul GG. The porous media model for the hydraulic system of a conifer tree: linking sap flux data to transpiration rate. *Ecol Model* 2006, 191:447–468.
- 212. Hölttä T, Cochard H, Nikinmaa E, Mencuccini M. Capacitive effect of cavitation in xylem conduits: results from a dynamic model. *Plant Cell Environ* 2009, 32:10–21.
- 213. Nikinmaa E, Sievänen R, Hölttä T. Dynamics of leaf gas exchange, xylem and phloem transport, water potential and carbohydrate concentration in a realistic 3-D model tree crown. *Ann Bot* 2014, 114:653–666.
- 214. Schiestl-Aalto P, Kulmala L, Mäkinen H, Nikinmaa E, Mäkelä A. CASSIA a dynamic model for predicting intra-annual sink demand and interannual growth variation in scots pine. *New Phytol* 2015, 206:647–659. doi:10.1111/nph.13275.
- 215. Steppe K, De Pauw DJW, Lemeur R, Vanrolleghem PA. A mathematical model linking tree sap flow dynamics to daily stem diameter fluctuations and radial stem growth. *Tree Physiol* 2005, 26:257–273.
- 216. De Pauw DJW, Steppe K, De Baets B. Identifiability analysis and improvement of a tree water flow and storage model. *Math Biosci* 2008, 211:314–332.
- 217. De Schepper V, Steppe K. Development and verification of a water and sugar transport model using measured stem diameter variations. *J Exp Bot* 2010, 61:2083–2099. doi:10.1093/jxb/erq018.
- 218. De Schepper V, Steppe K. Tree girdling responses simulated by a water and carbon transport model. *Ann Bot* 2011, 108:1147–1154. doi:10.1093/aob/ mcr068.
- 219. Daudet FA, Lacointe A, Gaudillere JP, Cruiziat P. Generalized Münch coupling between sugar and water fluxes for modelling carbon allocation as affected by water status. *J Theor Biol* 2002, 214:481–498. doi:10.1006/jtbi.2001.2473.
- 220. Hsiao TC, Acevedo E, Fereres E, Henderson DW. Stress metabolism: water stress, growth and osmotic adjustment. *Philos Trans R Soc Lond B Biol Sci* 1976, 273:479–500.
- 221. Wang Z, Quebedeaux B, Stutte GW. Osmotic adjustment: effect of water stress on carbohydrates in leaves, stems and roots of apple. *Aust J Plant Physiol* 1995, 22:747–754.
- 222. Bartlett MK, Scoffoni C, Sack L. The determinants of leaf turgor loss point and prediction of drought

tolerance of species and biomes: a global meta-analysis. *Ecol Lett* 2012, 15:393–405.

- 223. Cosgrove DJ. Growth of the plant cell wall. Nat Rev Mol Cell Biol 2005, 6:850-861. doi:10.1038/ nrm1746.
- 224. Hölttä T, Mäkinen H, Nöjd P, Mäkelä A, Nikinmaa E. A physiological model of softwood cambial growth. *Tree Physiol* 2010, 30:1235–1252. doi:10.1093/treephys/tpq068.
- 225. Lockhart JA. An analysis of irreversible plant cell elongation. J Theor Biol 1965, 8:264–275.
- 226. Cosgrove D. Biophysical control of plant cell growth. Annu Rev Plant Physiol 1986, 37:377-405.
- 227. Boyer JS, Silk WK. Hydraulics of plant growth. *Funct Plant Biol* 2004, 2004:761–773.
- 228. Hsiao TC. Plant responses to water stress. Annu Rev Plant Physiol 1973, 24:519–570.
- 229. Muller B, Pantin F, Génard M, Turc O, Freixes S, Piques M, Gibon Y. Water deficits uncouple growth from photosynthesis, increase C content, and modify the relationships between C and growth in sink organs. J Exp Bot 2011, 62:1715–1729. doi:10.1093/jxb/erq438.
- 230. Tardieu F, Granier C, Muller B. Water deficit and growth. Co-ordinating processes without an orchestrator? *Curr Opin Plant Biol* 2011, 14:283–289.
- 231. Cannell MGR, Thornley JHM. Modelling plant respiration: some guiding principles. *Ann Bot* 2000, 85:45–54.
- 232. McDowell NG. Mechanisms linking drought, hydraulics, carbon metabolism, and mortality. *Plant Physiol* 2011, 155:1051–1059.
- 233. Brando PM, Nepstad DC, Davidson EA, Trumbore SE, Ray D, Camargo P. Drought effects on litterfall, wood production and belowground carbon cycling in an Amazon forest: results of a throughfall reduction experiment. *Philos Trans R Soc B* 2008, 363:1839–1848. doi:10.1098/rstb.2007.0031.
- 234. Brzostek ER, Dragoni D, Schmid HP, Rahman AF, Sims D, Wayson CA, Johnson DJ, Phillips RP. Chronic water stress reduces tree growth and the carbon sink of deciduous hardwood forests. *Glob Chang Biol* 2014, 20:2531–2539. doi:10.1111/gcb.12528.
- 235. Fischer EM, Seneviratne SI, Vidale PL, Lüthi D, Schär C. Soil moisture–atmosphere interactions during the 2003 European summer heat wave. *J Climate* 2007, 20:5081–5099.
- 236. Lawlor DW, Tezara W. Causes of decreased photosynthetic rate and metabolic capacity in waterdeficient leaf cells: a critical evaluation of mechanisms and integration of processes. *Ann Bot* 2009, 103:561–579. doi:10.1093/aob/mcn244.
- 237. Bartlett MK, Zhang Y, Kreidler N, Sun S, Ardy R, Cao K, Sack L. Global analysis of plasticity in turgor

loss point, a key drought tolerance trait. *Ecol Lett* 2014, 17:1580–1590. doi:10.1111/ele.12374.

- 238. Hartmann H, Ziegler W, Kolle O, Trumbore S. Thirst beats hunger - declining hydration during drought prevents carbon starvation in Norway spruce saplings. *New Phytol* 2013, 200:340–349. doi:10.1111/nph.12331.
- 239. Gaylord ML, Kolb TE, Pockman WT, Plaut JA, Yepez EA, Macalady AK, Pangle RE, McDowell NG. Drought predisposes pinon-juniper woodlands to insect attacks and mortality. *New Phytol* 2013, 198:567–578.
- 240. Poyatos R, Aguadé D, Galiano L, Mencuccini M, Martínez-Vilalta J. Drought-induced defoliation and long periods of near-zero gas exchange play a key role in accentuating metabolic decline of Scots pine. *New Phytol* 2013, 220:388–401. doi:10.1111/ nph.12278.
- 241. Penuelas J, Filella I, Zhang XY, Llorens L, Ogaya R, Lloret L, Comas P, Estiarte M, Terradas J. Complex spatiotemporal phenological shifts as a response to rainfall changes. *New Phytol* 2004, 161:837–846. doi:10.1111/j.1469-8137.2004.01003.x.
- 242. Vico G, Thompson SE, Manzoni S, Molini A, Albertson JD, Almeida-Cortez JS, Fay PA, Feng X, Guswa AJ, Liu H, et al. Climatic ecophysiological and phenological controls on plant ecohydrological strategies in seasonally dry ecosystems. *Ecohydrology* 2015:660–681. doi:10.1002/eco.1533.
- 243. Manzoni S, Vico G, Thompson S, Beyer F, Weih M. Contrasting leaf phonological strategies optimize carbon gain under droughts of different duration. *Adv Water Resour* 2015, 84:37–51. doi:10.1016/j. advwatres.2015.08.001.
- 244. McDowell N, Pockman W, Allen C, Breshears DD, Cobb N, Kolb T, Sperry J, West A, Williams D, Yepez E. Mechanisms of plant survival and mortality during drought: why do some plants survive while others succumb to drought? *New Phytol* 2008, 178:719–739. doi:10.D1111/j.1469-8137D2008D02436.x.
- 245. McDowell NG, Beerling DJ, Breshears DD, Fisher RA, Raffa KF, Stitt M. The interdependent mechanisms underlying climate-driven vegetation mortality. *Trends Ecol Evol* 2011, 26:523–532.
- 246. Sevanto S, McDowell NG, Dickman LT, Pangle R, Pockman WT. How do trees die? A test of the hydraulic failure and carbon starvation hypotheses. *Plant Cell Environ* 2014, 37:153–161. doi:10.1111/ pce.1214.
- 247. Allen CD, Macalady AK, Chenchouni H, Bachelet D, McDowell NG, Vennetier M, Kitzberger T, Rigling A, Breshears DD, Hogg EH(T), et al. A global overview of drought and heat-induced tree mortality reveals emerging climate change risks for forests. *For Ecol Manage* 2010, 259:660–684.

- 248. Phillips OL, van der Heijden G, Lewis SL, López-González G, Aragao LEOC, Lloyd J, Malhi Y, Monteagudo A, Almeida S, et al. Drought-mortality relationships for tropical forests. *New Phytol* 2010, 187:631–646.
- 249. Choat B, Jansen S, Brodribb TJ, Cochard H, Delzon S, Bhaskar R, Bucci SJ, Feild TS, Gleason SM, Hacke UG, et al. Global convergence in the vulnerability of forests to drought. *Nature* 2012, 491:752–755.
- 250. Adams HD, Guardiola-Claramonte M, Barron-Gafford GA, Breshears DD, Villegas JC, Zou CB, Troch PA, Huxman TE. Temperature sensitivity of drought-induced tree mortality portends increased regional die-off under global change-type drought. *Proc Natl Acad Sci USA* 2009, 106:7063–7066.
- 251. Breshears DD, Myers OB, Meyer CW, Barnes FJ, Zou CB, Allen CD, McDowell NG, Pockman WT. Tree die-off in response to global change-type drought: mortality insights from a decade of plant water potential measurements. *Front Ecol Environ* 2009, 7:185–189.
- 252. Breshears DD, Adams HD, Eamus D, McDowell NG, Law DJ, Will RE, Williams AP, Zou CB. The critical amplifying role of increasing atmospheric moisture demand on tree mortality and associated regional dieoff. *Front Plant Sci* 2013, 4:266. doi:10.3389/ fpls.2013.00266.
- 253. McDowell NG, Fisher RA, Xu C, Domec JC, Hölttä T, Mackay DS, Sperry JS, Boutz A, Dickman L, Gehres N, et al. Evaluating theories of droughtinduced vegetation mortality using a multimodelexperiment framework. *New Phytol* 2013, 200:304–321.
- 254. Meir P, Mencuccini M, Dewar RC. Drought-related tree mortality: addressing the gaps in understanding and prediction. *New Phytol* 2015, 207:28–33.
- 255. Reed DE, Ewers BE, Pendall E. Impact of mountain pine beetle induced mortality on forest carbon and water fluxes. *Environ Res Lett* 2014, 9:105004. doi:10.1088/1748-9326/9/10/105004.
- 256. Monteith JL. Evaporation and environment. In: Fogg GE, ed. Symposium Society Experimental Biology, The State and Movement of Water in Living Organisms, vol. 19. London: Cambridge University Press; 1965, 205–224.
- 257. Bonan GB. Forests and climate change: forcings, feedbacks, and the climate benefits of forests. *Science* 2008, 320:1444–1449.
- 258. Allen RG, Pereira LS, Raes D, and Smith M. Crop evapotranspiration-Guidelines for computing crop water requirements, volume 300 of FAO Irrigation and drainage paper. FAO – Food and Agriculture Organization of the United Nations, 1998.
- 259. Drewry DT, Kumar P, Long S, Bernacchi C, Liang XZ, Sivapalan M. Ecohydrological responses of dense canopies to environmental variability: 1. Interplay

between vertical structure and photosynthetic pathway. *J Geophys Res* 2010, 115(G04022):2010. doi:10.1029/2010JG001340.

- 260. Tague CL, Band LE. RHESSys: Regional Hydro-Ecologic Simulation System-an object-oriented approach to spatially distributed modeling of carbon, water, and nutrient cycling. *Earth Interact* 2004, 8:1–42.
- 261. Tague CL, McDowell NG, Allen CD. An integrated model of environmental effects on growth, carbohydrate balance, and mortality of *Pinus ponderosa* forests in the southern rocky mountains. *PLoS One* 2013, 8:e80286.
- 262. Niu GY, Paniconi C, Troch PA, Scott RL, Durcik M, Zeng X, Huxman T, Goodrich DC. An integrated modelling framework of catchment-scale ecohydrological processes: 1. Model description and tests over an energy-limited watershed. *Ecohydrology* 2014, 7:427–439. doi:10.1002/eco.1362.
- 263. Shen C, Niu J, Phanikumar MS. Evaluating controls on coupled hydrologic and vegetation dynamics in a humid continental climate watershed using a subsurface-land surface processes model. *Water Resour Res* 2013, 49:2552–2572. doi:10.1002/ wrcr.20189.
- 264. Della Chiesa S, Bertoldi G, Niedrist G, Obojes N, Endrizzi S, Albertson JD, Wohlfahrt G, Hörtnagl L, Tappeiner U. Modelling changes in grassland hydrological cycling along an elevational gradient in the Alps. *Ecohydrology* 2014, 7:1453–1473. doi:10.1002/eco.1471.
- 265. Zhou X, Istanbulluoglu E, Vivoni ER. Modeling the ecohydrological role of aspect-controlled radiation on tree-grass-shrub coexistence in a semiarid climate. *Water Resour Res* 2013, 49:2872–2895.
- 266. van Wijk MT, Rodriguez-Iturbe I. Tree-grass competition in space and time: insights from a simple cellular automata model based on ecohydrological dynamics. *Water Resour Res* 2002, 38:18.1–18.15.
- 267. Baldocchi DD, Wilson KB. Modeling CO_2 and water vapor exchange of a temperate broadleaved forest across hourly to decadal time scales. *Ecol Model* 2001, 142:155–184.
- 268. Nouvellon Y, Rambal S, Seen DL, Moran MS, Lhomme JP, Begue A, Chehbouni AG, Kerr Y. Modelling of daily fluxes of water and carbon from shortgrass steppes. *Agr Forest Meteorol* 2000, 100:137–153.
- 269. Montaldo N, Rondena R, Albertson JD, Mancini M. Parsimonious modeling of vegetation dynamics for ecohydrologic studies of water-limited ecosystems. *Water Resour Res* 2005, 41(W10416):2005. doi:10.1029/2005WR004094.
- 270. Vertessy RA, Hatton TJ, Benyon RG, Dawes WR. Long-term growth and water balance predictions for a mountain ash (*Eucalyptus regnans*) forest

catchment subject to clear-felling and regeneration. *Tree Physiol* 1996, 16:221–232.

- 271. Cervarolo G, Mendicino G, Senatore A. A coupled ecohydrological-three-dimensional unsaturated flow model describing energy, H₂O and CO₂ fluxes. *Ecohydrology* 2010, 3:205–225. doi:10.1002/eco.111.
- 272. Battaglia M, Sands P, White D, Mummery D. CAB-ALA: a linked carbon, water and nitrogen model of forest growth for silvicultural decision support. *For Ecol Manage* 2004, 193:251–282.
- 273. Kirschbaum MUF, Keith H, Leuning R, Cleugh HA, Jacobsen KL, van Gorsel E, Raison J. Modelling net ecosystem carbon and water exchange of a temperate *Eucalyptus delegatensis* forest using multiple constraints. *Agr Forest Meteorol* 2007, 145:48–68.
- 274. Dufrêne E, Davi H, François C, le Maire G, Le Dantec V, Granier A. Modelling carbon and water cycles in a beech forest Part I: model description and uncertainty analysis on modelled NEE. *Ecol Model* 2005, 185:407–436.
- 275. Ogée J, Brunet Y, Loustau D, Berbigier P, Delzon S. MuSICA, a CO₂, water and energy multilayer, multileaf pine forest model: evaluation from hourly to yearly time scales and sensitivity analysis. *Glob Chang Biol* 2003, 9:697–717. doi:10.1046/j.1365-2486.2003.00628.
- 276. Wang L, Koike T, Yang K, Jackson TJ, Bindlish R, Yang D. Development of a distributed biosphere hydrological model and its evaluation with the southern great plains experiments (SGP97 and SGP99). J Geophys Res 2009, 114(D08107):2014. doi:10.1029/ 2008JD010800.
- 277. Govind A, Chen JM, Margolis H, Ju W, Sonnentag O, Giasson MA. A spatially explicit hydro-ecological modeling framework (BEPS-TerrainLab V2.0): model description and test in a boreal ecosystem in Eastern North America. *J Hydrol* 2009, 367:200–216.
- 278. Richards LA. Capillary conduction of liquids through porous mediums. *Physics* 1931, 1:318–333.
- 279. Celia MA, Bouloutas ET, Zarba RL. A general massconservative numerical solution for the unsaturated flow equation. *Water Resour Res* 1990, 26:1483–1496.
- 280. Simunek J, van Genuchten MT. Modeling nonequilibrium flow and transport processes using HYDRUS. *Vadose Zone J* 2008, 7:782–797.
- 281. Paniconi C, Putti M. A comparison of Picard and Newton iteration in the numerical solution of multidimensional variably saturated flow problems. *Water Resour Res* 1994, 30:3357–3374.
- 282. Schwinning S. The ecohydrology of roots in rocks. *Ecohydrology* 2010, 3:238–245. doi:10.1002/ eco.134.
- 283. Oleson KW, Lawrence DM, Bonan GB, Flanner MG, Kluzek E, Lawrence PJ, Levis S, Swenson SC,

Thornton PE. Technical description of version 4.0 of the Community Land Model (CLM). Technical Report NCAR/TN-478+STR, Natl. Cent. for Atmos. Res., Boulder, Colorado, 2010.

- 284. Huang M, Piao S, Sun Y, Ciais P, Cheng L, Mao J, Poulter B, Shi X, Zeng Z, Wang Y. Change in terrestrial ecosystem water-use efficiency over the last three decades. *Glob Chang Biol* 2015, 21:2366–2378. doi:10.1111/gcb.12873.
- 285. Gitelson AA, Gamon JA. The need for a common basis for defining light-use efficiency: implications for productivity estimation. *Remote Sens Environ* 2015, 156:196–201. doi:10.1016/j.rse.2014.09.017.
- 286. Anderson MC, Norman JM, Meyers TP, Diak GR. An analytical model for estimating canopy transpiration and carbon assimilation fluxes based on canopy light-use efficiency. *Agr Forest Meteorol* 2000, 101:265–289.
- 287. Istanbulluoglu E, Wang T, Wedin DA. Evaluation of ecohydrologic model parsimony at local and regional scales in a semiarid grassland ecosystem. *Ecohydrology* 2011, 5:121–142. doi:10.1002/eco.211.
- 288. Arora VK. Modelling vegetation as a dynamic component in soil-vegetation-atmosphere- transfer schemes and hydrological models. *Rev Geophys* 2002, 40:3-1–3-26. doi:10.1029/2001RG000103.
- 289. Friend AD, Stevens AK, Knox RG, Cannell MGR. A process-based, terrestrial biosphere model of ecosystem dynamics (Hybrid v3.0). *Ecol Model* 1997, 95:249–287.
- 290. Davi H, Barbaroux C, François C, Dufrêne E. The fundamental role of reserves and hydraulic constraints in predicting LAI and carbon allocation in forests. *Agr Forest Meteorol* 2009, 149:349–361.
- 291. Sala A, Woodruff DR, Meinzer FC. Carbon dynamics in trees: feast or famine? *Tree Physiol* 2012, 32:764– 775. doi:10.1093/treephys/tpr143.
- 292. Fatichi S, Leuzinger S, Körner C. Moving beyond photosynthesis: from carbon source to sink-driven vegetation modeling. *New Phytol* 2014, 201:1086–1095. doi:10.1111/nph.12614.
- 293. Manzoni S, Porporato A. Soil carbon and nitrogen mineralization: theory and models across scales. *Soil Biol Biochem* 2009, 41:1355–1379. doi:10.1016/j. soilbio.2009.02.031.
- 294. Parton WJ, Stewart JWB, Cole CV. Dynamics of C, N, P and S in grassland soils a model. *Biogeochemistry* 1988, 5:109–131.
- 295. Dickinson RE, Berry JA, Bonan GB, Collatz GJ, Field CB, Fung IY, Goulden M, Hoffmann WA, Jackson RB, Myneni R, et al. Nitrogen controls on climate model evapotranspiration. J Climate 2002, 15:278–294.
- 296. Kirschbaum MUF, Paul KI. Modelling C and N dynamics in forest soils with a modified version of

the CENTURY model. Soil Biol Biochem 2002, 34:341-354.

- 297. Xu-Ri, Prentice IC. Terrestrial nitrogen cycle simulation with a dynamic global vegetation model. *Glob Change Biol* 2008, 14:1745–1764. doi:10.1111/ j.1365-2486.2008.01625.x.
- 298. Zaehle S, Friend A. Carbon and nitrogen cycle dynamics in the O-CN land surface model: 1. Model description, site-scale evaluation, and sensitivity to parameter estimates. *Global Biogeochem Cycles* 2010, 24:GB1005. doi:10.1029/2009GB003521.
- 299. Orwin KH, Kirschbaum MUF, St John MG, Dickie IA. Organic nutrient uptake by mycorrhizal fungi enhances ecosystem carbon storage: a model-based assessment. *Ecol Lett* 2011, 14:493–502. doi:10.1111/j.1461-0248.2011.01611.x.
- 300. Wang G, Post WM, Mayes MA. Development of microbial-enzyme-mediated decomposition model parameters through steady-state and dynamic analyses. *Ecol Appl* 2013, 23:255–272.
- 301. Allison SD, Wallenstein MD, Bradford MA. Soilcarbon response to warming dependent on microbial physiology. *Nat Geosci* 2010, 3:336–340.
- 302. Wieder WR, Bonan GB, Allison SD. Global soil carbon projections are improved by modelling microbial processes. *Nat Clim Change* 2013, 3:909–912.
- 303. Manzoni S, Schimel JP, Porporato A. Responses of soil microbial communities to water stress: results from a meta-analysis. *Ecology* 2012, 93:930–938.
- 304. Li J, Wang G, Allison SD, Mayes MA, Luo Y. Soil carbon sensitivity to temperature and carbon use efficiency compared across microbial-ecosystem models of varying complexity. *Biogeochemistry* 2014, 119:67–84. doi:10.1007/s10533-013-9948-8.
- 305. Manzoni S, Trofymow JA, Jackson RB, Porporato A. Stoichiometric controls on carbon, nitrogen, and phosphorus dynamics in decomposing litter. *Ecol Monogr* 2010, 80:89–106.
- 306. Sinsabaugh RL, Manzoni S, Moorhead DL, Richter A. Carbon use efficiency of microbial communities: stoichiometry, methodology and modelling. *Ecol Lett* 2013, 16:930–939. doi:10.1111/ele.12113.
- 307. Wu L, McGechan MB, McRoberts N, Baddeley JA, Watson CA. SPACSYS: integration of a 3D root architecture component to carbon, nitrogen and water cycling—model description. *Ecol Model* 2007, 200:343–359.
- 308. Hinsinger P, Brauman A, Devau N, Gérard F, Jourdan C, Laclau JP, Le Cadre E, Jaillard B, Plassard C. Acquisition of phosphorus and other poorly mobile nutrients by roots. Where do plant nutrition models fail? *Plant Soil* 2011, 348:29–61. doi:10.1007/ s11104-011-0903-y.
- 309. Sterner RW, Elser JJ, Vitousek P. Ecological Stoichiometry: The Biology of Elements from Molecules to

the Biosphere. Princeton, NJ, USA: Princeton University Press; 2002.

- 310. Field HA, Mooney C. The photosynthesis-nitrogen relationship in wild plants. In: Givnish TJ, ed. On the *Economy of Plant Form and Function*. Cambridge UK: Cambridge University Press; 1986, 25–55.
- 311. Evans JR. Photosynthesis and nitrogen relationship in leaves of C3 plants. *Oecologia* 1989, 78:9–19.
- 312. Kattge J, Knorr W, Raddatz T. and C. Wirth. Quantifying photosynthetic capacity and its relationship to leaf nitrogen content for global-scale terrestrial biosphere models. *Glob Chang Biol* 2009, 15:976–991. doi:10.1111/j.1365-2486.2008.01744.x.
- 313. Niinemets U, Keenan TF, Hallik L. A worldwide analysis of within-canopy variations in leaf structural, chemical and physiological traits across plant functional types. *New Phytol* 2015, 205:973–993. doi:10.1111/nph.13096.
- 314. Welsh AH, Peterson AT, Altmann SA. The fallacy of averages. *Am Nat* 1988, 132:277–288.
- 315. Pappas C, Fatichi S, Rimkus S, Burlando P, Huber MO. The role of local scale heterogeneities in terrestrial ecosystem modeling. J Geophys Res Biogeosci 2015, 120:341–360. doi:10.1002/2014JG002735.
- 316. Dozier J, Frew J. Rapid calculation of terrain parameters for radiation modelling from digital elevation data. *IEEE Trans Geosci Remote Sens* 1990, 28:963-969.
- 317. Chen Y, Hall A, Liou KN. Application of threedimensional solar radiative transfer to mountains. J Geophys Res 2006, 111:D21111. doi:10.1029/ 2006JD007163.
- 318. Bertoldi G, Rigon R, Over TM. Impact of watershed geomorphic characteristics on the energy and water budgets. *J Hydrometeorol* 2006, 7:389–403.
- 319. Yetemen O, Istanbulluoglu EI, Flores-Cervantes JH, Vivoni ER, Bras RL. Ecohydrologic role of solar radiation on landscape evolution. *Water Resour Res* 2015, 51:1127–1157.
- 320. Ivanov VY, Bras RL, Vivoni ER. Vegetationhydrology dynamics in complex terrain of semiarid areas: 2. Energy-water controls of vegetation spatiotemporal dynamics and topographic niches of favorability. *Water Resour Res* 2008, 44:W03430. doi:10.1029/2006WR005595.
- 321. Fatichi S, Ivanov VY, Caporali E. A mechanistic ecohydrological model to investigate complex interactions in cold and warm water-controlled environments. 2. Spatiotemporal analyses. J Adv Model Earth Syst 2012, 4:M05003.
- 322. Band LE, Tague CL, Groffman P, Belt K. Forest ecosystem processes at the watershed scale: hydrological and ecological controls of nitrogen export. *Hydrol Process* 2001, 15:2013–2028. doi:10.1002/hyp.253.

- 323. Lin L, Webster JR, Hwang T, Band LE. Effects of lateral nitrate flux and instream processes on dissolved inorganic nitrogen export in a forested catchment: a model sensitivity analysis. *Water Resour Res* 2015, 51:2680–2695. doi:10.1002/2014WR015962.
- 324. Caracciolo D, Noto LV, Istanbulluoglu E, Fatichi S, Zhou X. Climate change and ecotone boundaries: insights from a cellular automata ecohydrology model in a mediterranean catchment with topography controlled vegetation pattern. *Adv Water Resour* 2014, 73:159–175.
- 325. Borgogno F, D'Odorico P, Laio F, Ridolfi L. Mathematical models of vegetation pattern formation in ecohydrology. *Rev Geophys* 2009, 47:RG1005. doi:10.1029/2007RG000256.
- 326. Rietkerk M, Boerlijst MC, van Langevelde F, HilleRisLambers R, van de Koppel J, Kumar L, Prins HHT, de Roos AM. Self-organization of vegetation in arid ecosystems. *Am Nat* 2002, 160:524–530.
- 327. Ursino M. Modeling banded vegetation patterns in semiarid regions: interdependence between biomass growth rate and relevant hydrological processes. *Water Resour Res* 2007, 43:W04412. doi:10.1029/2006WR005292.
- 328. Saco PM, Willgoose GR, Hancock GR. Ecogeomorphology of banded vegetation patterns in arid and semi-arid regions. *Hydrol Earth Syst Sci* 2007, 11:1717–1730.
- 329. Baudena M, D'Andrea F, Provenzale A. A model for soil-vegetation-atmosphere interactions in waterlimited ecosystems. *Water Resour Res* 2008, 44: W12429. doi:10.1029/2008WR007172.
- 330. Thompson S, Katul G, McMahon SM. Role of biomass spread in vegetation pattern formation within arid ecosystems. *Water Resour Res* 2008, 44: W10421. doi:10.1029/2008WR006916.
- 331. Accatino F, De Michele C, Vezzoli R, Donzelli D, Scholes RJ. Tree-grass co-existence in savanna: interactions of rain and fire. *J Theor Biol* 2010, 267:235-242.
- 332. Foti R, Ramírez JA. A mechanistic description of the formation and evolution of vegetation patterns. *Hydrol Earth Syst Sci* 2013, 17:63–84. doi:10.5194/ hess-17-63-2013.
- 333. Running SW, Coughlan JC. A general model of forest ecosystem processes for regional applications, I: hydrologic balance, canopy gas exchange, and primary production processes. *Ecol Model* 1988, 42:125–154.
- 334. Prentice IC, Cramer W, Harrison SP, Leemans R, Monsereud RA, Solomon AM. A global biome model based on plant physiology and dominance, soil properties and climate. *J Biogeogr* 1992, 19:117–134.
- 335. Prentice IC, Sykes MT, Cramer W. A simulation model for the transient effects of climate change on forest landscapes. *Ecol Modell* 1993, 65:51–70.

- 336. Lüdeke MKB, Badeck FW, Otto RD, Häger C, Dönges S, Kindermann J, Würth G, Lang T, Jäkel U, Klaudius A, et al. The Frankfurt Biosphere Model: a global process-oriented model of seasonal and longterm CO₂ exchange between terrestrial ecosystems and the atmosphere. I. Model description and illustrative results for cold deciduous and boreal forests. *Climate Res* 1994, 4:143–166.
- 337. Ruimy A, Dedieu G, Saugier B. TURC: a diagnostic model of continental gross primary productivity and net primary productivity. *Global Biogeochem Cycles* 1996, 10:269–285.
- 338. Bonan GB. Land-atmosphere interactions for climate system models-coupling biophysical, biogeochemical, and ecosystem dynamical processes. *Remote Sens Environ* 1995, 51:57–73.
- 339. Bonan GB, Levis S, Sitch S, Vertenstein M, Oleson KW. A dynamic global vegetation model for use with climate models: concepts and description of simulated vegetation dynamics. *Glob Chang Biol* 2003, 9:1543–1566. doi:10.1046/j.1529-8817.2003.00681.x.
- 340. Sitch S, Huntingford C, Gedney N, Levy PE, Lomas M, Piao L, Betts R, Cias P, Cox P, Friedlingstein P. Evaluation of the terrestrial carbon cycle, future plant geography and climate-carbon cycle feedbacks using five dynamic global vegetation models (DGVMs). *Glob Chang Biol* 2008, 14:2015–2039. doi:10.1111/ j.1365-2486.2008.01626.x.
- 341. Prinn RG. Development and application of Earth System Models. *Proc Natl Acad Sci USA* 2012, 110:3673-3680. doi:10.1073/pnas.1107470109.
- 342. Wenzel S, Cox PM, Eyring V, Friedlingstein P. Emergent constraints on climate carbon cycle feedbacks in the CMIP5 Earth system models. *J Geophys Res Biogeosci* 2014, 119:794–807. doi:10.1002/ 2013JG00259.
- 343. Dickinson RE, Henderson-Sellers A, Kennedy PJ. Biosphere-atmosphere transfer scheme (BATS) version 1E as coupled to the NCAR Community Climate Model. Technical Report NCAR/TN-387+STR, Natl. Cent. for Atmos. Res., Boulder, Colorado, 1993.
- 344. Thornton PE, Lamarque JE, Rosenbloom NA, Mahowald NM. Influence of carbon-nitrogen cycle coupling on land model response to CO₂ fertilization and climate variability. *Global Biogeochem Cycles* 2007, 21:GB4018. doi:10.1029/2006GB002868.
- 345. Prentice IC, Bondeau A, Cramer W, Harrison SP, Hickler T, Lucht W, Stich S, Smith B, Sykes MT. Dynamic global vegetation modeling: quantifying terrestrial ecosystem responses to large-scale environmental change. In: Canadell JP, Pataki DE, Pitelka LF, eds. *Terrestrial Ecosystems in a Changing World*. Berlin, Heidelberg: Springer; 2007.

- 346. Levis S. Modeling vegetation and land use in models of the Earth System. WIREs Clim Change 2010, 1:840-856. doi:10.1002/wcc.83.
- 347. Quillet A, Peng C, Garneau M. Toward dynamic global vegetation models for simulating vegetationclimate interactions and feedbacks: recent developments, limitations, and future challenges. *Environ Rev* 2010, 18:333–353. doi:10.1139/A10-016.
- 348. Medlyn BE, Duursma RA, Zeppel MJB. Forest productivity under climate change: a checklist for evaluating model studies. *WIREs Clim Change* 2011, 2:332–355. doi:10.1002/wcc.108.
- 349. Arain MA, Yuan F, Black TA. Soil-plant nitrogen cycling modulated carbon exchanges in a western temperate conifer forest in Canada. *Agr Forest Meteorol* 2006, 140:171–192.
- 350. Clark DB, Mercado LM, Sitch S, Jones CD, Gedney N, Best MJ, Pryor M, Rooney GG, Essery RLH, Blyth E, et al. The Joint UK Land Environment Simulator (JULES), model description Part 2: carbon fluxes and vegetation dynamics. *Geosci Model Dev* 2011, 4:701–722.
- 351. Niu GY, Yang ZL, Mitchell KE, Chen F, Ek MB, Barlage M, Kumar A, Manning K, Niyogi D, Rosero E, et al. The community Noah land surface model with multiparameterization options (Noah-MP):
 1. Model description and evaluation with local-scale measurements. *J Geophys Res* 2011, 116:D12109. doi:10.1029/2010JD015139.
- 352. Sitch S, Smith B, Prentice IC, Arneth A, Bondeau A, Cramer W, Kaplan JO, Levis S, Lucht W, Sykes MT, et al. Evaluation of ecosystem dynamics, plant geography and terrestrial carbon cycling in the LPJ dynamic global vegetation model. *Glob Chang Biol* 2003, 9:161–185.
- 353. Krinner G, Viovy N, de Noblet-Ducoudre N, Ogée J, Polcher J, Friedlingstein P, Ciais P, Sitch S, Prentice IC. A dynamic global vegetation model for studies of the coupled atmosphere-biosphere system. *Global Biogeochem Cycles* 2005, 19:GB1015. doi:10.1029/ 2003GB002199.
- 354. Levis S, Bonan GB, Vertenstein M, Oleson KW. The Community Land Model's Dynamic Global Vegetation Model (CLMDGVM): Technical description and user's guide. Tech. note, NCAR/TN-459+IA, Natl. Cent. for Atmos. Res., Boulder, CO, 2004.
- 355. Lawrence DM, Oleson KW, Flanner MG, Thornton PE, Swenson SC, Lawrence PJ, Zeng X, Yang ZL, Levis S, Sakaguchi K, et al. Parameterization improvements and functional and structural advances in version 4 of the Community Land Model. J Adv Model Earth Syst 2011, 3:M03001. doi:10.1029/ 2011MS000045.
- 356. Kucharik C, Foley J, Delire C, Fisher V, Coe M, Lenters J, Young-Molling C, Ramankutty N, Norman J, Gower S. Testing the performance of a dynamic

global ecosystem model: water balance, carbon balance, and vegetation structure. *Global Biogeochem Cycles* 2000, 14:795–825.

- 357. Yang X, Wittig V, Jain AK, Post W. Integration of nitrogen cycle dynamics into the Integrated Science Assessment Model for the study of terrestrial ecosystem responses to global change. *Global Biogeochem Cycles* 2009, 23:GB4029. doi:10.1029/ 2009GB003474.
- 358. Tian H, Chen G, Liu M, Zhang C, Sun G, Lu C, Xu X, Ren W, Pan S, Chappelka A. Model estimates of net primary productivity, evapotranspiration, and water use efficiency in the terrestrial ecosystems of the southern United States during 1895-2007. For Ecol Manage 2010, 259:1311–1327.
- 359. Thornton PE, Law BE, Gholz HL, Clark KL, Falge E, Ellsworth DS, Goldstein AH, Monson RK, Hollinger D, Falk M, et al. Modeling and measuring the effects of disturbance history and climate on carbon and water budgets in evergreen needleleaf forests. *Agr Forest Meteorol* 2002, 113:185–222.
- 360. Medvigy D, Wofsy SD, Munger JW, Hollinger DY, Moorcroft PR. Mechanistic scaling of ecosystem function and dynamics in space and time: ecosystem demography model version 2. *J Geophys Res* 2009, 114:G01002. doi:10.1029/2008JG000812.
- 361. Grant RF, Barr AG, Black TA, Margolis HA, Dunn AL, Metsaranta J, Wang S, McCaughey JH, Bourque CA. Interannual variation in net ecosystem productivity of Canadian forests as affected by regional weather patterns a Fluxnet-Canada synthesis. *Agr Forest Meteorol* 2009, 149:2022–2039.
- 362. Cox PM. Description of the TRIFFID Dynamic Global Vegetation Model. Technical Note 24, Hadley Centre, 2001.
- 363. Goll DS, Brovkin V, Parida BR, Reick CH, Kattge J, Reich PB, van Bodegom PM, Niinemets U. Nutrient limitation reduces land carbon uptake in simulations with a model of combined carbon, nitrogen and phosphorus cycling. *Biogeosciences* 2012, 9:3547– 3569. doi:10.5194/bg-9-3547-2012.
- 364. Sato H, Itoh A, Kohyama T. SEIB-DGVM: a new dynamic global vegetation model using a spatially explicit individual-based approach. *Ecol Model* 2007, 200:279–307.
- 365. Woodward FI, Lomas MR. Vegetation dynamics simulating responses to climatic change. *Biol Rev* 2004, 2004:643–670.
- 366. Smith B, Prentice IC, Sykes MT. Representation of vegetation dynamics in modelling of terrestrial ecosystems: comparing two contrasting approaches within European climate space. *Glob Ecol Biogeogr* 2001, 10:621–637.
- 367. Scheiter S, Langan L, Higgins SI. Next-generation dynamic global vegetation models: learning from

community ecology. New Phytol 2013, 198:957–969.

- 368. Keenan TF, Davidson E, Moffat A, Munger W, Richardson AD. Using model-data fusion to interpret past trends, and quantify uncertainties in future projections, of terrestrial ecosystem carbon cycling. *Glob Chang Biol* 2012, 18:2555–2569. doi:10.1111/j.1365-2486.2012.02684.x.
- 369. Carvalhais N, Reichstein M, Seixas J, Collatz GJ, Pereira JS, Berbigier P, Carrara A, Granier A, Montagnani L, Papale D, et al. Implications of the carbon cycle steady state assumption for biogeochemical modeling performance and inverse parameter retrieval. *Global Biogeochem Cycles* 2008, 22: GB2007. doi:10.1029/2007GB003033.
- 370. Pavlick R, Drewry DT, Bohn K, Reu B, Kleidon A. The Jena Diversity-Dynamic Global Vegetation Model (JeDi-DGVM): a diverse approach to representing terrestrial bio-geography and biogeochemistry based on plant functional trade-offs. *Biogeosciences* 2013, 10:4137–4177. doi:10.5194/bg-10-4137-2013 1058.
- 371. Pappas C, Fatichi S, Leuzinger S, Wolf A, Burlando P. Sensitivity analysis of a process-based ecosystem model: pinpointing parameterization and structural issues. *J Geophys Res Biogeosci* 2013, 118:505–528. doi:10.1002/jgrg.20035.
- 372. Wang YP, Houlton BZ, Field CB. A model of biogeochemical cycles of carbon, nitrogen, and phosphorus including symbiotic nitrogen fixation and phosphatase production. *Global Biogeochem Cycles* 2007, 21:GB1018. doi:10.1029/2006GB002797.
- 373. Yang X, Thornton PE, Ricciuto DM, Post WM. The role of phosphorus dynamics in tropical forests a modeling study using CLM-CNP. *Biogeosciences* 2014, 11:1667–1681. doi:10.5194/bg-11-1667-2014.
- 374. Zaehle S, Dalmonech D. Carbon-nitrogen interactions on land at global scales: current understanding in modelling climate biosphere feedbacks. *Curr Opin Environ Sustain* 2011, 3:311–320. doi:10.1016/j. cosust.2011.08.008.
- 375. Bugmann HKM, Yan X, Sykes MT, Martin P, Lindner M, Desanker PV, Cumming SG. A comparison of forest gap models: model structure and behaviour. *Clim Change* 1996, 34:289–313.
- 376. Snell RS, Huth A, Nabel JEMS, Bocedi G, Travis JMJ, Gravel D, Bugmann H, Gutiérrez AG, Hickler T, Higgins SI, et al. Using dynamic vegetation models to simulate plant range shifts. *Ecography* 2014, 37:1184–1197.
- 377. West GB, Enquist BJ, Brown JH. A general quantitative theory of forest structure and dynamics. *Proc Natl Acad Sci USA* 2009, 106:7040–7045. doi:10.1073/pnas.0812294106.

- 378. Moorcroft PR, Hurtt GC, Pacala SW. A method for scaling vegetation dynamics: the ecosystem demography model (ED). *Ecol Monogr* 2001, 71:557–585.
- 379. Purves DW, Lichstein JW, Strigul N, Pacala SW. Predicting and understanding forest dynamics using a simple tractable model. *Proc Natl Acad Sci USA* 2008, 105:17018–17022.
- 380. Strigul N, Pristinski D, Purves D, Dushoff J, Pacala S. Scaling from trees to forests: tractable macroscopic equations for forest dynamics. *Ecol Monogr* 2008, 78:523-545.
- 381. Kim Y, Knox RG, Longo M, Medvigy D, Hutyra LR, Pyle EH, Wofsy SC, Bras RL, Moorcroft PR. Seasonal carbon dynamics and water fluxes in an Amazon rainforest. *Glob Chang Biol* 2012, 18:1322– 1334. doi:10.1111/j.1365-2486.2011.02629.x.
- 382. Medvigy D, Moorcroft PR. Predicting ecosystem dynamics at regional scales: an evaluation of a terrestrial biosphere model for the forests of northeastern North America. *Philos Trans R Soc Lond Ser B* 2012, 367:222–235. doi:10.1098/rstb.2011.0253.
- 383. Dietze MC, Matthes JH. A general ecophysiological framework for modelling the impact of pests and pathogens on forest ecosystems. *Ecol Lett* 2014, 2014:1418–1426. doi:10.1111/ele.12345.
- 384. Zeppel MJB, Adams HD, Anderegg WRL. Mechanistic causes of tree drought mortality: recent results, unresolved questions and future research needs. *New Phytol* 2011, 192:800–803. doi:10.1111/j.1469-8137.2011.03960.x.
- 385. Franklin O, Aoki K, Seidl R. A generic model of thinning and stand density effects on forest growth, mortality and net increment. *Ann For Sci* 2009, 66:815– 815. doi:10.1051/forest/2009073.
- 386. Manusch C, Bugmann H, Heiri C, Wolf A. Tree mortality in dynamic vegetation models -a key feature for accurately simulating forest properties. *Ecol Model* 2012, 243:101–111.
- 387. Cox PM, Betts RA, Collins M, Harris PP, Huntingford C, Jones CD. Amazonian forest dieback under climate-carbon cycle projections for the 21st century. *Theor Appl Climatol* 2004, 78:137–156.
- 388. Malhi Y, Roberts JT, Betts RA, Killeen TJ, Li W, Nobre CA. Climate change, deforestation, and the fate of the Amazon. *Science* 2008, 319:169–172.
- 389. Malhi Y, Arag ao LEOC, Galbraith D, Huntingford C, Fisher R, Zelazowski P, Sitch S, McSweeney C, Meir P. Exploring the likelihood and mechanism of a climate-change-induced dieback of the Amazon rainforest. *Proc Natl Acad Sci USA* 2008, 106:20610– 20615. doi:10.1073/pnas.0804619106.
- 390. Nepstad DC, Stickler CM, Soares-Filho B, Merry F. Interactions among Amazon land use, forests and climate: prospects for a near-term forest tipping point. *Philos Trans R Soc B* 2008, 363:1737–1746.

- 391. Davidson EA, de Araújo AC, Artaxo P, Balch JK, Brown IF, Bustamante MMC, Coe MT, DeFries RS, Munger W, Keller M, et al. The Amazon basin in transition. *Nature* 2012, 481:321–328. doi:10.1038/ nature10717.
- 392. Markewitz D, Devine S, Davidson EA, Brando P, Nepstad DC. Soil moisture depletion under simulated drought in the Amazon: impacts on deep root uptake. *New Phytol* 2010, 187:592–607. doi:10.1111/ j.1469-8137.2010.03391.x.
- 393. Ivanov VY, Hutyra LR, Wofsy SC, Munger JW, Saleska SR, de Oliveira RC Jr, de Camargo PB. Root niche separation can explain avoidance of seasonal drought stress and vulnerability of overstory trees to extended drought in a mature Amazonian forest. *Water Resour Res* 2012, 48:W12507. doi:10.1029/ 2012WR011972.
- 394. Powell TL, Galbraith DR, Christoffersen BO, Harper A, Hewlley M, Imbuzeiro A, Rowland L, Almeida S, Brando PM, Lola da Costa AC, et al. Confronting model predictions of carbon fluxes with measurements of Amazon forests subjected to experimental drought. *New Phytol* 2013, 200:350–365. doi:10.1111/nph.12390.
- 395. Gatti LV, Gloor M, Miller JB, Doughty CE, Malhi Y, Domingues LG, Basso LS, Martinewski A, Correia CSC, Borges VF, et al. Drought sensitivity of Amazonian carbon balance revealed by atmospheric measurements. *Nature* 2014, 506:76. doi:10.1038/ nature12957.
- 396. Brando PM, Goetz SJ, Baccini A, Nepstad DC, Beck PSA, Christman MC. Seasonal and interannual variability of climate and vegetation indices across the Amazon. *Proc Natl Acad Sci USA* 2010, 107:14685– 14690. doi:10.1073/pnas.0908741107.
- 397. Fu R, Yin L, Li W, Arias PA, Dickinson RE, Huang L, Chakraborty S, Fernandes K, Liebmann B, Fisher R, et al. Increased dry-season length over southern Amazonia in recent decades and its implication for future climate projection. *Proc Natl Acad Sci USA* 2013, 110:18110–18115. doi:10.1073/pnas.1302584110.
- 398. Knox R, Bisht G, Wang J, Bras RL. Precipitation variability over the forest to non-forest transition in southwestern Amazonia. J Climate 2011, 24:2368–2377.
- 399. Butt N, de Oliveira PA, Costa MH. Evidence that deforestation affects the onset of the rainy season in Rondonia, Brazil. *J Geophys Res* 2011, 116:D11120. doi:10.1029/2010JD015174.
- 400. Khanna J, Medvigy D. Strong control of surface roughness variations on the simulated dry season regional atmospheric response to contemporary deforestation in Rondônia, Brazil. J Geophys Res Atmos 2014, 119:13067–13078. doi:10.1002/ 2014JD022278.

- 401. Myers-Smith IH, Forbes BC, Wilmking M, Hallinger M, Lantz T, Blok D, Tape KD, Macias-Fauria M, Sass-Klaassen U, Lévesque E, et al. Shrub expansion in tundra ecosystems: dynamics, impacts and research priorities. *Environ Res Lett* 2011, 6:045509. doi:10.1088/1748-9326/6/4/045509.
- 402. Loranty MM, Goetz SJ. Shrub expansion and climate feedbacks in Arctic tundra. *Environ Res Lett* 2012, 7:011005. doi:10.1088/1748-9326/7/1/011005.
- 403. Pomeroy JW, Bewley DS, Essery RLH, Hedstrom NR, Link T, Granger RJ, Sicart JE, Ellis CR, Janowicz JR. Shrub tundra snowmelt. *Hydrol Process* 2006, 20:923–941. doi:10.1002/hyp.6124.
- 404. Drake BG, Gonzàlez-Meler MA, Long SP. More efficient plants: a consequence of rising atmospheric CO₂? *Annu. Rev Plant Physiol Plant Mol Biol* 1997, 48:609–639.
- 405. Ainsworth EA, Rogers A. The response of photosynthesis and stomatal conductance to rising [CO₂]: mechanisms and environmental interactions. *Plant Cell Environ* 2007, 30:258–270. doi:10.1111/j.1365-3040.2007.01641.x.
- 406. Hickler T, Smith B, Prentice IC, Mjöfors K, Miller P, Arneth A, Sykes MT. CO₂ fertilization in temperate FACE experiments not representative of boreal and tropical forests. *Glob Chang Biol* 2008, 14:1531– 1542. doi:10.1111/j.1365-2486.2008.01598.x.
- 407. Körner C. Plant CO₂ responses: an issue of definition, time and resource supply. *New Phytol* 2006, 172:393–411. doi:10.1111/j.1469-8137.2006.01886.x.
- 408. Leuzinger S, Luo Y, Beier C, Dieleman W, Vicca S, Körner C. Do global change experiments overestimate impacts on terrestrial ecosystems? *Trends Ecol Evol* 2011, 26:236–241. doi:10.1016/j. tree.2011.02.011.
- 409. Gedney N, Cox PM, Bletts RA, Boucher O, Huntingford C, Stott PA. Detection of a direct carbon dioxide effect in continental river runoff records. *Nature* 2006, 439:835–838.
- 410. Betts RA, Boucher O, Collins M, Cox PM, Falloon PD, Gedney N, Hemming DL, Huntingford C, Jones CD, Sexton DMH, et al. Projected increase in continental runoff due to plant responses to increasing carbon dioxide. *Nature* 2007, 448:1037–1041. doi:10.1038/nature06045.
- 411. Huntingford C, Zelazowski P, Galbraith D, Mercado LM, Sitch S, Fisher R, Lomas M, Walker AP, Jones CD, Booth BBB, et al. Simulated resilience of tropical rainforests to CO₂-induced climate change. *Nat Geosci* 2013, 6:268–273. doi:10.1038/ngeo1741.
- 412. Schimel D, Stephens BB, Fisher JB. Effect of increasing CO₂ on the terrestrial carbon cycle. *Proc Natl Acad Sci USA* 2015, 112:436–441.
- 413. Körner C, Morgan JA, Norby RJ. CO₂ fertilization: when, where, how much?. In: *Terrestrial Ecosystems*

in a Changing World. Berlin, Germany: Springer; 2007.

- 414. Norby RJ, Zak DR. Ecological lessons from free-Air CO₂ enrichment (FACE) experiments. *Annu Rev Ecol Evol Syst* 2011, 42:181–203. doi:10.1146/annurevecolsys-102209-144647.
- 415. van der Sleen P, Groenendijk P, Vlam M, Anten NPR, Boom A, Bongers F, Pons TL, Terburg G, Zuidema PA. No growth stimulation of tropical trees by 150 years of CO₂ fertilization but water-use efficiency increased. *Nat Geosci* 2015, 8:24–28. doi:10.1038/ NGEO2313.
- 416. Medvigy D, Wofsy SC, Munger JW, Moorcroft PR. Responses of terrestrial ecosystems and carbon budgets to current and future environmental variability. *Proc Natl Acad Sci USA* 2010, 107:8275-8280. doi:10.1073/pnas.0912032107.
- 417. Paschalis A, Fatichi S, Katul GG, Ivanov VY. Crossscale impact of climate temporal variability on ecosystem water and carbon fluxes. *J Geophys Res Biogeosci* 2015, 120. doi:10.1002/2015JG003002.
- 418. Mutke J, Barthlott W. Patterns of vascular plant diversity at continental to global scales. *Biol Skrifter* 2005, 55:521–531.
- 419. Bonan GB, Levis S, Kergoat L, Oleson KW. Landscapes as patches of plant functional types: an integrating concept for climate and ecosystem models. *Global Biogeochem Cycles* 2002, 16:Pages 5-1-5-23. doi:10.1029/2000GB001360.
- 420. Fyllas NM, Gloor E, Mercado LM, Sitch S, Quesada CA, Domingues TF, Galbraith DR, Torre-Lezama A, Vilanova E, Ramírez-Angulo E, et al. Analysing Amazonian forest productivity using a new individual and trait-based model (TFS v.1). *Geosci Model Dev* 2014, 7:1251–1269. doi:10.5194/gmd-7-1251-2014.
- 421. Sakschewski B, von Bloh W, Boit A, Rammig A, Kattge J, Poorter L, Peñuelas J, Thonicke K. Leaf and stem economics spectra drive diversity of functional plant traits in a dynamic global vegetation model. *Glob Chang Biol* 2015, 21:2711–2725. doi:10.1111/ gcb.12870.
- 422. Verheijen LM, Aerts R, Brovkin V, Cavender-Bares J, Cornelissen JHC, Kattge J, van Bodegom PM. Inclusion of ecologically based trait variation in plant functional types reduces the projected land carbon sink in an Earth System Model. *Glob Chang Biol* 2015, 21:3074–3086. doi:10.1111/gcb.12871.
- 423. Wright IJ, Reich PB, Westoby M, Ackerly DD, Baruch Z, Bongers F, Cavender-Bares J, Chapin T, Cornelissen JHC, Diemer M, et al. The worldwide leaf economics spectrum. *Nature* 2004, 428:821–827.
- 424. Chave J, Coomes D, Jansen S, Lewis SL, Swenson NG, Zanne AE. Towards a worldwide wood economics spectrum. *Ecology Letters* 2009, 12:351–366. doi:10.1111/j.1461-0248.2009.01285.x.

- 425. Manzoni S, Vico G, Porporato A, Katul G. Biological constraints on water transport in the soil-plantatmosphere system. *Adv Water Resour* 2013, 51:292-304.
- 426. Reich PB, Walters MB, Ellsworth DS. From tropics to tundra: global convergence in plant functioning. *Proc Natl Acad Sci USA* 1997, 94:13730–13734.
- 427. Reich PB. The world-wide 'fast-slow' plant economics spectrum: a traits manifesto. *J Ecol* 2014, 102:275-301. doi:10.1111/1365-2745.12211.
- 428. ter Steege H, Pitman NCA, Sabatier D, Baraloto C, Salom ao RP, Guevara JE, Phillips OL, Castilho CV, Magnusson WE, Molino JF, et al. Hyperdominance in the Amazonian tree flora. *Science* 2013, 342:1243092. doi:10.1126/science.1243092.
- 429. Saxton KE, Rawls WJ. Soil water characteristic estimates by texture and organic matter for hydrologic solutions. *Soil Sci Soc Am J* 2006, 70:1569–1578. doi:10.2136/sssaj2005.0117.
- 430. Romano N, Nasta P, Severino G, Hopmans JW. Using bimodal lognormal functions to describe soil hydraulic properties. *Soil Sci Soc Am J* 2011, 75:468–480. doi:10.2136/sssaj2010.0084.
- 431. Yang X, Post WM, Thornton PE, Jain A. The distribution of soil phosphorus for global biogeochemical modeling. *Biogeosciences* 2013, 10:2525–2537. doi:10.5194/bg-10-2525-2013.
- 432. de Brogniez D, Ballabio C, Stevens A, Jones RJ, Montanarella L, van Wesemael B. A map of the topsoil organic carbon content of Europe generated by a generalized additive model. *Eur J Soil Sci* 2015, 66:121– 134. doi:10.1111/ejss.12193.
- 433. Dietze MC, Sala A, Carbone MS, Czimczik CI, Mantooth JA, Richardson AD, Vargas R. Nonstructural carbon in woody plants. *Annu Rev Plant Biol* 2014, 65:667–687. doi:10.1146/annurev-arplant-050213-040054.
- 434. Aubinet M, Grelle A, Ibrom A, Rannik Ü, Moncrie J, Foken T, Kowalski AS, Martin PH, Berbigier P, Bernhofer C, et al. Estimates of the annual net carbon and water exchange of forests: the EUROFLUX methodology. *Adv Ecol Res* 2000, 30:113–175.
- 435. Baldocchi D, Falge E, Gu L, Olson R, Hollinger D, Running S, Anthoni P, Bernhofer C, Davis K, Evans R, et al. FLUXNET: a new tool to study the temporal and spatial variability of ecosystem-scale carbon dioxide, water vapor, and energy flux densities. *Bull Am Meteorol Soc* 2001, 82:2415–2434.
- 436. Baldocchi DD. Assessing the eddy covariance technique for evaluating carbon dioxide exchange rates of ecosystems: past, present and future. *Glob Chang Biol* 2003, 9:479–492.
- 437. Baldocchi DD. Breathing of the terrestrial biosphere: lessons learned from a global network of carbon dioxide flux measurement systems. *Austral J Bot* 2008, 56:1–26.

- 438. Aubinet M, Vesala T, Papale D. Eddy Covariance: A Practical Guide to Measurement and Data Analysis. Springer Atmospheric Sciences. Dordrechet, The Netherlands: Springer Netherlands; 2012.
- 439. Robinson DA, Campbell CS, Hopmans JW, Hornbuckle BK, Jones SB, Knight R, Ogden F, Selker J, Wendroth O. Soil moisture measurements for ecological and hydrological watershed scale observatories: a review. Vadose Zone J 2008, 7:358–389. doi:10.2136/vzj2007.0143.
- 440. Vereecken H, Huisman JA, Bogena HR, Vanderborght J, Vrugt JA, Hopmans JW. On the value of soil moisture measurements in vadose zone hydrology: a review. *Water Resour Res* 2008, 44:W00D06. doi:10.1029/2008WR006829.
- 441. Fatichi S, Katul GG, Ivanov VY, Pappas C, Paschalis A, Consolo A, Kim J, Burlando P. Abiotic and biotic controls of soil moisture spatio-temporal variability and the occurrence of hysteresis. *Water Resour Res* 2015, 51:3505–3524. doi:10.1002/2014WR016102.
- 442. Falge E, Baldocchi D, Olson R, Anthoni P, Aubinet M, Bernhofer C, Burba G, Ceulemans R, Clement R. Gap filling strategies for defensible annual sums of net ecosystem exchange. *Agr Forest Meteorol* 2001, 107:43–69.
- 443. Moffat AM, Papale D, Reichstein M, Hollinger DY, Richardson AD, Barr AG, Beckstein C, Braswell BH, Churkina G, Desai AR, et al. Comprehensive comparison of gap-filling techniques for eddy covariance net carbon fluxes. *Agr Forest Meteorol* 2007, 147:209–232.
- 444. Wilson K, Goldstein A, Falge E, Aubinet M, Baldocchi D, Berbigier P, Bernhofer C, Ceulemans R, Dolman H, Field C, et al. Energy balance closure at FLUXNET sites. *Agr Forest Meteorol* 2002, 113:223-243.
- 445. Leuning R, van Gorsel E, Massman WJ, Isaac PR. Reflections on the surface energy imbalance problem. *Agr Forest Meteorol* 2012, 156:65–74. doi:10.1016/ j.agrformet.2011.12.002.
- 446. Liu C, Zhang X, Zhang Y. Determination of daily evaporation and evapotranspiration of winter wheat and maize by large-scale weighing lysimeter and micro-lysimeter. *Agr Forest Meteorol* 2002, 111:109–120.
- 447. Seneviratne SI, Lehner I, Gurtz J, Teuling AJ, Lang H, Moser U, Grebner D, Menzel L, Schro K, Vitvar T, et al. Swiss prealpine Rietholzbach research catchment and lysimeter: 32 year time series and 2003 drought event. *Water Resour Res* 2012, 48:W06526.
- 448. Lischke H, Löffler TJ. Intraspecific density dependence is required to maintain species diversity in spatio-temporal forest simulations with reproduction. *Ecol Model* 2006, 198:341–361. doi:10.1016/j. ecolmodel.2006.05.005.

- 449. Nepstad DC, Tohver IM, Ray D, Moutinho P, Cardinot G. Mortality of large trees and lianas following experimental drought in an Amazon forest. *Ecology* 2007, 88:2259–2269.
- 450. Brienen RJW, Phillips OL, Feldpausch TR, Gloor E, Baker TR, Lloyd J, Lopez-Gonzalez G, Monteagudo-Mendoza A, Malhi Y, Lewis SL, et al. Long-term decline of the Amazon carbon sink. *Nature* 2015, 519:344–348. doi:10.1038/nature14283.
- 451. Malhi Y, Wood D, Baker TR, Wright J, Phillips OL, Cochrane T, Meir P, Chave J, Almeida S, Arroyo L, et al. The regional variation of aboveground live biomass in old-growth Amazonian forests. *Global Change Biol* 2006, 12:1107–1138. doi:10.1111/ j.1365-2486.2006.01120.x.
- 452. Babst F, Alexander MR, Szejner P, Bouriaud O, Klesse S, Roden J, Ciais P, Poulter B, Frank D, Moore DJP, et al. A tree-ring perspective on the terrestrial carbon cycle. *Oecologia* 2014, 176:307–322. doi:10.1007/s00442-014-3031-6.
- 453. Fatichi S, Leuzinger S. Reconciling observations with modeling: the fate of water and carbon allocation in a mature deciduous forest exposed to elevated CO₂. *Agr Forest Meteorol* 2013, 174-175:144–157. doi:10.1016/j.agrformet.2013.02.005.
- 454. De Kauwe MG, Medlyn BE, Zaehle S, Walker AP, Dietze MC, Hickler T, Jain AK, Luo Y, Parton WJ, Prentice IC, et al. Forest water use and water use efficiency at elevated CO₂: a model-data intercomparison at two contrasting temperate forest FACE sites. *Glob Chang Biol* 2013, 19:1759–1779. doi:10.1111/ gcb.12164.
- 455. Zaehle S, Medlyn BE, De KMG, WalkerAP DMC, Hickler T, Luo Y, Wang YP, El-Masri B, Thornton P, et al. Evaluation of 11 terrestrial carbon-nitrogen cycle models against observations from two temperate free-Air CO₂ enrichment studies. *New Phytol* 2014, 202:803–822. doi:10.1111/nph.12697.
- 456. Medlyn BE, Zaehle S, De Kauwe MG, Walker AP, Dietze MC, Hanson PJ, Hickler T, Jain AK, Luo Y, Parton W, et al. Using ecosystem experiments to improve vegetation models. *Nat Clim Change* 2015, 5:528–534. doi:10.1038/nclimate2621.
- 457. Smith NG, Rodgers VL, Brzostek ER, Kulmatiski A, Avolio ML, Hoover DL, Koerner SE, Grant K, Jentsch A, Fatichi S, et al. Towards a better integration of biological data from precipitation manipulation experiments into Earth system models. *Rev Geophys* 2014, 52:412–434. doi:10.1002/ 2014RG000458.
- 458. Kayler ZE, De Boeck HJ, Fatichi S, Grünzweig JM, Merbold L, Beier C, McDowell N, Dukes JS. Experiments to confront the environmental extremes of climate change. *Front Ecol Environ* 2015, 13:219–225. doi:10.1890/140174.

- 459. Schimel D, Pavlick R, Fisher JB, Asner GP, Saatchi S, Townsend P, Miller C, Frankenberg C, Hibbard K, Cox P. Observing terrestrial ecosystems and the carbon cycle from space. *Glob Chang Biol* 2015, 2015:1762–1776. doi:10.1111/gcb.12822.
- 460. Vivoni ER, Rango A, Anderson CA, Pierini NA, Schreiner-McGraw AP, Saripalli S, Laliberte AS. Ecohydrology with unmanned aerial vehicles. *Ecosphere* 2014:5(130). doi:10.1890/ES14-00217.1.
- 461. Asner GP, Powell GVN, Mascaro J, Knapp DE, Clark JK, Jacobson J, Kennedy-Bowdoin T, Balaji A, Paez-Acosta G, Victoria E, et al. High-resolution forest carbon stocks and emissions in the Amazon. Proc Natl Acad Sci USA 2010, 107:16738–16742.
- 462. Asner GP, Clark JK, Mascaro J, Galindo García GA, Chadwick KD, Encinales DNA, Paez-Acosta G, Montenegro EC, Kennedy-Bowdoin T, Duque A, et al. High-resolution mapping of forest carbon stocks in the Colombian Amazon. *Biogeosciences* 2012, 2012:2683–2696.
- 463. Justice CO, Vermote E, Townshend JRG, Defries R, Roy DP, Hall DK, Salomonson VV, Privette JL, Riggs G, Strahler A, et al. The Moderate Resolution Imaging Spectroradiometer (MODIS): land remote sensing for global change research. *IEEEns on Geosciences and Remote Sensing* 1998, 36:1228–1249.
- 464. Guanter L, Zhang Y, Jung M, Joiner J, Voig M, Berry JA, Frankenberg C, Huete AR, Zarco-Tejada P, Lee JE, et al. Global and time-resolved monitoring of crop photosynthesis with chlorophyll fluorescence. *Proc Natl Acad Sci USA* 2014, 111:E1327–E1333. doi:10.1073/pnas.1320008111.
- 465. Myneni RB, Hoffman S, Knyazikhin Y, Privette JL, Glassy J, Tian Y, Wang Y, Song X, Zhang Y, Smith GR, et al. Global products of vegetation leaf area and fraction absorbed PAR from year one of MODIS data. *Remote Sens Environ* 2002, 83:214–231.
- 466. Yang Y, Shang S, Guan H, Jiang L. A novel algorithm to assess gross primary production for terrestrial ecosystems from MODIS imagery. *J Geophys Res* 2013, 118:590–605. doi:10.1002/jgrg.20056.
- 467. Serbin SP, Singh A, McNeil BE, Kingdon CC, Townsend PA. Spectroscopic determination of leaf morphological and biochemical traits for northern temperate and boreal tree species. *Ecol Appl* 2014, 24:1651–1669.
- 468. Knyazikhin Y, Schull MA, Stenberg P, Mottus M, Rautiainen M, Yang Y, Marshak A, Carmona PL, Kaufmann RK, Lewis P, et al. Hyperspectral remote sensing of foliar nitrogen content. *Proc Natl Acad Sci* USA 2012, 3:E185–E192. doi:10.1073/ pnas.1210196109.

- 469. Homolová L, Malenovsky Z, Clevers JGPW, García-Santos G, Schaepman ME. Review of optical-based remote sensing for plant trait mapping. *Ecol Complex* 2013, 15:1–16.
- 470. Brocca L, Hasenauer S, Lacava T, Melone F, Moramarco T, Wagner W, Dorigo W, Matgen P, Martínez-Fernández J, Llorens P, et al. Soil moisture estimation through ASCAT and AMSR-E sensors: an intercomparison and validation study across Europe. *Remote Sens Environ* 2011, 115:3390–3408.
- 471. Entekhabi D, Njoku EG, O'Neill PE, Kellogg KH, Crow WT, Edelstein WN, Entin JK, Goodman SD, Jackson TJ, Johnson J, et al. The soil moisture active passive (SMAP) mission. *Proc IEEE* 2010, 98:704– 716. doi:10.1109/JPROC.2010.2043918.
- 472. Cox PM, Pearson D, Booth BB, Friedlingstein P, Huntingford C, Jones CD, Luke CM. Sensitivity of tropical carbon to climate change constrained by carbon dioxide variability. *Nature* 2013, 494:341–344. doi:10.1038/nature11882.
- 473. Graven HD, Keeling RF, Piper SC, Patra PK, Stephens BB, Wofsy SC, Welp LR, Sweeney C, Tans PP, Kelley JJ, et al. Enhanced seasonal exchange of CO₂ by northern ecosystems since 1960. *Science* 2013, 341:1085–1089.
- 474. Frankenberg C, Pollock R, Lee RAM, Rosenberg R, Blavier JF, Crisp D, O'Dell CW, Osterman GB, Roehl C, Wennberg PO, et al. The Orbiting Carbon Observatory (OCO-2): spectrometer performance evaluation using pre-launch direct sun measurements. *Atmos Meas Tech* 2015, 8:301–313. doi:10.5194/amt-8-301-2015.
- 475. McDowell NG, Ryan MG, Zeppel MJB, Tissue DT. Improving our knowledge of drought-induced forest mortality through experiments, observations, and modeling. *New Phytol* 2013, 200:289–293.
- 476. Xu C, McDowell NG, Sevanto S, Fisher RA. Our limited ability to predict vegetation dynamics under water stress. *New Phytol* 2013, 200:298–300.
- 477. Fatichi S, Ivanov VY. Interannual variability of evapotranspiration and vegetation productivity. *Water Resour Res* 2014, 50:3275-3294. doi:10.1002/ 2013WR015044.
- 478. Hartig F, Dyke J, Hickler T, Higgins SI, O'Hara RB, Scheiter S, Huth A. Connecting dynamic vegetation models to data - an inverse perspective. J Biogeogr 2012, 39:2240-2252.
- 479. Dietze MC, LeBauer D, Kooper R. On improving the communication between models and data. *Plant Cell Environ* 2013, 36:1575–1585.