

## Minireview

# Targeting the Notch Pathway to Prevent Rejection

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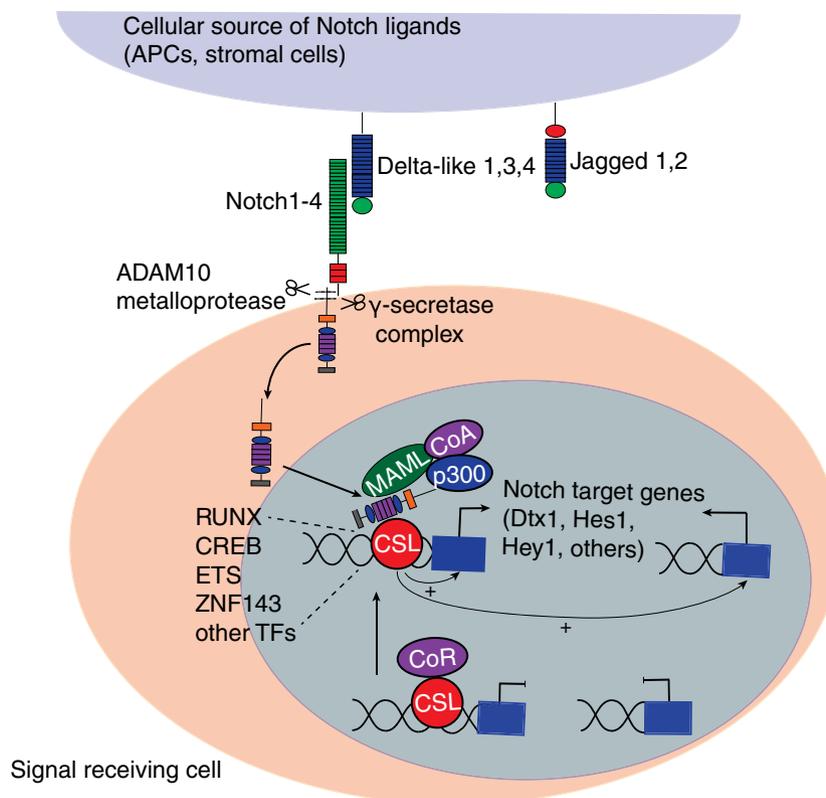
**Immune rejection is mediated by a complex interplay of cellular and humoral mechanisms. Current therapeutic strategies, which rely on global immunosuppression, can result in serious complications and are not completely effective. Notch signaling is a cell-to-cell communication pathway that plays an important role during T cell development and in the regulation of peripheral immune responses. Initial work, performed mainly through gain-of-function approaches, paradoxically identified Notch as an inducer of tolerance; however, recent studies using loss-of-function approaches in mouse models of transplant rejection and graft-versus-host disease have clarified an important role for Notch as a central mediator of T cell alloreactivity. Short-term inhibition of individual Notch ligands in the peritransplant period had long-lasting protective effects. In a vascularized heart allograft model, blockade of Delta-like Notch ligands dampened both cellular and humoral rejection. In this minireview, we summarize current knowledge about the role of Notch signaling during allograft rejection and provide an overarching mechanism through which Notch acts to promote T cell pathogenicity and allograft damage. We propose that targeting elements of the Notch pathway could provide a new therapeutic approach to prevent allograft rejection.**

**Abbreviations:** APC, antigen-presenting cell; CSL, CBF1/suppressor-of-hairless/Lag-1; DC, dendritic cell; DII1, Delta-like 1; DII3, Delta-like 3; DII4, Delta-like 4; DNMA1L, dominant negative MAML1; GVHD, graft-versus-host disease; ICN, intracellular Notch; IFN- $\gamma$ , interferon  $\gamma$ ; Jag1, Jagged 1; Jag2, Jagged 2; MAML, Mastermind-like; MAPK, mitogen-activated protein kinase; NF $\kappa$ B, nuclear factor  $\kappa$ B; Teff, effector T cells; TF, transcription factor; Th, T helper cell; TNF, tumour necrosis factor; Treg, regulatory T cells

First identified in *Drosophila* close to a century ago, the Notch pathway has received increasing recognition for its functions in mammalian biology and for its effects in health and disease (1,2). Notch plays an essential role at early stages of T cell development in the thymus because Notch ligands expressed by thymic epithelial cells induce commitment of thymocyte progenitors to the T lineage program (3). Notch also regulates subsets of innate lymphoid cells, B cells, and dendritic cells (DCs) (3). In addition, recent studies uncovered major functions of Notch signaling in the control of antigen-driven immune responses in the periphery (4,5). These effects of the Notch pathway are prominent in T cell alloimmunity and highly relevant in the context of allograft rejection (6–14). In this minireview, we introduce aspects of Notch regulation that are critical for its functions in immunobiology and discuss emerging evidence showing that Notch signaling could be an attractive new therapeutic target to prevent organ rejection.

## Introduction to Notch Signaling

Notch signaling is a highly conserved cell-to-cell communication pathway triggered by Notch ligand–receptor interactions between adjacent cells (Figure 1) (1,2). In mammals, four Notch receptors (Notch1–4) have been identified in addition to five Notch ligands of the Jagged (Jag1 and Jag2) and Delta-like (DII1, DII3, DII4) families. Jag1, Jag2, DII1 and DII4 have agonistic properties, whereas DII3 functions as a natural antagonist of the pathway (1). Notch ligand–receptor interactions induce sequential proteolytic cleavage of the receptor by an ADAM family metalloprotease (ADAM10) and by the  $\gamma$ -secretase complex, ultimately releasing intracellular Notch (ICN) into the cytoplasm (1,2). ICN migrates into the nucleus where it interacts with a DNA-binding transcription factor referred to as CSL (CBF1/suppressor-of-hairless/Lag-1) or RBP- $\kappa$  (encoded by *Rbpj*). On Notch activation, CSL and ICN nucleate a large transcription activation complex that recruits a member of the Mastermind-like (MAML) family and other coactivators to mediate transcriptional activation of Notch target genes (1,2,15). Recent studies in Notch-driven cancer cell lines detected binding of CSL and ICN at thousands of



**Figure 1: Overview of Notch signaling.** Mammalian Notch signaling is initiated by interactions between Notch receptors (Notch1–4) and Notch ligands (Delta-like 1, 3, and 4; Jagged 1 and 2). Ligand-receptor binding triggers two sequential proteolytic cleavages of the Notch receptor by the ADAM10 metalloprotease and by the  $\gamma$ -secretase complex, releasing ICN into the cytoplasm. On entry into the nucleus, ICN forms a transcriptional activation complex with the TF CSL, a member of the MAML family, and other coactivators such as p300. ICN/CSL transcriptional complexes often assemble adjacent to other TFs and can regulate Notch target gene expression proximally through promoter binding or distally through enhancer binding and long-range interactions. APC, antigen-presenting cell; CSL, CBF1/suppressor-of-hairless/Lag-1; ICN, intracellular Notch; MAML, Mastermind-like; TF, transcription factor.

genomic sites (16); however, only a fraction (<10%) of these sites appeared to be dynamically regulated by Notch signaling. Regulated sites were enriched for concomitant binding of specific transcription factors, suggesting context-specific regulation of the Notch target gene landscape by cooperating factors. Moreover, the majority of dynamic Notch-binding sites were located in distant elements with superenhancer features, suggesting that Notch is involved in long-range chromatin regulation (17). Additional studies about the molecular mechanisms of Notch-mediated transcriptional activation will be essential to understand the context-specific effects of the Notch pathway. Of note, little is known so far about the nature and regulation of Notch transcriptional targets in mature T cells.

Evolution selected multiple mechanisms to ensure tight control over Notch signaling because Notch receptors can deliver powerful signals with profound effects on cellular differentiation and function. Active ICN is rapidly targeted for degradation, ensuring that Notch signals are short lived in nature, unless receptors are repetitively

engaged (1,2). Notch receptors are relatively ubiquitous, although the *Notch1–4* paralogs are differentially expressed by specific cell types. In contrast, individual Notch ligands are expressed in a highly controlled fashion within defined anatomical niches. This regulatory arrangement ensures tight control over temporal and spatial delivery of Notch signals. Thymic epithelial cells, for example, express high levels of Dll4 under control of the *Foxn1* gene, generating an intrathymic niche that delivers strong Notch signals to T lineage progenitors while restricting normal T cell development to a single anatomical site (18). Evidence for restricted and regulated expression of Notch ligands within secondary lymphoid organs has also emerged (19). Jagged and Delta-like ligands were reported to induce different functional outcomes in several contexts (20). Based on these considerations, individual Notch ligands and receptors can have distinct biological effects. These distinct effects offer opportunities for the development of therapeutic targeting strategies that provide specific benefits and avoid the systemic side effects of pan-Notch inhibition (7,21,22).

## Early Work on Notch Signaling in T Cell Alloreactivity and Tolerance

Initial studies exploring a potential role for Notch signaling in mature T cell function and alloreactivity relied heavily on gain-of-function strategies. Lamb's group was the first to spark interest in a role for Notch as an inducer of tolerance (23). While studying T cell responses against a house dust mite protein, they engineered mouse DCs to overexpress the Notch ligand Jag1. Adoptive transfer of antigen-pulsed Jag1-overexpressing DCs led to antigen-specific hyporesponsiveness. Building on this concept, the Brenner group tested the ability of Jag1-overexpressing antigen-presenting cells (APCs) to modulate responses to alloantigens or viral antigens (24,25). Using Jag1-transduced Epstein–Barr virus–transformed lymphoblastoid cell lines, they observed decreased T cell reactivity and evidence for transferrable suppressive effects. Similar outcomes were described in an *in vivo* cardiac allograft model, in which Dallman and colleagues adoptively transferred mouse L cell fibroblasts engineered to overexpress MHC alloantigens and the Notch ligand Dll1 (26). Although Dll1-overexpressing L cells delayed allograft rejection in a CD8<sup>+</sup> T cell–dependent manner, it is unclear whether the effects were the result of direct engagement of Dll1 with Notch receptor in T cells. Similar observations were reported recently on *in vivo* transfer of a Jag1-transduced DC line in combination with CD40 blockade (27). Altogether, these studies suggested that inducing artificially high Notch signals in T cells could generate a state of antigen-specific tolerance.

In parallel, other laboratories observed that expression of specific Notch ligands could be induced by innate stimuli in professional APCs (20,28). In coculture systems, Delta-like or Jagged Notch ligands within APCs were reported to promote skewing of T cell differentiation toward the T helper type 1 (Th1) or Th2 lineage, respectively (20,29), although dichotomous inductive effects of Delta-like and Jagged Notch ligands were not detected in subsequent studies (30). Collectively, although helpful for recognizing an important role for Notch in T cell alloreactivity, the use of artificial *ex vivo* conditions and overexpression models led to conclusions that were contradictory and have to be interpreted with caution.

Subsequent *in vivo* loss-of-function studies of mature T cells identified an even broader range of effects of Notch on both CD4<sup>+</sup> and CD8<sup>+</sup> T cell reactivity and function (31–33), many of which are discussed in the “Mechanistic Considerations” section of this review.

## In Vivo Studies of Notch Signaling in Allograft Rejection

In recent years, several groups have used genetic and pharmacological loss-of-function approaches to evaluate the *in vivo* effects of Notch signaling in alloreactive T cell responses, in the settings of allograft rejection (Table 1) and graft-versus-host disease (GVHD) (Table 2) (6–9,11–14). An emerging consensus across these studies indicates that Notch signaling is a major proinflammatory pathway in T cell alloimmunity and that Notch inhibition can dampen both allograft rejection and GVHD. Consequently, the true *in vivo* functions of Notch signaling appear to be diametrically different from the tolerogenic effects first detected using artificial gain-of-function strategies (23–26). Furthermore, these studies identify Notch inhibition as a new promising therapeutic approach to mitigate the damaging consequences of T cell alloreactivity.

Riella and coworkers used monoclonal antibodies to target the Notch ligand Dll1 in an MHC-mismatched cardiac allograft transplantation model (9). Systemic anti-Dll1 antibodies delayed allograft rejection when provided in conjunction with costimulatory blockade in *Cd28*-deficient mice or in recipients treated with CTLA4-Ig fusion protein. Protection was associated with decreased production of IL-2, interferon  $\gamma$  (IFN- $\gamma$ ), IL-6 and IL-17 by donor-specific T cells but with increased production of the Th2 cytokines IL-4 and IL-5. In this model, the protective effects of anti-Dll1 antibodies were lost when transplantation was performed in STAT6-deficient recipients or on concomitant IL-4 neutralization, indicating that increased IL-4 production was important to delay rejection. This paper was the first to demonstrate a pathogenic effect of Notch signaling and a therapeutic benefit of Notch inhibition in allograft rejection *in vivo*, in stark contrast to earlier literature using artificial gain-of-function systems. Because this study was performed in the

**Table 1:** Modulation of Notch signaling in models of allograft rejection

Allograft rejection model	Outcome and suggested mechanism	Citation
MHC-mismatched heart BALB/c→B6 anti-Dll1	Delayed allograft rejection Decreased IFN- $\gamma$ , IL-2, IL-6 production Increased IL-4, IL-5, IL-13 production	(9)
MHC-mismatched heart BALB/c→B6 <i>Cd4-cre;ROSA<sup>D<sup>N</sup>MAML</sup></i> hosts anti-Dll1 and anti-Dll4	Delayed allograft rejection Decreased IFN- $\gamma$ production Decreased serum antibodies and complement deposition with systemic neutralization of Dll1 and Dll4	(14)

Dll, Delta-like; IFN- $\gamma$ , interferon  $\gamma$ .

**Table 2:** Modulation of Notch signaling in models of graft-versus-host disease

GVHD model	Outcome and suggested mechanism	Citation
B6→BALB/c major mismatch B6→B6xDBA/2 F1 <i>Cd4-cre;ROSA<sup>DNMAML</sup></i> donors <i>Cd4-cre;rbpj<sup>fl/fl</sup></i> donors	Protection from GVHD-induced mortality Decreased IFN- $\gamma$ , TNF- $\alpha$ , IL-17, IL-2 production Increased Treg numbers	(6)
B6→BALB/c major mismatch <i>Cd4-cre;notch1<sup>fl/fl</sup>;notch2<sup>fl/fl</sup></i> donors $\gamma$ -secretase inhibitors anti-Notch1 and anti-Notch2 anti-Dll1 and anti-Dll4	Protection from GVHD-induced mortality Decreased IFN- $\gamma$ , IL-2 production Increased Treg numbers	(7)
B6→BALB/c major mismatch B6→BALB/b minor mismatch <i>Cd4-cre;ROSA<sup>DNMAML</sup></i> hosts <i>Cd4-cre;rbpj<sup>fl/fl</sup></i> hosts	Protection from GVHD-induced mortality Blunted MAPK and NF $\kappa$ B signaling Increased expression of negative regulators of T cell signaling	(8)
B6→B6xBALB/c F1 major mismatch <i>Mx1-Cre;notch1<sup>fl/fl</sup></i> donors $\gamma$ -secretase inhibitors	Protection from bone marrow failure Decreased IFN- $\gamma$ , Gzmb production Decreased expression of T-bet	(11)
B6→BALB/c major mismatch B6→B6xDBA/2 F1 anti-Dll4	Protection from GVHD-induced mortality Decreased IFN- $\gamma$ , IL-17 production	(12)
B6→BALB/c major mismatch <i>Foxp3-cre;rbpj<sup>fl/fl</sup></i> donors <i>Foxp3-cre;notch1<sup>fl/fl</sup></i> donors	Protection from GVHD-induced mortality Increased Treg survival, numbers Decreased IFN- $\gamma$ production by Teff	(13)

Dll, Delta-like; GVHD, graft-versus-host disease; IFN- $\gamma$ , interferon  $\gamma$ ; MAPK, mitogen-activated protein kinase; NF $\kappa$ B, nuclear factor  $\kappa$ B; Teff, effector T cells; TNF- $\alpha$ , tumour necrosis factor  $\alpha$ ; Treg, regulatory T cells.

presence of costimulatory blockade and examined only the effect of Dll1 inhibition and not other Notch ligands, it was unclear if similar outcomes would be observed without costimulatory blockade or with more complete Notch inhibition. Because of the systemic nature of Dll1 inhibition, it could not be determined if the protective effects of anti-Dll1 antibodies were related to their direct effects on T cells and/or on other cell types.

To achieve a higher level of Notch inhibition in alloreactive T cells, Wood et al studied an MHC-mismatched heart allograft model in mice expressing the pan-Notch inhibitor dominant negative MAML1 (DNMAML) specifically in T cells (14). DNMAML blocks transcriptional activation downstream of all Notch ligands and receptors (15). In *Cd4-Cre;ROSA26<sup>DNMAML</sup>* mice, DNMAML expression first arises in CD4<sup>+</sup>CD8<sup>+</sup> thymocytes without interfering with early Notch-dependent stages of T cell development (34). Consequently, mature CD4<sup>+</sup> and CD8<sup>+</sup> T cells develop normally from double-positive thymocytes in these mice but cannot respond to Notch signals during subsequent T cell responses due to DNMAML expression. This strategy is highly effective in capturing the overall effects of Notch signaling in T cell immunity, regardless of the individual Notch ligands and receptors involved (6,8). DNMAML mice rejected MHC-mismatched hearts in a delayed fashion (14). Although the delay was relatively modest in the absence of other interventions, it was observed in the absence of costimulatory blockade, suggesting that complete Notch inhibition in T cells could achieve higher protection from rejection than the level of protection seen with partial Notch inhibition with anti-Dll1

antibodies (9,14). Importantly, on concomitant CD8 depletion prior to transplantation, DNMAML expression within host T cells led to markedly enhanced protection, with median allograft survival of >40 days. These findings suggested that CD4<sup>+</sup> alloreactive T cells were particularly sensitive to Notch inhibition. Mechanistically, DNMAML led to decreased production of both IFN- $\gamma$  and IL-4 by donor-reactive T cells, decreased immune cell infiltration and an increased ratio of regulatory T cells (Treg) to effector T cells (Teff) within the graft. DNMAML recipients also showed delayed appearance of donor-specific alloantibodies, suggesting a role for Notch in T cell help for allospecific B cell responses.

Building on their observations from studies using genetic pan-Notch inhibition in T cells, Wood et al assessed the impact of humanized anti-Dll1 and anti-Dll4 antibodies, alone or in combination, on allograft rejection (14). This approach was chosen because of the effects of Dll1 in transplant rejection (9) and the dominant role of Dll1/4 Notch ligands in GVHD (7,12). Anti-Dll1/4 antibodies had high therapeutic activity in allograft rejection. Both anti-Dll1 and anti-Dll4 antibodies by themselves induced significant protection, indicating that these two Notch ligands were involved nonredundantly in the rejection process. Combined administration of anti-Dll1 and anti-Dll4 antibodies was the most effective strategy tested, enabling long-term engraftment in CD8-depleted recipients and markedly delayed rejection even in CD8-replete hosts. Surprisingly, systemic Dll1/4 blockade provided a higher degree of protection from allograft rejection than DNMAML-mediated pan-Notch inhibition in T cells.

Enhanced protection was associated with a persistent decrease in donor-specific alloantibody titers, plasma cell numbers and complement deposition in the graft. These findings suggest that the therapeutic activity of anti-Dll1/4 antibodies is related both to their effects on T cells (preventing acute cellular rejection) and on the B cell response (preventing chronic rejection, at least in part through humoral mechanisms). Furthermore, long-term protection was observed on short-term Dll1/4 blockade in the peritransplant period, similar to findings in allogeneic bone marrow transplantation and GVHD (7,14).

Together, Dll1 and Dll4 Notch ligands play dominant roles in the regulation of alloimmunity, but the role of Jagged ligands remains unclear. To start addressing this question, Riella et al used a Jag2-specific antibody in mouse heart allograft rejection models (10). This antibody was previously shown to specifically bind Jag2 but was suggested to facilitate forward Notch signaling in an *in vitro* coculture system through unknown mechanisms (35). Jag2-specific antibodies induced accelerated rejection in two heterotopic heart transplantation models. Accelerated rejection was associated with complex immunological changes, including increased production of IL-6 and Th2 cytokines, and increased Treg expansion. These findings suggest that Jag2 can have a proinflammatory role in allograft rejection, but interpretation is challenging because the biochemical impact of the Jag2-specific antibody used in these studies is not fully characterized. Future work using genetic approaches and other pharmacological reagents could clarify the role of Jagged ligands in transplant rejection.

Although there are differences in experimental approaches, particularly in terms of global Notch inhibition in T cells versus selective systemic targeting of Notch pathway components, studies of allograft rejection and *in vivo* Notch inhibition delineate elements of an emerging consensus: (i) Notch signaling is a major pathway that promotes inflammation and opposes tolerance in allograft transplantation; (ii) Notch signaling controls alloreactive T cell immunity but may also regulate non-T cell subsets that contribute to the pathogenesis of organ rejection; (iii) targeting the Notch pathway has therapeutic potential to prevent allograft rejection, with short-term blockade of Delta-like Notch ligands in the peritransplant period capable of inducing long-term effects.

### ***In Vivo* Studies of Notch Signaling in GVHD**

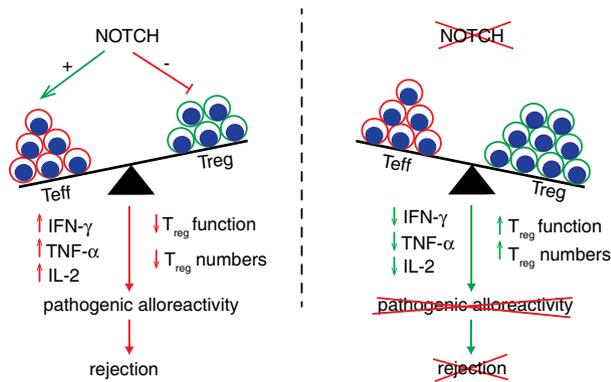
Although the focus of this minireview is on allograft rejection, it is useful for comparison to highlight other recent findings about the role of Notch signaling in GVHD (Table 2) (6–8,11,12). Using DNMA1L expression or *Rbpj* inactivation to block all canonical Notch signals in T cells, Zhang et al first reported major protective effects of Notch inhibition in mouse models of acute GVHD (6).

Notch inhibition led to markedly increased survival of transplant recipients. Notch-deprived alloreactive T cells showed decreased production of multiple inflammatory cytokines (including IFN- $\gamma$ , tumor necrosis factor  $\alpha$ , IL-17 and IL-4) and increased expansion of preexisting Treg (6,8). Decreased cytokine production was observed in both CD4<sup>+</sup> and CD8<sup>+</sup> T cells and was associated with features of acquired hyporesponsiveness in alloreactive T cells (8). Individual T cell effector functions were affected to a variable extent by Notch inhibition because *in vivo* T cell proliferation and expansion were preserved in irradiated recipients. T cell cytotoxic functions were also largely maintained in the absence of Notch signaling, leading to the preservation of potent graft-versus-tumor effects. Using a genetic strategy to inactivate the *Notch1* or the *Rbpj* gene only within Treg, Chatila's group reported that Notch negatively regulated Treg numbers and function *in vivo* and that Notch inhibition in Tregs alone conferred therapeutic benefits in acute GVHD (13). Consequently, Notch inhibition may exert beneficial immunomodulation in conventional CD4<sup>+</sup> and CD8<sup>+</sup> T cells as well as Treg.

Therapeutically,  $\gamma$ -secretase inhibitors were shown to be effective in a mouse model of alloimmune bone marrow injury (11); however, in acute GVHD models involving lethal irradiation, systemic pan-Notch inhibition with  $\gamma$ -secretase inhibitors was poorly tolerated because of on-target toxicity in the gut (7). To bypass this toxicity, the roles of individual Notch ligands and receptors were investigated using genetic models and paralog-specific monoclonal antibodies (7,12). Notch1/Notch2 receptors and Dll1 and Dll4 Notch ligands accounted for all effects of Notch signaling in alloreactive T cells during GVHD, with dominant roles for Notch1 and Dll4. Dll1 and Dll4 blockade emerged as the most promising therapeutic approach to prevent GVHD and avoid systemic side effects of pan-Notch inhibition. Interestingly, transient early Dll1 and Dll4 inhibition was essential and sufficient to confer long-term GVHD protection (7). Altogether, clear parallels are emerging between the functions of Notch signaling in acute GVHD and allograft rejection. In both cases, early Dll1 and Dll4-mediated Notch signals exert profound and durable proinflammatory effects such that transient Dll1 and Dll4 inhibition provides long-lasting therapeutic benefits.

### **Mechanistic Considerations**

The molecular mechanisms of Notch action in mature T cells remain under active investigation. The most relevant observations are and will continue to be derived from *in vivo* experiments that evaluate physiological levels of Notch signaling in defined immunological contexts. Along these lines of investigation, Notch was recently reported to regulate specific functions of CD4<sup>+</sup> and CD8<sup>+</sup> T cells, including *in vivo* survival and



**Figure 2: Emerging model of Notch signaling as a central regulator of alloreactivity versus tolerance.** Notch drives T cell pathogenicity during allotransplantation by enhancing pathogenic functions in Teff while decreasing numbers and beneficial immunosuppressive functions of FoxP3<sup>+</sup> Treg. Interfering with the Notch pathway can reverse this imbalance by dampening proinflammatory cytokine production by Teff and enhancing both Treg function and numbers. Importantly, short-term Notch inhibition in the peritransplant period can confer long-lasting immunological benefits. IFN- $\gamma$ , interferon  $\gamma$ ; Teff, effector T cells; TNF- $\alpha$ , tumour necrosis factor  $\alpha$ ; Treg, regulatory T cells.

metabolism, responsiveness to CD28-mediated costimulatory signals, and CD8<sup>+</sup> T cell differentiation (29,31,33, 36–38). An important overarching theme is that Notch does not appear to function as a lineage-specific regulator but rather as a regulator of T cell reactivity and function. In T cell alloimmunity, multiple investigators observed that Notch inhibition tips the balance between inflammatory Teff and suppressive Treg functions (Figure 2) (6,13,14). Notch-deficient Teff cells appear defective in their production of multiple inflammatory cytokines, whereas Notch-deficient Treg accumulate in higher absolute or relative numbers and may have enhanced suppressive ability. Furthermore, selective blockade of the Notch1 receptor in a fully MHC-mismatched model reduced Teff and increased Treg, significantly prolonging allograft survival (Riella, unpublished data). T<sub>regs</sub> not only were expanded in number but also had enhanced suppressive function in the absence of Notch1 signaling. The dual effects on both Teff and Treg likely account for the prolonged impact of transient Notch inhibition. Of note, key downstream effects of Notch signaling are likely to be mediated by canonical CSL/MAML-dependent transcriptional mechanisms (6,13), but the functionally essential targets of Notch signaling in Teff and Treg remain to be identified.

### Therapeutic Implications and Future Directions

Based on available preclinical data, we propose that Notch signaling is an attractive new therapeutic target to prevent allograft rejection. Short-term inhibition of Notch

signaling exerts a longstanding beneficial impact by dampening the alloimmune response, highlighting the promise of transient Notch inhibition strategies in the peritransplant period (7,14). Beyond allograft rejection, Notch inhibition could also be beneficial in other T cell-mediated immune disorders, including GVHD and autoimmunity (7,39,40). In practice, targeting individual Notch ligands and receptors with specific mAbs currently appears to be the most promising therapeutic approach to target Notch signaling in alloimmune cells and avoid the systemic side effects of pan-Notch inhibition (7,9,14). As for other strategies, an important challenge will be to translate these findings from preclinical mouse models into more advanced preclinical models (e.g. nonhuman primates) and into humans. Given that Notch is an ancient and highly conserved signaling pathway, it is tempting to speculate that key features of its effects will be conserved, although this needs to be investigated systematically. Promising observations such as enhanced Notch signaling in Teff during kidney transplant rejection in humans compared with nonrejectors provide optimism for this translational approach (Riella, unpublished data). Advances in our understanding of the immunobiological effects of Notch and carefully designed translational investigations could unravel the full therapeutic potential of Notch inhibition in allograft rejection and other immune-mediated disorders.

### Disclosure

The authors of this manuscript have no conflicts of interest to disclose as described by the *American Journal of Transplantation*.

### References

- Kopan R, Ilagan MX. The canonical Notch signaling pathway: Unfolding the activation mechanism. *Cell* 2009; 137: 216–233.
- Guruharsha KG, Kankel MW, Artavanis-Tsakonas S. The Notch signalling system: Recent insights into the complexity of a conserved pathway. *Nat Rev Genet* 2012; 13: 654–666.
- Radtke F, MacDonald HR, Tacchini-Cottier F. Regulation of innate and adaptive immunity by Notch. *Nat Rev Immunol* 2013; 13: 427–437.
- Amsen D, Helbig C, Backer RA. Notch in T cell differentiation: All things considered. *Trends Immunol* 2015; 36: 802–814.
- Osborne BA, Minter LM. Notch signalling during peripheral T-cell activation and differentiation. *Nat Rev Immunol* 2007; 7: 64–75.
- Zhang Y, Sandy AR, Wang J, et al. Notch signaling is a critical regulator of allogeneic CD4<sup>+</sup> T-cell responses mediating graft-versus-host disease. *Blood* 2011; 117: 299–308.
- Tran IT, Sandy AR, Carulli AJ, et al. Blockade of individual Notch ligands and receptors controls graft-versus-host disease. *J Clin Invest* 2013; 123: 1590–1604.
- Sandy AR, Chung J, Toubai T, et al. T cell-specific notch inhibition blocks graft-versus-host disease by inducing a hyporesponsive program in alloreactive CD4<sup>+</sup> and CD8<sup>+</sup> T cells. *J Immunol* 2013; 190: 5818–5828.

9. Riella LV, Ueno T, Batal I, et al. Blockade of Notch ligand delta1 promotes allograft survival by inhibiting alloreactive Th1 cells and cytotoxic T cell generation. *J Immunol* 2011; 187: 4629–4638.
10. Riella LV, Yang J, Chock S, et al. Jagged2-signaling promotes IL-6-dependent transplant rejection. *Eur J Immunol* 2013; 43: 1449–1458.
11. Roderick JE, Gonzalez-Perez G, Kuksin CA, et al. Therapeutic targeting of NOTCH signaling ameliorates immune-mediated bone marrow failure of aplastic anemia. *J Exp Med* 2013; 210: 1311–1329.
12. Mochizuki K, Xie F, He S, et al. Delta-like ligand 4 identifies a previously uncharacterized population of inflammatory dendritic cells that plays important roles in eliciting allogeneic T cell responses in mice. *J Immunol* 2013; 190: 3772–3782.
13. Charbonnier LM, Wang S, Georgiev P, Sefik E, Chatila TA. Control of peripheral tolerance by regulatory T cell-intrinsic Notch signaling. *Nat Immunol* 2015; 16: 1162–1173.
14. Wood S, Feng J, Chung J, et al. Transient blockade of delta-like Notch ligands prevents allograft rejection mediated by cellular and humoral mechanisms in a mouse model of heart transplantation. *J Immunol* 2015; 194: 2899–2908.
15. Maillard I, Weng AP, Carpenter AC, et al. Mastermind critically regulates Notch-mediated lymphoid cell fate decisions. *Blood* 2004; 104: 1696–1702.
16. Wang H, Zou J, Zhao B, et al. Genome-wide analysis reveals conserved and divergent features of Notch1/RBPJ binding in human and murine T-lymphoblastic leukemia cells. *Proc Natl Acad Sci U S A* 2011; 108: 14908–14913.
17. Wang H, Zang C, Taing L, et al. NOTCH1-RBPJ complexes drive target gene expression through dynamic interactions with superenhancers. *Proc Natl Acad Sci U S A* 2014; 111: 705–710.
18. Calderon L, Boehm T. Synergistic, context-dependent, and hierarchical functions of epithelial components in thymic microenvironments. *Cell* 2012; 149: 159–172.
19. Fasnacht N, Huang HY, Koch U, et al. Specific fibroblastic niches in secondary lymphoid organs orchestrate distinct Notch-regulated immune responses. *J Exp Med* 2014; 211: 2265–2279.
20. Amsen D, Blander JM, Lee GR, Tanigaki K, Honjo T, Flavell RA. Instruction of distinct CD4 T helper cell fates by different notch ligands on antigen-presenting cells. *Cell* 2004; 117: 515–526.
21. Ridgway J, Zhang G, Wu Y, et al. Inhibition of Dll4 signalling inhibits tumour growth by deregulating angiogenesis. *Nature* 2006; 444: 1083–1087.
22. Wu Y, Cain-Hom C, Choy L, et al. Therapeutic antibody targeting of individual Notch receptors. *Nature* 2010; 464: 1052–1057.
23. Hoyne GF, Le Roux I, Corsin-Jimenez M, et al. Serrate1-induced notch signalling regulates the decision between immunity and tolerance made by peripheral CD4(+) T cells. *Int Immunol* 2000; 12: 177–185.
24. Vigouroux S, Yvon E, Wagner HJ, et al. Induction of antigen-specific regulatory T cells following overexpression of a Notch ligand by human B lymphocytes. *J Virol* 2003; 77: 10872–10880.
25. Yvon ES, Vigouroux S, Rousseau RF, et al. Overexpression of the Notch ligand, Jagged-1, induces alloantigen-specific human regulatory T cells. *Blood* 2003; 102: 3815–3821.
26. Wong KK, Carpenter MJ, Young LL, et al. Notch ligation by Delta1 inhibits peripheral immune responses to transplantation antigens by a CD8+ cell-dependent mechanism. *J Clin Invest* 2003; 112: 1741–1750.
27. Lin Y, Chen W, Li J, et al. Overexpression of Jagged-1 combined with blockade of CD40 pathway prolongs allograft survival. *Immunol Cell Biol* 2015; 93: 213–217.
28. Schaller MA, Neupane R, Rudd BD, et al. Notch ligand Delta-like 4 regulates disease pathogenesis during respiratory viral infections by modulating Th2 cytokines. *J Exp Med* 2007; 204: 2925–2934.
29. Maekawa Y, Tsukumo S, Chiba S, et al. Delta1-Notch3 interactions bias the functional differentiation of activated CD4+ T cells. *Immunity* 2003; 19: 549–559.
30. Ong CT, Sedy JR, Murphy KM, Kopan R. Notch and presenilin regulate cellular expansion and cytokine secretion but cannot instruct Th1/Th2 fate acquisition. *PLoS ONE* 2008; 3: e2823.
31. Helbig C, Gentek R, Backer RA, et al. Notch controls the magnitude of T helper cell responses by promoting cellular longevity. *Proc Natl Acad Sci U S A* 2012; 109: 9041–9046.
32. Bailis W, Yashiro-Ohtani Y, Fang TC, et al. Notch simultaneously orchestrates multiple helper T cell programs independently of cytokine signals. *Immunity* 2013; 39: 148–159.
33. Laky K, Evans S, Perez-Diez A, Fowlkes BJ. Notch signaling regulates antigen sensitivity of naive CD4+ T cells by tuning co-stimulation. *Immunity* 2015; 42: 80–94.
34. Tu L, Fang TC, Artis D, et al. Notch signaling is an important regulator of type 2 immunity. *J Exp Med* 2005; 202: 1037–1042.
35. Elyaman W, Bassil R, Bradshaw EM, et al. Notch receptors and Smad3 signaling cooperate in the induction of interleukin-9-producing T cells. *Immunity* 2012; 36: 623–634.
36. Backer RA, Helbig C, Gentek R, et al. A central role for Notch in effector CD8(+) T cell differentiation. *Nat Immunol* 2014; 15: 1143–1151.
37. Maekawa Y, Ishifune C, Tsukumo S, Hozumi K, Yagita H, Yasutomo K. Notch controls the survival of memory CD4+ T cells by regulating glucose uptake. *Nat Med* 2015; 21: 55–61.
38. Mathieu M, Duval F, Daudelin JF, Labrecque N. The Notch signaling pathway controls short-lived effector CD8+ T cell differentiation but is dispensable for memory generation. *J Immunol* 2015; 194: 5654–5662.
39. Sandy AR, Stoolman J, Malott K, Pongtornpipat P, Segal BM, Maillard I. Notch signaling regulates T cell accumulation and function in the central nervous system during experimental autoimmune encephalomyelitis. *J Immunol* 2013; 191: 1606–1613.
40. Billiard F, Lobry C, Darrasse-Jeze G, et al. Dll4-Notch signaling in Flt3-independent dendritic cell development and autoimmunity in mice. *J Exp Med* 2012; 209: 1011–1028.