## Cytogenomic array detects a subset of myelodysplastic syndrome with increased risk that is invisible to conventional karyotype

Sarah M. Choi<sup>1\*</sup>, Steven B. Van Norman<sup>1\*</sup>, Dale L. Bixby<sup>2</sup>, and Lina Shao<sup>1#</sup>

<sup>1</sup>Department of Pathology, University of Michigan, Ann Arbor, MI <sup>2</sup>Department of Internal Medicine, University of Michigan, Ann Arbor, MI

\*SMC and SBV contributed equally to this work.

Running head: SNP-A detects an increased-risk subset of MDS

### Correspondence

Lina Shao, PhD, Michigan Medicine, Department of Pathology, 2800 Plymouth Road, Building 36, Room 1361-2, Ann Arbor, MI 48109-2800 Fax 734-232-5360 Email: linashao@med.umich.edu

KEYWORDS: myelodysplastic syndrome, cytogenomic array, SNP array, MDS, SNP-A

Funding information: Not applicable.

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/gcc.22783

### Abstract

Conventional karyotyping is essential standard practice in the initial evaluation of myelodysplastic syndrome (MDS) and is the most impactful single component of the Revised International Prognostic Scoring System (IPSS-R). While single nucleotide polymorphism array (SNP-A) has demonstrated the ability to detect chromosomal defects with greater sensitivity than conventional karyotype, widespread adoption is limited by the unknown additional prognostic impact of SNP-A analysis. Here we investigate the significance of additional SNP-A abnormalities in the setting of MDS and demonstrate differences in survival of patients with additional abnormalities, even those initially characterized as relatively lower risk either by cytogenetic score or IPSS-R. Our findings identify specific abnormalities, particularly *KMT2A* partial tandem duplication (*KMT2A*-PTD), that are invisible to conventional karyotype and potentially contribute to the poor prognosis of MDS patients. Furthermore, these results demonstrate the added value of SNP-A analysis in identifying patients who may benefit from more aggressive therapy, particularly those who would otherwise be classified into lower risk categories.

KEYWORDS: myelodysplastic syndrome, cytogenomic array, SNP array, MDS, SNP-A

### **INTRODUCTION**

Prognostic classification of myelodysplastic syndrome (MDS) relies heavily on cytogenetic abnormalities and is currently utilized to guide therapeutic decision making, including identifying appropriate candidates for bone marrow transplantation. Indeed, cytogenetic risk is the most heavily weighted component of the Revised International Prognostic Scoring System (IPSS-R), which categorizes patients into multiple risk groups of ascending associated poor prognosis.<sup>1</sup> While conventional karyotype is the gold standard for detection of genomic abnormalities in both diagnostic and prognostic settings, single nucleotide polymorphism arrays (SNP-As) have emerged as potential means of further categorizing prognostic risk beyond traditional karyotyping in many hematologic malignancies due to the assay's greater sensitivity in detecting unbalanced chromosomal defects and copy-neutral loss of heterozygosity (CN-LOH).<sup>2-6</sup> However, widespread adoption and incorporation into prognostic algorithms has not yet occurred despite evidence of the clinical significance of SNP-A in combination with already established karyotypic features.<sup>5-8</sup> We therefore sough to refine our understanding of the significance of additional SNP-A abnormalities and their impact on prognosis and ultimately risk of death. In particular, given the high frequency of MDS cases showing a normal karyotype,<sup>9,10</sup> we were especially interested in the potential impact of SNP-A in this group of patients.

### MATERIALS AND METHODS

This study was approved by the Institutional Review Board of the University of Michigan. We retrospectively reviewed 108 consecutive patients who underwent karyotyping and Thermo Fisher Cytoscan® array (SNP-A) analysis<sup>11</sup> for diagnosis/classification of a suspected myeloid neoplasm and identified 77 patients with a diagnosis of *de novo* MDS, excluding therapy-related cases. Among these, we identified cases for which additional abnormalities were detected by SNP-A analysis that were not identified using conventional karyotype. We reviewed each patient's electronic medical record including laboratory values at diagnosis (hemoglobin, absolute neutrophil count (ANC), platelet count, and bone marrow blast percentage). We then compared overall survival based on the presence or absence of additional cytogenomic abnormalities detected by SNP-A on groups that were stratified by cytogenetic

risk and IPSS-R score (Figure 1). Overall survival (OS) was calculated from date of diagnosis to date of death, censoring for patients alive at the completion of the study. Patients with both low-risk karyotype (very good-intermediate cytogenetic risk group) and consistent SNP-A results were compared to patients with similar low-risk karyotype but with additional SNP-A abnormalities. These groups were also stratified by the IPSS-R, and survival was compared in patients with and without additional SNP-A abnormalities. Unpaired t, Mann-Whitney, Chi-Square, and Fisher's tests were used as applicable to compare differences in characteristics between groups with and without additional abnormalities. Log-rank (Mantle-Cox) test was used to compare overall survival between the groups.

### **RESULTS AND DISCUSSION**

Of the 77 patients for whom both karyotype and SNP-A were performed (Table 1), 36 cases had additional abnormalities detected by SNP-A (47% of all cases; Table 2; Figure 2). Follow up time ranged from 1.5-85 months (Table 1). Deletions were the most common finding (29 instances), followed by CN-LOH (19 instances). The most prevalent single abnormality detected was *KMT2A(MLL)* partial tandem duplication (*KMT2A*-PTD) (5 cases). Additional abnormalities included cryptic deletions involving *TET2* (3), *RUNX1* (2), and *CUX1* (1). SNP-A detected a monosomy 7 in two cases whose conventional cytogenetics showed a normal karyotype after examining adequate number of metaphase cells.

Cases with and without additional SNP-A abnormalities showed similar overall distribution of morphologic categorization and no statistically significant differences in hemoglobin, ANC, platelet count, bone marrow blast percentage, or IPSS-R (Table 1; Supporting Information Figure S1).

Among matched cases with very good-intermediate cytogenetic risk, those that had additional abnormalities detected on SNP-A showed worse overall survival (median 35.4 months) than those that did not have additional abnormalities (median survival not reached) (Figure 3A; P= 0.010). Similar observations were seen when cases were stratified into matching IPSS-R categories, where very low-intermediate risk cases with additional SNP-A abnormalities showed worse overall survival (median 35.4 months) similar to cases of high-very high risk (median 31.3 months) compared to very low-intermediate risk cases without additional SNP-A abnormalities (median 62.6 months) (Figure 3B; P =0.020).

On the contrary, in cases with either poor-very poor cytogenetic risk or high-very high IPSS-R score, the presence of additional SNP-A abnormalities did not show a statistically significant impact on

overall survival though a trend towards poorer survival was observed (Figures 3A, P = 0.054 and 3B, P = 0.052). This finding suggests that the prognostic impact of SNP-A abnormalities may be primarily concentrated in cases that would otherwise be classified as lower risk either by cytogenetic score or IPPS-R.

We questioned whether or not incorporating additional SNP-A findings would impact the calculated IPSS-R. Taking into consideration cases with anomalies that were greater than 5 Mb excluding CN-LOH and small focal deletions and gains, only 2 cases changed IPSS-R (one from very low to intermediate; one from very low to low). In terms of our analysis, therefore, there was no impact on survival.

Based on our previous observation of a small cohort of low risk MDS cases of *KMT2A*-PTD which showed poor overall survival,<sup>12</sup> we questioned whether this single abnormality could be responsible for the survival differences of the cases with additional SNP-A abnormalities. When *KMT2A*-PTD cases were excluded from the analysis, the effects of additional SNP-A abnormalities when stratifying patients according to cytogenetic risk and IPSS-R were somewhat abrogated. A trend towards poorer survival was still noticeable, but no longer statistically significant (Figure 4A; P = 0.069 and Figure 4B; P = 0.063). Consequently, *KMT2A*-PTD may be at least partly responsible for the worse survival seen in patients with additional SNP-A abnormalities and otherwise very good-intermediate cytogenetic risk or very low-intermediate IPSS-R, though other abnormalities may also contribute to a lesser extent. Genomic locations of *KMT2A*-PTD in five cases are shown in Figure 5.

To summarize, our findings suggest that the presence of additional SNP-A abnormalities, detected in almost half of MDS cases, has further impact on prognosis and overall survival than that afforded by conventional karyotype analysis. Cases identified as very good-intermediate cytogenetic risk that have additional SNP-A abnormalities demonstrate overall survival approaching that of patients with poor-very poor cytogenetic risk. Similarly, cases identified as very low-intermediate risk by IPSS-R demonstrate overall survival more similar to patients with high-very high risk by IPSS-R. Although a significant component of these differences may be accounted for by the presence of *KMT2A*-PTD, the data imply that other additional abnormalities also impact the prognosis of MDS.

Although the types of abnormalities we noted in our study were largely similar to those reported in previous studies analyzing SNP-A in MDS patients,<sup>4-7</sup> the frequency of additional SNP-A abnormalities in our study of 47% was higher than previous reports, which showed a range of 18-39% of

cases with additional SNP-A abnormalities. The increased frequency we observed could potentially be due to higher resolution of our array platform and as well as our analysis. Coincident with this observation, *KMT2A*-PTD was only rarely reported in these prior studies, whereas in our study this abnormality was the most common additional SNP-A abnormality observed.

The data also demonstrate that the effect of additional SNP-A abnormalities is particularly profound in cases that have very good-intermediate karyotypic features or cases of otherwise low-intermediate risk by IPSS-R. Thus, the SNP-A may be more effectively utilized in these particular cases and, perhaps not surprisingly, is more limited in usefulness in cases that already have definitive poor risk features. Moreover, because a large proportion of cases in the very low-intermediate risk/very good-intermediate cytogenetic groups are patients with a normal karyotype, SNP-A is a potentially useful tool in further delineating risk subgroups within this substantial fraction of MDS cases. Interestingly, the survival of patients in our study was longer than those of prior studies that performed survival analysis,<sup>5,7</sup> which showed a median survival of 43-50 months in patients with favorable or normal karyotype without SNP-A abnormalities and 16-20 months in patients with additional SNP-A abnormalities. The relatively increased overall survival seen in our cohort compared to prior studies may potentially be attributed to either differences in patient population, clinical practice, or a consequence of relatively smaller cohort size.

The contribution of individual specific abnormalities to prognosis also remains an area of future investigation. Our data suggest *KMT2A*-PTD may be partially responsible for the poorer survival seen in MDS patients, who otherwise might be classified as having lower risk disease, though a definitive determination is limited by the small number of *KMT2A*-PTD. Additionally, two cases also identified CN-LOH as additional anomalies on chromosome arms 7q and 11q, respectively, which contain the genes *EZH2* on 7q and *CBL* on 11q, both recurrently mutated in MDS with suggested adverse prognostic impact.<sup>13-16</sup> Consequently, the relative contribution of *KMT2A*-PTD to prognosis requires further confirmation in larger sample study. Because patients with additional SNP-A abnormalities have poorer overall survival and relatively increased risk, the presence of these abnormalities may potentially identify these patients as candidates for more urgent therapeutic intervention including transplantation.

While conventional karyotype continues as expected standard practice in MDS diagnosis, the appropriate utilization of other related ancillary testing to provide a comprehensive genetic assessment is an important as-yet unresolved issue. For example, fluorescence in situ hybridization (FISH) analysis can

also detect additional abnormalities outside of karyotype,<sup>17,18</sup> however the National Comprehensive Cancer Network (NCCN) practice guidelines currently only recommend FISH in cases where standard cytogenetics cannot be obtained.<sup>19</sup> The American Society for Clinical Pathology also supports a similar recommendation.<sup>20</sup> The European Society for Medical Oncology (ESMO) practice guidelines slightly differ in this respect as they acknowledge a potential benefit of FISH in cytogenetically normal cases and thus recommend FISH in the setting of normal karyotype.<sup>21</sup> A similar algorithm could potentially be applied with respect to SNP-A by focusing its utilization in cases that would otherwise be designated of lower risk. Recent studies have attempted to compare the relative detection rates of abnormalities across different modalities and have confirmed overall relatively higher resolution in SNP-A<sup>22</sup> and next generation sequencing (NGS) platforms which complement standard cytogenetics and FISH analysis.<sup>23</sup> Outside of larger genomic alterations, the impact of point mutations, which have also been implicated in survival of MDS patients with further prognostic prognostic relevance, <sup>15,24</sup> remains another area for subsequent investigation. As clinical standard of care does not yet include testing for point mutations, we were unable to ascertain their significance in the context of additional SNP-A abnormalities, but this is an area in which further analysis is warranted.

In conclusion, these findings demonstrate the potential prognostic and therapeutic impact of the cytogenomic array, with particular utility in MDS cases that would otherwise be classified as very low-intermediate risk.

### REFERENCES

- 1. Greenberg PL, Tuechler H, Schanz J, et al. Revised international prognostic scoring system for myelodysplastic syndromes. *Blood*. 2012;120(12):2454-2465.
- 2. Li MM, Monzon FA, Biegel JA, et al. A multicenter, cross-platform clinical validation study of cancer cytogenomic arrays. *Cancer Genet*. 2015;208(11):525-536.
- Maciejewski JP, Tiu RV, O'Keefe C. Application of array-based whole genome scanning technologies as a cytogenetic tool in haematological malignancies. *Br J Haematol.* 2009;146(5):479-488.
- 4. Stevens-Kroef MJ, Olde Weghuis D, Elldrissi-Zaynoun N, et al. Genomic array as compared to karyotyping in myelodysplastic syndromes in a prospective clinical trial. *Genes Chromosomes Cancer*. 2017;56(7):524-534.
- 5. Thiel A, Beier M, Ingenhag D, et al. Comprehensive array CGH of normal karyotype myelodysplastic syndromes reveals hidden recurrent and individual genomic copy number alterations with prognostic relevance. *Leukemia*. 2011;25(3):387-99.
- Mohamedali A, Gäken J, Twine NA, et al. Prevalence and prognostic significance of allelic imbalance by single-nucleotide polymorphism analysis in low-risk myelodysplastic syndromes. *Blood*. 2007;110(9):3365-3373.
- Tiu RV, Gondek LP, O'Keefe CL, et al. Prognostic impact of SNP array karyotyping in myelodysplastic syndromes and related myeloid malignancies. *Blood*. 2011;117(17):4552-4560.
- Starczynowski DT, Vercauteren S, Telenius A, et al. High-resolution whole genome tiling path array CGH analysis of CD34+ cells from patients with low-risk myelodysplastic syndromes reveals cryptic copy number alterations and predicts overall and leukemia-free survival. *Blood*. 2008;112(8):3412-3424.
- 9. Deeg HJ, Scott BL, Fang M, et al. Five-group cytogenetic risk classification, monosomal karyotype, and outcome after hematopoietic cell transplantation for MDS or acute leukemia evolving from MDS. *Blood*. 2012;120(7):1398-1408.
- Schanz J, Tuchler H, Sole F, et al. New comprehensive cytogenetic scoring system for primary myelodysplastic syndromes (MDS) and oligoblastic acute myeloid leukemia after MDS derived from an international database merge. *J Clin Oncol.* 2012;30(8):820-829.

- Wang Y, Miller S, Roulston D, Bixby D, Shao L. Genome-wide single-nucleotide polymorphism array analysis improves prognostication of acute lymphoblastic leukemia/lymphoma. *J Mol Diagnostics*. 2016;18(4):595-603.
- Choi SM, Dewar R, Burke PW, Shao L. Partial tandem duplication of KMT2A (MLL) may predict a subset of myelodysplastic syndrome with unique characteristics and poor outcome. *Haematologica*. 2018;103(3):e131-e134.
- 13. Martin I, Such E, Navarro B, et al. Prognostic impact of gene mutations in myelodysplastic syndromes with ring sideroblasts. *Blood Cancer J*. 2017;7(12):630.
- Xu L, Gu Z-H, Li Y, et al. Genomic landscape of CD34+ hematopoietic cells in myelodysplastic syndrome and gene mutation profiles as prognostic markers. *Proc Nat Acad Sci*. 2014;111(23):8589-8594.
- Bejar R, Stevenson K, Abdel-Wahab O, et al. Clinical effect of point mutations in myelodysplastic syndromes. *N Engl J Med.* 2011;364(26):2496-2506.
- Hou H-A, Tsai C-H, Lin C-C, et al. Incorporation of mutations in five genes in the revised International Prognostic Scoring System can improve risk stratification in the patients with myelodysplastic syndrome. *Blood Cancer J.* 2018;8(4):39.
- Braulke F, Jung K, Schanz J, et al. Molecular cytogenetic monitoring from CD34+ peripheral blood cells in myelodysplastic syndromes: first results from a prospective multicenter German diagnostic study. *Leuk Res.* 2013;37(8):900-906.
- Coleman JF, Theil KS, Tubbs RR, Cook JR. Diagnostic yield of bone marrow and peripheral blood FISH panel testing in clinically suspected myelodysplastic syndromes and/or acute myeloid leukemia: a prospective analysis of 433 cases. *Am J Clin Pathol*. 2011;135(6):915-920.
- 19.(U.S.) NCCN. Myelodysplastic Syndrome (Version 2.2019).<a href="https://www.nccn.org/professionals/physician\_gls/PDF/mds.pdf">https://www.nccn.org/professionals/physician\_gls/PDF/mds.pdf</a>.
- 20. Pathology ASfC. 2016; <u>http://www.choosingwisely.org/clinician-lists/american-society-clinical-</u>pathology-fish-for-myelodyplastic-syndrome-related-abnormalities/.
- Fenaux P, Haase D, Sanz GF, Santini V, Buske C, Group EGW. Myelodysplastic syndromes: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2014;25 Suppl 3:iii57-69.

----r Manuscrip Autho

- 22. Makishima H, Rataul M, Gondek LP, et al. FISH and SNP-A karyotyping in myelodysplastic syndromes: improving cytogenetic detection of del(5q), monosomy 7, del(7q), trisomy 8 and del(20q). *Leuk Res.* 2010;34(4):447-453.
- 23. Mukherjee S, Sathanoori M, Ma Z, et al. Addition of chromosomal microarray and next generation sequencing to FISH and classical cytogenetics enhances genomic profiling of myeloid malignancies. *Cancer Genet*. 2017;216-217:128-141.
- 24. Haferlach T, Nagata Y, Grossmann V, et al. Landscape of genetic lesions in 944 patients with myelodysplastic syndromes. *Leukemia*. 2014;28(2):241-247.

### **Figure Legends**

FIGURE 1 Study design. MDS cases for which both karyotyping and cytogenomic array (SNP-A) was performed were stratified by 1) cytogenetic risk and 2) Revised International Prognostic Scoring System (IPSS-R). They were then further stratified based on the presence or absence of additional SNP-A abnormalities. Survival analysis was performed.

FIGURE 2 Frequency of additional SNP-A abnormalities. The number of instances of each type of additional abnormality, which were not detected by conventional karyotype, are depicted for each chromosome.

FIGURE 3 The presence of additional SNP-A abnormalities negatively impacts survival in patients with very good-intermediate cytogenetic risk and very low-intermediate IPSS-R. Survival analysis of patients stratified by cytogenetic risk (A; P<0.0001) and by IPSS-R (B; P =0.002) was performed comparing cases with and without additional SNP-A abnormalities. (A) Cases with very good-intermediate cytogenetic risk with additional SNP-A abnormalities showed worse overall survival than those that did not have additional abnormalities (P = 0.010; black and red groups). SNP-A abnormalities had no statistically significant survival impact in cases with additional SNP-A abnormalities showed worse overall survival risk (P =0.054; green and blue groups). (B) Very low-intermediate IPSS-R cases with additional SNP-A abnormalities showed worse overall survival compared to those without additional abnormalities (P =0.020; black and red groups). SNP-A abnormalities had no statistically significant survival compared to those without additional abnormalities (P =0.020; black and red groups). SNP-A abnormalities had no statistically significant survival impact in cases with high-very high IPSS-R score (P =0.052; green and blue groups).

FIGURE 4 *KMT2A*-PTD significantly impacts survival of patients with additional SNP-A abnormalities. (A) Patients with very good-intermediate karyotype showed worse survival in the presence of additional SNP-A abnormalities (P = 0.010), an effect which was abrogated but not completely eliminated when cases of *KMT2A*-PTD were excluded (P = 0.069). (B) Similar survival effects were seen in cases with very low-intermediate IPSS-R with (P = 0.020) and without (P = 0.063) *KMT2A*-PTD.

FIGURE 5 Genomic locations and graphic display of array results for *KMT2A*-PTD. Patient 7 had gain of 11q23.3 at the other homologue of chromosome 11.



----SCTD uthor Manu



Stratified by cytogenetic risk

Α

в



### Very good-intermediate karyotype

No additional SNP-A abnormalities N=33
 Additional SNP-A abnormalities N=26

### Poor-very poor karyotype

No additional SNP-A abnormalities N=8
 Additional SNP-A abnormalities N=5

Stratified by IPSS-R



### Very low-intermediate IPSS-R

### High-very high IPSS-R

- No additional SNP-A abnormalities N=15
- Additional SNP-A abnormalities N=16



20 40 60 80 100

Time

60 80

40 Time

20 Ó



			C	ytogenetic	risk group		
		Very go	od-intermediate		Poo	or-very poor	
	Total	No additional SNP- A abnormalities	Additional SNP- A abnormalities	Р	No additional SNP-A abnormalities	Additional SNP- A abnormalities	Р
Number of patients	77	33	26		8	10	
Age		66 (59-74)	71 (64-78)	0.18	64.4 (60.6-75.5)	71.2 (62.9-80.0)	0.36
Sex	77			0.59			0.64
Male	50	20	18		6	6	
Female	27	13	8		2	4	
Initial diagnosis	77			0.29			0.46
MDS-SLD	8	3	4		1	0	
MDS-MLD	35	17	10		4	4	
MDS-EB1	12	6	2		2	2	
MDS-EB2	22	7	10		1	4	
IPSS-R		3 (2.5-4.9)	2.75 (1.5-4.25)	0.16	6 (4.25-7.6)	7.25 (6-8.5)	0.14
ANC, k/uL		1.6 (0.8-3.4)	1.7 (1.0-2.5)	0.79	0.9 (0.3-2.3)	0.8 (0.5-1.4)	0.85
Hgb, g/dL		9.8 (8.1-11.5)	9.6 (8.0-10.9)	0.73	9.0 (7.6-9.9)	9.1 (8.0-9.5)	0.56
Plt, k/uL		75.5 (53-191)	118 (66-180)	0.6	65 (30-110)	43 (18-59)	0.37
BM blast percent, %		2.3 (0.9-6.1)	2.0 (1.0-4.3)	0.95	1.8 (0.3-6.7)	3.4 (1.3-9.8)	0.24
Therapy				0.8			NA
Supportive Hypomethylating		15	12		2	4	
agent		18	12		5	7	
Transplant		9	5		3	0	
Other		5	6		0	0	
months	ie,	32.6	26.5		21.7	7.2	

				<b>IPSS</b> -	·R		
		Very low-inter	mediate risk		High-ver	ry high	
		No additional SNP- A abnormalities	Additional SNP- A abnormalities	Р	No additional SNP-A abnormalities	Additional SNP- A abnormalities	Р
Number of patients	77	26	20		18	13	
Age		67.9 (59.7-75.0)	74.0 (62.8-79.8)	0.22	62.5 (59.5-71.4)	69.1 (65.4-76.3)	0.14
Sex	77			0.35			>0.99
Male	50	15	15		12	8	
Female	27	11	5		6	5	
Initial diagnosis	77			0.16			0.69

MDS-SLD	8	5	3		0	0	
MDS-MLD	35	16	11		4	4	
MDS-EB1	12	4	1		5	2	
MDS-EB2	22	1	5		9	7	
IPSS-R		2.75 (2-3.5)	2 (1.5-3)	0.3	6 (5.5-6.5)	6.25 (5.6-7.9)	0.3
ANC, k/uL		1.6 (0.8-3.3)	2 (1.1-2.6)	0.37	1.2 (0.4-3.1)	0.9 (0.5-1.9)	0.83
Hgb, g/dL		10.1 (8.6-12.3)	10.1 (8.1-11.3)	0.66	8.3 (7.5-9.8)	9.0 (7.6-9.4)	0.86
Plt, k/uL		87.5 (69.5-164)	118 (69.8-189)	0.62	44 (28-81)	45 (24-122)	0.98
BM blast percent, %		1.6 (0.6-2.5)	1.9 (0.8-2.1)	0.77	8 (1.5-10)	7.4 (2.1-12)	0.58
Therapy				0.99			0.19
Supportive		14	10		3	6	
agent		13	9		10	10	
Transplant		5	3		7	2	
Other		4	3		1	3	
Median follow up time months	е,	46.2	26.5		24.2	10.3	

*Median value (25th-75th percentile) displayed; absolute neutrophil count (ANC), hemoglobin (Hgb), platelet count (Plt), bone marrow (BM), NA (unable to perform), "Other" therapy (therapies not listed above)* 

# Author Manuscript

### TABLE 2 Additional abnormalities detected by cytogenomic array

				SNP-A results			
Patient	Karyotype	Cytogene tic risk	Cytogenetic risk after array (anomalies >5 Mb but excluding CN-LOH and small focal deletions, duplications)	Array results	%	Size (Mb)	S C V k
1	46,XY[20]	Good	Good	arr[hg19] 4q11q35.2(52,686,799-190,921,709)x2 hmz	90	138.2	T
2	47,XXX?c[20]	Good	Good	arr[hg19] 21q11.2q22.3(14,386,012-48,084,820)x2 hmz	30-90	33.7	T
				arr[hg19] Xp22.33q28(168,546-155,233,731)x3	100	155.1	Ŋ
3	46,XY[16]	Good	Intermediate	arr[hg19] 15q14q22.2(35,437,654-61,211,671)x1-2	15	25.8	T
4	46,XY[20]	Good	Poor	arr[hg19] 6q23.2q23.3(134,365,000-136,607,455)x1	100	2.2	$\uparrow$
				arr[hg19] 7p22.3q36.3(43,360-159,119,707)x1-2	7	159.1	$\top$
				arr[hg19] 11q13.1q25(65,577,515-134,942,626)x2 hmz	100	69.4	T
5	46,XX[20]	Good	Good	arr[hg19] 7q35q36.3(144,958,661-159,119,220)x2 hmz	90	14.2	T
				arr[hg19] 11q23.3(118,338,293-118,354,345)x2-3	45	16 kb	T
6	46,XY[20]	Good	Good	arr[hg19] 11q12.2q25(60,804,709-134,942,626)x2 hmz	90	74.1	T
				arr[hg19] 11q23.3(118,338,293-118,349,247)x2-3	>50	11 kb	T
7	46,XX[20]	Good	Good	arr[hg19] 11q23.3(118,123,516-118,470,527)x2-4	80	347 kb	+
				arr[hg19] 11q24.3(128,408,210-128,699,707)x2-3	80	291 kb	T
8	46,XY[20]	Good	Good	arr[hg19] 1p36.33p34.1(882,802-45,000,436)x2 hmz	90	44.1	
9	46,XY[20]	Good	Good	arr[hg19] 3q11.1q29(93,735,022-197,851,260)x2 hmz	35	104.1	
				arr[hg19] 4q24(106,130,009-106,190,922)x1-2	50	61 kb	
10	46,XX[20]	Good	Good	arr[hg19] 7q21.3q36.3(97,735,123-159,119,220)x2 hmz	50	61.4	T
				arr[hg19] 21q11.2q22.3(15,867,134-48,084,820)x2 hmz	50	32.2	T
11	46,XX[20]	Good	Good	arr[hg19] 8q11.23q24.3(53,704,149-146,292,734)x2 hmz	15	92.6	T
12	46,XY[20]	Good	Good	arr[hg19] 4q22.1q35.2(92,145,040-190,921,709) x2 hmz	10	98.8	T
13	46,XY[20]	Good	Poor	arr[hg19] 7p22.3q36.3(43,360-159,119,707)x1-2	12	159.1	T
14	46,XY[20]	Good	Good	arr[hg19] 3q21.3q29(126,531,213-197,851,260)x2 hmz	85	71.3	T
				arr[hg19] 7q22.1(99,829,321-102,058,793)x1-2	80	2.2	T
				arr[hg19] 21q22.12(36,294,421-37,432,271)x1-2	85	1.1	T
15	46,XX[20]	Good	Good	arr[hg19] 11q23.3(118,338,521-118,355,688)x2-3	30	17 kb	T
16	46,XY[20]	Good	Good	arr[hg19] 4q24(105,995,910-106,227,999)x1-2	90	0.2	╞
17	46,XY[20]	Good	Good	arr[hg19] 21q22.12(36,002,849-37,408,933)x1-2	80	1.4	1

							_
18	46,XY[20]	Good	Good	arr[hg19] 9q34.3(139,101,277-139,734,766)x1	80	0.6	
				arr[hg19] 11q23.3(118,335,185-118,359,052)x3	80	24 kb	T
19	46,XY[20], NUP98	Good	Good	arr[hg19] 5q35.3(176,650,787-176,768,901)x2-3	80	0.1	
	rearrangement (80% by FISH)			arr[hg19] 11p15.4(3,764,205-3,832,210)x2-3	80	68 kb	
20	46,XY,del(5)(q15q33)[4]/4	Good	Good	arr[hg19] 1p36.33p31.3(903,425-65,250,982)x2 hmz	60	64.3	1
	6,XY[1]			arr[hg19] 5q15q33.2(93,274,523-154,360,732)x1-2	40	64.1	Ŋ
21	46,XY,del(5)(q15q31)[19]/	Good	Good	arr[hg19] 3p13p11.1(70,310,611-88,552,092)x1-2	60	18.2	
	46,XY[1]			arr[hg19] 5q15q31.1(95,982,582-131,916,380)x1-2	70	35.9	) J
22	46,XX,del(5)(q13q33)[12]/	Good	Good	arr[hg19] 5q14.2q34(81,856,536-160,672,001)x1-2	50	78.8	У
	40, 40			arr[hg19] 12p13.31p13.1(10,000,550-13,258,017)x1-2	30	3.3	
24	46,XY,del(13)(q12q14)[7]/ 46,XY[18]	Intermedi ate	Intermediate	arr[hg19] 4q24(105,942,532-106,564,759)x1-2	95	0.6	
				arr[hg19] 13q13.1q14.3(33,109,828-53,700,736)x1-2	25	20.6	3
				arr[hg19] Xq28 or Yq12(154,941,868-155,233,731 or 59,044,874-59,336,737)x1-2	95	0.3	
26	47,XY,+8[18]/46,XY[2]	Intermedi	Intermediate	arr[hg19] 8p23.3q24.3(158,048-146,295,771)x2-3	30	146.1	Ŋ
		ate		arr[hg19] 11q13.2q25(67,015,468-134,942,626)x2 hmz	50	69.9	T
27	47,XX,+8[14]/46,XX[6]	Intermedi	Intermediate	arr[hg19] 3q26.2(168,582,060-170,230,667)x2-3	90	1.6	T
		ate		arr[hg19] 8p23.3q11.1(158,048-47,126,524)x2-3	65	47	
				arr[hg19] 8q11.1q11.21(47,127,862-51,456,633)x1-2	90	4.3	
				arr[hg19] 8q11.21q24.3(51,456,754-146,295,771)x2-3	65	94.8	
29	44,XX,add(4)(q23),-	Very poor	Very poor	arr[hg19] 4q25(109,354,361-111,191,990)x1	100	1.8	
	p11.2)del(6)(p12p21),der(6			arr[hg19] 4q32.3q35.2(167,790,246-190,957,473)x1	100	23.2	J
	)t(6;13)(p21;q14)del(6)(p1 2p21),der(11)dup(11)(q13q 13)del(11)(q23q23),-			arr[hg19] 5p15.33p15.2(113,576-13,611,558)x1	100	13.5	T
	13,add(14)(p11.2),del(18)( q21q23)[cp13]/45,sl,+mar[			arr[hg19] 5q11.2q35.1(53,519,660-171,907,198)x1	100	118.4	1
	7]			arr[hg19] 6p24.1p22.1(11,797,999-27,746,178)x1	100	15.9	
				arr[hg19] 6p22.1p21.1(30,105,444-44,326,337)x3	100	14.2	Ŋ
				arr[hg19] 6p21.1p12.1(44,326,504-56,764,165)x1	100	12.4	Ŋ
				arr[hg19] 6p12.1q27(56,768,218-170,919,482)x3	100	114.2	Ŋ
				arr[hg19] 11p11.12q25(51,126,723-134,938,470)cx	100	83.8	T
				arr[hg19] 12p13.2p11.21(11,658,944-31,485,751)x1	100	19.8	Ŋ
				arr[hg19] 13q11q34(19,436,286-115,107,733)cx	100	95.7	T
				arr[hg19] 17q22(54,260,365-55,515,281)x1	100	1.3	T
				arr[hg19] 18q21.2q23(52,763,256-78,014,123)x1	100	25.3	Ŋ
30	43,XY,der(3)t(3;16)(p12;q	Very poor	Very poor	arr[hg19] 3p21.2p12.2(50,699,382-83,166,160)x1-2	65	32.5	Y

		13),der(5)t(3;5)(p12;q13),- 12,der(13)t(12;13)(q12;q3			arr[hg19] 3p11.2q22.3(87,417,120-136,702,414)x1-2	10	49.3	
		4),der(17)t(17;20)(p13;p11 .2),-			arr[hg19] 5q13.2q35.3(72,110,523-180,719,789)x1-2	65	108.6	Y
		20,idic(22)(p11.2)[cp18]/4 6,XY[2]			arr[hg19] 8q24.13q24.21(126,229,398-130,825,360)x2-3	65	4.6	+
					arr[hg19] 12p13.33q12(173,786-46,058,033)x1-2	65	45.9	Y
					arr[hg19] 12q13.11q13.12(48,829,668-49,813,922)x1-2	65	1	+
					arr[hg19] 12q21.1q21.2(75,107,892-76,608,109)x1-2	65	1.5	┢
$\mathbf{\Sigma}$					arr[hg19] 14q11.2q32.33(20,511,672-107,285,437)x2 hmz	90	86.8	
					arr[hg19] 16p11.2(28,689,085-32,922,512)x1-2	65	4.2	T
					arr[hg19] 17p13.3p13.1(9,474-8,172,907)x1-2	65	8.2	Y
_					arr[hg19] 19q13.32q13.33(47,126,613-49,500,959)x1-2	65	2.4	1
)					arr[hg19] 20p12.1(15,681,353-17,022,497)x1-2	65	1.4	
2					arr[hg19] 20p11.21q11.21(22,735,537-29,871,042)x1-2	65	7.1	+
1					arr[hg19] 20q11.21(29,874,663-31,364,166)x3	100	1.5	T
7					arr[hg19] 20q11.21q13.33(31,382,491-62,897,159)x1-2	65	31.5	T
					arr[hg19] 22q11.1q13.33(16,888,899-51,197,838)x2-3	55	34.3	Y
_	31	47,XY,t(1;3)(p13;q21),del( 4)(q21q25),del(11)(q14q24	Very poor	Very poor	arr[hg19] 4q21.23q25(84,749,459-109,483,856)x1-2	60	24.7	Y
		),+19,del(20)(q11.2q13.3)[			arr[hg19] 11q14.2q24.1(86,711,530-123,487,591)x1-2	60	36.8	Y
۲.		4)			arr[hg19] 17q11.2(29,262,000-30,466,769)x1-2	60	1.2	
					arr[hg19] 19p13.3q13.43(260,911-58,956,888)x2-3	20	58.7	Y
					arr[hg19] 20q11.21q13.31(31,062,502-56,290,652)x1-2	50	25.2	Y
2					arr[hg19] 21q22.12(36,202,439-36,282,500)x2-3	100	80 kb	
					arr[hg19] Xp22.2(15,644,219-16,702,011)x1-2	50	1.1	
_	32	44-	Very poor	Very poor	arr[hg19] 3p26.3p12.3(61,891-74,491,151)cx	80	74.4	Y
5		46,X Y,der(2)ins(2;6)(q23; p24p12)add(2)(q23),der(3) t(3;12)(p24;p13),add(4)(q1 2),der(5;22)(p10;q10),-			arr[hg19] 4q12q32.2(57,131,170-163,425,170)cth	80	106.3	
		6,der(12)t(3;12)t(?6;12)(q1 2;q24),der(19)dup(19)(q13			arr[hg19] 5q11.2q12.1(54,136,001-60,742,205)x1-2	90	6.6	Y
_		.1q13.4)add(19)(q13.4),+m			arr[hg19] 5q12.1q35.3(61,997,673-177,260,317)x1-2	90	115.3	Y
		44,sl,add(X)(p11.2),-			arr[hg19] 6p25.3p24.2(156,974-10,880,159)x1-2	80	10.7	┢
		der(2)ins(2;6)add(2),+add(2),+add(2),+add(2),+add(2),-add(2),-add(2),+add(2)			arr[hg19] 16q22.3q23.2(73,602,589-80,595,329)x1-2	80	7	┢
5		der(3)t(3;12),+3,+6,-7,- der(12)t(3;12)t(6:12)+add(			arr[hg19] 17p13.3p11.2(18,900-16,402,114)x2 hmz	90	16.4	┢
		12)(p13),add(19)(p13),add			arr[hg19] 19q13.11q13.43(34,783,132-58,956,888)x2-3	80	24.2	Y
-		(?21)(p11.2),+del(?22)(q11		1		1		4

				arr[hg19] 21q22.13q22.3(39,246,697-43,050,829)x2-3	30	3.8	Τ
				arr[hg19] 22q11.1q13.1(16,888,899-38,450,184)x2-3	80	21.6	Y
				arr[hg19] 22q13.1q13.33(39,235,339-51,197,838)x2-3	80	12	Y
33	44,XY,t(1;12)(q21;q24.1),- 3,add(3)(p11),der(5)t(3;5)( p13;q23),-6,-	Very poor	Very poor	arr[hg19] 1q21.3q22(153,368,019-155,017,913)x1-2	30	1.7	Y
	7,+11,der(11)t(3;11)(q21;q 13)[15]/46,XY[5]			arr[hg19] 3p26.1q29(4,103,600-197,851,936)cx	30	193.8	Y
				arr[hg19] 5q23.2q35.3(124,723,524-180,719,789)x1-2	30	56	Y
				arr[hg19] 6p25.3p22.3(156,974-24,311,197)x1-2	30	24.2	Y
				arr[hg19] 6p22.2p22.1(26,553,570-28,222,528)x1-2	30	1.7	Y
				arr[hg19] 6p21.1p12.3(43,395,395-47,462,571)x2-3	30	4.1	
				arr[hg19] 6p12.3q27(47,467,934-170,919,482)x1-2	30	123.5	
				arr[hg19] 7p21.3p21.1(8,299,324-16,722,353)x1-2	30	8.4	
				arr[hg19] 7p13p12.1(44,817,005-50,784,997)x1-2	30	6	
				arr[hg19] 7q11.21(63,083,343-66,898,842)x1-2	30	3.8	
				arr[hg19] 7q35q36.3(144,075,389-159,119,707)x1-2	30	15	
				arr[hg19] 17p13.3p11.2(18,900-17,027,255)x2 hmz	40	17	
57	46,X,del(Y)(q11.23)[8]/46, XY[12]	Good	Good	arr[hg19] Yp11.31q11.221(2,650,140-19,576,531)x1-2	40	16.9	
				arr[hg19] Yq11.221q11.23(19,585,828-28,799,937)x0-1	40	8.7	T
69	45-46,XY,dic(3;5)(5pter-	Very poor	Very poor	arr[hg19] 3p22.2p12.1(37,299,213-83,644,799)x1-2	70	46.4	Y
	>5q11.2::3p12- >3q29::3p22-			arr[hg19] 5q11.1q12.3(49,430,268-64,262,486)x1-2	20-55	14.8	Y
	>3pter),del(7)(q21q36),+0- 1mar[cp18]/46,XY[2]			arr[hg19] 5q12.3q13.3(66,243,349-76,743,838)x1-2	25-70	10.5	Y
	······[···············			arr[hg19] 5q14.3q35.3(85,913,148-180,719,789)x1-2	70	94.8	Y
				arr[hg19] 7q21.13q36.3(88,745,758-159,119,707)x1-2	70	70.4	Y
70	46~49,XX,-	Very poor	Very poor	arr[hg19] 2q33.1q37.3(201,635,517-242,783,384)x1-2	15	41.2	Y
	2,del(5)(q15q55),- 6,del(6)(p23p24),-7,-13,-			arr[hg19] 5q13.3q35.3(73,921,819-180,719,789)x1-2	15	106.8	Y
	15,+16,del(16)(q12q24),+1 7,add(17)(p11.2),add(17)(p			arr[hg19] 6p22.3p22.1(18,309,604-28,848,258)x4-6	amp	10.5	
	12),-18,-19,- 21,+1~2r,+3~5mar[cp10]/4 6,XX[10]			arr[hg19] 13q11q34(19,436,286-115,107,733)x1-2	15	95.7	Y
				arr[hg19] 19p13.2(10,856,592-12,040,283)x4-7	amp	1.2	
				arr[hg19] 19p13.2p13.12(13,477,560-15,274,712)x2-5	amp	1.8	
				arr[hg19] 21q21.1(19,629,216-20,241,815)x4-7	amp	0.6	
				arr[hg19] 21q22.12q22.3(35,848,786-48,097,372)x2-3	40	12.2	$\square$
73	45,XX,der(4)t(4;?13)(q35; q14),-13,- 16,der(17)t(?16;17)(p11.2;	Very poor	Very poor	arr[hg19] 1p36.33p32.1(849,466-59,863,870)x2-3	80	59	Y

		p11.2),+der(?)t(?;1)(?;p31)						
		8,dic(8;10)(q26;q2?2)ins(1			arr[hg19] 1p31.1(71,990,325-73,466,675)x2-3	85	1.5	Y
		0;1)(q26;p31p36),- 16.der(17)t(?16:17)[cp2]			arr[hg19] 4q35.2(190,712,389-190,957,473)x1-2	85	0.3	Ŷ
					arr[hg19] 8q23.1q24.3(108,384,827-146,292,734)x2 hmz	85	37.9	
					arr[hg19] 13q21.31q21.33(65,565,962-69,826,807)x1-2	85	4.3	
					arr[hg19] 13q33.1(103,508,960-103,531,681)x1-2	85	23 kb	
					arr[hg19] 13q33.1(103,592,015-104,256,190)x1-2	85	0.7	
					arr[hg19] 13q33.2q33.3(105,264,673-109,888,382)x1-2	85	4.6	
					arr[hg19] 16q11.2q24.3(46,503,572-90,155,062)x1	100	43.7	Y
					arr[hg19] 17p13.3p11.2(525-17,988,254)x1	95	18	Y
,	74	45,XY,del(5)(q22q33),der(	Very poor	Very poor	arr[hg19] 5q21.3q33.3(106,203,054-157,533,436)x1-2	30	51.3	Ŷ
		17;20)(q10;p10)[3]/46,sl,d el(7)(q11.2q36),+8[5]/44,sl			arr[hg19] 7p22.3q36.3(43,360-159,119,707)x1-2	15	159.1	Y
		,-7[6]/85- 94 slx2 add(11)(q12)x2 +1			arr[hg19] 8p23.3q24.3(158,048-146,295,771)x2-3	10	146	Y
		-2mar,4-			arr[hg19] 11q13.4(70,735,084-74,545,922)x1-2	10	3.8	
		11dmin[cp4]/46,XY[2]			arr[hg19] 11q13.4q14.1(74,950,287-80,597,434)x2-3	10	5.7	
						10	11.0	L
					an[ng19] 11q14.1q14.3(80,885,357-92,080,784)x1-2	10	5.4	L
					aff[ng19] 11q22.1q22.3(97,942,172-105,515,945)x1-2	10	3.4	L
					arr[ng19] 11q22.3q23.3(104,772,138-117,129,522)x1-2	10	12.4	L
					arr[hg19] 11q23.3(117,940,196-118,570,397)x2-3	80	0.6	
					arr[hg19] 11q23.3q24.2(118,719,566-126,066,194)x1-2	10	7.4	
					arr[hg19] 11q24.2q25(126,201,307-134,938,470)x3-5	amp	8.7	
					arr[hg19] 17p13.3p11.2(525-21,722,139)x1-2	30	21.7	Ŷ
					arr[hg19] 20q11.22q13.33(32,359,017-62,915,555)x1-2	30	32.6	Y
					arr[hg19] 21q21.3q22.3(30,253,288-48,097,372)x2-3	10	17.8	
	76	46- 48 XX del(1)(p22p36 1) de	Very poor	Very poor	arr[hg19] 1p21.1p13.3(103,620,493-108,243,851)x1-2	70	4.6	Ŷ
		l(2)(p24),del(3)(p21),-			arr[hg19] 3q26.33(180,626,349-180,728,459)x1-2	70	0.1	
		5,der(6)t(1;6)(p13;q25),der (7)add(7)(p21)del(7)(q32q			arr[hg19] 5p15.33p14.1(113,576-24,705,254)x1-2	80	24.6	
		36),+8,+8,+8,del(11)(q22q			arr[hg19] 5p14.1p13.2(26,602,764-36,629,258)x1-2	80	10	
		12,+13,del(13)(q12q14)x2,			arr[hg19] 5q14.3(86,075,402-91,536,195)x1-2	80	5.5	
		add(16)(q11.2),add(17)(p1 3),add(19)(p13),-21, +0-			arr[hg19] 5q21.1q32(99,149,791-149,316,740)x1-2	80	50.2	
		2mar[cp20]			arr[hg19] 5q33.1q34(151,060,202-163,263,177)x1-2	80	12.2	
					arr[hg19] 6q11.1q13(62,118,669-73,010,280)x1-2	80	10.9	Y
					arr[hg19] 6q24.2q25.1(144,387,634-152,252,413)x1-2	80	7.9	Y
					arr[hg19] 7p21.3p21.1(11,371,004-17,035,173)x1-2	80	5.7	Y
					arr[hg19] 7p14.1(37.771.693-40.371.298)x1-2	80	2.6	Y

		arr[hg19] 7q33q36.3(134,801,961-159,119,707)x1-2	80	24.3	J
		arr[hg19] 8p23.3q24.3(158,048-146,295,771)x3-4	pentas omy	146.1	J
		arr[hg19] 12p12.3q22(15,277,112-95,085,949)cx	80	79.8	
		arr[hg19] 13q11q13.1(19,436,286-32,565,539)x2-3	70	13.1	J
		arr[hg19] 13q13.1q21.31(32,576,910-64,735,038)x1-2	70	32.2	J
		arr[hg19] 13q21.31q34(64,738,525-115,107,733)x2-3	70	50.4	J
		arr[hg19] 16q12.1(48,341,743-50,857,833)x1-2	80	0.4	
		arr[hg19] 16q12.1q24.3(51,570,940-90,155,062)x1-2	80	2.5	
		arr[hg19] 16q11.2q12.1(46,580,413-47,022,778)x1-2	80	38.6	
		arr[hg19] 17p13.3p11.2(18,900-20,697,797)x2 hmz	80	20.7	

Copy neutral loss of heterozygosity (CN-LOH), partial tandem duplication (PTD), amplification (amp)



GCC\_22783\_Figure 1.tif

---Author Manuscrip



GCC\_22783\_Figure 2.tif



GCC\_22783\_Figure 3\_2.tif



GCC\_22783\_Figure 4.tif

			_	40.1		_	_		_
,87.98		1000	11000	180-30	,100		71380	,80.96	_
-				KMT2A					
0.8		1	5						
Patient 18				e Pres	4.244	9			
11 <mark>2</mark> -		41		a free		23	P	÷	1
Patient 15									
Patient 7	2		64	1º	18.9	5		÷.	2
64 6.4		- b	10			111		- P	÷
Patient 6			1		1.0				
10	2	- 10		Sam	See	- 22-3		- 6	

GCC\_22783\_Figure 5.tif

### TABLE 1 Clinical characteristics of study group

<

			С	ytogenetic risl	k group		
		Very go	od-intermediate		Poo	or-very poor	
	Total	No additional SNP- A abnormalities	Additional SNP- A abnormalities	Р	No additional SNP-A abnormalities	Additional SNP- A abnormalities	Р
Number of patients	77	33	26		8	10	
Age		66 (59-74)	71 (64-78)	0.18	64.4 (60.6-75.5)	71.2 (62.9-80.0)	0.36
Sex	77			0.59			0.64
Male	50	20	18		6	6	
Female	27	13	8		2	4	
Initial diagnosis	77			0.29			0.46
MDS-SLD	8	3	4		1	0	
MDS-MLD	35	17	10		4	4	
MDS-EB1	12	6	2		2	2	
MDS-EB2	22	7	10		1	4	
IPSS-R		3 (2.5-4.9)	2.75 (1.5-4.25)	0.16	6 (4.25-7.6)	7.25 (6-8.5)	0.14
ANC, k/uL		1.6 (0.8-3.4)	1.7 (1.0-2.5)	0.79	0.9 (0.3-2.3)	0.8 (0.5-1.4)	0.85
Hgb, g/dL		9.8 (8.1-11.5)	9.6 (8.0-10.9)	0.73	9.0 (7.6-9.9)	9.1 (8.0-9.5)	0.56
Plt, k/uL		75.5 (53-191)	118 (66-180)	0.6	65 (30-110)	43 (18-59)	0.37
BM blast percent, %		2.3 (0.9-6.1)	2.0 (1.0-4.3)	0.95	1.8 (0.3-6.7)	3.4 (1.3-9.8)	0.24
Therapy				0.8			NA
Supportive Hypomethylating		15	12		2	4	
agent		18	12		5	7	
Transplant		9	5		3	0	
Other		5	6		0	0	
Median follow up time months	•,	32.6	26.5		21.7	7.2	
_		Very low-inter	mediate risk	IPSS-R	High-ver	w high	
5		No additional SNP- A abnormalities	Additional SNP- A abnormalities	P	No additional SNP-A abnormalities	Additional SNP- A abnormalities	Р
Number of patients	77	26	20		18	13	
Age		67.9 (59.7-75.0)	74.0 (62.8-79.8)	0.22	62.5 (59.5-71.4)	69.1 (65.4-76.3)	0.14
Sex	77			0.35			>0.99
Male	50	15	15		12	8	
Female	27	11	5		6	5	
Initial diagnosis	77		0.16				0.69
MDS-SLD	8	5	3		0	0	

				IPSS	5-R		
		Very low-inter	mediate risk		High-ver	ry high	
5		No additional SNP- A abnormalities	Additional SNP- A abnormalities	Р	No additional SNP-A abnormalities	Additional SNP- A abnormalities	Р
Number of patients	77	26	20		18	13	
Age		67.9 (59.7-75.0)	74.0 (62.8-79.8)	0.22	62.5 (59.5-71.4)	69.1 (65.4-76.3)	0.14
Sex	77			0.35			>0.99
Male	50	15	15		12	8	
Female	27	11	5		6	5	
Initial diagnosis	77			0.16			0.69
MDS-SLD	8	5	3		0	0	
MDS-MLD	35	16	11		4	4	
MDS-EB1	12	4	1		5	2	
MDS-EB2	22	1	5		9	7	
IPSS-R		2.75 (2-3.5)	2 (1.5-3)	0.3	6 (5.5-6.5)	6.25 (5.6-7.9)	0.3
ANC, k/uL		1.6 (0.8-3.3)	2 (1.1-2.6)	0.37	1.2 (0.4-3.1)	0.9 (0.5-1.9)	0.83
Hgb, g/dL		10.1 (8.6-12.3)	10.1 (8.1-11.3)	0.66	8.3 (7.5-9.8)	9.0 (7.6-9.4)	0.86
Plt, k/uL		87.5 (69.5-164)	118 (69.8-189)	0.62	44 (28-81)	45 (24-122)	0.98
BM blast percent, %		1.6 (0.6-2.5)	1.9 (0.8-2.1)	0.77	8 (1.5-10)	7.4 (2.1-12)	0.58
Therapy				0.99			0.19
Supportive		14	10		3	6	

Hypomethylating agent	13	9	10	10
Transplant	5	3	7	2
Other	4	3	1	3
Median follow up time, months	46.2	26.5	24.2	10.3

*Median value (25th-75th percentile) displayed; absolute neutrophil count (ANC), hemoglobin (Hgb), platelet count (Plt), bone marrow (BM), NA (unable to perform), "Other" therapy (therapies not listed above)* 

## -Author Manuscrip

### TABLE 2 Additional abnormalities detected by cytogenomic array

-

$\bigcirc$		SNP-A results				SNP-A and karyotype comparison			
Patient	Karyotype	Cytogene tic risk	Cytogenetic risk after array (anomalies >5 Mb but excluding CN-LOH and small focal deletions, duplications)	Array results	%	Size (Mb)	SNP-A concordant with karyotype	Additional abnormality	Additional karyotypic abnormality not in SNP-A
<u> </u>	46,XY[20]	Good	Good	arr[hg19] 4q11q35.2(52,686,799-190,921,709)x2 hmz	90	138.2		4q CN-LOH	
2	47,XXX?c[20]	Good	Good	arr[hg19] 21q11.2q22.3(14,386,012-48,084,820)x2 hmz	30-90	33.7		21q CN-LOH	
				arr[hg19] Xp22.33q28(168,546-155,233,731)x3	100	155.1	Yes		
3	46,XY[16]	Good	Intermediate	arr[hg19] 15q14q22.2(35,437,654-61,211,671)x1-2	15	25.8		15q deletion	
4	46,XY[20]	Good	Poor	arr[hg19] 6q23.2q23.3(134,365,000-136,607,455)x1	100	2.2		6q deletion	
				arr[hg19] 7p22.3q36.3(43,360-159,119,707)x1-2	7	159.1		-7	
				arr[hg19] 11q13.1q25(65,577,515-134,942,626)x2 hmz	100	69.4		11q CN-LOH	
5	46,XX[20]	Good	Good	arr[hg19] 7q35q36.3(144,958,661-159,119,220)x2 hmz	90	14.2		7q CN-LOH	
σ				arr[hg19] 11q23.3(118,338,293-118,354,345)x2-3	45	16 kb		11q gain (KMT2A- PTD)	
6	46,XY[20]	Good	Good	arr[hg19] 11q12.2q25(60,804,709-134,942,626)x2 hmz	90	74.1		11q CN-LOH	
$\leq$				arr[hg19] 11q23.3(118,338,293-118,349,247)x2-3	>50	11 kb		homozygous KMT2A-PTD	
7	46,XX[20]	Good	Good	arr[hg19] 11q23.3(118,123,516-118,470,527)x2-4	80	347 kb		11q gain (KMT2A- PTD)	
				arr[hg19] 11q24.3(128,408,210-128,699,707)x2-3	80	291 kb		11q24.3 gain	
8	46,XY[20]	Good	Good	arr[hg19] 1p36.33p34.1(882,802-45,000,436)x2 hmz	90	44.1		1p CN-LOH	
9	46,XY[20]	Good	Good	arr[hg19] 3q11.1q29(93,735,022-197,851,260)x2 hmz	35	104.1		3q CN-LOH	
$\bigcirc$				arr[hg19] 4q24(106,130,009-106,190,922)x1-2	50	61 kb		4q deletion including TET2	
10	46,XX[20]	Good	Good	arr[hg19] 7q21.3q36.3(97,735,123-159,119,220)x2 hmz	50	61.4		7q CN-LOH	
				arr[hg19] 21q11.2q22.3(15,867,134-48,084,820)x2 hmz	50	32.2		21q CN-LOH	
11	46,XX[20]	Good	Good	arr[hg19] 8q11.23q24.3(53,704,149-146,292,734)x2 hmz	15	92.6		8q CN-LOH	
12	46,XY[20]	Good	Good	arr[hg19] 4q22.1q35.2(92,145,040-190,921,709) x2 hmz	10	98.8		4q CN-LOH	
13	46,XY[20]	Good	Poor	arr[hg19] 7p22.3q36.3(43,360-159,119,707)x1-2	12	159.1		-7	
14	46,XY[20]	Good	Good	arr[hg19] 3q21.3q29(126,531,213-197,851,260)x2 hmz	85	71.3		3q CN-LOH	
				arr[hg19] 7q22.1(99,829,321-102,058,793)x1-2	80	2.2		7q deletion including CUX1	
$\leq$				arr[hg19] 21q22.12(36,294,421-37,432,271)x1-2	85	1.1		21q deletion including RUNX1	
15	46,XX[20]	Good	Good	arr[hg19] 11q23.3(118,338,521-118,355,688)x2-3	30	17 kb		11q gain (KMT2A- PTD)	

_	16	46,XY[20]	Good	Good	arr[hg19] 4q24(105,995,910-106,227,999)x1-2	90	0.2		4q deletion including TET2																
+-	17	46,XY[20]	Good	Good	arr[hg19] 21q22.12(36,002,849-37,408,933)x1-2	80	1.4		21q deletion including RUNX1																
C	18	46,XY[20]	Good	Good	arr[hg19] 9q34.3(139,101,277-139,734,766)x1	80	0.6		9q deletion (constitutional?)																
	_				arr[hg19] 11q23.3(118,335,185-118,359,052)x3	80	24 kb		11q gain (KMT2A- PTD)																
-	19	46,XY[20], NUP98	Good	Good	arr[hg19] 5q35.3(176,650,787-176,768,901)x2-3	80	0.1		5q gain																
	_	rearrangement (80% by FISH)			arr[hg19] 11p15.4(3,764,205-3,832,210)x2-3	80	68 kb		11p gain associated with NUP98-NSD1 translocation																
	20	46,XY,del(5)(q15q33)[4]/4	Good	Good	arr[hg19] 1p36.33p31.3(903,425-65,250,982)x2 hmz	60	64.3		1p CN-LOH																
10		0,A1[1]			arr[hg19] 5q15q33.2(93,274,523-154,360,732)x1-2	40	64.1	Yes																	
U.	21	46,XY,del(5)(q15q31)[19]/	Good	Good	arr[hg19] 3p13p11.1(70,310,611-88,552,092)x1-2	60	18.2		3p deletion																
		46,XY[1]			arr[hg19] 5q15q31.1(95,982,582-131,916,380)x1-2	70	35.9	Yes																	
	22	46,XX,del(5)(q13q33)[12]/	Good	Good	arr[hg19] 5q14.2q34(81,856,536-160,672,001)x1-2	50	78.8	Yes																	
					arr[hg19] 12p13.31p13.1(10,000,550-13,258,017)x1-2	30	3.3		12p deletion including ETV6																
	24	46,XY,del(13)(q12q14)[7]/ 46,XY[18]	Intermedi ate	Intermediate	arr[hg19] 4q24(105,942,532-106,564,759)x1-2	95	0.6		4q deletion including TET2																
					arr[hg19] 13q13.1q14.3(33,109,828-53,700,736)x1-2	25	20.6	Yes																	
$\mathcal{A}$	)				arr[hg19] Xq28 or Yq12(154,941,868-155,233,731 or 59,044,874-59,336,737)x1-2	95	0.3		Xq28 or Yq12 deletion (not specific)																
	26	47,XY,+8[18]/46,XY[2]	Intermedi	Intermediate	arr[hg19] 8p23.3q24.3(158,048-146,295,771)x2-3	30	146.1	Yes																	
			ate		arr[hg19] 11q13.2q25(67,015,468-134,942,626)x2 hmz	50	69.9		11q CN-LOH																
	27	47,XX,+8[14]/46,XX[6]	Intermedi	Intermediate	arr[hg19] 3q26.2(168,582,060-170,230,667)x2-3	90	1.6		3q gain																
			ate		arr[hg19] 8p23.3q11.1(158,048-47,126,524)x2-3	65	47																		
_												1									arr[hg19] 8q11.1q11.21(47,127,862-51,456,633)x1-2	90	4.3		8q deletion (+8 actually had small deletion near centromere)
					arr[hg19] 8q11.21q24.3(51,456,754-146,295,771)x2-3	65	94.8																		
	29	44,XX,add(4)(q23),-	Very poor	Very poor	arr[hg19] 4q25(109,354,361-111,191,990)x1	100	1.8		4q25 deletion																
		$p_{11.2}(6)(p_{12})(0,12)(p_{21})$ $p_{11.2}(6)(p_{12})(p_{21})(p_{12})(p_{1$			arr[hg19] 4q32.3q35.2(167,790,246-190,957,473)x1	100	23.2	Yes		add(14p) not observed															
		(0,13)(p21;q14)del(0)(p1) 2p21),der(11)dup(11)(q13q) 13)del(11)(q23q23),- 13 add(14)(p112) del(18)(							arr[hg19] 5p15.33p15.2(113,576-13,611,558)x1	100	13.5		monosomy 5 as 5p deletion, 5q deletion and marker												
_		q21q23)[cp13]/45,sl,+mar[			arr[hg19] 5q11.2q35.1(53,519,660-171,907,198)x1	100	118.4																		
	2	7]			arr[hg19] 6p24.1p22.1(11,797,999-27,746,178)x1	100	15.9		6p22p24 deletion																
					arr[hg19] 6p22.1p21.1(30,105,444-44,326,337)x3	100	14.2	Yes																	
$\checkmark$					arr[hg19] 6p21.1p12.1(44,326,504-56,764,165)x1	100	12.4	Yes																	
1					arr[hg19] 6p12.1q27(56,768,218-170,919,482)x3	100	114.2	Yes																	
_					arr[hg19] 11p11.12q25(51,126,723-134,938,470)cx	100	83.8		11 abnormalities complex																

				arr[hg19] 12p13.2p11.21(11,658,944-31,485,751)x1	100	19.8	Yes		
				arr[hg19] 13q11q34(19,436,286-115,107,733)cx	100	95.7		13q chromothripisis	
				arr[hg19] 17q22(54,260,365-55,515,281)x1	100	1.3		17q deletion	
$\bigcirc$				arr[hg19] 18q21.2q23(52,763,256-78,014,123)x1	100	25.3	Yes		
30	43,XY,der(3)t(3;16)(p12;q	Very poor	Very poor	arr[hg19] 3p21.2p12.2(50,699,382-83,166,160)x1-2	65	32.5	Yes		
	13),der(5)t(3;5)(p12;q13),- 12,der(13)t(12;13)(q12;q3 4),der(17)t(17;20)(p13;p11 .2),-			arr[hg19] 3p11.2q22.3(87,417,120-136,702,414)x1-2	10	49.3		3p11.2q22.3 deletion	16q and 13q abnormalities balanced
1	20,idic(22)(p11.2)[cp18]/4			arr[hg19] 5q13.2q35.3(72,110,523-180,719,789)x1-2	65	108.6	Yes		
$\bigcirc$	6,XY[2]			arr[hg19] 8q24.13q24.21(126,229,398-130,825,360)x2-3	65	4.6		8q gain	
10				arr[hg19] 12p13.33q12(173,786-46,058,033)x1-2	65	45.9	Yes		
()				arr[hg19] 12q13.11q13.12(48,829,668-49,813,922)x1-2	65	1		12q13 deletion	
				arr[hg19] 12q21.1q21.2(75,107,892-76,608,109)x1-2	65	1.5		12q21 deletion	
				arr[hg19] 14q11.2q32.33(20,511,672-107,285,437)x2 hmz	90	86.8		14q CN-LOH	
				arr[hg19] 16p11.2(28,689,085-32,922,512)x1-2	65	4.2		16p deletion	
				arr[hg19] 17p13.3p13.1(9,474-8,172,907)x1-2	65	8.2	Yes		
				arr[hg19] 19q13.32q13.33(47,126,613-49,500,959)x1-2	65	2.4		19q deletion	
$\mathcal{O}$				arr[hg19] 20p12.1(15,681,353-17,022,497)x1-2	65	1.4		chromosome 20 abnormalities complex	
				arr[hg19] 20p11.21q11.21(22,735,537-29,871,042)x1-2	65	7.1			
				arr[hg19] 20q11.21(29,874,663-31,364,166)x3	100	1.5			
				arr[hg19] 20q11.21q13.33(31,382,491-62,897,159)x1-2	65	31.5			
				arr[hg19] 22q11.1q13.33(16,888,899-51,197,838)x2-3	55	34.3	Yes		
31	47,XY,t(1;3)(p13;q21),del( 4)(q21q25),del(11)(q14q24	Very poor	Very poor	arr[hg19] 4q21.23q25(84,749,459-109,483,856)x1-2	60	24.7	Yes		t(1;3) not observed
$\frown$	),+19,del(20)(q11.2q13.3)[ 4]			arr[hg19] 11q14.2q24.1(86,711,530-123,487,591)x1-2	60	36.8	Yes		
$\bigcirc$	.1			arr[hg19] 17q11.2(29,262,000-30,466,769)x1-2	60	1.2		17q deletion involving NF1	
				arr[hg19] 19p13.3q13.43(260,911-58,956,888)x2-3	20	58.7	Yes		
				arr[hg19] 20q11.21q13.31(31,062,502-56,290,652)x1-2	50	25.2	Yes		
<u> </u>			arr[hg19] 21q22.12(36,202,439-36,282,500)x2-3	100	80 kb		21q gain (partial duplication of RUNX1)		
$\square$				arr[hg19] Xp22.2(15,644,219-16,702,011)x1-2	50	1.1		Xp deletion involving ZRSR1	
32	44- 46.XY.der(2)ins(2:6)(a23·	Very poor	Very poor	arr[hg19] 3p26.3p12.3(61,891-74,491,151)cx	80	74.4	Yes		
$\triangleleft$	p24p12)add(2)(q23),der(3) t(3;12)(p24;p13),add(4)(q1 2),der(5;22)(p10;q10),- 6 dpr(12)((21)2)((21)2)(21)			arr[hg19] 4q12q32.2(57,131,170-163,425,170)cth	80	106.3		add(4q) chromothripisis	chromosomes 2 and 12 abnormalities to be balanced
	6,der(12)t(3;12)t(?6;12)(q1 2;q24),der(19)dup(19)(q13			arr[hg19] 5q11.2q12.1(54,136,001-60,742,205)x1-2	90	6.6	Yes		

	.1q13.4)add(19)(q13.4),+m ar[cp17]/43- 44,sl,add(X)(p11.2),-			arr[hg19] 5q12.1q35.3(61,997,673-177,260,317)x1-2	90	115.3	Yes		
				arr[hg19] 6p25.3p24.2(156,974-10,880,159)x1-2	80	10.7		6p deletion	
	der(2)ins(2;6)add(2),+add( 2)(q32),-			arr[hg19] 16q22.3q23.2(73,602,589-80,595,329)x1-2	80	7		16q deletion	
$\bigcirc$	der(3)t(3;12),+3,+6,-7,-			arr[hg19] 17p13.3p11.2(18,900-16,402,114)x2 hmz	90	16.4		17p CN-LOH	
	12)(p13),add(19)(p13),add			arr[hg19] 19q13.11q13.43(34,783,132-58,956,888)x2-3	80	24.2	Yes		
	(?21)(p11.2),+del(?22)(q11 .2q13)mar[cp3]			arr[hg19] 20q11.23q13.32(34,882,014-58,360,984)x1-2	20	23.5		20q deletion	
<u> </u>				arr[hg19] 21q22.13q22.3(39,246,697-43,050,829)x2-3	30	3.8		21q gain	
15				arr[hg19] 22q11.1q13.1(16,888,899-38,450,184)x2-3	80	21.6	Yes		
$\bigcirc$				arr[hg19] 22q13.1q13.33(39,235,339-51,197,838)x2-3	80	12	Yes		
33	44,XY,t(1;12)(q21;q24.1),-	Very poor	Very poor	arr[hg19] 1q21.3q22(153,368,019-155,017,913)x1-2	30	1.7	Yes		chromosome
()	3,add(3)(p11),der(5)t(3;5)( p13;q23),-6,-								11 abnormalities
	7,+11,der(11)t(3;11)(q21;q			arr[ha10] 3p26 1a20/4 103 600 107 851 036) cy	30	103.8	Vac		to be balanced
	15)[15]/40,741[5]			arr[hg10] 5222 2225 2(124 722 524 180 710 780)×1.2	30	193.8	Vac		
				$an[lng19] 5425.2455.5(124, 725, 924-100, 717, 767) \times 1-2$	30	30	I es		
				atr[ng19] 6p25.3p22.3(156,9/4-24,311,19/)X1-2	30	24.2	Yes		
				arr[hg19] 6p22.2p22.1(26,553,570-28,222,528)x1-2	30	1.7	Yes		
				arr[hg19] 6p21.1p12.3(43,395,395-47,462,571)x2-3	30	4.1		loss of 6 and 7 complex	+11
				arr[hg19] 6p12.3q27(47,467,934-170,919,482)x1-2	30	123.5			
				arr[hg19] 7p21.3p21.1(8,299,324-16,722,353)x1-2	30	8.4			
>				arr[hg19] 7p13p12.1(44,817,005-50,784,997)x1-2	30	6			
				arr[hg19] 7q11.21(63,083,343-66,898,842)x1-2	30	3.8			
				arr[hg19] 7q35q36.3(144,075,389-159,119,707)x1-2	30	15			
0				arr[hg19] 17p13.3p11.2(18,900-17,027,255)x2 hmz	40	17		17p CN-LOH	
57	46,X,del(Y)(q11.23)[8]/46, XY[12]	Good	Good	arr[hg19] Yp11.31q11.221(2,650,140-19,576,531)x1-2	40	16.9		Yp gain [del(Y) to be idic(Yq)]	
$\bigcirc$				arr[hg19] Yq11.221q11.23(19,585,828-28,799,937)x0-1	40	8.7			
69	45-46,XY,dic(3;5)(5pter-	Very poor	Very poor	arr[hg19] 3p22.2p12.1(37,299,213-83,644,799)x1-2	70	46.4	Yes	5q deletion complex	
	>3q29::3p22-			arr[hg19] 5q11.1q12.3(49,430,268-64,262,486)x1-2	20-55	14.8	Yes		
	>3pter),del(7)(q21q36),+0- 1mar[cp18]/46.XY[2]			arr[hg19] 5q12.3q13.3(66,243,349-76,743,838)x1-2	25-70	10.5	Yes		
<b>—</b>				arr[hg19] 5q14.3q35.3(85,913,148-180,719,789)x1-2	70	94.8	Yes		
				arr[hg19] 7q21.13q36.3(88,745,758-159,119,707)x1-2	70	70.4	Yes		
70	46~49,XX,-	Very poor	Very poor	arr[hg19] 2q33.1q37.3(201,635,517-242,783,384)x1-2	15	41.2	Yes		
	2,del(5)(q15q33),- 6,del(6)(p23p24),-7,-13,-			arr[hg19] 5q13.3q35.3(73,921,819-180,719,789)x1-2	15	106.8	Yes		
$\leq$	15,+16,del(16)(q12q24),+1 7.add(17)(p11.2) add(17)(p			arr[hg19] 6p22.3p22.1(18,309,604-28,848,258)x4-6	amp	10.5		6p amplification	
	12),-18,-19,- 21,+1~2r,+3~5mar[cp10]/4 6,XX[10]			arr[hg19] 13q11q34(19,436,286-115,107,733)x1-2	15	95.7	Yes		loss of 7, 15, abnormalities on

									chromosome 16 and 17 not seen by array			
+				arr[hg19] 19p13.2(10,856,592-12,040,283)x4-7	amp	1.2		19p amplification				
$\bigcirc$				arr[hg19] 19p13.2p13.12(13,477,560-15,274,712)x2-5	amp	1.8		19p amplification				
				arr[hg19] 21q21.1(19,629,216-20,241,815)x4-7	amp	0.6		21q amplification				
				arr[hg19] 21q22.12q22.3(35,848,786-48,097,372)x2-3	40	12.2		21q gain				
73	45,XX,der(4)t(4;?13)(q35; q14),-13,- 16,der(17)t(?16;17)(p11.2; p11.2),+der(?)t(?;1)(?;p31)	Very poor	Very poor	arr[hg19] 1p36.33p32.1(849,466-59,863,870)x2-3	80	59	Yes		the subclone - 8 and dic(8;10) not observed			
	[18]/44-45,XX,- 8 dic(8:10)(a26:a2?2)ins(1			arr[hg19] 1p31.1(71,990,325-73,466,675)x2-3	85	1.5	Yes					
( )	0;1)(q26;p31p36),-			arr[hg19] 4q35.2(190,712,389-190,957,473)x1-2	85	0.3	Yes					
	16, der(17)t(?16;17)[cp2]			arr[hg19] 8q23.1q24.3(108,384,827-146,292,734)x2 hmz	85	37.9		8q CN-LOH				
				arr[hg19] 13q21.31q21.33(65,565,962-69,826,807)x1-2	85	4.3		loss of 13 complex				
				arr[hg19] 13q33.1(103,508,960-103,531,681)x1-2	85	23 kb						
				arr[hg19] 13q33.1(103,592,015-104,256,190)x1-2	85	0.7						
				arr[hg19] 13q33.2q33.3(105,264,673-109,888,382)x1-2	85	4.6						
				arr[hg19] 16q11.2q24.3(46,503,572-90,155,062)x1	100	43.7	Yes					
$(\mathbf{U})$				arr[hg19] 17p13.3p11.2(525-17,988,254)x1	95	18	Yes					
74	45,XY,del(5)(q22q33),der(	Very poor	Very poor	Very poor	Very poor	Very poor	arr[hg19] 5q21.3q33.3(106,203,054-157,533,436)x1-2	30	51.3	Yes		
	el(7)(q11.2q36),+8[5]/44,sl			arr[hg19] 7p22.3q36.3(43,360-159,119,707)x1-2	15	159.1	Yes					
	,-7[6]/85- 94.slx2.add(11)(q12)x2.+1			arr[hg19] 8p23.3q24.3(158,048-146,295,771)x2-3	10	146	Yes					
	-2mar,4-			arr[hg19] 11q13.4(70,735,084-74,545,922)x1-2	10	3.8						
<u> </u>	11unin[cp4]/40,A1[2]	dmin[cp4]/46,XY[2]					arr[hg19] 11q13.4q14.1(74,950,287-80,597,434)x2-3	10	5.7		add(11q) complex	near tetraploid clone not observed
$\frown$				arr[hg19] 11q14.1q14.3(80,885,537-92,686,784)x1-2	10	11.8						
$\bigcirc$				arr[hg19] 11q22.1q22.3(97,942,172-103,313,943)x1-2	10	5.4						
				arr[hg19] 11q22.3q23.3(104,772,138-117,129,522)x1-2	10	12.4						
				arr[hg19] 11q23.3(117,940,196-118,570,397)x2-3	80	0.6						
				arr[hg19] 11q23.3q24.2(118,719,566-126,066,194)x1-2	10	7.4						
-				arr[hg19] 11q24.2q25(126,201,307-134,938,470)x3-5	amp	8.7						
				arr[hg19] 17p13.3p11.2(525-21,722,139)x1-2	30	21.7	Yes					
				arr[hg19] 20q11.22q13.33(32,359,017-62,915,555)x1-2	30	32.6	Yes					
				arr[hg19] 21q21.3q22.3(30,253,288-48,097,372)x2-3	10	17.8		21q gain				
76	46- 48 VV dol(1)( $-22-26(1)$ ) $\frac{1}{2}$	Very poor	Very poor	arr[hg19] 1p21.1p13.3(103,620,493-108,243,851)x1-2	70	4.6	Yes					
	l(2)(p24),del(3)(p21),-			arr[hg19] 3q26.33(180,626,349-180,728,459)x1-2	70	0.1		3q deletion				

	5, der(6)t(1;6)(p13;q25), der	arr[hg19] 5p15.33p14.1(113,576-24,705,254)x1-2	80	24.6		loss of 5 complex	
	(7)(add(7)(p21)del(7)(q32q)) 36),+8,+8,+8,del(11)(q22q)	arr[hg19] 5p14.1p13.2(26,602,764-36,629,258)x1-2	80	10			
	23),- 12.+13.del(13)(q12q14)x2.	arr[hg19] 5q14.3(86,075,402-91,536,195)x1-2	80	5.5			
$\bigcirc$	add(16)(q11.2),add(17)(p1	arr[hg19] 5q21.1q32(99,149,791-149,316,740)x1-2	80	50.2			
	2mar[cp20]	arr[hg19] 5q33.1q34(151,060,202-163,263,177)x1-2	80	12.2			
		arr[hg19] 6q11.1q13(62,118,669-73,010,280)x1-2	80	10.9	Yes		
<u> </u>		arr[hg19] 6q24.2q25.1(144,387,634-152,252,413)x1-2	80	7.9	Yes		
1		arr[hg19] 7p21.3p21.1(11,371,004-17,035,173)x1-2	80	5.7	Yes		
$\bigcirc$		arr[hg19] 7p14.1(37,771,693-40,371,298)x1-2	80	2.6	Yes		
10		arr[hg19] 7q33q36.3(134,801,961-159,119,707)x1-2	80	24.3	Yes		
0,		arr[hg19] 8p23.3q24.3(158,048-146,295,771)x3-4	pentas omy	146.1	Yes		
		arr[hg19] 12p12.3q22(15,277,112-95,085,949)cx	80	79.8		loss of 12 complex	
		arr[hg19] 13q11q13.1(19,436,286-32,565,539)x2-3	70	13.1	Yes		
		arr[hg19] 13q13.1q21.31(32,576,910-64,735,038)x1-2	70	32.2	Yes		
		arr[hg19] 13q21.31q34(64,738,525-115,107,733)x2-3	70	50.4	Yes		
		arr[hg19] 16q12.1(48,341,743-50,857,833)x1-2	80	0.4		add(16q) complex	
$(\mathbf{U})$		arr[hg19] 16q12.1q24.3(51,570,940-90,155,062)x1-2	80	2.5			
		arr[hg19] 16q11.2q12.1(46,580,413-47,022,778)x1-2	80	38.6			
>		arr[hg19] 17p13.3p11.2(18,900-20,697,797)x2 hmz	80	20.7		17p CN-LOH	loss of 21 not observed

Copy neutral loss of heterozygosity (CN-LOH), partial tandem duplication (PTD), amplification (amp)