1	
2	DR. JOSEPH DIBATTISTA (Orcid ID : 0000-0002-5696-7574)
3	DR. JONATHAN CHANG (Orcid ID : 0000-0002-3811-1254)
4	
5	
6	Article type : Original Research
7	Š
8	
9	Article Type: Original Article
10	Title: Ice ages and butterflyfishes: phylogenomics elucidates the ecological and evolutionary
11	history of reef fishes in an endemism hotspot
12	
13	Running Head: Evolution of endemic butterflyfish
14	Joseph D. DiBattista ^{a,b,c*} , Michael E. Alfaro ^d , Lori Sorenson ^d , John H. Choat ^e , Jean-Paul A.
15	Hobbs ^c , Tane H. Sinclair-Taylor ^a , Luiz A. Rocha ^f , Jonathan Chang ^d , Osmar J. Luiz ^g , Peter F.
16	Cowman ^h , Matt Friedman ^{i,j} , and Michael L. Berumen ^a
17	
18	^a Red Sea Research Center, Division of Biological and Environmental Science and Engineering,
19	King Abdullah University of Science and Technology, Thuwal 23955, Saudi Arabia
20	
21	^b Australian Museum Research Institute, Australian Museum, 1 William St, Sydney, NSW 2010,
22	Australia
23	
24	^c School of Molecular and Life Sciences, Curtin University, Perth, WA 6845, Australia
25	
	This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the <u>Version of Record</u> . Please cite this article as <u>doi:</u> 10.1002/ECE3.4566

26	^d Department of Ecology and Evolutionary Biology, University of California Los Angeles, Los
27	Angeles, CA 90095, USA
28	
29	^e School of Marine and Tropical Biology, James Cook University, Townsville, QLD 4811,
30	Australia
31	
32	^f Section of Ichthyology, California Academy of Sciences, San Francisco, CA 94118, USA
33	
34	^g Research Institute for the Environment and Livelihoods, Charles Darwin University, Darwin,
35	NT 0810, Australia
36	
37	^h ARC Centre of Excellence for Coral Reef Studies, James Cook University, Townsville,
38	Australia
39	
40	ⁱ Department of Earth Sciences, University of Oxford, South Parks Road, Oxford, OX1 3AN, UK
41	
42	^j Museum of Paleontology and Department of Earth and Environmental Sciences, Ann Arbor, MI
43	48109, USA
44	
45	
46	*Correspondence: Australian Museum Research Institute, Australian Museum, 1 William St,
47	Sydney, NSW 2010, Australia
48	Phone: +61 0 478 514 097
49	E-mail: josephdibattista@gmail.com
50	
51	Abstract
52	For tropical marine species, endemism hotspots occur in peripheral areas furthest from the center
53	of diversity, but the evolutionary processes that lead to their origin remains elusive. We test
54	several hypotheses related to the evolution of peripheral endemics by sequencing ultraconserved
55	element (UCE) loci to produce a genome-scale phylogeny of 47 butterflyfish species (family

56 Chaetodontidae) that includes all shallow-water butterflyfish from the coastal waters of the 57 Arabian Peninsula (i.e. Red Sea to Arabian Gulf) and their close relatives. Bayesian tree building methods produced a well-resolved phylogeny that elucidated the origins of butterflyfishes in this 58 59 endemism hotspot. We show that UCEs, often used to resolve deep evolutionary relationships, 60 represent an important tool to assess the mechanisms underlying recently diverged taxa. Our analyses indicate that unique environmental conditions in the coastal waters of the Arabian 61 Peninsula probably contributed to the formation of endemic butterflyfishes. Older endemic 62 species are also associated with narrow versus broad depth ranges, suggesting that adaptation to 63 deeper coral reefs in this region occurred only recently (< 1.75 Ma). Even though deep reef 64 environments were drastically reduced during extreme low sea level stands of glacial ages, 65 shallow reefs persisted, and as such there was no evidence supporting mass extirpation of fauna 66 in this region. 67

- 68
- 69
- 70
- 71

72 Keywords: biogeographic barriers; *Chaetodon*; coral reef; glaciation events; Pleistocene;

73 ultraconserved elements

74 Introduction

75 Explaining the underlying factors responsible for the diversity of species accumulated at 76 endemism hotspots remains a difficult problem in the field of biogeography. Recent research has 77 identified the importance of peripheral regions from tropical oceans in generating and exporting 78 biological diversity, thus intermittently seeding adjacent seas (DiBattista et al. 2010; Eble et al. 2011; Gaither et al. 2010, 2011; Malay & Paulay 2010; Skillings et al. 2010; Bowen et al. 2013; 79 80 DiBattista et al. 2013), however, direct tests of this assumption are rare. Renewed interest in the 81 Red Sea to Arabian Gulf (or Persian Gulf) region provides a new opportunity to explore hypotheses associated with how endemics are formed in peripheral areas, and its potential 82 contribution to the species richness of marine biodiversity hotspots. The Red Sea is a semi-83 84 enclosed basin located at the north-western corner of the Indian Ocean and harbors one of the highest levels of endemism for marine organisms (12.9% for fishes, 12.6% for polychaetes, 8.1% 85

for echinoderms, 16.5% for ascidians, and 5.8% for scleractinian corals; DiBattista et al. 2016a). 86 87 The level of endemism among well-characterized groups in the Red Sea, such as the shore fishes, exceeds those of all other peripheral endemic hotspots identified for the Indian Ocean (DiBattista 88 et al. 2016a). Although many of these Red Sea endemics extend their distribution into the 89 adjacent Gulf of Aden and Arabian Sea (Kemp 1998; DiBattista et al. 2016a,b), it is not clear 90 whether they are paleo-endemics (old lineages restricted due to range contraction), neo-endemics 91 (young lineages at site of origin), or "ecological" endemics (old or young lineages restricted 92 range due to species ecology; see Cowman et al. 2017) and where, when, and how this 93 94 diversification occurred.

The Red Sea has a unique geological and paleoclimactic history that may have played a 95 role in its high levels of endemism (see DiBattista et al. 2016b for review). In brief, the Red Sea 96 basin was formed by episodes of sea floor spreading 41 to 34 Ma (Girdler & Styles 1974), 97 followed by intermittent connections to the Mediterranean Sea in the north (~14 Ma to 5 Ma; 98 Hubert-Ferrari et al. 2003), and a more recent connection to the Gulf of Aden in the south 99 100 through the Strait of Bab al Mandab (~5 Ma to present; Bailey 2009). The Strait is a narrow channel (29 km) with a shallow sill (137 m) that constitutes the only connection between the Red 101 Sea and the Indian Ocean (Bailey 2009). Water exchange is regulated by Indian Ocean monsoon 102 103 patterns (Smeed 1997; Raitsos et al. 2013) but was historically minimal or absent during reduced 104 sea levels caused by glacial periods of the Pleistocene (Rohling et al. 2009), including the most 105 recent glacial maximum (20 to 15 ka; Siddall et al. 2003; Ludt & Rocha 2015). Restricted water 106 flow resulted in increased salinity within the Red Sea (Biton et al. 2008), leading some to 107 suggest that there was complete extirpation of Red Sea fauna during these periods (Klausewitz 108 1989). The "Pleistocene extirpation" hypothesis, wherein all Red Sea fauna were eliminated during the last glacial maximum (~18 ka) and subsequently re-populated via more recent 109 110 colonization events, remains controversial and untested with modern comparative approaches (DiBattista et al. 2016b), although similar geological events may have occurred in the 111 Mediterranean Sea (Bianchi et al. 2012). Thus, despite some agreement on the broad strokes of 112 its geologic history, little consensus has emerged on the processes that shaped the Arabian 113 114 Peninsula's present day marine biodiversity, their influence on biodiversity in adjacent regions, and the role of historical closures of the Strait of Bab al Mandab. 115

116 Butterflyfishes and bannerfishes, brightly colored reef fishes in the family Chaetodontidae, are a potential model system for elucidating the origins, maintenance, and 117 evolutionary history of Red Sea endemics and their influence on species richness in adjacent 118 marine regions. The family is diverse (17 species in the Red Sea and >130 species in the greater 119 Indo-West Pacific; Allen et al. 1998) and phylogentically well-resolved compared to other reef 120 fish families (Cowman 2014). A high proportion of the Chaetodontidae species found in the 121 122 coastal waters of the Arabian Peninsula are endemic (32%; DiBattista et al. 2016a). Although recent molecular phylogenies of chaetodontids have helped to clarify many aspects of their 123 124 evolutionary history (Fessler & Westneat 2007; Hsu et al. 2007; Bellwood et al. 2010; Cowman & Bellwood 2011; Cowman & Bellwood 2013; Hodge et al. 2014), a lack of sampling of 125 Arabian Peninsula species has impeded our understanding of the diversification in this region. 126 The evolution of endemic species has been linked to ecological traits, such as reductions 127 in dispersal ability and changes in body size (i.e. the island rule; reviewed by Lomolino 2005; 128 Whittaker & Fernandez-Palacios 2007). For reef fishes, certain traits associated with dispersal 129 130 ability are linked to geographic range size. For example, large, gregarious, and nocturnal species tend to have larger range sizes than small, solitary, and strictly diurnal species (Luiz et al. 2012; 131 Luiz et al. 2013). Moreover, dispersal ability can potentially influence clade diversification: to 132 133 successfully colonize and establish populations in peripheral areas, tropical fish species must be 134 good dispersers (Hobbs et al. 2012). Following diversification in peripheral areas, newly formed lineages may evolve traits less conducive to dispersal, thus becoming endemic to the area where 135 136 it originated, as often occurs in the evolution of insular terrestrial endemics (Whittaker & 137 Fernandez-Palacios 2007). We therefore predict that butterflyfishes endemic to the Arabian Peninsula region will have smaller body sizes, higher sociability, and reduced dispersal ability 138 compared to their widespread congeners. Broadly speaking, endemic species tend to be 139 140 ecological specialists and thus adapted to the environmental condition in which they arose (McKinney 1997). We therefore additionally predict that these endemics will have a higher level 141 of ecological specialization than widespread species. For reef fishes, habitat specialization is 142 often defined by the depth range where they occur and the number of different habitats that they 143 144 exploit (e.g. coral reefs, rocky reefs, seagrass beds, mangroves; Luiz et al. 2012). Dietary specialization is often defined by the proportion of different food categories targeted (Pratchett 145

2014). We predict that butterflyfishes endemic to the Arabian Peninsula region will have higher
dietary specialization and reliance on corals for food given recent origins alongside their coral
rich habitat (Renema *et al.* 2016). We choose to focus on adult versus larval ecological traits
because more information about the former is available, and has been shown to correlate with
past (Ottimofiore *et al.* 2017) and present (Luiz *et al.* 2013) geographic range size.

The aims of this study are threefold. First, we aim to reconstruct the phylogeny and 151 152 evolutionary timescale for Red Sea to Arabian Gulf butterflyfishes in order to test whether these peripheral areas intermittently seed the broader Indo-West Pacific with biodiversity 153 ("evolutionary incubator" hypothesis). Outcomes that would allow rejection of this hypothesis 154 include a lack of evidence supporting Arabian Peninsular endemic fish lineages giving rise to 155 Indo-West Pacific fish lineages as well as restricted ancestral ranges expanding into this broader 156 157 region. Second, we look to test the extent to which butterflyfish maintained a continuous presence in the Red Sea during the major environmental fluctuations of the Pleistocene 158 ("Pleistocene extirpation" hypothesis). Outcomes that would allow rejection of this hypothesis 159 160 include a lack of evidence supporting Arabian Peninsular endemic fish originating after the glacial cycles of the Pleistocene, as well as colonization events dated only before or after this 161 epoch. Third, we aim to test whether species endemic to the coastal waters of the Arabian 162 163 Peninsula non-randomly associate with particular ecological traits ("ecological trait" hypothesis), 164 which may be important in explaining patterns of diversification in this region. The expectation 165 here is that endemic fishes are more specialized and thus better adapted to local conditions than 166 their widespread congeners. Outcomes that would allow rejection of this hypothesis include a 167 lack of association between endemism and any of the ecological traits considered here.

168

169 Materials and Methods

170 Materials

171 Site location, sampling date, and museum voucher information (where available) for each

specimen are outlined in Table S1 available as electronic supplementary material. All

butterflyfish species included in this study and their geographic distribution are listed in Table 1.

As our primary objective is to reconstruct the evolutionary history of butterflyfishes known to

175 occur in the Red Sea and adjacent gulfs or seas, we concentrated our sampling efforts on those

species and their closest relatives. Although five major Chaetodontidae lineages were sampled,
Chaetodon Clade CH1 (*Chaetodon robustus* and *C. hoefleri*, restricted to the Altantic; Cowman
& Bellwood 2013), and multiple bannerfish genera (*Amphichaetodon, Chelmon, Chelmonops, Coradion, Hemitaurichthys*, and *Johnrandallia*) without species represented in the Red Sea were
not sampled in this study. Two species of the *Prognathodes* genus were included to facilitate
fossil calibration, but were not included in the biogeographic analyses due to their Atlantic
distributions (see below).

In total, we sampled 47 chaetodontid species (35% of the entire family), which includes 183 all regional endemics and wide-ranging species found in the Arabian Peninsula region, save Roa 184 *jayakari*, a rare deepwater species distributed from the Red Sea to coastal India; we were unable 185 to secure a tissue sample as part of this study. Eight of these species have not previously been 186 sampled in phylogenetic studies of the family (Fessler & Westneat 2007; Bellwood 2010; 187 Cowman & Bellwood 2011; Hodge et al. 2014). Tissues were preserved in a saturated salt-188 DMSO solution or 95% ethanol prior to processing. This research was carried out under the 189 190 general auspices of King Abdullah University of Science and Technology's (KAUST) 191 arrangements for marine research with the Saudi Arabian Coast Guard and the Presidency of Meteorology and Environment. The animal use protocol was approved by KAUST's Biosafety 192 193 and Ethics Committee (KAUST does not provide specific approval number).

194

195 Phylogenomics approach

196 We employ the sequence capture method of ultraconserved elements (UCEs) to produce millions 197 of reads in parallel from multiple butterflyfish specimens collected from the Gulf of Aqaba in the 198 west (Red Sea) to the Hawaiian Archipelago in the east (Pacific Ocean). UCEs are a class of 199 highly conserved and abundant nuclear markers distributed throughout the genomes of most 200 organisms (Bejerano et al. 2004; Siepel et al. 2005; Reneker et al. 2012). These markers do not 201 intersect paralogous genes (Derti et al. 2006), do not have retro-element insertions (Simons et al. 202 2006), have a range of variant sites (i.e. evolving on different time-scales; Faircloth et al. 2012), and have been used to reconstruct phylogenies across vertebrates (Bejerano et al. 2004; Faircloth 203 204 et al. 2012, 2013; McCormack et al. 2013; Smith et al. 2014; Sun et al. 2014), including fishes at 205 both shallow (Mcgee et al. 2016) and deep (Faircloth et al. 2013; Harrington et al. 2016; Alfaro

et al. 2018) phylogenetic scales.

207

208 DNA library preparation and next-generation sequencing

209 DNA was extracted with DNeasy Blood and Tissue kits (Qiagen, Valencia, CA), which included 210 an RNAse A treatment step. Each extracted sample was visualised by gel electrophoresis to assess DNA quality. Total DNA from each extracted aliquot was quantified using a Qubit 211 dsDNA HS Assay Kit (Invitrogen, Carlsbad, CA), 1.2 µg of DNA per individual sample was 212 fragmented by sonication to 500 base pairs (bp) using a Covaris S2 sonicator (Covaris Inc. 213 214 Woburn, MA) and used for UCE library prep. In brief, we end-repaired, adenylated, and ligated fragmented DNA to Illumina TruSeq-style adapters, which included custom sequence tags to 215 barcode each individual sample (Faircloth & Glenn, 2012). Following an 18-cycle PCR to 216 amplify indexed libraries for enrichment, we created pools by combining 62.5 ng of eight 217 individual libraries. Each pool was concentrated to 147 ng per µl using a vacuum centrifuge. We 218 219 then followed an established workflow for target enrichment (Gnirke et al. 2009) with 220 modifications specified in Faircloth et al. (2012). Specifically, we enriched each pool, targeting 221 UCE loci and their flanking sequence, using synthetic RNA capture probes (MyBaits, Mycroarray, Inc., Ann Arbor, MI). We combined the enriched, indexed pools at equimolar ratios 222 223 prior to sequencing. The two final pooled libraries were each run paired-end (150 bp sequencing) 224 on independent lanes of an Illumina HiSeq2000 (v3 reagents) at the King Abdullah University of Science and Technology (KAUST) Bioscience Core Lab (BCL). Detailed methods of library 225 enrichment, post-enrichment PCR, and validation using relative qPCR may be found at 226 227 http://ultraconserved.org/#protocols.

228

229 Sequence read quality control, assembly, and UCE identification

- 230 We removed adapter contamination and low quality bases with illumiprocessor (Faircloth, 2013),
- a parallel wrapper to Trimmomatic (Bolger *et al.* 2014). To assemble the trimmed dataset, we
- used the PHYLUCE pipeline (version 8ca5884; Faircloth 2016) with the
- phyluce_assembly_assemblo_trinity.py wrapper script for Trinity (version 1.5.0; Grabherr *et al.*
- 234 2011). We matched assembled contigs to enriched UCE loci by aligning contigs from each
- species to our UCE probes using the phyluce_assembly_match_contigs_to_probes.py script with

the LASTZ assembler (Harris 2007). We stored these match results into a SQLite relational

237 database after excluding contigs that matched multiple UCE loci and UCE loci whose probes

238 matched multiple contigs.

We used phyluce_align_seqcap.py to align UCE loci with MAFFT (Katoh et al. 2002; 239 Katoh & Standley 2013). Following alignment, we end- and internally-trimmed alignments with 240 GBLOCKS (Castresana 2000) to improve phylogenetic inference by removing poorly-aligned or 241 highly divergent sites (Talavera & Castresana 2007). We selected loci that were present in at 242 least 75% of our specimens and concatenated the alignments into a PHYLIP-formatted matrix 243 for phylogenetic analysis. We included previously published UCE data for three species in our 244 alignment to represent Acanthomorpha outgroup lineages and more accurately calibrate the 245 phylogeny (see below). 246

247

248 Phylogenetic analysis of concatenated UCE data: evaluation of the "evolutionary incubator"
249 and "Pleistocene extirpation" hypotheses

We fully partitioned our concatenated alignment by UCE locus and performed Bayesian analyses of the dataset with ExaBayes (Aberer *et al.* 2014) and two independent runs, sampling every 500 generations. We used the autostopping convergence criteria of an average standard deviation of split frequencies of less than 5% and visualised the log-likelihood of each chain to ensure convergence in Tracer version 1.6 (Rambaut *et al.* 2014).

We estimated divergence times using MCMCTREE in the PAML package on the 255 256 Bayesian consensus topology. We used the likelihood approximation approach following the 257 two-step procedure described by Dos Reis & Yang (2011) by first estimating a mean substitution 258 rate for the entire alignment with BASEML under a strict molecular clock and then using this estimate to set the rgene_prior in MCMCTREE. We used a single, unpartitioned alignment for 259 260 computational tractability, with an HKY85 model, five categories for the gamma distribution of 261 rate heterogeneity, an rgene_gamma prior for the gamma distribution describing gene rate 262 heterogeneity of (2, 371.0575, 1) and a sigma2_gamma prior of (2, 5, 1). We adopted a calibration strategy that builds on Harrington et al. (2016) by including more proximal 263 264 acanthomorph outgroups to Chaetodontidae and their immediate relatives. We constrained six nodes on the basis of fossil information using hard lower and soft upper bounds outlined in 265

Figure S1 (available as Supplementary Material). We assigned a minimum amount of prior 266 267 weight for ages below the lower bound (1e-200) and 5% prior weight for ages higher than the upper bound. Briefly, we link a series of carangimorph, syngnathiform, holocentroid, and 268 269 lampridiform fossils to the sequences of acanthomorph outgroup fossils as per Harrington et al. (2016). This resulted in the following outgroup node calibrations: acanthuroids versus all other 270 taxa (lower bound: 54.17 Ma; upper bound: 70.84 Ma); acanthurids versus zanclids (lower 271 272 bound: 49.0 Ma; upper bound: 62.7 Ma), *Naso* versus *Acanthurus* (lower bound: 49.0 Ma; upper 273 bound: 57.22 Ma), Chaetodontidae versus Pomacanthidae (lower bound: 29.62 Ma; upper bound: 59.26 Ma), and the total-group Chaetodon versus Prognathodes (lower bound: 7 Ma; upper 274 bound: 47.5 Ma). Further justification for calibrations are available as electronic supplementary 275 material (Appendix S2). 276

277

278 Ancestral biogeographic range estimation: evaluation of the "evolutionary incubator" and

279 "Pleistocene extirpation" hypotheses

280 We estimated ancestral distribution patterns for chaetodontid lineages using the pruned timecalibrated phylogeny analysed with the R package BioGeoBEARS (Matzke 2013), which allows 281 several models of biogeographic evolution to be compared via likelihood inference, and the 282 283 ability to incorporate a parameter allowing for founder-event speciation. For these analyses, we 284 coded each taxon based on presence/absence in nine discrete geographical areas: Gulf of Aqaba, rest of the Red Sea, Djibouti and Gulf of Aden, Socotra, South Oman, Arabian Gulf, Gulf of 285 Oman and Pakistan, rest of Indian Ocean, and Pacific Ocean. The discrete coding of geographic 286 287 areas adjacent to the Arabian Peninsula enable a fine-scale investigation of the ancestral 288 biogeography of that region for our taxa of interest. Presence/absence and geographical range data for each taxon were obtained from a combination of DiBattista et al. (2016a) and FishBase 289 290 (Froese & Pauly 2011). Prognathodes spp. (a Chaetodontidae genus) were not considered in this 291 part of the analysis given that these two taxa are restricted to tropical Atlantic waters. 292 We constrained our biogeographic analyses to prohibit colonization events between the Red Sea and Indian/Pacific Ocean regions before 5 Ma reflecting the time when a more 293 294 permanent connection was formed via the Strait of Bab al Mandab (Bailey 2009). Our 295 BioGeoBEARS analysis evaluated the DEC, DIVALIKE, and BAYAREALIKE models with

and without the jump (J) parameter (Matzke 2013). These models describe biogeographic
scenarios where dispersal, extinction, cladogenesis, vicariance, and founder events are
differentially invoked to explain present day distributional patterns. In our case, we were
interested in whether the range-restricted endemics from the coastal waters of the Arabian
Peninsula represent ancient relicts, new colonization events, and/or a source of biodiversity (at
some point in the past) for the broader Indo-West Pacific.

302

303 *Comparative trait analysis: evaluation of the "ecological trait" hypothesis*

In order to determine whether particular species-level traits were associated with the evolution of 304 endemism in this subset of Chaetodontidae species, we fitted a phylogenetic generalized linear 305 model (function 'phyloglm' in R package 'phylolm' [Ho et al. 2016]) that assumed "regional 306 307 endemism" (i.e. endemic to the coastal waters of the Arabian Peninsula; DiBattista et al. 2016a) as the binomial response variable and a suite of ecological traits as the predictive fixed factors. 308 For model selection, we performed a backward stepwise procedure for PGLM's (function 309 310 'phylostep' in R package 'phylolm' [Ho et al. 2016]), which entailed sequential optimization by removing non-influential fixed-effect terms from the full model based on Akaike information 311 criteria (AIC). Full details on the methods and data sources are provided in Table S2 as 312 313 electronic supplementary material. We also explore interactions among the predictive traits using 314 a regression tree approach (De'ath & Fabricius 2000; function 'rpart' in R package 'rpart' [Therneau et al. 2015]) 315

316 Among the predictive variables considered were: maximum body size (total length = 317 TL)(Allen et al. 1998; Kuiter 2002), depth range inhabited (Allen et al. 1998), social structure (three categories ordered from low to high sociability: solitary, pair formation, and group 318 319 formation; Allen et al. 1998; Kuiter 2002; Yabutu & Berumen 2013), habitat breadth (estimated 320 as the sum value of all habitat types inhabited: C = coral, R = rocky, D = deep reef, S = sediment, 321 R = rubble, CO = coastal, CA = algal beds; Allen *et al.* 1998; Kuiter 2002), and dietary reliance on coral reefs (four categories ordered from low to high reliance: planktivore, benthic 322 invertivore, facultative corallivore, and obligate corallivore; Cole & Pratchett 2013). We also 323 324 included the phylogenetic age of species (Myr) as an additional fixed factor to test if species traits are influenced by time of divergence from sister taxa. For phylogenetic age, we evaluate 325

for each species (regional endemic and widespread) whether we sampled its closest sister species 326 327 by comparing our phylogeny with those published previously (Cowman & Bellwood 2011) and other published accounts (Kuiter 2002). The ecological traits were selected because they are 328 329 associated with specialization, fitness, and range expansion in butterflyfishes, and thus may help 330 to explain patterns of evolution in fish endemic to the coral reefs of the Arabian Peninsula. We do note this may be an oversimplification given that our categories are coarse and biased towards 331 332 adult versus larval traits, which are themselves data deficient. Previous work, however, has demonstrated that traits associated with the successful recruitment of reef fish is more important 333 334 than traits associated with dispersal in determining differentiation between habitats (Gaither et al. 2015; Keith et al. 2015). 335

- 336
- 337 **Results**

338 UCE sequences

Reads, contigs, and UCE loci per individual are outlined in Table S3 (available as electronic

supplementary material). In summary, we sequenced a total of 153.31 million reads, with a mean

of 1.55 million reads per sample from 47 focal taxa (excluding outgroups; also see Table 1).

Overall, we assembled a mean of 12,969 contigs (95 CI, min = 10,593, max = 15,345) and 901

343 UCE loci per sample (95 CI, min = 871, max = 932).

344

Phylogenetic reconstruction and timing of divergence to evaluate the "evolutionary incubator"
and "Pleistocene extirpation" hypotheses

Following assembly, alignment, trimming, and filtering out loci that were present in fewer than

348 75 specimens (for a 75% complete dataset), we retained 971 alignments with a mean length of

515.6 bp. The concatenated supermatrix contained 500,642 bp with 52,680 informative sites and

350 was 83.3% complete based on the proportion of non-gap sequences. The following samples were

351 excluded from further analysis due to the low number of loci recovered:

- 352 *Chaetodon_interruptus*1a, *Chaetodon_lineolatus*1a, *Chaetodon_lunula*1a, and *Chaetodon*
- 353 *ulietensis*1a (for full details see Table S1); however, tissue replicates were retained for two of the

four species listed here (*C. lineolatus* and *C. lunula*).

355 Our Bayesian and maximum likelihood analyses produced a fully resolved topology that

shared key points of congruence with prior multi-locus studies of butterflyfishes (Fessler & 356 357 Westneat 2007; Hsu et al. 2007; Bellwood et al. 2010; Cowman & Bellwood 2011; Hodge et al. 2014; see Figure S2). Although direct comparisons to previous phylogenies are difficult because 358 359 these are missing many of the regional endemics (e.g. Chaetodon dialeucos, C. gardineri, C. leucopleura, C. nigropunctatus, C. pictus, C. triangulum, Heniochus intermedius), and contain 360 less sequence data and data overlap (e.g. six loci and 73% complete matrix; Hodge et al. 2014), 361 362 where there was overlap in the data sets the tips of the tree displayed similar topologies (Figure S3). In our case, however, almost every node in the tree was strongly supported (posterior 363 probabilities of 1.0; Figure 1 and 2). 364

By only considering a single representative sample per species on our chronogram 365 (Figure 2), we found that the majority of Red Sea to Arabian Gulf butterflyfish diverged from 366 their closest relatives in the last five million years (4.17 Ma to 1.16 Ma), with an average lineage 367 age of 2.39 Ma. In comparison to previous fossil calibrated studies of Chaetodontidae (Cowman 368 & Bellwood 2011; Hodge et al. 2014), the mean ages and 95% highest posterior density (HPD) 369 370 estimates are more restricted, but for the most part overlap with previous estimates (Figure S3). In terms of the topology, although our phylogenetic sampling is restricted, it still captures crown 371 nodes and age estimates of major chaetodontid lineages (with the exception of the bannerfish 372 373 lineage), as well as subclades containing Red Sea to Arabian Gulf species and their most recent common ancestors (Figure S2). Most of the clades included species pairs diverging with non-374 375 overlapping distributions dating back 2 to 1 Ma. This divergence does not appear to coincide 376 with the timing of the emergence of apparent geographic (and geological) barriers such as the 377 Strait of Bab al Mandab (Figure 2 and 3). Regional endemics appear to have diverged earliest 378 from ancestors that gave rise to the clades including *Chaetodon larvatus* and *Chaetodon* 379 semilarvatus. At least one entire subclade of CH4 was comprised of regional endemics 380 (Chaeoton dialeucos, C. nigropunctatus, and C. mesoleucos). The split between the 381 butterflyfishes (Chaetodon, Prognathodes) and bannerfishes (Heniochus, Forcipiger) was much older, with a mean of 28.7 Ma (95% HPD: 40.0-18.26; Figure 2 and Figure S1), indicating an 382 ancient split between these highly divergent body forms. 383 384

Ancestral range reconstruction to evaluate the "evolutionary incubator" and "Pleistocene
extirpation" hypotheses

Model comparison revealed the DEC+J model as the most likely (LnL = -250.79, AIC weight = 0.76) and the DIVALIKE+J model as the second most likely (LnL = -252.76, AIC weight = 0.11; Table 2 and Figure 4). The importance of the J parameter, which models long-distance or "jump" dispersal, is that ancestral ranges often comprise one area rather than several areas. The addition of the J parameter resulted in a significantly better model fit for DEC models when compared via a likelihood ratio test (LRT: D = 8.67, p = 0.0032).

393 Under the DEC+J model, Chaetodontidae have their crown origins in the Indo-West 394 Pacific, with subsequent dispersal to include the Arabian Peninsula and lineages leading to the base of Chaetodon and the bannerfish clade (Forcipiger/Heniochus; Figure 4). Within the CH2 395 396 clade, diversification was restricted to the Pacific Ocean with subsequent dispersal to the Indian Ocean (Chaetodon madagaskariensis, C. punctatofasciaticus, C. unimaculatus), and three of the 397 species have dispersed as far as Socotra (Chaetodon guttatissimus, C. kleinii, C. trifasciatus). 398 399 Only one species within CH2 diverged in the Gulf of Aden and subsequently colonized the Red Sea (*Chaetodon paucifasciatus*). The age of *C. paucifasciatus* is relatively young (1.5 Ma, HPD: 400 0.8 - 2.3 Ma), suggesting a similar timeline for its occupation of the Red Sea from the Gulf of 401 Aden. 402

403 In the CH3 clade, three species are present in the Red Sea that were restricted to the Arabian Peninsula (Chaetodon austriacus, C. melapterus, C. larvatus). In the case of sister pair 404 405 C. austriacus and C. melapterus, the reconstruction suggests that speciation occurred by 406 vicariance within the Red Sea. Although posterior probabilities make the details of this split 407 equivocal, the most likely scenario is a split between the Gulf of Aqaba and the Red Sea, where C. austriacus subsequently recolonized the entire Red Sea but C. melapterus expanded out to the 408 409 Gulf of Aden, Arabian Sea, and Arabian Gulf, but no further. The extended history of the clade, 410 although not completely sampled (see Figure S2), suggests that a widespread ancestor expanded into the Red Sea with subsequent vicariance between the Pacific Ocean, Indian Ocean, and 411 Arabian Peninsula sites. Indeed, *Chaetodon larvatus* appears to originate in Djibouti and the 412 Gulf of Aden followed by dispersal into the Red Sea and South Oman. *Chaetodon trifascialis*, on 413 the other hand, maintained connections across the Indo-West Pacific with subsequent range 414

415 expansion into the Red Sea.

The CH4 clade has been the most successful in terms of butterflyfish colonizing the Red 416 Sea. Eight extant species from CH4 are distributed in at least some parts of the Red Sea 417 (Chaetodon auriga, C. fasciatus, C. leucopleura, C. lineolatus, C. melannotus, C. mesoleucos, C. 418 419 pictus, C. semilarvatus), four of which are restricted to the Arabian Peninsula (Chaetodon fasciatus, C. mesoleucos, C. pictus, C. semilarvatus). Moreover, the reconstruction identified a 420 mix of origin states for CH4 species found in the Red Sea. Both C. fasciatus and C. leucopleura 421 have their origins within the Red Sea, whereas C. lineolatus and C. mesoleucos have their 422 423 reconstructed origins at Socotra. The origins of C. semilarvatus are placed in South Oman, whereas the origins of C. pictus are placed in the Gulf of Oman. With the exception of C. 424 lineolatus, a widespread Indo-West Pacific species, all CH4 lineages have reconstructed origins 425 426 in the Arabian Peninsula and Indian Ocean region, and subsequent dispersal was limited from this region. *Chaetodon lineolatus* appears to be the only species in CH4 to originate in the 427 Arabian Peninsula and then disperse across the broader Indo-West Pacific. However, unsampled 428 429 taxa from this clade are more widely distributed across the Indian and Pacific Oceans (Figure S2). 430

Three taxa of the bannerfish clade are also present in the Red Sea (*Heniochus intermedius, H. diphreutes, Forcipiger flavissimus*), with *H. intermedius* considered a Red Sea to Gulf of Aden endemic. Despite these taxa only being representative of a small proportion of the entire bannerfish clade, the reconstruction suggests a widespread ancestor that diverged in the Arabian Peninsula (*H. intermedius*) with subsequent (successful) colonization of the broader Indo-West Pacific (*F. flavissimus* and *H. diphreutes*).

437

438 Correlational trait analysis to evaluate the "ecological trait" hypothesis

Based on the best-fit PGLM, depth range and phylogenetic age were negatively correlated with endemism, with depth range being a stronger predictor than phylogenetic age (Table 3, Figure 5 and 6). Exploring these relationships using a regression tree approach reveals that the effect of phylogenetic age is dependent on depth range. Endemic species from the Arabian Peninsula region are therefore more likely to be younger than widespread ones, but only for those species with depth ranges extending to mesophotic reefs (depth range > 27 m; Figure 5 and 6). Endemism was not correlated with any of the other factors in the analysis for the butterflyfishesconsidered here (electronic supplementary material, Table S2 and S4).

447

448 Discussion

449 This study used 901 loci to successfully generate a genome-scale phylogeny of bannerfishes and butterflyfishes endemic to the coastal reefs of the Arabian Peninsula. This is the first time this 450 451 genomic method has been applied to species-level phylogenetic analyses of a reef fish group. Our phylogeny, which includes all shallow water chaetodontid species found in the Red Sea to 452 453 Arabian Gulf and their close relatives distributed throughout the Indo-West Pacific, provides divergence times with narrow confidence intervals and biogeographic insight into this endemism 454 hotspot. Reconstructing the evolutionary history of these fishes with their widespread relatives 455 456 does not appear to support the evolutionary incubator hypothesis. That is, despite generating significant biodiversity in the form of endemic species, these peripheral areas of the Arabian 457 Peninsula do not appear to have exported significant biodiversity to the central Indo-West 458 459 Pacific. In fact, potentially only three species with reconstructed origins in the Arabian Peninsula (*Chaetodon lineolatus, Heniochus diphreutes, Forcipiger flavissimus*) appear to subsequently 460 disperse to the Indo-West Pacific. Our phylogenetic analyses also revealed that most endemic 461 462 species originated prior to and persisted through the major environmental fluctuations of the 463 Pleistocene, which does not support the Pleistocene extirpation hypothesis. The ecological traitbased analyses revealed that the evolution of Red Sea to Arabian Gulf endemic butterflyfishes 464 465 was associated with specialization to shallow reef habitat and, to a lesser extent, species' 466 phylogenetic age.

467

468 Evaluating the "evolutionary incubator" hypothesis

The Red Sea, Gulf of Aden, Arabian Sea, and Arabian Gulf are all peripheral to the broader Indo-West Pacific biogeographic region and potentially produce/contribute new reef fish species to the centre (see Bowen *et al.* 2013; Hodge *et al.* 2014). Temporally, the Red Sea to Arabian Gulf butterflyfish assemblage (16 species in total) is made up of recently diverged lineages, with ages ranging from 4.17 Ma (*Forcipiger flavissimus*) to 1.16 Ma (*C. austriacus/C. melapterus* split). In a few cases, the Red Sea to Gulf of Aden endemics appear to have diverged as the 475 earliest lineage of that clade (e.g. C. larvatus and C. semilarvatus; Figure 2 and 3). Indeed, the 476 "oldest" endemic butterflyfish lineage in our study, C. larvatus (2.86 Ma, 4.3-1.6 Ma 95% HPD), appeared in the late Pliocene, and diverged from an Indo-West Pacific lineage that later gave rise 477 478 to species allopatric between the two ocean basins (C. triangulum in the Indian Ocean and C. baronessa in the Pacific Ocean). The ancestral range reconstruction of these Arabian Peninsula 479 endemics demonstrates consistent colonization routes to the Red Sea and Arabian Sea via the 480 481 Indian Ocean from the east (Figure 4), but with few examples of reciprocal expansion from the Arabian Peninsula back to the Indian/Pacific Ocean. For example, both C. larvatus and C. 482 483 semilarvatus appear to have historically diverged in Djibouti/Gulf of Aden and South Oman, respectively, successfully colonized the Red Sea, but not established further south and east based 484 on present day distributions. Similar reconstruction results were obtained for the regional 485 486 endemic *C. pictus* (Red Sea to Gulf of Oman), which showed apparent historical divergence in the Gulf of Oman and only recent colonization of the southern limits of the Red Sea. 487

Other endemics appear to have historically diverged within the Red Sea (C. fasciatus) or 488 489 adjacent Djibouti and Gulf of Aden (C. paucifasciatus) but not colonized any further to the 490 southeast. Although equivocal based on the probabilistic uncertainty of nodes in the ancestral range reconstruction of the most likely model (DEC+J), there are a number of competing 491 492 explanations for how C. austriacus and C. melapterus diverged from each other within the 493 coastal waters of the Arabian Peninsula (also see Waldrop et al. 2016), particularly since C. 494 *melapterus* is the only species in this complex present in the Arabian Gulf. The most likely 495 explanation is based on present day distributions (Figure 3c): C. austriacus is largely restricted to 496 the northern and central Red Sea (with rare records in the southern Red Sea and outside of the 497 Red Sea), whereas C. melapterus is most abundant within or adjacent to the Arabian Gulf (also with rare records in the southern Red Sea) – these bodies of water show opposite trends in terms 498 499 of productivity, sea surface temperature, and temporal patterns of environmental variation 500 (Raitsos et al. 2013; Pous et al. 2015). These environmental conditions are additionally significantly different from the rest of the Indian Ocean, and thus the unique conditions in the 501 Red Sea and Arabian Gulf may help explain how endemics evolved, or at least, concentrated and 502 503 persisted in these peripheral locations.

504

Despite a lack of supporting evidence for the evolutionary incubator hypothesis, a clear

505 pattern emerges that the unique environmental conditions in these peripheral seas may have 506 contributed to the formation of endemic species as outlined above. For example, some butterflyfish subclades are comprised entirely of regional endemics (e.g. *Chaetodon dialeucos*, 507 508 *C. mesoleucos*, and *C. nigropunctatus*), which provides further evidence that coral reef habitat 509 surrounding the Arabian Peninsula may have generated a number of new taxa. Moreover, C. *dialeucos*, an Omani species, shows geographical divergence with the remaining taxa in its group 510 511 (Figure 3), which all went on to colonize the Red Sea and the Arabian Gulf and must have 512 therefore encountered contrasting environments at the western and eastern margins of their 513 range. The shallow Arabian Gulf started to fill with seawater approximately 14,000 years ago 514 after being dry during the last glacial maximum prior to that (Lambeck 1996), suggesting that it was seeded by successive waves of colonization from coastal Oman. The same process would 515 516 have been ongoing at the western margin of the C. dialeucos range, except that the conditions encountered in the Red Sea would have contrasted to those in the Arabian Gulf (DiBattista et al. 517 2016b). So, while there is some evidence to suggest vicariance at the scale of the Arabian 518 Peninsula (i.e. diversification of most taxa occurred in the Plio-Pleistocene), a stronger scenario 519 520 is that natural selection driven by the major differences in environment and habitat within the area probably played an important role in the formation of endemic species assemblages (e.g. 521 522 Gaither et al. 2015). Thus, even though the distribution of some of the butterflyfishes considered 523 in the present study does stop abruptly at the entrance of the Strait of Hormuz (Chaetodon collare, C. pictus, and C. gardneri), it does not support the argument for geographically-driven 524 525 allopatry. Indeed, all of these species have a different distributional response near the other end 526 of their distribution at the Strait of Bab al Mandab, which includes stopping before the Straits or 527 extending through the Straits into the southern Red Sea (Figure 3). The alternative is that the incumbent widespread butterflyfish may have restricted the Red Sea to Arabian Gulf endemics 528 529 from expanding further via competitive exclusion.

The current environment of the Red Sea is spatially structured with major contrasts in the cool oligotrophic waters of the northern region compared to the much higher temperatures and productivity of the southern region (i.e. Farasan Islands in Saudi Arabia to the east and Dhalak Archipelago in Eritrea to the west) (Raitsos *et al.* 2013; Racault *et al.* 2015). This shift in environmental conditions is most clearly demonstrated in the differences in life history traits associated with reef fish species that occur in both areas, but is also seen in abundance estimates

across these gradients (DiBattista *et al.* 2016a; Roberts *et al.* 2016). Such putative selection

537 gradients are most evident in corals, which show signatures of local adaptation to divergent

- environmental conditions (D'Angelo *et al.* 2015).
- 539

540 Evaluating the "Pleistocene extirpation" hypothesis

541 The second hypothesis that we tested in this study was the Pleistocene extirpation hypothesis, 542 which predicts that all Red Sea fauna were eliminated during the last glacial maxima (~18 ka) 543 and were only re-populated via recent colonization events (see Hemleben et al. 1996; Biton et al. 2008). The number of species diverging at early stages in the Pleistocene dispute the argument 544 that Red Sea fauna did not survive complete closure or restriction of water flow at the Strait of 545 Bab al Mandab (Figure 2). Although it clearly does not coincide with a single vicariance event 546 given the variability in the splitting dates between closely related species (Figure 3; see 547 548 Michonneau 2015 for invertebrate examples) and ancestral range reconstruction favoring +J549 parameter models (i.e. founder events between non-adjacent ocean regions; see Table 2), glaciations likely played a role in their separation. Moreover, even though almost all sister 550 species have small areas of overlap at their range edge, which is usually associated with 551 552 allopatric speciation, in our case these do not coincide with geographical boundaries (i.e. 553 vicariant chokepoints) such as the Strait of Bab al Mandab (see Figure 3; Lambeck 1996; 554 DiBattista et al. 2016b). In fact, the non-congruent age and distribution of the endemic species 555 indicate a series of variable events, which may reflect localized patterns of habitat and 556 environmental change as outlined in the previous section. The best example is the relatively 557 young clade of Arabian Peninsula endemics: C. dialeucos, C. nigropunctatus, and C. mesoleucos (crown node age 2.0 Ma; 2.9-1.2 Ma 95% HPD). This group appears to have been influenced by 558 559 boundaries presented by the Omani coastline across areas where there are known changes in the 560 upwelling regime (Shi et al. 2000; McIlwain et al. 2005). This is in sharp contrast to the Indo-West Pacific parrotfishes, where present day species boundaries support the notion of allopatric 561 562 divergence (Choat et al. 2012), and endemics appear to have diverged into one or more 563 subsequent endemics (i.e. secondary endemism; Rotondo et al. 1981) based on sympatrically distributed sister-species pairs (highlighted in Choat et al. 2012). Moreover, Red Sea endemics 564

from most other families of reef fish appear to have equal proportions of allopatrically and
sympatrically distributed sister-species (Hodge *et al.* 2014), which is not the case for the
butterflyfishes.

568 The diversification of these butterflyfishes occurred at a time when the coral assemblages 569 of the world's reefs underwent a major change in coral composition and growth forms. The global proportion of staghorn coral occurrences in coral assemblages persisted throughout most 570 of the Cenozoic but increased substantially during the Pliocene and especially the Quaternary 571 (Renema et al. 2016). Indeed, the rapidly growing branching acroporid corals offered different 572 573 structural components in terms of shelter and feeding/foraging modes when compared to massive 574 corals such as poritids that dominated Miocene reefs more than 5 Ma. Thus, the chaetodontids of the Arabian Peninsula (particularly the corallivorous species) were exposed to a much more 575 576 dynamic environment than the widespread Indo-West Pacific species (Coles 2003) because of their close association with sensitive coral genera that proliferated in the region. 577

578

579 Evaluating the "ecological trait" hypothesis

580 The third hypothesis that we test here is whether ecological traits are linked to the evolution of endemism among butterflyfishes in the Red Sea to Arabian Gulf. We found a negative, 581 582 significant relationship between endemism and depth range and, to a lesser extent, phylogenetic 583 age for these butterflyfishes (Figure 5 and 6). The relationship between a narrow versus broad 584 depth range and endemism supports the view that endemic species tend to be more specialized to 585 local resources than widespread species (Hawkins et al. 2000). The majority of regional 586 endemics in this study had depth ranges that did not extend deeper than 25 meters (Figure 6), 587 despite the availability of light dependent coral habitat extending beyond that (Kahng et al. 2010). The broad range of ages represented by these shallow water specialists suggests that 588 589 adaptation to shallow reefs occurred multiple times across a relatively wide time frame (i.e. 1.3 590 to 3.3 Myr). On the other hand, speciation of endemics with a preference for deep reefs seems to 591 be a recent phenomenon, as deeper depth ranges were strongly associated with young age (less than 1.75 Myr; Figure 6). 592

593

594 Comments on incomplete sampling and biogeographic biases

595 The goal of this study was to reconstruct the evolutionary history of Red Sea to Arabian Gulf 596 butterflyfishes. As is the case with all phylogenetic and biogeographic reconstructions, our results have to be interpreted in light of the taxa that are not sampled, both extant and extinct. 597 598 Indeed, the inclusion of missing taxa has the potential to alter lineage relationships and their age 599 estimates, whereas their geographic distribution may alter the most likely biogeographic scenarios reconstructed across the tree (see discussion in Cowman & Bellwood 2013). Here, we 600 601 have been able to sample all Red Sea to Arabian Gulf butterflyfishes (save one species, Roa *jayakari*), and their close relatives from the Indian and Pacific Oceans, across four major 602 603 chaetodontid lineages (Figure S2). From a temporal perspective, the topology and ages estimated 604 for the genomic scale UCE data overlap with previous studies (Figures S2 and S3). Moreover, our sampling of eight species that have not previously been included in phylogenetic studies of 605 the Chaetodontidae family means that for 13 out of the 17 Arabian Peninsula species, we are 606 confident that we have sampled their direct sister lineage. Two of the outstanding three species 607 (Chaetodon melannotus, C. trifascialis) are wide ranging Indo-West Pacific taxa that are 608 609 reconstructed to have dispersed to the Arabian Peninsula (Figure 4). The most likely sister species of *C. melannotus* is *C. ocellicaudus* (Kuiter 2002; also see Figure S2), a west Pacific 610 species not sampled in our dataset. In the case of C. trifascialis, it is placed as the sister lineage 611 612 of a subclade of CH3 containing 10 species distributed across the Indian and Pacific Oceans, of 613 which we sampled four (Figure S2; Cowman & Bellwood 2011). The final outstanding species, *Chaetodon leucopleura*, is placed as a sister species to *Chaetodon gardineri*. Both species have 614 615 not previously been sampled in phylogenetic studies, but are recognized to be closely related to a 616 third species, Chaetodon selene (widespread in the west Pacific, Kuiter 2002), which was not 617 sampled in our UCE dataset. In each of these three cases, and more broadly across the family, the inclusion of unsampled species would increase the influence of the Indian Ocean and Pacific 618 619 Ocean in the ancestral estimation of biogeographic ranges. As such, it would act to strengthen 620 our conclusion that even though the Red Sea and adjacent gulfs and seas have been important for the generation of endemic species, they have had little contribution to the wider Indo-West 621 Pacific diversity of butterflyfishes. 622

623

624 Conclusion

It appears that the unique environmental conditions in the coastal waters of the Arabian 625 Peninsula may have contributed to the formation of endemic butterflyfishes; however, there is a 626 lack of evidence for endemics contributing significant species richness to adjacent seas (i.e. 627 628 evolutionary incubator hypothesis). Moreover, even with catastrophic environmental instability 629 experienced by the Red Sea and coastal environments of the Arabian Peninsula due to sea level changes associated with glacial cycles (Ludt & Rocha 2015), there is no evidence for a massive 630 631 extirpation of butterflyfish fauna in the region (i.e. Pleistocene extirpation hypothesis; also see DiBattista et al. 2016b). The broad range of phylogenetic ages among endemic, shallow water 632 633 butterflyfishes supports the view that species may have survived in isolated refugia within the Red Sea (DiBattista et al. 2016b). None of the dispersal-related traits were associated with 634 endemism, suggesting that factors other than those related to species intrinsic dispersal potential 635 636 may be limiting dispersal into the greater Indian Ocean (e.g. coastline geography, oceanographic barriers). 637

638

639 Acknowledgements

This work was supported by the KAUST Office of Competitive Research Funds (OCRF) under 640 Award No. CRG-1-2012-BER-002 and baseline research funds to M.L.B.; National Geographic 641 642 Society Grant 9024-11 to J.D.D.; National Science Foundation grant DEB-0842397 to M.E.A.; 643 California Academy of Sciences funding to L.A.R; Australia Research Council Discovery ECR Award DE170100516 to P.F.C. For support in Socotra, we kindly thank the Ministry of Water 644 645 and Environment of Yemen, staff at the Environment Protection Authority (EPA) Socotra, and 646 especially Salah Saeed Ahmed, Fouad Naseeb and Thabet Abdullah Khamis, as well as Ahmed 647 Issa Ali Affrar from Socotra Specialist Tour for handling general logistics. For logistic support elsewhere, we thank Eric Mason at Dream Divers, Nicolas Prévot at Dolphin Divers and the 648 649 crew of the M/V Deli in Djibouti, David Pence, the KAUST Coastal and Marine Resources Core 650 Lab and Amr Gusti, the Administration of the British Indian Ocean Territory and Chagos Conservation Trust, the Ministry of Agriculture and Fisheries in Oman including Abdul Karim, 651 the Ministère de la Pêche et des Résources Halieutiques – Madagascar, the Western Australia 652 653 Department of Fisheries, Parks Australia, as well as the University of Milano-Bicocca Marine Research and High Education Centre in Magoodhoo, the Ministry of Fisheries and Agriculture, 654

Republic of Maldives, and the community of Maghoodhoo, Faafu Atoll. For specimen 655 collections, we thank David Bellwood, Brian Bowen, John Burns, Darren Coker, Richard 656 Coleman, Joshua Copus, Joshua Drew, Iria Fernandez-Silva, Michelle Gaither, Brian Greene, 657 658 Elliott Jessup, Randy Kosaki, Jason Leonard, Keo Lopes, Sarah Longo, Cassie Lyons, Jennifer 659 McIlwain, Gerrit Nanninga, David Pence, Mark Priest, Richard Pyle, Frédéric Ramahatratra, Mark Royer, Pablo Saenz-Agudelo, Anne Sheppard, Charles Sheppard, Jacqueline Troller, 660 661 Daniel Wagner, Robert Whitton, and members of the Reef Ecology Lab at KAUST. For assistance with bench work at KAUST we thank Craig Michell. We also acknowledge important 662 663 contributions from Robert J. Toonen, John E. Randall, Jo-Ann C. Leong, David Catania for assistance with specimen archiving, and the KAUST Bioscience Core Laboratory with 664 Sivakumar Neelamegam and Hicham Mansour for their assistance with Illumina sequencing. 665 Special thanks to Scott Partridge for the use of his illustrations of the Chaetodon genus. 666 667 References 668 Aberer AJ, Kobert K, Stamatakis A (2014) ExaBayes: Massively parallel Bayesian tree inference 669 670 for the whole-genome era. *Molecular Biology and Evolution* **31**, 2553–2556. 671 Allen GR, Steene R, Allen M. (1998) A Guide to Angelfishes and Butterflyfishes. 250 p. 672 673 Odyssey Publishing, Sydney. 674 Alfaro ME, Faircloth BC, Harrington RC, Sorenson L, Friedman M, Thacker CE, Oliveros CH, 675 676 Černý D, Near TJ (2018) Explosive diversification of marine fishes at the Cretaceous– 677 Palaeogene boundary. Nature Ecology & Evolution DOI: 10.1038/s41559-018-0494-6. 678 679 Bailey G (2009) The Red Sea, Coastal Landscapes, and Hominin Dispersals in *The Evolution of* 680 Human Populations in Arabia Paleoenvironments, Prehistory and Genetics. Ed Petraglia MD, Springer Science. 681 682 683 Bejerano G, Haussler D, Blanchette M (2004) Into the heart of darkness: large-scale clustering

of human non-coding DNA. *Bioinformatics* **20**, i40-i48.

685

- Bellwood DR, Klanten S, Cowman PF, Pratchett MS, Konow N, van Herwerden L (2010)
- 687 Evolutionary history of the butterflyfishes (f: Chaetodontidae) and the rise of coral feeding
- fishes. Journal of Evolutionary Biology 23, 335-349.
- 689
- Bianchi CN, Morri C, Chiantore M, Montefalcone M, Parravicini V, Rovere A (2012)
- Mediterranean Sea biodiversity between the legacy from the past and a future of change. In *Life in the Mediterranean Sea: a look at habitat changes*, 1-55.
- 693
- Biton E, Gildor H, Peltier WR (2008) Red Sea during the Last Glacial Maximum: Implications
 for sea level reconstruction. *Paleoceanography* 23.
- 696
- Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: A flexible trimmer for Illumina Sequence
 Data. *Bioinformatics* 30, 2114-2120.
- 699
- Bowen BW, Rocha LA, Toonen RJ, Karl SA (2013) The origins of tropical marine biodiversity. *Trends in Ecology and Evolution* 28, 359-366.
- 702
- Castresana J (2000) Selection of conserved blocks from multiple alignments for their use in
 phylogenetic analysis. *Molecular Biology and Evolution* 17, 540-552.
- 705
- 706 Choat JH, Klanten OS, van Herwerden L, Robertson DR, Clements KD (2012) Patterns and
- processes in the evolutionary history of parrotfishes (Family Labridae). *Biological Journal of the*
- 708 Linnean Society **107**, 529-557.
- 709
- 710 Cole AJ, Pratchett MS (2014) Diversity in diet and feeding behaviour of butterflyfishes: reliance
- on reef corals versus reef habitats. In: Pratchett MS, Berumen ML, Kapoor BG (eds) *Biology of*
- 712 *Butterflyfishes*. CRC Press, Boca Raton, FL, USA, pp. 107-139.
- 713
- Cole AJ, Pratchett MS, Jones GP (2008) Diversity and functional importance of coral-feeding

- fishes on tropical coral reefs. *Fish and Fisheries* **9**, 286-307.
- 716
- Coles SL (2003) Coral species diversity and environmental factors in the Arabian Gulf and the
- Gulf of Oman: a comparison to the Indo-Pacific region. *Atoll Research Bulletin* **507**, 1-19.
- 719
- 720 Cowman PF (2014) Historical factors that have shaped the evolution of tropical reef fishes: A
- review of phylogenies, biogeography, and remaining questions. *Frontiers in Genetics* **5**, 1–15.
- 722
- Cowman PF, Bellwood DR (2011) Coral reefs as drivers of cladogenesis: expanding coral reefs,
 cryptic extinction events, and the development of biodiversity hotspots. *Journal of Evolutionary Biology* 24, 2543-2562.
- 726
- Cowman PF, Bellwood DR (2013) The historical biogeography of coral reef fishes: Global
 patterns of origination and dispersal. *Journal of Biogeography* 40, 209-224.
- 729
- 730 Cowman PF, Parravicini V, Kulbicki M, Floeter SR (2017) The biogeography of tropical reef
- fishes: endemism and provinciality through time. *Biological Reviews* **92**, 2112-2130.
- 732
- Crosby MP, Reese ES, Berumen ML (2013) Butterflyfishes as ambassadors of coral reefs.
 Chapter 10 In Pratchett, Berumen, Kapoor *Biology of Butterflyfishes*.
- 735
- 736 D'angelo C, Hume BC, Burt J, Smith EG, Achterberg EP, Wiedenmann J (2015) Local
- adaptation constrains the distribution potential of heat-tolerant *Symbiodinium* from the
- 738 Persian/Arabian Gulf. *The ISME Journal* 9, 2551-2560.
- 739
- 740 De'ath G, Fabricius KE (2000) Classification and regression trees: a powerful yet simple
- technique for ecological data analysis. *Ecology* **81**, 3178-3192.
- 742
- 743 Derti A, Roth FP, Church GM, Wu CT (2006) Mammalian ultraconserved elements are
- strongly depleted among segmental duplications and copy number variants. *Nature*

- 745 *Genetics* **38**, 1216-1220.
- 746
- 747 DiBattista JD, Wilcox C, Craig MT, Rocha LA, Bowen BW (2010) Phylogeography of the
- 748 Pacific Blueline Surgeonfish, Acanthurus nigroris, reveals high genetic connectivity and a
- ryptic endemic species in the Hawaiian Archipelago. Journal of Marine Biology 2011.
- 750
- 751 DiBattista JD, Berumen ML, Gaither MR, Rocha LA, Eble JA, Choat JH, Craig MT,
- 752 Skillings DJ, Bowen BW (2013) After continents divide: comparative phylogeography of reef
- fishes from the Red Sea and Indian Ocean. *Journal of Biogeography* **40**, 1170-1181.
- 754

755 DiBattista JD, Roberts M, Bouwmeester J, Bowen BW, Coker DF, Lozano-Cortés DF,

- Choat JH, Gaither MR, Hobbs JP, Khalil M, et al. (2016a) A review of contemporary patterns of
- rendemism for shallow water reef fauna in the Red Sea. *Journal of Biogeography* **43**, 423-439.
- 758

759 DiBattista JD, Choat JH, Gaither MR, Hobbs JP, Lozano-Cortés DF, Myers RF,

- Paulay G, Rocha LA, Toonen RJ, Westneat M, et al. (2016b) On the origin of endemic species in
- the Red Sea. *Journal of Biogeography* **43**, 13-30.
- 762

763 Dos Reis M, Yang Z (2011) Approximate likelihood calculation on a phylogeny for

Bayesian estimation of divergence times. *Molecular Biology and Evolution* **28**, 2161-2172.

765

Eble JA, Toonen RJ, Sorenson L, Basch LV, Papastamatiou YP, Bowen BW (2011) Escaping

paradise: larval export from Hawaii in an Indo-Pacific reef fish, the yellow tang (*Zebrasoma*

- *flavescens*). *Marine Ecology Progress Series* **428**, 245.
- 769
- Faircloth BC (2013) illumiprocessor: a trimmomatic wrapper for parallel adapter and quality
 trimming. DOI: 10.6079/J9ILL.
- 772

Faircloth BC. (2016) PHYLUCE is a software package for the analysis of conserved genomic

774 loci. *Bioinformatics* **32**, 786-788.

- 775
- Faircloth BC, Glenn TC (2012) Not all sequence tags are created equal: designing and validating
 sequence identification tags robust to indels. *PLOS ONE* 7, e42543.
- 778
- 779 Faircloth BC, McCormack JE, Crawford NG, Harvey MG, Brumfield RT, Glenn
- TC (2012) Ultraconserved elements anchor thousands of genetic markers spanning multiple
- 781 evolutionary timescales. *Systematic Biology* sys004.

a dh

- 782
- Faircloth BC, Sorenson L, Santini F, Alfaro ME (2013) A phylogenomic perspective on the
- radiation of ray-finned fishes based upon targeted sequencing of ultraconserved elements
 (UCEs). *PLOS ONE* 8, e65923.
- 786
- 787 Fessler JL, Westneat MW (2007) Molecular phylogenetics of the butterflyfishes
- (Chaetodontidae): taxonomy and biogeography of a global coral reef fish family. *Molecular Phylogenetics and Evolution* 45, 50-68.
- 790

Froese R, Pauly D (2011) FishBase: World Wide Web electronic publication. Available at:
www.fishbase.org (accessed February, 2016).

793

Gaither MR, Toonen RJ, Robertson DR, Planes S, Bowen BW (2010) Genetic evaluation of

marine biogeographical barriers: perspectives from two widespread Indo-Pacific snappers

796 (Lutjanus kasmira and Lutjanus fulvus). Journal of Biogeography **37**, 133-147.

- 797
- 798 Gaither MR, Jones SA, Kelley C, Newman SJ, Sorenson L, Bowen BW (2011)

799 High connectivity in the deepwater snapper *Pristipomoides filamentosus* (Lutjanidae) across the

800 Indo-Pacific with isolation of the Hawaiian Archipelago. *PLOS ONE* **6**, e28913.

- 801
- Gaither MR, Bernal MA, Coleman RR, Bowen BW, Jones SA, Simison WB, Rocha LA (2015)

803 Genomic signatures of geographic isolation and natural selection in coral reef fishes. *Molecular*

804 *Ecology* **24**, 1543-1557.

000	
806	Girdler RW, Styles P (1974) Two-stage Red Sea floor spreading. Nature 247, 7-11.
807	
808	Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, Adiconis X, Fan
809	L, Raychowdhury R, Zeng Q, et al. (2011) Full-length transcriptome assembly from RNA-seq
810	data without a reference genome. Nature Biotechnology 29, 644-652.
811	
812	Harrington RC, Faircloth BC, Eytan RI, Smith WL, Near TJ, Alfaro ME, Friedman M (2016)
813	Phylogenomic analysis of carangimorph fishes reveals flatfish asymmetry arose in a blink of the
814	evolutionary eye. BMC Evolutionary Biology 16, 224.
815	
816	Harris RS (2007) Improved pairwise alignment of genomic DNA. Ph.D. Thesis. Pennsylvania
817	State University.
818	
819	Hawkins JP, Roberts CM, Clark V (2000) The threatened status of restricted-range coral reef fish
820	species. Animal Conservation 3, 81-88.
821	
822	Ho LST, Ane C, Lachlan R, Tarpinian K, Feldman R, Ho MLST (2016) Package 'phylolm'.
823	
824	Hobbs JPA, Jones GP, Munday PL, Connolly SR, Srinivasan M (2012) Biogeography and
825	the structure of coral reef fish communities on isolated islands. Journal of Biogeography 39,
826	130-139.
827	
828	Hodge JR, Herwerden L, Bellwood DR (2014) Temporal evolution of coral reef fishes: global
829	patterns and disparity in isolated locations. Journal of Biogeography 41, 2115-2127.
830	
831	Hsu KC, Chen JP, Shao KT (2007) Molecular phylogeny of Chaetodon (Teleostei:
832	Chaetodontidae) in the Indo-West Pacific: evolution in geminate species pairs and species
833	groups. Raffles Bulletin of Zoology 14, 77-86.
834	

835	Hubert-Ferrari A, King G, Manighetti I, Armijo R, Meyer B, Tapponnier P (2003)
836	Long-term elasticity in the continental lithosphere; modelling the Aden Ridge propagation and
837	the Anatolian extrusion process. Geophysical Journal International 153, 111-132.
838	
839	Kahng SE, Garcia-Sais JR, Spalding HL, Brokovich E, Wagner D, Weil E, Hinderstein L,
840	Toonen RJ (2010) Community ecology of mesophotic coral reef ecosystems. Coral Reefs 29,
841	255-275.
842	()
843	Katoh K, Misawa K, Kuma K, Miyata T (2002) MAFFT: a novel method for rapid multiple
844	sequence alignment based on fast Fourier transform. Nucleic Acids Research 30, 3059-3066.
845	
846	Katoh K, Standley DM. 2013 MAFFT multiple sequence alignment software version 7:
847	improvements in performance and usability. Molecular Biology and Evolution 30, 772-780.
848	
849	Kemp J (1998) Zoogeography of the coral reef fishes of the Socotra Archipelago. Journal of
850	<i>Biogeography</i> 25 , 919-933.
851	
852	Keith SA, Woolsey ES, Madin JS, Byrne M, Baird AH (2015) Differential establishment
853	potential of species predicts a shift in coral assemblage structure across a biogeographic barrier.
854	<i>Ecography</i> 38 , 1225-1234.
855	
856	Klausewitz W (1989) Evolutionary history and zoogeography of the Red Sea ichthyofauna.
857	Fauna of Saudi Arabia 10, 310-337.
858	
859	Kuiter RH (2002) Butterflyfishes, Bannerfishes and their Relatives: A Comprehensive Guide to
860	Chaetodontidae and Microcanthidae. TMC Publishing, Chorleywood, UK.
861	
862	Lambeck K (1996) Shoreline reconstructions for the Persian Gulf since the last glacial
863	maximum. Earth and Planetary Science Letters 142, 43-57.
864	

Lomolino MV (2005) Body size evolution in insular vertebrates: generality of the island rule. *Journal of Biogeography* 32, 1683-1699.

867

Ludt WB, Rocha LA (2015) Shifting seas: the impacts of Pleistocene sea-level fluctuations on the evolution of tropical marine taxa. *Journal of Biogeography* **42**, 25-38.

870

.

- Luiz OJ, Madin JS, Robertson DR, Rocha LA, Wirtz P, Floeter SR (2012) Ecological traits
- 872 influencing range expansion across large oceanic dispersal barriers: insights from tropical

Atlantic reef fishes. *Proceedings of the Royal Society B* **279**, 1033-1040.

874

Luiz OJ, Allen AP, Robertson DR, Floeter SR, Kulbicki M, Vigliola L, Becheler R,

876 Madin JS (2013) Adult and larval traits as determinants of geographic range size among tropical

reef fishes. *Proceedings of National Academy of Sciences U.S.A.* **110**, 16498-16502.

878

Malay MCMD, Paulay G (2010) Peripatric speciation drives diversification and distributional
pattern of reef hermit crabs (Decapoda: Diogenidae: *Calcinus*). *Evolution* 64, 634-662.

881

882 Matzke NJ (2013) Probabilistic historical biogeography: new models for founder-event

speciation, imperfect detection, and fossils allow improved accuracy and model-testing.

884 Frontiers in Biogeography 4, 210.

885

886 McCormack JE, Harvey MG, Faircloth BC, Crawford NG, Glenn TC, Brumfield RT (2013) A

phylogeny of birds based on over 1,500 loci collected by target enrichment and high-throughput
sequencing. *PLOS ONE* 8, e54848.

889

890 McGee MD, Faircloth BC, Borstein SR, Zheng J, Darrin Hulsey C, Wainwright PC, Alfaro ME

- 891 (2016) Replicated divergence in cichlid radiations mirrors a major vertebrate innovation.
- 892 *Proceedings of the Royal Society B* **283**.
- 893

McIlwain JL, Claereboudt MR, Al-Oufi HS, Zaki S, Goddard JS (2005) Spatial variation in age

895	and growth of the kingfish (Scomberomorus commerson) in the coastal waters of the Sultanate of
896	Oman. Fisheries Research 73, 283-298.

- 897
- McKinney ML (1997) Extinction vulnerability and selectivity: combining ecological and paleontological views. *Annual Review of Ecology and Systematics* **28**, 495-516.
- 900
- 901 Michonneau F (2015) Cryptic and not-so-cryptic species in the complex "Holothuria
- 902 (*Thymiosycia*) *imaptiens*" (Forsskål, 1775)(Echinodermata: Holothuroidea: Holothuriidae).
- 903 *bioRxiv*. DOI: 10.1101/014225.

904

Ottimofiore, E, Albouy, C, Leprieur, F, Descombes, P, Kulbicki, M, Mouillot, D, Parravicini, V,
Pellissier L (2017) Responses of coral reef fishes to past climate changes are related to lifehistory traits. *Ecology and Evolution* 7, 1996-2005.

908

Pous S, Lazure P, Carton X (2015) A model of the general circulation in the Persian Gulf and in
the Strait of Hormuz: Intraseasonal to interannual variability. *Continental Shelf Research* 94, 5570.

912

913 Pratchett MS (2014) Feeding preferences and dietary specialisation among obligate coral-

- feeding butterflyfishes. In: Pratchett MS, Berumen ML, Kapoor BG (eds) *Biology of*
- 915 *Butterflyfishes*. CRC Press, Boca Raton, FL, USA, pp. 140–179.
- 916
- 917 Racault MF, Raitsos DE, Berumen ML, Brewin RJ, Platt T, Sathyendranath S, Hoteit I (2015)

918 Phytoplankton phenology indices in coral reef ecosystems: application to ocean-color

observations in the Red Sea. *Remote Sensing of Environment* **160**, 222-234.

- 920
- Raitsos DE, Pradhan Y, Brewin RJW, Stenchikov G, Hoteit I (2013) Remote sensing the
- phytoplankton seasonal succession of the Red Sea. *PLOS ONE* **8**, e64909.
- 923

924 Rambaut A, Suchard MA, Xie D, Drummond AJ (2014) Tracer v1.6,

925

http://beast.bio.ed.ac.uk/Tracer.

- 926
- 927 Reneker J, Lyons E, Conant GC, Pires JC, Freeling M, Shyu CR, Korkin D (2012) Long
- 928 identical multispecies elements in plant and animal genomes. *Proceedings of the National*
- 929 Academy of Sciences U.S.A. 109, E1183-E1191.
- 930
- 831 Roberts MB, Jones GP, McCormick MI, Munday PL, Neale S, Thorrold S, Robitzch VS,
- Berumen ML (2016) Homogeneity of coral reef communities across 8 degrees of latitude in the
- 933 Saudi Arabian Red Sea. *Marine Pollution Bulletin* **105**, 558-565.
- 934

Rohling EJ, Grant K, Bolshaw M, Roberts AP, Siddall M, Hemleben C, Kucera M (2009)

- 936 Antarctic temperature and global sea level closely coupled over the past five glacial cycles.
- 937 *Nature Geoscience* **2**, 500-504.

.

- 938
- 939 Schuster F (2002) Oligocene and Miocene examples of *Acropora*-dominated
- 940 palaeoenvironments: Mesohellenic Basin (NW Greece) and northern Gulf of Suez (Egypt).
- 941 *Proceedings of the 9th International Coral Reef Symposium Bali* **1**, 199-204.
- 942
- Shi W, Morrison JM, Bohm E, Manghnani V (2000) The Oman upwelling zone during 1993,
 1994 and 1995. *Deep-Sea Research II* 47, 1227-1247.
- 945
- 946 Siddall M, Rohling EJ, Almogi-Labin A, Hemleben C, Meischner D, Schmelzer I, Smeed
- DA (2003) Sea-level fluctuations during the last glacial cycle. *Nature* **423**, 853-858.
- 948
- 949 Siepel A, Bejerano G, Pedersen JS, Hinrichs AS, Hou M, Rosenbloom K, Clawson H,
- 950 Spieth J, Hillier LW, Richards S, et al. (2005) Evolutionarily conserved elements in vertebrate,
- insect, worm, and yeast genomes. *Genome Research* **15**, 1034-1050.
- 952
- Simons C, Pheasant M, Makunin IV, Mattick JS (2006) Transposon-free regions in
- mammalian genomes. *Genome Research* **16**, 164-172.

- 956 Skillings DJ, Bird CE, Toonen RJ (2010) Gateways to Hawai'i: genetic population 957 structure of the tropical sea cucumber Holothuria atra. Journal of Marine Biology 2011. and the 958 Smeed D (1997) Seasonal variation of the flow in the strait of Bah al Mandab. Acta 959 Oceanologica 20, 773-781. 960 961 Smith BT, McCormack JE, Cuervo AM, Hickerson MJ, Aleixo A, Cadena CD, Pérez-962 963 Emán J, Burney CW, Xie X, Harvey MG, et al. (2014) The drivers of tropical speciation. Nature 515, 406-409. 964 965 Sun K, Meiklejohn KA, Faircloth BC, Glenn TC, Braun EL, Kimball RT (2014) The evolution 966 of peafowl and other taxa with ocelli (eyespots): a phylogenomic approach. Proceedings of the 967 Royal Society B 281, 20140823. 968 969 Talavera G, Castresana J (2007) Improvement of phylogenies after removing divergent and 970 ambiguously aligned blocks from protein sequence alignments. Systematic Biology 56, 564-577. 971 972 973 Therneau T, Atkinson B, Ripley B (2015) 'rpart'. R package version 4.1-10. 974 Whittaker RJ, Fernández-Palacios JM (2007) Island biogeography: ecology, evolution, and 975 976 conservation. Oxford University Press. 977 Yabuta S, Berumen ML (2013) Social structures and spawning behaviour of Chaetodon 978 979 butterflyfishes. In Biology of Butterflyfishes, 200-225. 980 981 982 983
- 984 Figure Legends

Figure 1. Inferred phylogeny of Red Sea to Arabian Gulf butterflyfish species, including some
of closest their congeners, based on ExaBayes analysis of ultraconserved element (UCE) data.
Yellow dots on node labels indicate a posterior probability of 1, whereas grey dots indicate a
posterior probability of less than 1 but greater than 0.6. Clades based on Bellwood et al. (2010)
and Cowman & Bellwood (2011) are indicated. Records for each species are mapped onto the
topology as follows: red = Red Sea to Arabian Gulf, green = rest of Indian Ocean, and blue =
Pacific Ocean (PO).

992

Figure 2. A fossil calibrated chronogram for select Chaetodontidae species based on analysis of
ultraconserved element (UCE) data. The time scale is calibrated in millions of years before
present. Node ages are presented as median node heights with 95% HPD intervals represented by
bars. Significant geological events in the coastal waters of the Arabian Peninsula are temporally
indicated by red dashed lines.

998

Figure 3. Distributions, range overlap, and ages of divergence in eight clades of butterflyfish
from the *Chaetodon* genus that contain species inhabiting the Red Sea to Arabian Gulf region.
Clade structure and node ages (median node heights with 95% HPD intervals represented by
bars) were extracted from Figure 2.

1003

1004 Figure 4. Ancestral range estimations inferred using the DEC + J model based on a time 1005 calibrated Bayesian phylogeny of Chaetodontidae species. States at branch tips indicate the current geographical distributions of taxa, whereas states at nodes indicate the inferred ancestral 1006 1007 distributions before speciation (middle) and after (corner). The regions considered in this analysis include: Gulf of Agaba, rest of Red Sea, Djibouti and Gulf of Aden, Socotra, South 1008 1009 Oman, Arabian Gulf, Gulf of Oman and Pakistan, rest of Indian Ocean, and Pacific Ocean. 1010 Abbreviations: Plio. = Pliocene; Ple. = Pleistocene. Significant vicariance in the Red Sea to 1011 Arabian Gulf region are indicated by red dashed lines.

1012

Figure 5. Estimated probability of endemism among Red Sea to Arabian Gulf butterflyfish
species, including some of their closest congeners, as a function of depth range. Different line

1015 types represent variability in estimated species phylogenetic age extracted from Figure 2 (see1016 legend).

1017

Figure 6. The classification of species-level traits associated with endemism among the Red Sea
to Arabian Gulf butterflyfishes (a). Data on the leaves (represented by squares) provide the
probability of endemism (top) and the percentage of all observations in the node (bottom). The

1021 right panel shows the prediction surface (b).

1022

1023 Data accessibility

1024 Data associated with this manuscript are available under NCBI BioProject PRJNA484421,

available at https://www.ncbi.nlm.nih.gov/bioproject/PRJNA484421.

1026

1027 Authors' contributions

-

- 1028 J.D.D., M.E.A., L.A.R., J.H.C., and M.L.B. designed the study; J.D.D., L.S., J.P.A.H., T.H.S.,
- 1029 L.A.R., and M.L.B. collected samples; J.D.D. and L.S. generated the UCE libraries; J.D.D.,
- 1030 M.E.A., J.C., O.J.L., and P.F.C. analysed and interpreted data; M.F. calibrated tree
- 1031 reconstructions; J.D.D. wrote the manuscript with input from all co-authors.

Author

Table 1. Species distribution and clade designation from Bellwood et al. (2010) and Cowman & Bellwood (2011) for all

1033 Chaetodontidae samples used in this study. Colours in the table header match the colors used to denote species distributions in Figure

1034 1. Asterisks indicate regional endemics for the purposes of our correlational trait analysis. The letters below each region indicate the

1035 geographic groupings used for BioGeoBEARS analysis. Although Chaetodon leucopleura, Chaetodon melapturus, and Chaetodon

1036 *pictus* are listed as being present in the Red Sea, this is based on rare records at their southern limits. Similarly, we have only sampled

1037 *C. pictus* (and not *Chaeotdon vagabundus*) at Socotra (DiBattista *et al.* 2017), and rare records of *Chaetodon austriacus* in the Gulf of

1038 Aden and South Oman likely represent waifs.

Species	Geographic distribution								
	Gulf of	Rest of Red	Djibouti and	Socotra	South Oman	Arabian	Gulf of	Rest of	Pacific
	Aqaba	Sea	Gulf of	(D)	(E)	Gulf	Oman and	Indian	Ocean
	(A)	(B)	Aden			(F)	Pakistan	Ocean	(I)
Clade 4									
Chaetodon auriga	\checkmark			\checkmark	\checkmark		\checkmark	\checkmark	\checkmark
Chaetodon auripes									\checkmark
Chaetodon collare				\checkmark	\checkmark			\checkmark	\checkmark
Chaetodon decussatus								\checkmark	
Chaetodon dialeucos*					\checkmark				
Chaetodon falcula								\checkmark	
Chaetodon fasciatus*	\checkmark	\checkmark							
Chaetodon gardineri*					\checkmark		\checkmark	\checkmark	
Chaetodon leucopleura		\checkmark						\checkmark	
Chaetodon lineolatus	\checkmark	\checkmark						\checkmark	\checkmark
Chaetodon lunula				\checkmark	\checkmark			\checkmark	\checkmark
Chaetodon melannotus	\checkmark							\checkmark	\checkmark
Chaetodon mesoleucos*	\checkmark	\checkmark							
Chaetodon nigropunctatus*						\checkmark			
Chaetodon oxycephalus								\checkmark	\checkmark
Chaetodon pictus*		\checkmark		\checkmark	\checkmark				
Chaetodon semilarvatus*	\checkmark	\checkmark			\checkmark				
Chaetodon vagabundus				\checkmark				\checkmark	\checkmark

Clade 3

Chaetodon austriacus* Chaetodon baronessa	\checkmark	\checkmark	\checkmark		\checkmark				
Chaetodon bennetti Chaetodon larvatus*	\checkmark	\checkmark	\checkmark		\checkmark			\checkmark	
Chaetodon lunulatus Chaetodon melapterus* Chaetodon plebeius		\checkmark	\checkmark	\checkmark	\checkmark	\checkmark	\checkmark		
Chaetodon speculum Chaetodon triangulum									
Chaetodon trifascialis Chaetodon trifasciatus Chaetodon zanzibariensis	\checkmark	\checkmark	イイ	$\sqrt{1}$	\checkmark			シン	V
<u>Clade 2</u> Chaetodon guttatissimus Chaetodon interruptus				\checkmark				$\sqrt{1}$	
Chaetodon kleinii Chaetodon madagaskariensis Chaetodon mertensii Chaetodon pauoifaraiatus*	al	al	al	V				$\sqrt{1}$	
Chaetodon paucijaseumis Chaetodon pelewensis Chaetodon trichrous Chaetodon unimaculatus Chaetodon xanthurus	v	v	,						\sim \sim \sim \sim \sim \sim
Bannerfishes									v
Forcipiger flavissimus Forcipiger longirostris		\checkmark		$\sqrt{1}$	I	1	I	$\sqrt{1}$	
Heniochus acuminatus Heniochus diphreutes Heniochus intermedius*	$\sqrt[n]{\sqrt{1}}$	$\sqrt[n]{\sqrt{2}}$	$\frac{1}{\sqrt{2}}$	$\sqrt[n]{}$	N	Ν	N	$\sqrt[n]{}$	$\sqrt[n]{\sqrt{1-1}}$
1039									
1040									
1041									
1042									

استعاد

Table 2. Akaike information criterion (AIC) model testing based on distribution patterns for butterflyfish lineages using the timecalibrated phylogeny analysed with the R module BioGeoBEARS, where d represents the dispersal parameter, e represents the
extinction parameter, and j represents founder-event speciation. For these models we coded each taxon based on presence/absence in
nine discrete geographical areas: a) Gulf of Aqaba, b) rest of Red Sea, c) Djibouti and Gulf of Aden, d) Socotra, e) South Oman, f)
Arabian Gulf, g) Gulf of Oman and Pakistan, h) rest of Indian Ocean and i) Pacific Ocean. Grey shading indicates the favoured model
based on AIC scores.

la	Ln Likelihood	Number of parameters	d	e	j	AIC	AIC weight
DEC	-255.13	2	0.06	0	0	514.25	0.03
DEC+J	-250.79	3	0.05	0	0.04	507.58	0.76
DIVALIKE	-253.88	2	0.07	0.04	0	511.76	0.09
DIVALIKE+J	-252.76	3	0.06	0.02	0.03	511.52	0.11
BAYAREALIKE	-259.86	2	0.05	0.18	0	523.71	0
BAYAREALIKE+J	-255.48	3	0.04	0.08	0.06	516.96	0.01

Table 3. Summary of the final (best) phylogenetic, linear multi-regression model, based on estimated probability of endemism as a response variable, selected after the backward stepwise phylostep procedure. Coefficients in bold indicate significance (p < 0.05).

		Estimate	Std. err.	z value	<i>p</i> value	_
	(Intercept)	6.170	2.506	2.461	0.013	
	Depth range	-1.423	0.543	-2.620	0.008	
	Phylogenetic age	-1.209	0.694	-1.742	0.061	
70						

























log Depth Range (m)



H. intermedius

(b)

