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Identification of *CFTR* Variants in Latino Patients with Cystic Fibrosis from the Dominican Republic and Puerto Rico

Short Running Title: CFTR Variants in the Dominican Republic and Puerto Rico

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#### **ABSTRACT**

**Background:** In cystic fibrosis (CF), the spectrum and frequency of *CFTR* variants differ by geography and race/ethnicity. *CFTR* variants in White patients are well-described compared to Latino patients. No studies of *CFTR* variants have been done in patients with CF in the Dominican Republic or Puerto Rico.

**Methods:** *CFTR* was sequenced in 61 Dominican Republican patients and 21 Puerto Rican patients with CF and >60 mmol/L sweat chloride. The spectrum of *CFTR* variants was identified and the proportion of patients with 0, 1, or 2 *CFTR* variants identified was determined. The functional effects of identified *CFTR* variants were investigated using clinical annotation databases and computational prediction tools.

Results: Our study found 10% of Dominican patients had two *CFTR* variants identified compared to 81% of Puerto Rican patients. No *CFTR* variants were identified in 69% of Dominican patients and 10% of Puerto Rican patients. In Dominican patients, there were 19 identified *CFTR* variants, accounting for 25/122 disease alleles (20%). In Puerto Rican patients, there were 16 identified CFTR variants, accounting for 36/42 disease alleles (86%) in Puerto Rican patients. Thirty *CFTR* variants were identified overall. The most frequent variants for Dominican patients were p.Phe508del, p.Ala559Thr and for Puerto Rican patients were p.Phe508del, p.Arg1066Cys, p.Arg334Trp, p.I507del.

**Conclusions:** In this first description of the *CFTR* variants in patients with CF from the Dominican Republic and Puerto Rico, there was a low detection rate of This article is protected by copyright. All rights reserved.

two *CFTR* variants after full sequencing with the majority of patients from the Dominican Republic without identified variants.

### **INTRODUCTION**

Although the majority of the 80,000 people worldwide with cystic fibrosis (CF) are White, an increasing proportion of patients are of other races and ethnicities. In the United States (U.S.), the percentage of patients with CF who are Latino increased from 5.6% to 8.7% over the past 15 years<sup>1</sup>. The increase in the Latino CF population is important as these patients have increased morbidity and mortality compared to the White CF population<sup>2,3</sup>.

Latino patients not only have a different clinical course than White patients, but they also have different *CFTR* variants<sup>4</sup>. Latino patients are also more likely to have CFTR Class IV-V or uncharacterized variants whereas White patients are more likely to have Class I-III variants. Latino patients are more likely to have one or no *CFTR* variants identified, in part due to CF genetic panels and newborn screens having lower sensitivity to variants that are more common in the Latino population<sup>5</sup>.

CFTR variants are population-specific and the spectrum of known CFTR variants is based largely on investigations of White populations<sup>6</sup>. Even among investigations of Latino populations, there have been limited efforts to describe

the genetic profile of CF in the Caribbean<sup>7</sup>. There is considerable genetic heterogeneity between Latino populations and within the Caribbean<sup>8</sup>.

In this study, full genetic sequencing of *CFTR* was done in Dominican and Puerto Rican patients with CF to describe the spectrum of *CFTR* variants. The proportion of patients with 0, 1, or 2 *CFTR* variants identified was determined. The functional impact of each identified *CFTR* variant was classified based on clinical databases and deleteriousness prediction algorithms.

### **METHODS**

# **Study Population**

This was a cross-sectional study of *CFTR* variants in patients with CF in the Dominican Republic and Puerto Rico. All patients had a diagnosis of CF made by their clinical doctor based on the presence of clinical CF symptoms and a positive sweat chloride concentration (>60 mmol/L) based on Cystic Fibrosis Foundation guidelines<sup>9</sup>. Patients with an intermediate sweat chloride concentration (<60 mmol/L) were not included in our study. Patients six years of age and older were recruited from CF clinics in the Dominican Republic and Puerto Rico in 2017. Consent and assent were obtained from patients and their guardians as appropriate. This study was approved by the Western Institutional Review Board.

At the time of recruitment, *CFTR* variants identified through prior genotyping, sweat chloride concentration, demographic data, pancreatic

sufficiency status, and pulmonary function percent predicted based on Global Lung Initiative (GLI) was recorded for each patient. Blood was drawn for genetic analysis.

# Whole genome sequencing analysis

DNA was isolated from whole blood using the Wizard® Genomic DNA Purification kits (Promega, Fitchburg, WI). DNA samples were quantified by fluorescence using the Quant-iT PicoGreen dsDNA assay (ThermoFisher Scientific, Waltham, MA) on a Spectramax fluorometer (Molecular Devices, Sunnyvale, CA). DNA samples were sequenced as part of the Trans-Omics for Precision Medicine (TOPMed) whole genome sequencing (WGS) program<sup>9</sup>. WGS was performed at the Northwest Genomics Center on a HiSeqX system (Illumina, San Diego, CA) using a paired-end read length of 150 bp, to a minimum of 30x mean genome coverage. Details on DNA sample handling, quality control, library construction, clustering and sequencing, read processing and sequence data quality control are previously described<sup>10</sup>. Variant calls were obtained from TOPMed data freeze 8 VCF files. The term "variant" is used in place of "mutation" or "polymorphism" Variants with a minimal depth of coverage of 10 reads were included in our analyses.

Variants were annotated in TOPMed using the WGSA pipeline<sup>12</sup>. *CFTR* variants were annotated with reference to the NM\_000492.3 transcript. Genetic variants in *CFTR* were extracted (ENSG00000001626; ENST00000003084) from chr7:117,465,784-117,715,971, which included segments 15kb upstream of the

CFTR transcription start site and 47kb downstream of the last exon. Sequences aligned to hs38DH 1000 Genomes GRCh38/hg38 reference assembly using BWA-MEM were received as CRAM files from TOPMed<sup>13</sup>. Chromosome 7 sequence reads were extracted from the CRAM files using Samtools v1.9<sup>14</sup>. Copy number variation was detected using a bin size of 500 with CNVnator v0.3.3<sup>15</sup>. Structural variation in chromosome 7, including deletions, duplications, inversions and translocations, were detected with LUMPY express v0.2.13<sup>16</sup>. The sequencing quality of variants that did not have value "PASS" in the FILTER field from TOPMed was checked by manual inspection of the sequencing reads alignment using Integrative Genome Viewer. Other possible FILTER values include CEN (variant overlaps with centromeric region), SVM (variant failed SVM filter) and DISC (variant with high mendelian or duplicate genotype discordance [3/5% or more])<sup>10</sup>.

Phased genotypes from TOPMed data freeze 8 were used to determine whether two variants are in *cis* or *trans* (see section below). These were statistically phased by applying Eagle 2.4 (Dec 13, 2017) to the whole panel of 137,977 samples included in TOPMed freeze 8. Phasing was done in 1 Mb chunks with 0.1 Mb overlap. The entire CFTR locus (chr7:117,465,784 - 117,715,971) falls within a single chunk. Phasing was limited to variants which pass all filters and starts with minDP10 genotypes to restrict to high quality genotypes. Phasing imputes any missing genotypes. Statistical phasing has limited accuracy for very rare variants (those seen in fewer than 5 individuals in the panel).

# **Clinical annotations of variants**

To determine the clinical impact of *CFTR* variants identified by sequencing, variants were first compared to the Clinical and Functional Translation of CFTR (CFTR2) database <sup>12</sup>. The CFTR2 database provides functional classifications for variants with clinical and laboratory evidence of phenotypic consequence. These classifications include: "CF-causing", "varying clinical consequence", "unknown significance" and "non-CF-causing". CFTR variants identified by sequencing that were not listed in the CFTR2 database were analyzed to identify common variants, defined by an allele frequency >3% on Genome Aggregation Database (gnomAD) in the general non-CF population<sup>9</sup>. Variants that are common in a non-CF population are unlikely to be diseasecausing. All identified variants that were not in the CFTR2 database or had an allele frequency <3% were cross-referenced with two variant databases, ClinVar and Invitae, to determine the functional impact of the variant. ClinVar is a publicly available repository of genotype-phenotype investigations<sup>20</sup>. Invitae is a clinical genetic sequencing laboratory<sup>21</sup>. Variants in the ClinVar and Invitae databases were annotated as "pathogenic", "likely pathogenic", "uncertain significance" or "benign".

Genetic variants without clinical annotations in the CFTR2, ClinVar, or Invitae databases were analyzed for deleteriousness using three computational prediction algorithms: Combined Annotation Dependent Depletion (CADD), FATHMM-XF, and Rare Exome Variant Ensemble Learner (REVEL)<sup>22–24</sup>.

Variants predicted to be deleterious had a scaled C-score ranking from CADD >16 or were predicted to be likely damaging by FATHMM-XF or had a REVEL score greater than 0.5.

Variants were categorized into five functional classifications based on databases and computational predictions: 1) CF-disease causing variant, 2) variant of varying clinical consequence, 3) variant of uncertain significance, 4) variant predicted to be deleterious, and 5) Non-CF-disease causing or likely benign variant (Figure 1).

The *CFTR* variants and genotype for each patient were determined. In patients with two variants, the phased genotype (variants in *cis* or *trans*) was assessed using BCFtools<sup>25</sup>. Patients were categorized as fully identified *CFTR* genotype (two variants in *trans*) versus those who were not (with two variants in *cis*, one variant, or no variants).

# **RESULTS**

Genotyping results prior to recruitment

Our study population consisted of 82 patients diagnosed with CF from the Dominican Republic (N = 61) and Puerto Rico (N = 21). At recruitment, 3% of Dominican patients had two identified *CFTR* variants, 3% had one variant, and 93% had not been genetically tested for CF. Among Puerto Rican patients, 57% of patients had two identified *CFTR* variants, 24% had one variant, 5% had no variants, and 14% had not been tested.

At the time of recruitment, Dominican patients were a median age of 10.6 years old and Puerto Rican patients were 15.4 years old. The majority of Dominican patients (86.9%) and Puerto Rican patients (81.0%) were pancreatic insufficient. The average FEV<sub>1</sub> percent predicted was 91.7% in Dominican patients and was 83.4% in Puerto Rican patients.

Whole genome sequencing results

There were 1568 CFTR variants identified by whole genome sequencing in our study population (Figure 1). No structural variation or copy number variation was detected in the CFTR region (ENSG0000001626, chr7:117,465,784-117,715,971). Of the 1568 variants identified, 29 variants were functionally classified in the CFTR2 database: 16 CF-disease causing variants, 4 varying clinical consequence variants, 2 variants of uncertain significance, and 7 non-CF-causing variants. Of the 1539 CFTR variants not present in the CFTR2 database, 397 were identified as common variants in the general population, therefore were interpreted as benign and not analyzed further. Of the remaining 1142 variants, functional classification using the ClinVar and Invitae databases was determined in 30 variants: 1 CF-disease causing variant, 4 variants of uncertain significance, and 25 likely benign variants. There was no functional classification description for the 1112 remaining variants, so they were further annotated using three functional prediction algorithms. Seven variants were predicted to be deleterious by at least one computational prediction tool. Three of these seven variants were removed after manual inspection of the sequencing reads alignment suggested these were sequencing or alignment errors.

Overall, we identified 30 *CFTR* variants: 16 CF-disease causing variants (Table 1), 4 variants of varying clinical consequence, 6 variants of uncertain significance, and 4 variants predicted to be deleterious (Table 2). The most frequent known disease-causing variants for Dominican patients were p.Phe508del (10%) and p.Ala559Thr (3%). The most frequent known disease-causing variants for Puerto Rican patients were p.Phe508del (33%), p.Arg1066Cys (33%), p.Arg334Trp (14%), and p.Ile507del I (14%).

Only 10% of Dominican patients had two CF-disease causing variants in trans compared to 81% of Puerto Rican patients (Table 3). Both Dominican patients (10%) and Puerto Rican patients (10%) had multiple *CFTR* variants in *cis*. Eleven percent of Dominican patients had only one *CFTR* variant identified; no Puerto Rican patients had only one *CFTR* variant identified. No variants were identified in 69% of Dominican patients and in 10% of Puerto Rican patients.

The 30 identified *CFTR* variants accounted for 25/122 disease alleles (20%) in Dominican patients and 36/42 disease alleles (86%) in Puerto Rican patients.

#### DISCUSSION

In this first genetic description of *CFTR* variants in Dominican and Puerto Rican patients with CF, we found that there was a low rate of patients having two

CFTR variants identified after full sequencing. The spectrum of CFTR variants differed between the two populations, which are of the same ethnicity in close geographic proximity. In the overall CF population in the U.S., 86% of patients have at least one copy of p.Phe508del. In contrast, in our study, only 9.8% of Dominican patients and 33% of Puerto Rican patients had at least one copy of p.Phe508del<sup>1</sup>. The most frequent variants we found in Puerto Rican patients occurred at low rates in the general CF population in the U.S.: p.lle507del is the 15<sup>th</sup> most common variant occurring in 0.8% of the general CF population but was observed in 14% of Puerto Rican patients in our study; p.Arg334Trp is the 25<sup>th</sup> most common variant occurring in 0.3% of the general CF population but was observed in 14% of Puerto Rican patients and 3% of Dominican patients in our study. P.Arg1066Cys was observed in a third of Puerto Rican patients in our study but is not in the top 25 most common variants in the general CF population. P.Ala559Thr was observed in 3% of Dominican patients in our study was not in the top 25 variants of the general CF population<sup>1</sup>.

The spectrum of *CFTR* variants varies between Latino populations across the world and also varied between the two specific Latino populations we studied: Dominicans and Puerto Ricans. In patients with CF in Spain, the most frequent *CFTR* variants were p.Phe508del (52%), p.Gly542x (8%), p.Asn1303Lys (3%), and 3849+10kbC→T (2%)<sup>26</sup>. In Latino patients with CF from across the U.S., the most frequent variants were p.Phe508del (37%), p.Gly542x (11%), and p.Arg334Trp (11%)<sup>27</sup>, but most frequent variants differed across the U.S. In the Southwestern U.S., the most frequent *CFTR* variants were

p.Phe508del (47%), p.Gly542x (5%), and 3849+10kbC→T (3%)<sup>28</sup>, which was similar to the pattern observed in Southern California: p.Phe508del (52%), p.Gly542x (4%), 3849+10kbC→T (4%), p.Ser549Asn (2%)<sup>29</sup>. In Illinois, the most frequent variants were p.Phe508del (52%), 3849+10kbC→T (7%), p.Phe311del (7%)<sup>4</sup>. P.Phe508del was observed at a lower frequency in Dominican patients (10%) and Puerto Rican patients (33%) in this study than in the Latino populations described above. 3849+10kbC→T, a frequent variant in all referenced Latino populations, was not found in any patient in our study, which may be due to low frequencies in other Hispanic populations (2-3%). P.Gly542X, one of the most frequent variants in Latino populations, was not present at all in Dominicans and observed only in 10% of Puerto Rican patients. The unique spectrum of CFTR variants in Dominican and Puerto Rican patients may be due to their heterogeneous genetic background, with a higher proportion of African ancestry than in Latino populations from the mainland U.S.8. Our findings highlight the need for investigating population-specific *CFTR* variants.

In this comprehensive genetic analysis of patients with clinically confirmed CF, 81% of Puerto Rican patients had disease-causing *CFTR* variants identified on both chromosomes compared to only 10% of the Dominican patients. Over two-thirds of Dominican patients had no identifiable variant in *CFTR* compared to 10% of Puerto Rican patients. We were surprised at the high proportion of Dominican patients without any identifiable *CFTR* variants. In contrast, sequencing analysis in other Latino populations with CF have reported much higher detection rate (~95%) of *CFTR* variants<sup>4,26</sup>. All the patients included in our

study had clinical evidence of CFTR dysfunction with symptoms consistent with CF and a sweat chloride concentration of >60 mmol/L. Analysis of nasal potential difference and functional analysis of the CFTR channel may increase our understanding of CFTR function in patients lacking *CFTR* variants. Other studies have described patients with symptoms of CF and elevated sweat chloride concentrations but without evidence of *CFTR* variants. Patients without 2 *CFTR* variants in *trans* may have variants in other genes such as the epithelial sodium channel (ENaC) or may have defective pathways that lead to CFTR dysfunction<sup>32</sup>.

We were also surprised that 10% of Dominican and 10% of Puerto Rican patients had multiple variants found on only one chromosome (i.e., in *cis*). Genotype phase is not routinely analyzed in clinical sequencing of *CFTR*, so deleterious effects of different variants in *cis* may be more common in the general CF population than currently understood.

Understanding the spectrum and frequency of *CFTR* variants in diverse populations is important for improving CF genetic panels and newborn screening programs. Genetic panels and newborn screening programs are generally developed based on variant frequencies observed in the White population and have lower sensitivity (i.e., higher false negative rate) when applied to a Latino population. The ACMG/ACOG-recommended CFTR 23 variant panel, offered to pregnant women, has a 76% detection rate in White patients with CF, while the detection rate is only 48% in Latino patients with CF<sup>5,33</sup>. The ACOG panel would

have detected only 7 of the 30 variants that we identified in our study (5 variants in Dominicans, 6 variants in Puerto Ricans). Using the ACOG panel in our patients, 73% of patients would have no variants identified and 15% would have had only 1 variant identified.

Genetic screens and newborn screens should be sensitive to the target population and include the prevalent *CFTR* variants for all racial and ethnic groups to minimize false negative diagnoses. Detection via newborn screening is important as patients diagnosed via newborn screen demonstrate improved lung function and nutritional status compared to those not detected on a newborn screen<sup>34</sup>. In the Illinois newborn screen, for example, Latino infants were more likely to have undefined variants and twice as likely to have only one variant identified compared to White infants<sup>4</sup>. Latino patients have both more rare and novel *CFTR* variants so newborn or genetic screens will always be less effective for Latino patients if they do not include sequencing<sup>35,36</sup>.

CFTR genetic variant identification and functional classification have become increasingly valuable not only for CF phenotype prediction but also for identifying those patients who would benefit from CFTR modulator therapies<sup>37</sup>. CFTR modulators target specific CFTR variants, which occur more frequently in White patients compared to minorities. As a result, only a third of Latino patients qualify compared to three-quarters of White patients<sup>38</sup>. This is consistent with our study's findings that only 5 of 82 patients (3 Dominicans and 2 Puerto Ricans) were eligible for CFTR modulator pharmacotherapy. Only two of the 30 variants

we identified were eligible CFTR modulator targets: p.Phe508del and p.Arg74Trp. The most common variant in Puerto Rican patients, p.Arg1066Cys, is not approved for CFTR modulators. To combat this disparity in access to lifealtering pharmacogenetic therapies, the first step is to identify *CFTR* variants in CF populations, as we have done in this study, and then to describe the functional implications of the identified variants and investigate the protein response to CFTR modulators. The final step is to include Latino and other non-Latino non-White patients in clinical trials of CFTR modulators, as minorities are under-represented in the majority of CF pharmacotherapy clinical trials<sup>39</sup>.

In silico prediction algorithms have been used to identify likely disease-contributing *CFTR* variants, but the utility of predictive algorithms is controversial as they cannot differentiate between variants that caused severe, moderate, or minimal reduction in CFTR function<sup>40–42</sup>. Our study similarly found inconsistent predictions as the algorithms predicted 5 variants to be deleterious but were annotated by CFTR2 as "non-CF-causing" (Supplemental Table 1). Additionally, we identified three variants that were predicted to be deleterious but were removed after manual inspection of the sequencing reads alignment.

Although we sequenced the majority of known patients with CF over 6 years old in both Puerto Rico and the Dominican Republic, our study was limited by a small number of patients. To fully understand *CFTR* variants in these populations, a genetic analysis of the general population of the Dominican

Republic and Puerto Rico is needed. Our study identified multiple variants in *cis*, but our study was not designed to genotype parents to confirm genotype phase.

Our study results indicate that the spectrum of *CFTR* variants in an unstudied CF population cannot be inferred from another CF population, even if the racial and ethnic background is similar. Genetic panels and even genome sequencing have limitations in identifying *CFTR* variants in Latino patients with CF. Understanding the spectrum of *CFTR* variants in all populations with CF is the first step towards developing effective CF treatment for all patients. Studies of cystic fibrosis and pharmacotherapies need to include more racially diverse populations in order to make precision medicine socially precise.

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# **Figure**

Figure 1 Flowchart for assignment of CFTR variant categories.

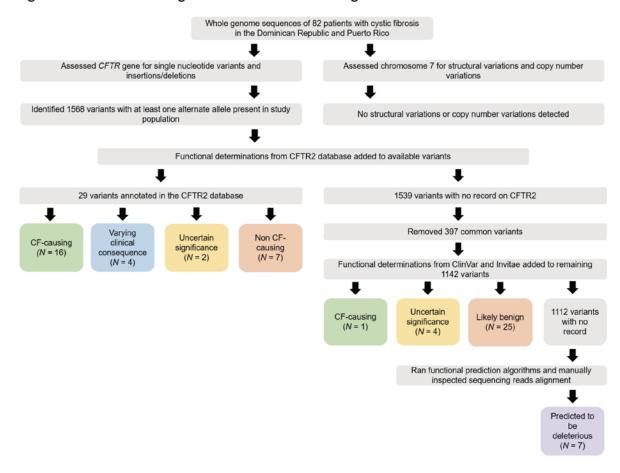


Table 1. Known disease-causing *CFTR* variants

| # | Protein name | cDNA name         | Position<br>(GRCh38) | Reference<br>allele | Alternate<br>allele | Occurrences, n (%) |         |
|---|--------------|-------------------|----------------------|---------------------|---------------------|--------------------|---------|
|   |              |                   |                      |                     |                     | D.R.               | P.R.    |
| 1 | p.Phe508del  | c.1521_1523delCTT | 7:117559590          | ATCT                | A                   | 6 (10%)            | 7 (33%) |
| 2 | p.Arg1066Cys | c.3196C>T         | 7:117611637          | C                   | T                   |                    | 7 (33%) |
| 3 | p.Arg334Trp  | c.1000C>T         | 7:117540230          | C                   | T                   | 1 (2%)             | 3 (14%) |
| 4 | p.Ile507del  | c.1519_1521delATC | 7:117559586          | TATC                | T                   |                    | 3 (14%) |
| 5 | p.Gly542*    | c.1624G>T         | 7:117587778          | G                   | T                   |                    | 2 (10%) |
| 6 |              | c.1680-886A>G     | 7:117589467          | A                   | G                   |                    | 2 (10%) |

| 7  | p.Asn1303Lys | c.3909C>G   | 7:117652877 | C  | G |        | 2 (10%) |
|----|--------------|-------------|-------------|----|---|--------|---------|
| 8  | p.Arg553*    | c.1657C>T   | 7:117587811 | C  | T | 1 (2%) | 1 (5%)  |
| 9  |              | c.2988+1G>A | 7:117606754 | G  | A | 1 (2%) | 1 (5%)  |
| 10 | p.Ala559Thr  | c.1675G>A   | 7:117587829 | G  | A | 2 (3%) |         |
| 11 |              | c.164+1G>A  | 7:117504364 | G  | A |        | 1 (5%)  |
| 12 | p.Tyr1092*   | c.3276C>A   | 7:117611717 | C  | A |        | 1 (5%)  |
| 13 |              | c.3368-2A>G | 7:117614611 | A  | G |        | 1 (5%)  |
| 14 | p.Thr1220fs  | c.3659delC  | 7:117627711 | AC | A |        | 1 (5%)  |
| 15 | p.Ile148fs   | c.442delA   | 7:117531067 | CA | C | 1 (2%) |         |
| 16 | p.Arg709*    | c.2125C>T   | 7:117592292 | C  | T | 1 (2%) |         |

D.R.: Dominican Republic

P.R.: Puerto Rico

Table 2. Potentially disease-causing variants

| # | Protein name                             | -DNA                    | Position    | Reference | Alternate<br>Allele | Occurrences, n (%) |        |
|---|--|-------------------------|-------------|-----------|---------------------|--------------------|--------|
| # | Protein name                             | cDNA name               | (GRCh38)    | Allele    |                     | D.R.               | P.R.   |
|   | Variants of varying clinical consequence |                         |             |           |                     |                    |        |
| 1 | p.Arg74Trp                               | c.220C>T                | 7:117509089 | C         | T                   | 1 (2%)             | 1 (5%) |
| 2 |  | c.1210-<br>34TG[11]T[5] | 7:117548628 | GTT       | G                   | 2 (3%)             |        |
| 3 |  | c.1210-<br>34TG[12]T[5] | 7:117548630 | T         | G                   | 1 (2%)             |        |
| 4 | p.Asp1270Asn                             | c.3808G>A               | 7:117642528 | G         | A                   |                    | 1 (5%) |
|   | Variants of uncertain significance       |                         |             |           |                     |                    |        |
| 1 |  | c226G>T                 | 7:117479869 | G         | T                   | 1 (2%)             |        |
| 2 | p.Val201Met                              | c.601G>A                | 7:117535269 | G         | A                   |                    | 1 (5%) |
| 3 | p.Ser589Asn                              | c.1766G>A               | 7:117590439 | G         | A                   | 1 (2%)             |        |
| 4 | p.Tyr1014Cys                             | c.3041A>G               | 7:117610571 | A         | G                   | 1 (2%)             |        |
| 5 | p.Arg1158Gln                             | c.3473G>A               | 7:117627526 | G         | A                   | 1 (2%)             |        |
| 6 | p.Asp1445Asn                             | c.4333G>A               | 7:117666998 | G         | A                   | 1 (2%)             |        |

|   | Varia                  | ants predicted to be | deleteriou | s |        |  |
|---|------------------------|----------------------|------------|---|--------|--|
| 1 | <br>n.49-<br>4832T>G   | 7:117470859          | T          | G | 3 (5%) |  |
| 2 | <br>c.367-<br>4084T>G  | 7:117711674          | T          | G | 3 (5%) |  |
| 3 | <br>c.1585-<br>1361A>G | 7:117586378          | A          | G | 1 (2%) |  |
| 4 | <br>n2799A>G           | 7:117710954          | A          | G | 1 (2%) |  |

D.R.: Dominican Republic

P.R.: Puerto Rico

Table 3. Type and frequency of *CFTR* genotypes.

| CFTR Genotype                      | Dominican Republic, $N(\%)$ | Puerto Rico, N (%) |
|------------------------------------|-----------------------------|--------------------|
| 2 CF-disease causing, <i>trans</i> | 6 (10%)                     | 17 (81%)           |
| 2 CF-disease causing, <i>cis</i>   | 1 (2%)                      | 1 (5%)             |
| 2 VVCC, 1 VUS, cis                 |                             | 1 (5%)             |
| 1 VUS, 1 predicted, cis            | 1 (2%)                      |                    |
| 2 predicted, cis                   | 3 (5%)                      |                    |
| 1 VVCC, 1 VUS, cis                 | 1 (2%)                      |                    |
| 1 CF-causing                       | 1 (2%)                      |                    |
| 1 VVCC                             | 3 (5%)                      |                    |
| 1 VUS                              | 2 (3%)                      |                    |
| 1 predicted                        | 1 (2%)                      |                    |
| No variants                        | 42 (69%)                    | 2 (10%)            |

CFTR variants were categorized using three functional annotation databases as well as three computational tools for predicting deleteriousness. Cis and trans describe the relationship between two or more variants; cis refers to variants on the same gene copy while trans describes variants on different gene copies. VVCC: variant of varying clinical consequence. VUS: variant of uncertain significance.