

Supporting Information

for Adv. Healthcare Mater., DOI: 10.1002/adhm.201700980

Biomaterials-Based Approaches to Tumor Spheroid and Organoid Modeling

Pradip Shahi Thakuri, Chun Liu, Gary D. Luker, and Hossein Tavana*



Copyright WILEY-VCH Verlag GmbH & Co. KGaA, 69469 Weinheim, Germany, 2016.

Supporting Information

Biomaterials-Based Approaches to Tumor Spheroid and Organoid Modeling

Pradip Shahi Thakuri, Chun Liu, Gary D. Luker, Hossein Tavana*

Table S1. A summary of natural and synthetic materials used for 3D tumor modeling as spheroids and organoids, types of cancer cells used in each study, and major conclusion of the studies are shown.

| BIOMATERIAL | 3D TUMOR MODEL | OUTCOMES | REFEREN CE | | |
|---|--|--|---------------|--|--|
| Natural materials for tumor spheroid cultures | | | | | |
| Collagen | Mono-culture of breast cancer cells MDA-MB-231 | Reduced spheroid invasion in stiffer matrices; paclitaxel resistance | 47 | | |
| | Co-culture of breast cancer cells MCF7 and mammary fibroblasts | Formation of tight clusters with distinct border, apical-basal polarity, and lumen | 49 | | |
| | Co-culture of colon cancer cells LS174T and cancer associated fibroblasts | Co-culture promoted LS174T spheroid invasion in collagen matrix | 50 | | |
| | Co-culture of liver cancer cells HEPG2 and fibroblasts NIH 3T3 | Co-culture spheroids were highly resistant to doxorubicin compared to the mono-culture cancer cells spheroids | 53 | | |
| | Mono-culture of colon cancer cells HT-29 | Upregulation of HIF-1α and VEGF angiogenic factors | 51 | | |
| | Mono-culture of breast cancer cells MDA-MB-231 | Upregulation of HIF-1α and VEGF-A; delayed upregulation of both markers at lower cell density | 40 | | |
| | Mono-culture of ovarian cancer cells SK-OV-3 | Mimicked oxygen gradients by 3D spatial localization of cells in a six-layered scaffold | 54 | | |
| Laminin rich ECM (IrECM) | Mono-culture of various prostate and breast cancer cells | Malignant sub-types displayed disorganized, proliferative and non-polar colonies and were distinguishable from non-malignant cells based on their morphology; Gene expression of malignant cancer with distinct morphology frequently clustered together | 56,57,58 | | |
| | Mono-culture of Lewis lung carcinoma LLC1 | Cytoskeleton arrangement without formation of stress fibers; LLC1 clusters had marked differences in metabolic, MAPK, cell adhesion, and immune response genes compared to the 2D culture of the cells | 60 | | |
| | Co-culture of prostate cancer cells PC-3 and bone stromal cells; Pre-formed breast cancer spheroids in IrECM | Increased α_6 - and/or β_1 -integrin in the co- culture compared to the mono-culture of PC-3 cells; Blocking $\beta1$ integrin inhibited the growth of the spheroids | 70, 66 | | |

WILEY-VCH

| Alginate | Mono-culture spheroids of hepatocarcinoma cells | Preserved acini, apical morphogenesis, stem cell markers and β-catenin signaling; Wnt/β-catenin signaling pathway activity promoted spheroid formation and maintaining cancer cells stemness; Cells in spheroid culture were highly tumorigenic in mouse compared to cells cultured in 2D | 71 | | |
|---|---|---|-----|--|--|
| | Mono-culture of oral cancer cells | High level of pro-angiongneic maker IL-8 but moderate alteration of VEGF expression | 72 | | |
| | Mono-culture of hepatocarcinoma cells | Actin reorganization to facilitate spheroid formation; Cell in spheroids expressed tight junctions, canaliculi like structures, showed microvilli on their surfaces and were arranged in trabecular form | 73 | | |
| | Mono-culture of prostate cancer cells PC-3 | Enriched expression of stem cell maker genes NANOG, OCT4, CD44, and CD133 | 74 | | |
| Chitosan | Mono-culture of colon cancer and hepatocarcinoma cells | Upregulated cancer stem cell genes (OCT4, NANOG, CD133, CD44), epithelial maker EpCAM, and non-canonical Wnt-STAT3 signaling in CD44 hepatocellular carcinoma cells | 79 | | |
| Chitosan-Alginate (CA) | Mono-culture of Hepatocarcinoma cells | High expression of Glypican-3 in spheroids compared to 2D cultures; cells were more tumorigenic, formed large tumors, and expressed pro-angiogenic growth factor such as IL-8, bFGF and VGF | 83 | | |
| | Mono-culture of glioblastoma cells U-87 MG | Increased in vivo angiogenic capability of CA pre-cultured cells | 85 | | |
| Hyaluronic acid (HA) | Mono-culture of prostate patient derived cells | Continued expression of androgen receptor in long term culture; resistance to docetaxel treatment | 94 | | |
| | Mono-culture of prostate cancer cells LNCaP | Higher mRNA level of E-cadherin, and integrins α_5 and β_1 | 95 | | |
| | Mono-culture of prostate cancer cells LNCaP | Spheroids showed cortically organized F- actin, and increased protein and mRNA expression of pro-angiongenic factors VEGEF ₁₆₅ and IL-8 | 98 | | |
| Chitosan- hyaluronan (CH) | Mono-cultures of A549 and H1299 small cell lung cancers | Strong upregulation of N-cadherin, vimentin, fibronectin, anti-apoptotic genes BCRC5 and BCL2, EMT-related transcription factor TWIST1, and cancer stem cell genes CD44, CD133, SOX2, NANOG, POU5F1 | 99 | | |
| Silk | Osteosacroma | Level of Cyclin B, E2F1, Ki67, and PcNA were similar to those in a SCID mouse model | 104 | | |
| | Breast cancer MDA-MB- 231 cells | Spheroids displayed proliferation gradients of cells and growth that followed Gompertz law; Upregulated IL-8 and VEGF markers | 105 | | |
| Synthetic materials for tumor spheroid cultures | | | | | |
| RGD | Mono-cultures of ovarian | Proliferation dependent on integrin | 109 | | |

WILEY-VCH

| | II 0\/ N/7 0 | 1. 1 | |
|-------------------------|---------------------------------|--|----------|
| functionalized | cancer cells OV-MZ-6 and SKOV-3 | binding capacity; Significantly | |
| PEG hydrogels | SKUV-3 | upregulated α_3 , α_5 , β_1 integrins and MMP-9 levels; resistance to paclitaxel | |
| PEG-fibrinogen | Mono-cultures of breast | MCF-7, SK-BR-3 cells formed compact | 110, 111 |
| i Lo-iibi iilogeli | cancer cells MCF-7, SK- | spheroids in large range of hydrogel | 110, 111 |
| | BR-3, and MDA-MB-231; | stiffness; MDA-MB-231 cells showed | |
| | Mono-cultures of breast | elongated morphology in softer matrices | |
| | and prostate cancers | but round spheroids at higher stiffness; | |
| | | Cells in PEG-fibrinogen microspheres | |
| | | showed loss of apico-basal polarity, | |
| | | cellular and nuclear atypia, increased | |
| | | disorganization, elevated nuclear | |
| | | cytoplasmic ratio and nuclear volume | |
| | | density, and reduced length of cell-cell | |
| Cyctoine | Mono-culture of liver cancer | junctions | 113 |
| Cysteine responsive PEG | cells HEPG2 | Recovered HEPG2 spheroid secreted higher level of urea and albumin | 113 |
| hydrogel | Cells FIEFG2 | compared to the 2D culture of cells; Level | |
| nyuroger | | of secreted albumin was similar to the | |
| | | physiologic level in the body | |
| HA-PEGDA, | Layered co-culture of | Prostate cancer spheroid preserved PSA | 114 |
| HA-SH/PEGDA | uterine with ESS1 | and EGFR in the co-culture; Cells in | |
| | endometrial stromal | uterine cancer spheroid expressed | |
| | sarcoma cells or prostate | mucin1 and estrogen-induced gene 121 | |
| | cancer cell with HS27A | protein in the co-culture | |
| | bone marrow stroma cells | 5 1 20 12 1 1 1 1 1 1 | 0.1 |
| | PDX cells (MDA PCa 183 | Preserved epithelial phenotype of the | 94 |
| | and MDA PCa118b) | native tumors; Resistance to docetaxel compared to the spheroid that were | |
| | | generated from bone metastatic prostate | |
| | | cancer cell line (C42B) | |
| PEG-DEX ATPS | Mono-culture of breast | Spheroids showed normal growth over | 124 |
| | cancer cells MDA-MB-157 | time, secreted and deposited major ECM | |
| | | proteins such as collagen I, fibronectin, | |
| | | and laminin; showed proliferation | |
| | | gradients, size and density dependent | |
| | | hypoxia, expressed stem cell markers (CD24, CD133, NANOG) and displayed | |
| | | hypoxia mediated docorubicin resistance | |
| Polycaprolactone | Mono-culture of TC-71 | Spheroids preserved major marker such | 127 |
| (PCL) | Ewing sarcoma cells | as CD99 ⁺ , keratin ⁻ and smooth muscle | 121 |
| (- 0_) | | actin; Significantly upregulated phospho- | |
| | | IGF-1R | |
| PLGA | Mono-culture of ovarian | Expressed E-cadherin and proliferated in | 137 |
| | cancer cells HO1980 | the microsphere | |
| | Mono-culture of | High expression of angiogenic factors, | 138 |
| | glioblastoma cells U-251 | and resistance to doxorubicin; resistance | |
| | | to apoptosis (low caspase activity) by upregulating apoptosis-resistance | |
| | | proteins such as survivin and BCL-2 | |
| PLG | Mono-culture of oral | 3D PLG pre-cultured OSCC-3 cells | 141 |
| | squamous cell carcinoma | contained more blood vessels relative to | • • • |
| | OSCC-3 | the density of blood vessels in tumors | |
| | | formed by implanting 2D pre-cultured | |
| | | cells; tumors formed from 3D PLG pre- | |
| | | cultured spheroids expressed higher α_5 - | |
| Th' | Mana automa (CP) | integrin receptors | 4.45 |
| Thermoresponsive | Mono-culture of liver cancer | Enhanced albumin secretion and urea | 145 |

WILEY-VCH

| hydrogels | cells HepG2 | synthesis over a three-week culture period | | | | |
|--|---------------------|--|-----|--|--|--|
| Biomaterials for Tumor organoid cultures | | | | | | |
| PEG-Matrigel hydrogel | Mammary carcinoma | Enhanced the stiffness of the Matrigel for 50 50 to 4000 Pa; Functionalizing PEG with adhesive peptides promoted migratory capacity of mammary carcinoma | 184 | | | |
| Recombinant matrix | Intestine organoids | Precisely controlled biochemical and biomechanical cues for intestinal organoids | 185 | | | |