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Article type : Original Research

**Title:** Biodiversity improves the ecological design of sustainable biofuel systems

**Running Head:** Ecological design of algal biofuel systems

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**Abstract**

For algal biofuels to become a commercially viable and sustainable means of decreasing greenhouse gas emissions, growers are going to need to design feedstocks that achieve at least three characteristics simultaneously: attain high yields; produce high quality biomass; and remain stable through time. These three qualities have proven difficult to achieve simultaneously under the ideal conditions of the lab, much less under field conditions (e.g., outdoor culture ponds) where feedstocks are exposed to highly variable conditions and the crop is vulnerable to

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30 invasive species, disease, and grazers. Here we show that principles from ecology can be used to  
31 improve the design of feedstocks and to optimize their potential for ‘multifunctionality’. We  
32 performed a replicated experiment to test these predictions under outdoor conditions. Using 80  
33 ponds of 1,100 L each, we tested the hypotheses that polycultures would outperform  
34 monocultures in terms of the following functions: biomass production, yield of biocrude from  
35 biomass, temporal stability, resisting population crashes, and resisting invasions by unwanted  
36 species. Overall, species richness improved stability, biocrude yield, and resistance to invasion.  
37 While this suggests that polycultures could outperform monocultures on average, invasion  
38 resistance was the only function where polycultures outperformed the best single species in the  
39 experiment. Due to tradeoffs among different functions that we measured, no species or  
40 polyculture was able to maximize all functions simultaneously. However, diversity did enhance  
41 the potential for multifunctionality – the most diverse polyculture performed more functions at  
42 higher levels than could any of the monocultures. These results are a key finding for ecological  
43 design of sustainable biofuel systems because they show that while a monoculture may be the  
44 optimal choice for maximizing short-term biomass production, polycultures can offer a more  
45 stable crop of the desired species over longer periods of time.

46

## 47 **Introduction**

48 In both conventional agriculture and biofuel cultivation, researchers seek to identify  
49 species with superior potential for producing food or fuel. Although many species perform well  
50 under ideal conditions, when grown at larger scales those crops are often unable to attain high  
51 biomass yields, produce biomass that is favorable for fuel production, remain stable through time  
52 despite fluctuating conditions, resist population crashes caused by disease and pests, and resist  
53 invasion by nuisance species. Successful crops must meet all of these criteria simultaneously –  
54 they need to achieve multifunctionality. Algae are a promising source of renewable biofuels, but  
55 the challenge of achieving multifunctionality has limited the commercial-scale cultivation of  
56 algal feedstocks in open ponds (Department of Energy, 2010; National Research Council, 2012).  
57 Under conditions used for mass cultivation, algae have low productivity and lipid content  
58 relative to their potential (Sheehan, Dunahay, Benemann, & Roessler, 1998; Williams & Laurens,  
59 2010); exhibit low temporal stability and frequent crashes (Beyter et al., 2016); and are invaded  
60 by pathogens and unwanted species (McBride et al., 2014; Smith et al., 2015). Intensive

61 agricultural practices have not overcome the problems faced by algal feedstock cultivation  
62 (National Research Council, 2012) and would likely exacerbate environmental problems if  
63 implemented at large scales (Foley et al., 2005; Wiens, Fargione, & Hill, 2011). Given the  
64 failures of this approach, we need to develop alternative algal feedstocks that can achieve  
65 multifunctionality under the conditions used in large-scale cultivation.

66 A potential strategy for achieving multifunctionality would be to cultivate algae as multi-  
67 species polycultures rather than monocultures. Numerous experiments have shown that diversity  
68 increases the potential for multifunctionality by communities (Byrnes et al., 2014; Lefcheck et al.,  
69 2015). Biodiversity enhances multifunctionality when biological tradeoffs mean that no single  
70 species is capable of maximizing all of the different functions, but certain combinations of  
71 species are able to perform more functions at higher levels simultaneously than species can  
72 individually (Lefcheck et al., 2015). Although biodiversity can improve the performance of a  
73 single function compared to monocultures (Cardinale et al., 2011), multifunctionality does not  
74 require that polycultures outperform the best single species for any given function (i.e.,  
75 transgressive overyielding). Thus, the effect of biodiversity on multifunctionality is distinct from  
76 the positive effects of biodiversity on productivity (Hooper et al., 2005), temporal stability  
77 (Gross et al., 2014; Hautier et al., 2015), and resistance to invasive species and pathogens  
78 (Mitchell, Tilman, & Groth, 2002; Shea & Chesson, 2002). Based on this body of evidence,  
79 numerous papers in the last decade have proposed that multi-species polycultures of algae could  
80 be used to improve several aspects of multifunctionality in biofuel cultivation, including  
81 productivity (Shurin et al., 2013), biomass characteristics (Newby et al., 2016; Stockenreiter,  
82 Graber, Haupt, & Stibor, 2011), temporal stability (Beyter et al., 2016; Nalley, Stockenreiter, &  
83 Litchman, 2014), and resisting causes of population crashes (Smith & McBride, 2015; Smith et  
84 al., 2015).

85 To date, there have been few tests of the hypothesis that biodiversity can improve the  
86 cultivation of algal feedstocks, and nearly all have been constrained to laboratory-scale  
87 experiments. The few laboratory experiments that have tested this hypothesis have shown that,  
88 compared to the average monoculture, diverse cultures of algae may (Liu, 2016; Shurin et al.,  
89 2013; Stockenreiter et al., 2013; Stockenreiter, Haupt, Seppälä, Tamminen, & Spilling, 2016) or  
90 may not (Narwani, Lashaway, Hietala, Savage, & Cardinale, 2016) exhibit higher total cell  
91 biovolume or lipid content, but do exhibit more stable production through time (Narwani et al.,

92 2016). Although laboratory experiments suggest that biodiversity could improve  
93 multifunctionality in algal biofuel feedstock cultivation, it is unknown whether those findings are  
94 applicable to conditions in the field where conditions are often less favorable.

95 The performance exhibited by a mono- or polyculture under laboratory conditions does  
96 not necessarily translate to outdoor cultivation. In particular, large outdoor cultures of algae  
97 exhibit sudden catastrophic population ‘crashes’ due to environmental fluctuations, disease, pests,  
98 and invasive species. Several studies have demonstrated the feasibility of growing algal  
99 polycultures in open outdoor ponds (Beyter et al., 2016; Bhattacharjee & Siemann, 2015; Cho et  
100 al., 2017; Sturm, Peltier, Smith, & deNoyelles, 2012), but because these studies did not  
101 simultaneously evaluate the performance for each of those same species when grown as  
102 monocultures, it is not possible to isolate the effect of species richness (as opposed to species  
103 identity) on the performance of the cultures. As a result, the hypothesis that biodiversity  
104 improves several aspects of algal biofuel cultivation remains untested under outdoor conditions.

105 Here we present the results of an experiment designed to test a set of hypotheses about  
106 how algal polycultures impact feedstock cultivation under field conditions. Based on predictions  
107 from the literature and evidence from ecological experiments, we aimed to test eight hypotheses  
108 in our study (see Table 1 for a summary of the hypotheses). We hypothesized that compared to  
109 monocultures, polycultures would: (H1) increase biomass production; (H2) increase the  
110 proportion of biomass that can be converted to bio-crude oil; (H3) increase temporal stability;  
111 (H4) decrease the magnitude of crash events; (H5) delay crash events; (H6) decrease the  
112 abundance of invasive algae; and (H7) delay the impact of invasive algae. For each of these  
113 hypotheses we defined quantitative measures of performance (hereafter called ‘functions’) and  
114 asked whether polycultures outperformed monocultures on average, whether polycultures  
115 outperformed all of their component species, and whether any polycultures outperformed the  
116 single best species used in the experiment (transgressive overyielding). Additionally, because all  
117 of these functions are important for the overall performance of a crop, we asked whether  
118 polycultures could maintain more functions at higher levels of performance than could  
119 monocultures (multifunctionality, H8). We grew four species of green microalgae as mono- and  
120 polycultures in outdoor open ponds for 10 weeks. Although polycultures did not consistently  
121 improve biomass production or stability compared to the best single species, polycultures did  
122 delay invasions by unwanted algae longer than the best monocultures could. Moreover, select

123 polycultures performed more functions at higher levels of performance than even the best  
124 monocultures.

## 125 **Materials and methods**

126 Species selection. The species selected for this experiment were freshwater green  
127 microalgae that 1) were part of the Department of Energy's Aquatic Species Program, 2) are  
128 widespread throughout the United States (Environmental Protection Agency, 2012), and 3) are  
129 known to contribute to enhanced biomass production (Fritschie, Cardinale, Alexandrou, &  
130 Oakley, 2014), stability (Narwani et al., 2016) and feedstock quality (Hietala et al., 2017) in our  
131 own prior laboratory experiments. Based on these prior lab-based experiments, we ranked each  
132 species and polyculture in terms of its: mean biomass concentration, mean stability of biomass  
133 through time (mean divided by standard deviation), and the mean higher heating value (HHV) of  
134 biocrude produced from hydrothermal liquefaction (HTL) of biomass. We compared all possible  
135 sets of four species (see Experimental Design) based on their overall performance (Supporting  
136 Information). Based on this ranking, we selected four species for this experiment:

137 *Ankistrodesmus falcatus* (A), *Chlorella sorokiniana* (B), *Scenedesmus acuminatus* (D), and  
138 *Selenastrum capricornutum* (F). We employ the same species codes as our previous work for  
139 consistency (Godwin et al., 2017a; Godwin et al., 2017b; Hietala et al., 2017; Narwani et al.,  
140 2016).

141 Study site. The experiment was performed at the University of Michigan's Edwin S.  
142 George Reserve near Pinckney, MI, USA (42.47°N, 84.00°W). This reserve is situated among  
143 mixed land uses (forest, row crops, and pasture) and is predominantly covered by temperate  
144 forest and wetlands. The plot for the present experiment was located 43 m from a fenced area  
145 containing 9 large ponds (Fig. S3). The ponds are each 30 m in diameter and are separated by a  
146 border of mowed grass. The littoral vegetation is predominantly *Typha* and the ponds contained  
147 both macrophytes and phytoplankton. Prior to the experiment, we removed all vegetation from  
148 the plot and a surrounding buffer zone of 5 m. The ground was covered with permeable fabric  
149 and a layer of wood chips to stabilize the soil and prevent growth of vegetation during our study.

150 Experimental design. The design of the experiment included: each of the four species as  
151 monocultures, all pairs of species as 2-species polycultures, and the 4-species polyculture. As  
152 summarized in Fig. 1, each monoculture was replicated 7 times, each two-species polyculture  
153 was replicated 6 times, the four-species polyculture was replicated 8 times, and 8 units served as

154 controls without any inoculum of the focal species. To account for spatial effects (e.g. proximity  
155 to the ponds), we divided the plot into four spatial blocks and then used a partially balanced  
156 complete block design (Kuehl, 2000) to assign the treatments to the experimental units with  
157 every treatment replicated 1-2 times within each block. When randomizing the assignment of  
158 inoculation treatments to the experimental units, we included the constraint that adjacent ponds  
159 within blocks would not have the same treatment. The complete experimental layout is illustrated  
160 in Fig. S3.

161 Experimental ponds and setup. The ponds used for the experiment were circular cattle  
162 tanks made of black fiberglass-reinforced polyethylene (Fig. 1). Ponds were maintained at a  
163 depth of 50 cm, which corresponds to a volume of 1,100 L. Each pond was continuously mixed  
164 and aerated by four 30 cm air diffusers that delivered 35 L min<sup>-1</sup> of air to each pond. Prior to  
165 inoculation, ponds were cleaned with high-pressure water and rinsed with concentrated  
166 hydrochloric acid to remove any mineral deposits and biofilm organisms. On the day of  
167 inoculation, each pond was scrubbed with sodium hypochlorite solution (0.33% w/v), drained,  
168 rinsed again with sodium hypochlorite solution for five minutes, and then rinsed with treated  
169 water (see Water supply and growth medium). Immediately prior to filling, the ponds were  
170 saturated with 70% ethanol and allowed to dry.

171 Water supply and growth medium. We used Bold-3N medium (Bold, 1949) as the growth  
172 medium in ponds because it contains high concentrations of inorganic nutrients needed to  
173 support high population densities of algae (8.82 mmol L<sup>-1</sup> nitrate and 1.76 mmol L<sup>-1</sup>  
174 phosphate) and mimics the high-nutrient conditions used for commercial production. Water for  
175 the experiment was pumped from a groundwater well located at the Reserve. This water  
176 contained a high concentration of calcium hardness (>5,000 µeq L<sup>-1</sup>), which has potential to  
177 precipitate phosphate in the Bold-3N medium from solution. To avoid this problem, we removed  
178 the calcium from the groundwater using a zeolite ion-exchange resin, periodically recharged with  
179 sodium chloride. We monitored the effectiveness of this system by titration (Hach Company, kit  
180 HA-71A) and only used water with hardness below 200 µeq L<sup>-1</sup>. After the softening step, water  
181 was filtered through a 10 µm woven mesh filter and disinfected using a 200 watt flow-through  
182 UV lamp (Aquaneering). Treated water was dispensed through clean hoses that were treated  
183 daily with sodium hypochlorite to prevent contamination by algae or other organisms. Ponds  
184 were filled with treated water before adding the components of Bold-3N medium.

185 Macronutrients, in the form of inorganic salts, were added directly to the ponds. Micronutrients  
186 were added as a single concentrated solution that had been sterilized using a 0.2  $\mu\text{m}$  filter.

187 Inoculation process. Prior to inoculating the 1,100 L experimental ponds, we established  
188 12 L ‘inoculum cultures’ at the field site. These mono- and polycultures were grown in 20 L  
189 polyethylene buckets that had been sanitized as described for the ponds. The buckets were filled  
190 with 12 L of Bold-3N medium, covered with transparent polyethylene lids, and continually  
191 aerated with air delivered via a single air diffuser. We inoculated the 12 L inoculum cultures  
192 with laboratory-grown stocks of each algal species, using a substitutive design for polycultures in  
193 which the total biomass added to each 12 L inoculum culture was constant at 1,050 mg dry mass,  
194 regardless of species richness. In the polycultures, the biomass of each species was equal to  
195 1,050 mg divided by the species richness. The 12 L inoculum cultures were positioned adjacent  
196 to the ponds plot and were exposed to full sunlight for between 8 and 17 days. One 12 L  
197 inoculum culture was prepared for each experimental pond, including eight controls that received  
198 no algae. We sampled the 12 L inoculum cultures at the end of their incubation and detected no  
199 algae in the control units and only the appropriate species in the experimental treatments.

200 After sanitizing and filling each of the 1,100 L experimental ponds, we inoculated them  
201 with the entire contents of the corresponding 12 L inoculum culture. The eighty experimental  
202 ponds were established over a period of nine days. We began inoculating the ponds on 24 May,  
203 working in numeric order as shown in Fig. S3, and finished on 2 June. After inoculation, two  
204 ponds (pond 23 treatment F and pond 24 treatment DF) showed evidence of unwanted calcium  
205 phosphate precipitation, likely due to undetected hard water. Those two ponds were drained,  
206 cleaned, filled with medium, and re-inoculated on 17 June.

207 Sampling. The ponds were sampled via an opaque polyethylene sampling tube  
208 originating at the center of each pond and terminating outside the pond. This sampling tube was  
209 installed prior to filling the ponds and allowed for samples to be collected without a researcher  
210 having any contact with the pond, reducing the risk of any potential contamination. At the time  
211 of sampling, compressed air was injected into the bottom of each sampling tube, creating an air  
212 lift that delivered the contents of the pond into the sampling containers. Beginning on 7 June  
213 (week 1), we sampled the ponds every 7 days until 10 August (week 10). On each sampling date,  
214 we collected a total of 3.5 L from each pond. Following each sampling event, we added  
215 additional treated water to replace evaporative losses and maintain the total culture volume at

216 1,100 L. Due to low rainfall during the experiment the volume of the ponds never exceeded  
217 1,100 L. The 3.5 L sample taken on each date was used to measure the following variables.

218 Algal biomass and species composition. We measured the biomass of algae in each pond  
219 by filtering duplicate subsamples onto dried and pre-weighed glass fiber filters (Merck-Millipore  
220 AP40, 47 mm diameter) using low vacuum pressure (<200 mm Hg). Filter samples were rinsed  
221 and dried to constant mass at 60 °C. Filter blanks were included at the beginning and end of each  
222 sampling event. The mass change from the blanks was used to correct the mass change from the  
223 pond samples. The temporal stability of algal biomass was quantified as the inverse of the  
224 coefficient of variation (mean divided by the standard deviation) from weeks 2 through 10.  
225 Although this measure of stability represents the overall temporal variability of the culture, it  
226 does not necessarily reflect rapid changes in biomass that are characteristic of population crashes.  
227 Therefore, we defined biomass crashes as a proportional loss of biomass during a one-week  
228 period and computed the maximum crash magnitude measured in each pond. This approach  
229 allows for quantitative comparisons in terms of both the magnitude of crashes observed over a  
230 time period and the length of time prior to a crash event.

231 We preserved samples for algal identification and abundance by adding phosphate-  
232 buffered formaldehyde to a concentration of 1%. For each sampling date, we enumerated algae  
233 in each sample by microscopy using a hemacytometer and quantified the proportion of algal cells  
234 that were not the treatment species for that pond. Over the course of the experiment, a total of  
235 nine invader species of algae were observed in at least one sample.

236 Hydrothermal liquefaction. We used hydrothermal liquefaction (HTL) to convert algal  
237 biomass into biocrude, which is a precursor of renewable transportation fuels (Savage, 2012).  
238 Unlike direct lipid extraction, HTL does not require high lipid content in the algae and instead,  
239 converts whole wet biomass to biocrude (Valdez, Nelson, Wang, Lin, & Savage, 2012). We  
240 performed HTL on algal biomass samples from weeks 1 through 8. To concentrate biomass for  
241 HTL, we settled a 2.5 L sample in the dark for 7 days. The samples were further concentrated by  
242 decanting and centrifugation (Hietala et al., 2017; Narwani et al., 2016). The concentrated  
243 biomass was dried at 60 °C until mass was constant. The full procedure for HTL follows that of  
244 our previous work (Hietala et al., 2017). In short, the dried biomass samples were mixed with  
245 deionized water to 5% solids content and subjected to HTL at 350°C for 20 minutes. Biocrude  
246 was separated from the other products using dichloromethane and then dried under nitrogen to



247 evaporate residual solvent. For each reaction, we calculated biocrude yield as the mass of  
248 biocrude product divided by the mass of algae used for HTL.

249 Data analysis – linear models. We used general linear mixed models to analyze the  
250 effects of initial algal species richness and composition on: (H1) mean biomass concentration  
251 ( $\text{mg L}^{-1}$ ); (H2) biocrude yield (g biocrude per g dry algae); (H3) temporal stability of biomass as  
252  $\text{CV}^{-1}$  through time; (H4) maximum proportion of invader algae observed in each pond, and (H6)  
253 maximum crash magnitude observed in each pond (% reduction in 7 days) For each parameter,  
254 the full model consisted of the fixed effects of species richness (SR), species composition  
255 (Combo, nested in SR), and week. Spatial block and pond identity were initially included as  
256 random effects and retained when they significantly improved the Akaike information criterion  
257 (AIC). We then removed non-significant terms stepwise until reaching the minimal adequate  
258 model that contained effects of species richness, species composition, and time. Temporal  
259 stability, maximum crash magnitude, and maximum proportion of invading algae are all  
260 measures with only one value for each pond; thus, the effects of time and pond were not included  
261 in the statistical models. Control treatment ponds were used to measure the progress of algal  
262 invasions in the absence of any inoculum treatment, but were excluded from all statistical  
263 analyses so that the effects of species richness and species composition were not affected by this  
264 treatment. All linear model analyses were performed in R using the package lme4 (R Core Team  
265 2015). When we found significant effects of species richness or species composition, we  
266 performed post-hoc tests using Tukey's honest significant difference method in the R package  
267 lmerTest. We used the post-hoc tests to compare the performance among levels of species  
268 richness and between each polyculture and the monocultures (Table 1).

269 Data analysis – logistic models for crash and invasion timing. The proportion of ponds in  
270 each treatment that exceeded the median crash magnitude (55%) on or before each date was  
271 modeled using logistic regression. We also used logistic regression to analyze the proportion of  
272 ponds in each treatment with at least 1% proportional representation of invading algae on or  
273 before each sampling date. The logistic regressions included categorical fixed effects of species  
274 richness and species composition and a continuous fixed effect of time. Logistic regressions were  
275 performed in R using the function "glm". Post-hoc comparisons for logistic regressions were  
276 performed as for the linear models.

277 Data analysis – multifunctionality. Because no single species or polyculture is likely to  
278 optimize all aspects of system performance (Godwin et al., 2017b; Hietala et al., 2017; Shurin et  
279 al., 2013), we sought to characterize tradeoffs among species and determine if polycultures can  
280 mitigate these tradeoffs if they perform more functions at higher levels of performance than  
281 monocultures do (multifunctionality, Byrnes et al., 2014; Lefcheck et al., 2015). We quantified  
282 the capacity for monocultures and polycultures to exhibit multifunctionality using a threshold  
283 approach similar to the one developed in the field of biodiversity-function (Byrnes et al., 2014;  
284 Lefcheck et al., 2015). The threshold approach compares different species compositions based on  
285 how many functions they perform at or above an arbitrary level of performance (i.e. thresholds).  
286 We used seven separate functions to describe the overall performance of each inoculation  
287 treatment: mean biomass; mean biocrude yield; mean temporal stability of biomass; mean  
288 maximum crash magnitude in 10 weeks; crash timing (based on logistic regression coefficients  
289 for  $\geq 55\%$  crash magnitude); mean maximum proportion of invaders in the ponds; and invasion  
290 timing (based on logistic regression coefficients for  $\geq 1\%$  invader algae). To allow for  
291 comparisons among various functions, we standardized performance for each function as the  
292 rank of each treatment relative to the other inoculation treatments (control ponds were excluded).  
293 Ranks were assigned such that the poorest performer was rank 1/11 and the best performer was  
294 rank 11/11 = 1. Because the number of species compositions was the same for each function, we  
295 set performance thresholds between 0 and 1 in increments of 1/11. We then tallied the number of  
296 functions that each monoculture or polyculture performed above each threshold.

297 The threshold approach has some known drawbacks that we sought to avoid. A recent  
298 paper showed that when the number of functions performed above a threshold is used as a  
299 dependent variable for regression, there can be artifacts that arise due to chance rather than  
300 biological effects (Gamfeldt & Roger, 2017). Thus, a ‘null’ model is required to detect biological  
301 effects against the background chance of an artifact. To generate a null model, we used  
302 randomization tests to assess the significance of differences in multifunctionality between mono-  
303 and polycultures at each threshold. For each performance threshold, we compared the number of  
304 functions performed by: the mean two-species polyculture and the mean monoculture, the four-  
305 species polyculture and the mean monoculture, the best two-species polyculture and the best  
306 monoculture, and the four-species polyculture and the best monoculture (Fig. S4). We then  
307 compared the observed differences to a null model based on randomized performance ranks. For

308 each comparison, we used the null model to generate a distribution of differences based on  
309 randomized performance ranks (n=10,000 iterations). This randomization method takes into  
310 account that there were different numbers of species compositions for monocultures, two-species  
311 polycultures, and the four-species polyculture.

## 312 **Results**

313 The original hypotheses for the experiment are summarized in Table 1. This table also  
314 provides a summary of findings from our experiment, and can serve as a reference guide for  
315 readers as we summarize all of the results.

316 Fig. 2a shows that in contrast to hypothesis H1, polycultures did not yield more biomass  
317 than monocultures (Table 2). The monoculture of *Selenastrum* (F) achieved the highest mean  
318 biomass concentration throughout the experiment (224 mg L<sup>-1</sup>), outperforming all of the other  
319 monocultures and polycultures. None of the polycultures significantly outperformed the mean of  
320 their component species, their best component species, or the best overall species (all p>0.05).

321 H2 was supported by a significant effect of species richness on biocrude yields (Fig. 2b, Table 2).  
322 Biocrude yield – measured by convention as the wt% of biomass – was significantly higher in  
323 the 2- (mean 30.4%) and 4-species polycultures (32.2%) than the monocultures (27.3%).

324 *Chlorella* (B) exhibited the highest biocrude yields among the monocultures, and none of the  
325 polycultures exhibited significantly higher biocrude yields than the best species. Consistent with  
326 H3, there was a significant positive effect of species richness on the temporal stability of biomass  
327 (Fig. 2c, Table 2). The effect of species richness on stability was due to the increased stability of  
328 the four-species polyculture relative to the monocultures. However, none of the polycultures  
329 exhibited significantly higher stability than the most stable monoculture (*Chlorella*, B).

330 Fig. 2d shows that contrary to H4, there was no significant effect of species richness on  
331 the magnitude of biomass crashes (Table 2). The monoculture of *Selenastrum* (F) had the  
332 smallest crash magnitudes and was nearly matched by polycultures AF and ABDF. The median  
333 maximum crash magnitude was a 55% reduction in biomass in one week. The 33<sup>rd</sup> and 67<sup>th</sup>  
334 percentiles occurred at magnitudes of 45% and 60%, respectively. Contrary to H5, species  
335 richness did not significantly delay large crash events compared to either the mean of the  
336 monocultures or to the best single species (Fig. 3a). Despite finding a significant effect of species  
337 richness on crash timing (Table 2), the 2-species polycultures tended to experience crashes  
338 earlier in the experiment than the monocultures, and the ability of the 4-species polycultures to

339 delay crashes was marginally significant (post-hoc  $p=0.063$ ). Among the monocultures,  
340 *Selenastrum* (F) was most resistant to crashes, but did exhibit one large crash beginning on week  
341 5.

342 Consistent with H6, we found that the maximum proportion of invading algae decreased  
343 with species richness (Fig. 2e, Table 2). The 4-species polyculture had significantly less invading  
344 algae than the monocultures and 2-species polycultures ( $p<1\times 10^{-4}$ ), but did not significantly  
345 outperform the best species (*Selenastrum*, F). Consistent with H7, we found that species richness  
346 significantly delayed invasion by unwanted species of algae (Fig. 3b, Table 2). The 2-species  
347 polycultures outperformed the monocultures at delaying invasion and the 4-species polyculture  
348 outperformed both the 2-species polycultures and the monocultures (all  $p<0.001$ ). The 4-species  
349 polyculture significantly outperformed the best single species (*Chlorella*, B) at delaying invasion  
350 ( $p<0.02$ ). Ponds inoculated with the 4-species polyculture remained below 1% invader algae  
351 until the last sampling, when two of the eight replicates were invaded. All of the species  
352 inoculation treatments offered some invasion resistance compared to the control ponds, which  
353 were rapidly colonized over the first week of the experiment.

354 Consistent with H8, we found that certain polycultures maintained more functions at  
355 higher levels of performance than any of their component species did as monocultures. Fig. 4a  
356 shows that no mono- or polyculture maximized all of the functions measured in the study.  
357 Among the monocultures, *Selenastrum* (F) had superior performance in terms of biomass  
358 production and reducing crash magnitudes, but its performance ranks for biocrude yield and  
359 invasion timing were each below the median level of performance. In contrast, the two-species  
360 polycultures AF, BD, and BF offered relatively high performance ranks for all of the functions.  
361 The four-species polyculture had high performance ranks for all functions except biomass  
362 production. Fig. 4b shows that for both the monocultures and the two-species polycultures, the  
363 mean number of functions performed above the threshold decreases steadily with increasing  
364 thresholds. The four-species polyculture maintained more functions at higher levels of  
365 performance than the monocultures did. When the performance threshold was between 40 and  
366 80% of the maximum, the four-species polyculture performed significantly more functions above  
367 the threshold than the mean monoculture or mean two-species polyculture. Moreover, even when  
368 compared to the best monoculture at each threshold, the four-species polycultures showed  
369 superior multifunctionality (Fig. 4c). Notably, the best of the monocultures performed only four

370 functions above the 60<sup>th</sup> percentile threshold, but the four-species polyculture performed six out  
371 of seven functions above the 70<sup>th</sup> percentile threshold.

372

### 373 **Discussion**

374 Laboratory experiments have suggested that algal diversity could improve several aspects  
375 of biofuel feedstock cultivation, but this prediction has not been rigorously tested under field  
376 conditions. We experimentally tested eight hypotheses for how polycultures influence  
377 performance of outdoor biofuel feedstock cultivation (Table 1). In many cases our findings were  
378 contrary to *a priori* predictions that were based on previously published literature, and  
379 polycultures did not consistently outperform the best single species. The two well-supported  
380 hypotheses were that polycultures would decrease invasion by unwanted algae, and that  
381 polycultures would improve overall multifunctionality of the feedstocks. In the following  
382 sections we assess each of these hypotheses and discuss what our findings mean for the prospect  
383 of large-scale biofuel feedstock cultivation.

384 Biomass production and biocrude yield. Our experiment contradicts the prediction that  
385 algal diversity increases the production of biomass (H1). In our experiment most of the  
386 polycultures actually produced less biomass than the average monoculture (Fig. 2a) – results that  
387 are consistent with the recent laboratory experiment that used the same species pool (Narwani et  
388 al., 2016). The poor performance of polycultures in the laboratory experiment was attributable to  
389 competition among species of algae. A similar explanation is likely for the present experiment;  
390 nutrient concentrations remained high, but *Chlorella* and *Selenastrum* dramatically attenuated  
391 light, which suggests that light was a limiting resource (Fig. S5). In both the present experiment  
392 and the laboratory mesocosms, the 2-species polycultures AF and BF produced more biomass  
393 than the most diverse polyculture, but polycultures collectively underperformed relative to the  
394 best species. This finding is a contrast to the predominantly positive effects of species richness  
395 observed in biodiversity-function experiments (Cardinale et al., 2011; Hooper et al., 2005) and in  
396 several experiments performed in the context of algal biofuels (Shurin et al., 2013; Stockenreiter  
397 et al., 2011; Stockenreiter et al., 2016). This contradiction could be due the limited taxonomic  
398 diversity and functional variation used in our experiment: previous studies that reported positive  
399 effects of species richness on the production of biomass or biovolume included algae from a  
400 greater variety of taxonomic groups (e.g. diatoms, cyanobacteria, and chrysophytes) (Shurin et

401 al., 2013; Stockenreiter et al., 2016). Further experiments will be needed to determine whether  
402 our findings are specific to the species pool that we used or are representative of the culture  
403 conditions used in our study.

404 While we did not find an effect of diversity on total biomass, our experiment did support  
405 the hypothesis that species richness increases the yield of biocrude per mass of algal feedstock  
406 (H2, Fig. 2b). A similar effect has been observed in previous studies that found increased  
407 biocrude yield (Hietala et al., 2017) in polycultures compared to monocultures as well as  
408 increased lipid content (Stockenreiter et al., 2011; Stockenreiter et al., 2013), which would lead  
409 to increased biocrude yield. The positive effect of biodiversity on biocrude yield of algal  
410 feedstocks could potentially enhance overall production of biofuel from a culture, but it remains  
411 unclear whether those effects would offset or overcome a potential decrease in the total biomass  
412 production by polycultures compared to monocultures.

413 Stability and Crashes. Consistent with H3, biodiversity increased stability of biomass  
414 through time compared to the average monoculture in our experiment; however, polycultures did  
415 not outperform the best monoculture. Although the effect of biodiversity on stability is well  
416 documented in natural ecosystems and in experiments (Gonzalez & Loreau, 2009; Gross et al.,  
417 2014), it has only recently become a focus in the application of biodiversity to biofuels (Beyter et  
418 al., 2016; Narwani et al., 2016). Narwani and others (2016) found a similar positive effect of  
419 diversity on stability when they subjected laboratory cultures to weekly fluctuations in water  
420 temperature. The magnitude of weekly temperature change in that experiment was smaller than  
421 the daily temperature oscillations observed in the outdoor ponds of this experiment (Fig. S6).  
422 However, both experiments found that species richness tended to decrease biomass but increase  
423 the temporal stability of biomass. For instance, polyculture AF exhibited 19% less biomass and  
424 33% higher stability than the most productive species (*Selenastrum*, F), but 53% higher biomass  
425 and 5% higher stability than the most stable species (*Chlorella*, B). This suggests that certain  
426 polycultures might offer a compromise between production and stability.

427 Our findings did not support the hypotheses that diversity would minimize and delay  
428 crash events relative to the monocultures (H4 & H5). Polycultures did not significantly  
429 outperform the best species (*Selenastrum*, F) in terms of minimizing or delaying crashes. Crash  
430 events are particularly important for large-scale outdoor cultivation since the culture typically  
431 needs to be re-established, which is a major expense in terms of resources and lost productivity

432 (National Research Council, 2012). Resistance to crash events will be a key metric for  
433 identifying mono- and polycultures that are suitable for outdoor cultivation. A previous study  
434 found that, compared to the mean monoculture, polycultures were less likely to exhibit low  
435 biomass yields over time (Narwani et al., 2016). Yet, we are unaware of any other experiments  
436 that have attempted to quantify the impact of species richness on sudden crash events in biofuel  
437 feedstock cultures. Because crash events are less likely to occur under laboratory conditions,  
438 these findings underscore the importance of testing the hypothesized benefits of biodiversity  
439 under conditions that mimic commercial production.

440 Lifecycle assessment (LCA) is a tool that can evaluate the impact of feedstock cultivation  
441 on the overall energy balance and resource requirements of a hypothetical algal bio-refinery. Our  
442 results suggest that describing the growth of algae in open ponds will require a more realistic  
443 approach than is typically used for LCAs. In particular, most LCAs are performed under ‘steady-  
444 state’ assumptions where the culture is in continual production for a large fraction of the year.  
445 Recently, some LCAs have adopted models of productivity that incorporate effects of seasonality  
446 and geography (Davis et al., 2012). While this represents an improvement over models that  
447 assume invariant productivity, our experiment shows the importance of sudden crash events and  
448 invasions for outdoor cultivation. In addition to the loss of production output, these catastrophic  
449 events will often require a cultivation pond to be drained and restarted, which increases demands  
450 on energy and resources. Our experiment shows that the risk of culture crash is not uniform  
451 through time and that this risk differs substantially among types of feedstocks. Since both small  
452 fluctuations and large crashes are inevitable consequences of cultivating algae outdoors, these  
453 aspects of temporal instability need to be explicitly represented in models describing algal  
454 cultivation.

455 Invasion By Algae. Our experiment supports the hypotheses that increased species  
456 richness can decrease and delay invasion by unwanted species of algae (H6 & H7). As  
457 monocultures, *Chlorella* (B) and *Selenastrum* (F) delayed invasions and decreased invader  
458 prevalence relative to the other monocultures, but both of these species were susceptible to  
459 invasions. However, most of the ponds inoculated with both *Chlorella* and *Selenastrum* remained  
460 free of invader algae for 10 weeks (BF and ABDF). The superior resistance to invasion by  
461 certain polycultures is a key finding because it suggests that biodiversity could help overcome a  
462 major challenge for cultivation at large scales.

463 Biodiversity can increase resistance to invasion when the resident species utilize the  
464 available resources to the extent that invader species are unable to establish and grow (Shea &  
465 Chesson, 2002). Inorganic nutrients (N and P) remained at high concentrations throughout our  
466 experiment, but both *Chlorella* and *Selenastrum* attenuated available light when they were  
467 dominant (Fig. S5). This suggests that competition for light could limit the success of invaders.  
468 Light availability is a function of the concentration of biomass in algal cultures due to self-  
469 shading (Kenny & Flynn, 2015), which means that dense cultures should be invaded more slowly  
470 than cultures with low biomass or have recently undergone a crash. This is an important finding  
471 because it underscores the importance of delaying and decreasing crashes for preventing invasion  
472 by unwanted algae.

473 Multifunctionality. Our experiment supports the hypothesis that polycultures can  
474 maintain more functions at higher levels of performance than monocultures (H8). Although none  
475 of the monocultures performed more than four of the seven functions above the 60<sup>th</sup> percentile,  
476 the 4-species polyculture maintained six functions above the 70<sup>th</sup> percentile. However, these  
477 benefits for multifunctionality occur because of strong tradeoffs among species and polycultures.  
478 For example, the four-species polyculture exhibited high performance ranks for most functions,  
479 but this benefit was offset by poor performance in biomass production. It appears that such  
480 tradeoffs are common when designing polycultures for biofuel feedstock cultivation (Godwin et  
481 al., 2017b; Hietala et al., 2017; Narwani et al., 2016; Shurin et al., 2013), so identifying  
482 polycultures that can come closer to optimizing multiple functions should be a priority for future  
483 research.

484 Superior multifunctionality by polycultures is an important finding because the best  
485 monocultures – *Chlorella* and *Selenastrum* – each had poor performance for at least one function.  
486 For example, *Chlorella* tended to experience crashes earlier in the experiment and was more  
487 likely to be invaded than other species combinations, and *Selenastrum* had lower biocrude yields  
488 and was invaded earlier compared to other species combinations. Thus, picking a monoculture  
489 means that at least one function will be below the median performance rank. In contrast, picking  
490 the polyculture AF, BD, or BF would result in all seven functions being performed at or above  
491 the median rank. A key consequence of these performance tradeoffs is that choosing a feedstock  
492 based on any single function (e.g. biomass production) will likely result in undesirable



493 performance in terms of other functions, but polycultures are more likely to perform all functions  
494 at a high level.

495 The benefits of multifunctionality become more even important when we consider other  
496 aspects of the biofuel production lifecycle that were not examined in our present study.  
497 Specifically, other work has shown that biodiversity can improve nutrient-use efficiency  
498 (Godwin et al., 2017b; Shurin, Mandal, & Abbott, 2014), nutrient recycling (Godwin et al.,  
499 2017a), lipid content (Stockenreiter et al., 2016), and biocrude characteristics (Hietala et al.,  
500 2017) in algal biofuel systems. Each of these aspects of multifunctionality is essential for algae-  
501 based biofuels to be both economically feasible and sustainable. The practical importance of  
502 multifunctionality is reflected in the numerous lifecycle assessments that have quantified how  
503 each of these functions impacts the balance of energy and greenhouse gases over the lifecycle  
504 (Frank, Elgowainy, Han, & Wang, 2013; Orfield et al., 2014; Quinn & Davis, 2015). The  
505 potential for polycultures to improve multifunctionality suggests that polycultures could be  
506 designed to improve performance across the biofuel lifecycle and overcome biological tradeoffs  
507 exhibited by monocultures.

508 Large-scale cultivation will require feedstocks that are not only stable under outdoor  
509 cultivation, but can also be harvested at a high rate through time. The rate of productivity (mass  
510 per area per time) has a large influence on the feasibility of future algal biofuel systems (Quinn  
511 & Davis, 2015). Although we did not harvest the algae continuously or periodically as would be  
512 required to accurately estimate productivity (Kenny & Flynn, 2015), we did quantify the  
513 maximum growth rates of the species compositions during the first two weeks of the experiment  
514 (Supporting Information). Fig. S7 shows that the species compositions that exhibited highest  
515 mean biomass also exhibited highest growth rates. Specifically, compositions B, F, and BF grew  
516 more quickly ( $0.52 - 0.54 \text{ d}^{-1}$ ) than did compositions A and AD ( $0.30 - 0.34 \text{ d}^{-1}$ ). These estimates  
517 of maximum growth rates suggest that *Selenastrum*, *Chlorella*, and their bi-culture (BF) could  
518 achieve high productivity under outdoor conditions. Further experiments will be required to  
519 determine which species compositions and harvesting regimes lead to the highest and most stable  
520 productivity under realistic conditions.

521 Despite the growing body of literature highlighting the potential for biodiversity to  
522 improve algal biofuel production, our study is one of a small number that have experimentally  
523 tested these predictions in field conditions. Although the studies performed to date were by no

524 means exhaustive, the collective evidence from these experiments suggests that the effects of  
525 biodiversity on biomass production are likely smaller than what has been forecast. At the same  
526 time, there is a growing body of evidence suggesting that biodiversity has other benefits besides  
527 biomass production, including temporal stability, resistance to unwanted pest species, more  
528 efficient use of nutrients, and greater levels of multifunctionality. The practical importance of  
529 biodiversity and multifunctionality will depend upon how these functions impact the long-term  
530 balance of energy and cost in commercial-scale cultivation. Determining the net impact of  
531 biodiversity will require (1) additional experiments that directly test the hypothesized benefits of  
532 biodiversity under relevant conditions and (2) more realistic LCAs that use empirical data from  
533 these experiments to evaluate the performance of different feedstocks in terms of energy return  
534 on energy invested and other metrics of environmental sustainability.

535

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541

### 542 **Conflict of Interest**

543 There are no conflicts of interest.

544

### 545 **References**

- 546 Beyter, D., Tang, P.-Z., Becker, S., Hoang, T., Bilgin, D., Lim, Y. W., Peterson, T. C., Mayfield,  
547 S., Haerizadeh, F., Shurin, J. B., Bafna, V., McBride, R., & Löffler, F. E. (2016).  
548 Diversity, productivity, and stability of an industrial microbial ecosystem. *Applied and*  
549 *Environmental Microbiology*, 82(8), 2494-2505.
- 550 Bhattacharjee, M., & Siemann, E. (2015). Low algal diversity systems are a promising method  
551 for biodiesel production in wastewater fed open reactors. *Algae*, 30(1), 67-79.
- 552 Bold, H. C. (1949). The morphology of *Chlamydomonas chlamydogama* sp. Nov. *Bulletin of the*  
553 *Torrey Botanical Club*, 76, 101-108.

554 Byrnes, J. E. K., Gamfeldt, L., Isbell, F., Lefcheck, J. S., Griffin, J. N., Hector, A., Cardinale, B.  
555 J., Hooper, D. U., Dee, L. E., & Duffy, E. J. (2014). Investigating the relationship  
556 between biodiversity and ecosystem multifunctionality: Challenges and solutions.  
557 *Methods in Ecology and Evolution*, 5(2), 111-124.

558 Cardinale, B. J., Matulich, K. L., Hooper, D. U., Byrnes, J. E., Duffy, E., Gamfeldt, L.,  
559 Balvanera, P., O'Connor, M. I., & Gonzalez, A. (2011). The functional role of producer  
560 diversity in ecosystems. *American Journal of Botany*, 98(3), 572-592.

561 Cho, D. H., Choi, J. W., Kang, Z., Kim, B. H., Oh, H. M., Kim, H. S., & Ramanan, R. (2017).  
562 Microalgal diversity fosters stable biomass productivity in open ponds treating  
563 wastewater. *Scientific Reports*, 7(1), 1979.

564 Davis, R., Fishman, D., Frank, E. D., Wigmosta, M. S., Aden, A., Coleman, A. M., Pienkos, P.  
565 T., Skaggs, R. J., Venteris, E. R., & Wang, M. Q. (2012). *Renewable diesel from algal*  
566 *lipids: An integrated baseline for cost, emissions, and resource potential from a*  
567 *harmonized model*, National Renewable Energy Laboratory (NREL), Golden, CO.

568 Department of Energy, United States. (2010). *National algal biofuels technology roadmap*,  
569 Office of Energy Efficiency and Renewable Energy Biomass Program. Washington, D.C.

570 Environmental Protection Agency, United States,. (2012). 2007 National Lakes Assessment.  
571 Retrieved from [https://www.epa.gov/national-aquatic-resource-surveys/national-lakes-](https://www.epa.gov/national-aquatic-resource-surveys/national-lakes-assessment-2007-results)  
572 [assessment-2007-results](https://www.epa.gov/national-aquatic-resource-surveys/national-lakes-assessment-2007-results)

573 Foley, J. A., DeFries, R., Asner, G. P., Barford, C., Bonan, G., Carpenter, S. R., Chapin, F. S.,  
574 Coe, M. T., Daily, G. C., Gibbs, H. K., Helkowski, J. H., Holloway, T., Howard, E. A.,  
575 Kucharik, C. J., Monfreda, C., Patz, J. A., Prentice, I. C., Ramankutty, N., & Snyder, P. K.  
576 (2005). Global consequences of land use. *Science*, 309(5734), 570-574.

577 Frank, E. D., Elgowainy, A., Han, J., & Wang, Z. (2013). Life cycle comparison of hydrothermal  
578 liquefaction and lipid extraction pathways to renewable diesel from algae. *Mitigation and*  
579 *Adaptation Strategies for Global Change*, 18(1), 137-158.

580 Fritschie, K. J., Cardinale, B. J., Alexandrou, M. A., & Oakley, T. H. (2014). Evolutionary  
581 history and the strength of species interactions: Testing the phylogenetic limiting  
582 similarity hypothesis. *Ecology*, 95(5), 1407-1417.

583 Gamfeldt, L., & Roger, F. (2017). Revisiting the biodiversity-ecosystem multifunctionality  
584 relationship. *Nature Ecology & Evolution*, 1(7), 168.

585 Godwin, C. M., Hietala, D., Lashaway, A., Narwani, A., Savage, P. E., & Cardinale, B. J.  
586 (2017a). Algal polycultures enhance coproduct recycling from hydrothermal liquefaction.  
587 *Bioresource Technology*, 224, 630-638.

588 Godwin, C. M., Hietala, D., Lashaway, A., Narwani, A., Savage, P. E., & Cardinale, B. J.  
589 (2017b). Ecological stoichiometry meets ecological engineering: Using polycultures to  
590 enhance the multifunctionality of algal biocrude systems. *Environmental Science &*  
591 *Technology*, 51(19), 11450-11458.

592 Gonzalez, A., & Loreau, M. (2009). The causes and consequences of compensatory dynamics in  
593 ecological communities. *Annual Review of Ecology, Evolution, and Systematics*, 40(1),  
594 393-414.

595 Gross, K., Cardinale, B. J., Fox, J. W., Gonzalez, A., Loreau, M., Polley, H. W., Reich, P. B., &  
596 van Ruijven, J. (2014). Species richness and the temporal stability of biomass production:  
597 A new analysis of recent biodiversity experiments. *The American Naturalist*, 183(1), 1-12.

598 Hautier, Y., Tilman, D., Isbell, F., Seabloom, E. W., Borer, E. T., & Reich, P. B. (2015).  
599 Anthropogenic environmental changes affect ecosystem stability via biodiversity. *Science*,  
600 348(6232), 336-340.

601 Hietala, D. C., Koss, C. K., Narwani, A., Lashaway, A. R., Godwin, C. M., Cardinale, B. J., &  
602 Savage, P. E. (2017). Influence of biodiversity, biochemical composition, and species  
603 identity on the quality of biomass and biocrude oil produced via  
604 hydrothermal liquefaction. *Algal Research*, 26, 203-214.

605 Hooper, D. U., Chapin F., Ewel, J., Hector, A., Inchausti, P., Lavorel, S., Lawton, J., Lodge, D.,  
606 Loreau, M., & Naeem, S. (2005). Effects of biodiversity on ecosystem functioning: A  
607 consensus of current knowledge. *Ecological Monographs*, 75(1), 3-35.

608 Kenny, P., & Flynn, K. J. (2015). In silico optimization for production of biomass and biofuel  
609 feedstocks from microalgae. *Journal of Applied Phycology*, 27, 33-48.

610 Kuehl, R. O. (2000). *Design of experiments: Statistical principles of research design and*  
611 *analysis* (Second ed.). Pacific Grove, CA: Duxbury/Thomson Learning.

612 Lefcheck, J. S., Byrnes, J. E., Isbell, F., Gamfeldt, L., Griffin, J. N., Eisenhauer, N., Hensel, M.  
613 J., Hector, A., Cardinale, B. J., & Duffy, J. E. (2015). Biodiversity enhances ecosystem  
614 multifunctionality across trophic levels and habitats. *Nature Communications*, 6.

615 Liu, J. (2016). Interspecific biodiversity enhances biomass and lipid productivity of microalgae  
616 as biofuel feedstock. *Journal of Applied Phycology*, 28(1), 25-33.

617 McBride, R. C., Lopez, S., Meenach, C., Burnett, M., Lee, P. A., Nohilly, F., & Behnke, C.  
618 (2014). Contamination management in low cost open algae ponds for biofuels production.  
619 *Industrial Biotechnology*, 10(3), 221-227.

620 Mitchell, C. E., Tilman, D., & Groth, J. V. (2002). Effects of grassland plant species diversity,  
621 abundance, and composition on foliar fungal disease. *Ecology*, 83(6), 1713-1726.

622 Nalley, J. O., Stockenreiter, M., & Litchman, E. (2014). Community ecology of algal biofuels:  
623 Complementarity and trait-based approaches. *Industrial Biotechnology*, 10(3), 191-201.

624 Narwani, A., Lashaway, A. R., Hietala, D. C., Savage, P. E., & Cardinale, B. J. (2016). Power of  
625 plankton: Effects of algal biodiversity on biocrude production and stability.  
626 *Environmental Science & Technology*, 50(23), 13142-13150.

627 National Research Council, Committee on the Sustainable Development of Algal Biofuels.,  
628 (2012). *Sustainable development of algal biofuels in the United States*: The National  
629 Academies Press.

630 Newby, D. T., Mathews, T. J., Pate, R. C., Huesemann, M. H., Lane, T. W., Wahlen, B. D.,  
631 Mandal, S., Engler, R. K., Feris, K. P., & Shurin, J. B. (2016). Assessing the potential of  
632 polyculture to accelerate algal biofuel production. *Algal Research*, 19, 264-277.

633 Orfield, N. D., Fang, A. J., Valdez, P. J., Nelson, M. C., Savage, P. E., Lin, X. N., & Keoleian, G.  
634 A. (2014). Life cycle design of an algal biorefinery featuring hydrothermal liquefaction:  
635 Effect of reaction conditions and an alternative pathway including microbial regrowth.  
636 *ACS Sustainable Chemistry & Engineering*, 2(4), 867-874.

637 Quinn, J. C., & Davis, R. (2015). The potentials and challenges of algae based biofuels: A  
638 review of the techno-economic, life cycle, and resource assessment modeling.  
639 *Bioresour Technol*, 184, 444-452.

640 Savage, P. E. (2012). Algae under pressure and in hot water. *Science*, 338(6110), 1039-1040.

641 Shea, K., & Chesson, P. (2002). Community ecology theory as a framework for biological  
642 invasions. *Trends in Ecology & Evolution*, 17(4), 170-176.

643 Sheehan, J., Dunahay, T., Benemann, J. R., & Roessler, P. (1998). *A look back at the U.S.*  
644 *Department of Energy's Aquatic Species Program: Biodiesel from algae*. Golden,  
645 Colorado: National Renewable Energy Laboratory, United States Department of Energy.

646 Shurin, J. B., Abbott, R. L., Deal, M. S., Kwan, G. T., Litchman, E., McBride, R. C., Mandal, S.,  
647 & Smith, V. H. (2013). Industrial - strength ecology: Trade - offs and opportunities in  
648 algal biofuel production. *Ecology Letters*, *16*(11), 1393-1404.

649 Shurin, J. B., Mandal, S., & Abbott, R. L. (2014). Trait diversity enhances yield in algal biofuel  
650 assemblages. *Journal of Applied Ecology*, *51*(3), 603-611.

651 Smith, V. H., & McBride, R. C. (2015). Key ecological challenges in sustainable algal biofuels  
652 production. *Journal of Plankton Research*, *37*(4), 671-682.

653 Smith, V. H., McBride, R. C., Shurin, J. B., Bever, J. D., Crews, T. E., & Tilman, G. D. (2015).  
654 Crop diversification can contribute to disease risk control in sustainable biofuels  
655 production. *Frontiers in Ecology and the Environment*, *13*(10), 561-567.

656 Stockenreiter, M., Graber, A.-K., Haupt, F., & Stibor, H. (2011). The effect of species diversity  
657 on lipid production by micro-algal communities. *Journal of Applied Phycology*, *24*(1),  
658 45-54.

659 Stockenreiter, M., Haupt, F., Graber, A.-K., Seppälä, J., Spilling, K., Tamminen, T., Stibor, H.,  
660 & Buschmann, A. (2013). Functional group richness: Implications of biodiversity for  
661 light use and lipid yield in microalgae. *Journal of Phycology*, *49*(5), 838-847.

662 Stockenreiter, M., Haupt, F., Seppälä, J., Tamminen, T., & Spilling, K. (2016). Nutrient uptake  
663 and lipid yield in diverse microalgal communities grown in wastewater. *Algal Research*,  
664 *15*, 77-82.

665 Sturm, B. S. M., Peltier, E., Smith, V., & deNoyelles, F. (2012). Controls of microalgal biomass  
666 and lipid production in municipal wastewater-fed bioreactors. *Environmental Progress &*  
667 *Sustainable Energy*, *31*(1), 10-16.

668 Valdez, P. J., Nelson, M. C., Wang, H. Y., Lin, X. N., & Savage, P. E. (2012). Hydrothermal  
669 liquefaction of *Nannochloropsis* sp.: Systematic study of process variables and analysis  
670 of the product fractions. *Biomass and Bioenergy*, *46*, 317-331.

671 Wiens, J. J., Fargione, J., & Hill, J. (2011). Biofuels and biodiversity. *Ecological Applications*,  
672 *21*(4), 1085-1095.

673 Williams, P. J. I. B., & Laurens, L. M. L. (2010). Microalgae as biodiesel & biomass feedstocks:  
674 Review & analysis of the biochemistry, energetics & economics. *Energy &*  
675 *Environmental Science*, *3*(5), 554.

676

677 Data Statement. The entire dataset is available in the Supporting Information.

678

679 **Tables**

680 Table 1. List of hypotheses regarding the potential for biodiversity to improve algal feedstock  
681 cultivation. Each prediction is evaluated in terms of (1) the average at each level of species  
682 richness, (2) the capacity for at least one polyculture to outperform its best component species  
683 and (3) the to capacity for at least one polyculture to outperform the best single species examined  
684 in the experiment. 'X' Means that the prediction was not supported. ✓ Means that the prediction  
685 was supported and was statistically significant ( $p < 0.05$ ).

<u>Hypotheses.</u> Compared to monocultures, certain polycultures:	Compared to...		
	...the mean of their component species	...their best component species	...the best species in the experiment
H1: Increase biomass	X	X	X
H2: Increase biocrude yield	✓	X	X
H3: Increase stability	✓	X	X
H4: Decrease crash magnitude	X	X	X
H5: Delay crashes	X	X	X
H6: Decrease invasive algae	✓	X	X
H7: Delay invasive algae	✓	✓	✓
H8: Maintain more functions at ... . . .	✓	✓	✓

686

687

688 Table 2. Results of general linear models and logistic regression models used to test hypotheses H1 through H7. Effects of species  
 689 combination (Combo) were nested in species richness (SR). Subscripts are used to display the numerator and denominator degrees of  
 690 freedom (e.g.  $F_{i,j}$ ). <sup>b</sup> Denotes spatial block was included as a random effect in the minimum adequate model and <sup>p</sup> denotes pond  
 691 identity was included as a repeated measures random effect in the minimum adequate model.  
 692

Hypotheses	Response variable	SR	SR Combo	Time	SR*Time	Combo*Time
H1	Biomass <sup>p</sup>	$F_{2,59}=1.91,$ p=0.16	$F_{8,59}=2.77,$ p<0.02	$F_{1,61}=6.20,$ p<0.02	$F_{2,61}=0.17,$ p=0.85	$F_{8,61}=3.37,$ p<0.01
H2	Biocrude yield <sup>b,p</sup>	$F_{2,80}=7.29,$ p<0.02	$F_{8,82}=3.58,$ p<0.02	$F_{1,83}=18.62,$ p<0.001	$F_{2,77}=0.70,$ p=0.50	$F_{8,73}=0.65,$ p=0.74
H3	Stability	$F_{2,61}=5.45,$ p<0.01	$F_{8,61}=4.66,$ p<0.001	NA	NA	NA
H4	Maximum crashes <sup>b</sup>	$F_{2,61}=1.34,$ p=0.27	$F_{8,61}=4.14,$ p<0.001	NA	NA	NA
H5	Crash timing	p<0.02	p<1x10 <sup>-9</sup>	p<1x10 <sup>-15</sup>	p=0.29	p=0.56
H6	Maximum invader algae <sup>b</sup>	$F_{2,58}=12.20,$ p<1x10 <sup>-4</sup>	$F_{8,58}=10.88,$ p<1x10 <sup>-8</sup>	NA	NA	NA
H7	Invasion timing (>1%)	p<1x10 <sup>-15</sup>	p<1x10 <sup>-15</sup>	p<1x10 <sup>-15</sup>	p=0.8	NA

693



694 **Figure Captions**

695 Fig. 1. Photographs of the experimental 1,100L outdoor ponds located at the Edwin S. George  
696 Reserve near Pinckney, MI, USA. Inset text summarizes the experimental design and replication.

697  
698 Fig. 2. Box-whisker charts showing the mean biomass (a), biocrude yield (b), biomass stability  
699 (c), maximum biomass crash magnitude (d), and maximum proportion of invader algae (e) for  
700 replicate ponds in each inoculation treatment. Blue styling denotes monocultures, orange denotes  
701 2-species polycultures, green denotes the 4-species polyculture, and grey denotes the control  
702 treatment (no inoculum). The horizontal lines in each box represent the median, the edges of the  
703 box represent the 25<sup>th</sup> and 75<sup>th</sup> percentiles, the whiskers represent the maximum and minimum  
704 values, and filled circles represent values more than twice the interquartile distance from the  
705 median. In panels a-c the dashed horizontal lines show the mean for each level of species  
706 richness. In panel d the dashed line represents the overall median. Species codes are listed in Fig.  
707 1.

708  
709 Fig. 3. Percent of replicate ponds that crashed on or before each date (a). The threshold for  
710 crashes was a 55% reduction in biomass in 7 days, which was the median for the maximum crash  
711 size observed in each pond. Occurrence of invader algae in the ponds through time (b). Error  
712 bars represent one standard error of the mean for replicate ponds (n=6 to 8 each).

713  
714 Fig. 4. Heatmap showing performance tradeoffs among the monocultures and polycultures (a).  
715 All performance ranks are ordered to that warmer colors represent more desirable performance.  
716 Plots of the number of functions performed above each threshold, with separate styling for the  
717 mean at each level of species richness (b) and the best performer at each level of species richness  
718 (c). Error bars in panel b denote the standard error. In panel c, lines were jittered vertically to  
719 improve readability: monocultures were moved up slightly and the 4-species culture was moved  
720 down slightly with respect to the 2-species polycultures. In panels b and c, \* symbols indicate  
721 significantly higher performance of the 4-species polyculture compared to the monocultures  
722 ( $p < 0.05$ ), as determined by randomization tests (see Methods, Fig. S4).



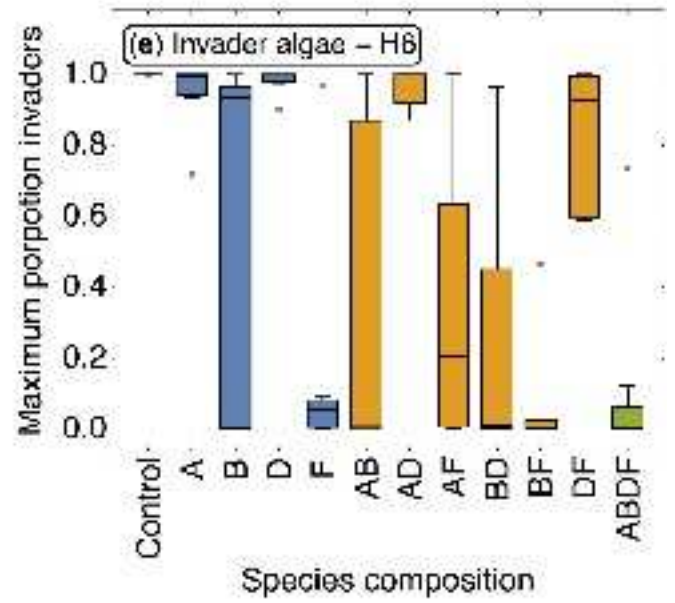
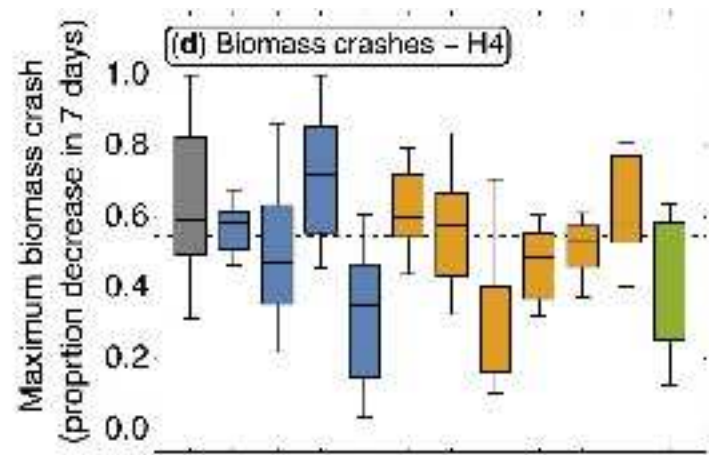
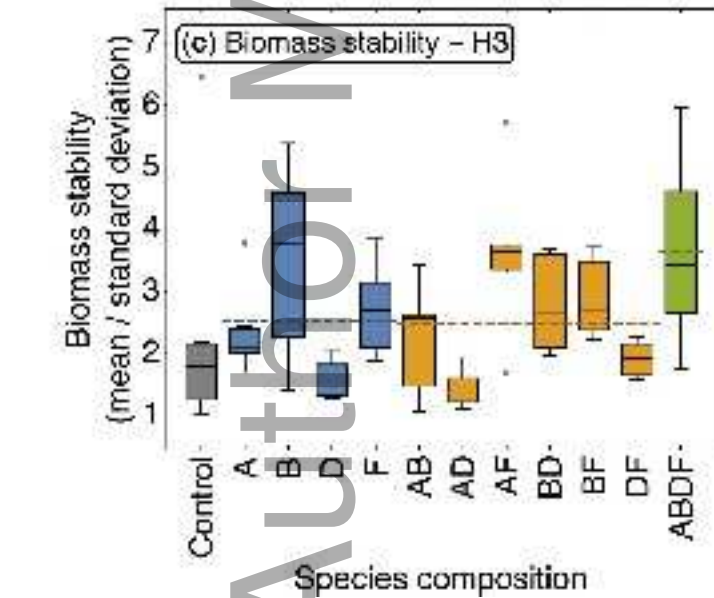
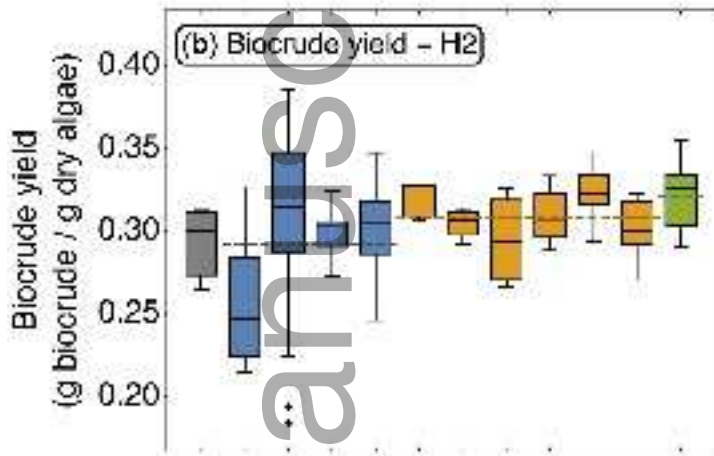
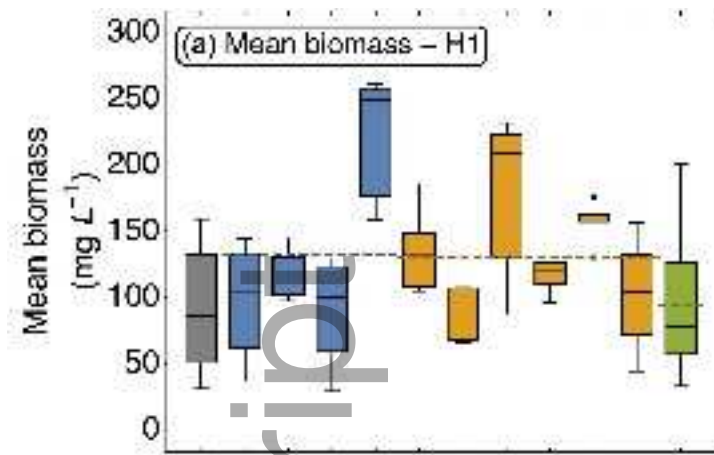
**Inoculation treatments  
and replication:**

A x 7	6 pairs x 6
B x 7	ABDF x 8
D x 7	Control x 8
F x 7	

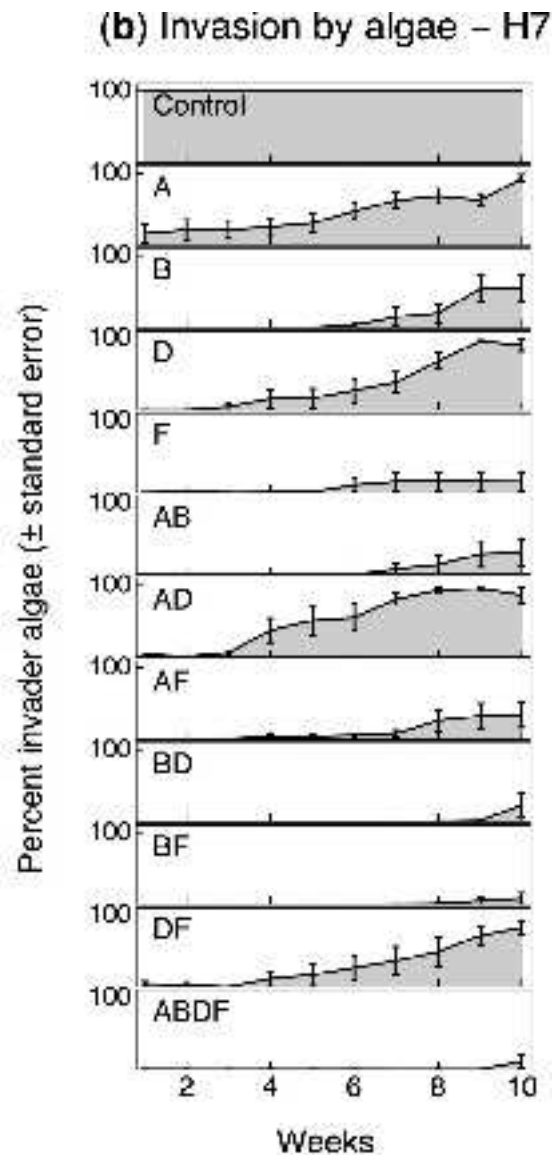
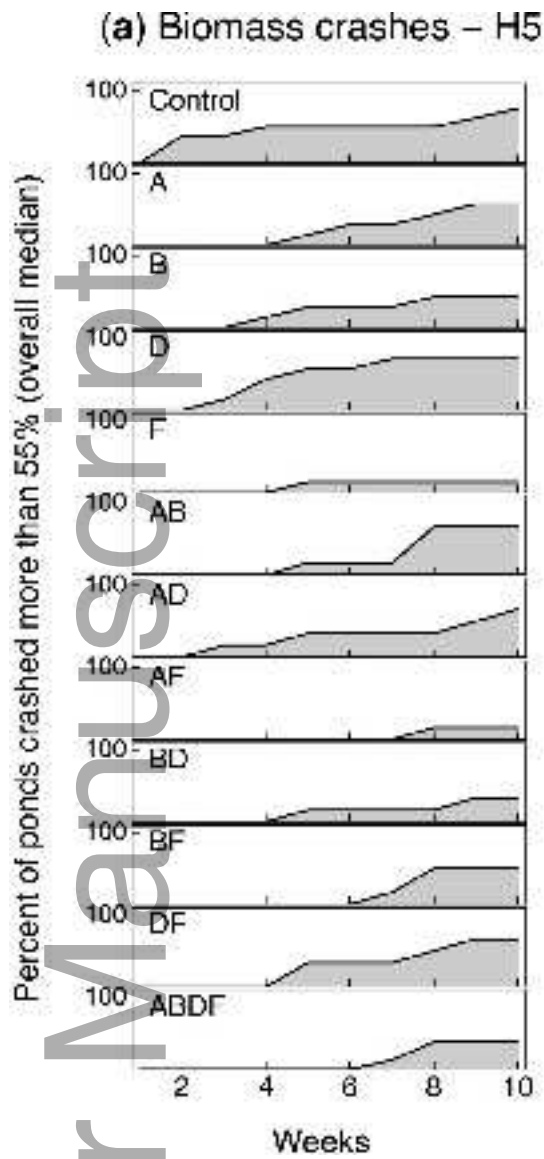


A=*Ankistrodesmus falcatus*    B=*Chlorella sorokiniana*  
D=*Scenedesmus acuminatus*    F=*Selenastrum capricornutum*

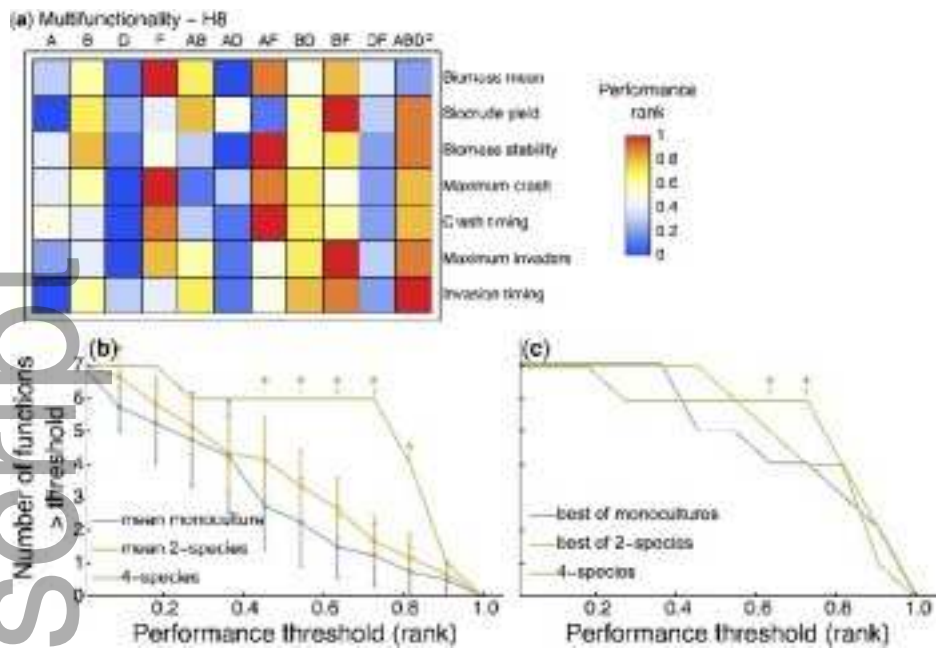
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gccb\_12524\_f2.jpeg



gcb\_12524\_f3.jpeg



gcb\_12524\_f4.jpeg