

Sensor-Mediated Granular Sludge Reactor for Nitrogen Removal and Reduced Aeration Demand using a Dilute Wastewater

Zerihun A. Bekele¹, Jeseth Delgado Vela², Charles B. Bott³, and Nancy G. Love^{1,*}

¹Department of Civil Engineering and Environmental Engineering, University of Michigan, 1351 Beal Ave, Ann Arbor, MI 48109, USA (Email: zerualem@umich.edu, nglove@umich.edu)

² Current address: Department of Civil and Environmental Engineering, Howard University, 2300 Sixth St NW, Washington, DC 20059, USA (Email: jeseth.delgadovela@howard.edu)

³Hampton Roads Sanitation District, 1434 Air Rail Ave, Virginia Beach, Virginia 23455, USA (Email: cbott@hrsdc.com)

*Corresponding Author: Department of Civil and Environmental Engineering, University of Michigan, 183 EWRE Building, 1351 Beal Avenue, Ann Arbor, MI 48109, USA (Email: nglove@umich.edu)

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1002/WER.1296](https://doi.org/10.1002/WER.1296)

This article is protected by copyright. All rights reserved

1 Abstract

2 A sensor-mediated strategy was applied to a lab-scale granular sludge reactor (GSR) to
3 demonstrate that energy efficient inorganic nitrogen removal is possible with a dilute
4 mainstream wastewater. The GSR was fed a dilute wastewater designed to simulate an A-
5 stage mainstream anaerobic treatment process. DO, pH, and ammonia/nitrate sensors
6 measured water quality as part of a real-time control strategy that resulted in low-energy
7 nitrogen removal. At a low COD ($0.2 \text{ kg/m}^3/\text{day}$) and ammonia ($0.1 \text{ kg-N/m}^3/\text{day}$) load, the
8 average degree of ammonia oxidation was $86.2 \pm 3.2\%$ and total inorganic nitrogen removal
9 was $56.7 \pm 2.9\%$ over the entire reactor operation. Aeration was controlled using a DO
10 setpoint, with and without residual ammonia control. Under both strategies, maintaining a
11 low bulk oxygen level (0.5 mg/L) and alternating aerobic/anoxic cycles resulted in a higher
12 level of nitrite accumulation and supported shortcut inorganic nitrogen removal by
13 suppressing nitrite oxidizing bacteria. Furthermore, coupling a DO setpoint aeration strategy
14 with residual ammonia control resulted in more stable nitrification and improved aeration
15 efficiency. The results show that sensor-mediated controls, especially coupled with a DO
16 setpoint and residual ammonia controls, are beneficial for maintaining stable aerobic granular
17 sludge.

18 Key words: NOB suppression, aeration control, partial nitrification/anammox, mainstream N
19 removal

20 1. INTRODUCTION

21 There is great interest in systems that are energy neutral or positive to also achieve
22 resource recovery using means that meet stringent effluent standards. The A-B (adsorption-
23 biooxidation) process targets this end point. The A-stage is dedicated to maximizing carbon
24 capture for later energy production and is commonly deployed using high rate activated
25 sludge (HRAS) (Jimenez et al., 2015) or chemically enhanced primary treatment (CEPT)
26 (Diamantis et al., 2013). Recently, the anaerobic membrane biofilm reactor (AnMBR) has
27 been proposed as a viable A-stage technology that could become an energy efficient option
28 (Smith et al., 2014). The B-stage is focused on energy efficient nutrient (commonly nitrogen)
29 management (Jetten, Horn, & van Loosdrecht, 1997; Wan, Gu, Zhao, & Liu, 2016). Most of
30 the energy expense in an A-B process occurs due to aeration in the B-stage, where the
31 remaining carbon and nitrogen (N) are removed. In some cases, it is possible for the energy

32 expense in the B-stage to negate the energy gained in the A-stage, making A-B process
33 inefficient for energy recovery (Zhou et al., 2013).

34 Processes that require less oxygen and do not require external substrate are desirable to
35 minimize aeration demand in the B-stage. Traditionally, N removal is done via complete
36 nitrification (first by ammonium oxidizing bacteria (AOB), then by nitrite oxidizing bacteria
37 (NOB)) followed by heterotrophic denitrification via ordinary heterotrophic organisms
38 (OHOs). This process has a high aeration demand and requires a higher theoretical oxygen
39 demand (ThOD) to nitrogen (ThOD/N) ratio (> 5) compared to other novel processes
40 (Daigger, 2014). In particular, A-stage processes such as HRAS and CEPT are less efficient
41 at removing nitrogen (de Graaff et al., 2016; Miller et al., 2016), resulting in an effluent
42 ThOD/N ratio < 5 . This ratio is insufficient for N removal, because additional carbon will be
43 lost during aeration for nitrification. In this case, exogenous electron donor may be needed to
44 achieve high levels of N removal.

45 Compared to HRAS and CEPT, the AnMBR A-stage process may be better suited to
46 achieve an overall A-B process energy efficiency. The effluent from mainstream anaerobic
47 treatment typically contains organic carbon (45-145 mg COD/L), ammonium (19-53 mg
48 N/L), dissolved methane (40-140 mg ThOD/L) and sulfide (0-145 mg ThOD/L) (Delgado
49 Vela et al., 2015). Hence, if we consider dissolved methane, sulfide and ammonium as
50 potential electron donors in addition to organic carbon for N removal in a downstream B-
51 stage system, it will be enough for complete N-removal. Methane and sulfide in the AnMBR
52 effluent are possible electron donors for N removal with nitrite/nitrate as electron acceptors
53 via denitrifying anaerobic methane oxidation (DAMO) (Raghoebarsing et al., 2006) and
54 sulfide oxidation (Souza & Foresti, 2013). Among these options, pursuing N removal via
55 nitrite is preferred as it uses less electron donor and less aeration for nitrification. Ammonia
56 can be used as electron donor for anaerobic ammonia oxidation (anammox), which requires
57 nitrite as an e^- acceptor (Strous, Jetten, Heijnen, & Kuenen, 1998). However, any of these
58 approaches requires an operational strategy that reliably allows nitrification and minimizes loss
59 of electron donors. Assuming this can be done, then partial nitrification and anammox (PN/A)
60 becomes the most attractive B-stage N removal option, as its low aeration demand and no
61 organic carbon demand (Winkler et al., 2012).

62 Besides targeting for efficient N removal processes, the use of advanced biofilm systems
63 such as aerobic granular sludge reactor (GSR) as a prospective B-stage technology has
64 significant advantages over conventional activated sludge systems (Sarma, Tay, & Chu,

65 2017). Noted advantages of aerobic GSR include: the presence of different redox zones
66 within the granules that support a diverse microbial ecology; high rates of settleability; and a
67 high biomass retention which is ideal for slow growing N-removing bacteria (de Kreuk,
68 Heijnen, & van Loosdrecht, 2005; Liu, Yang, & Tay, 2003; Szabó et al., 2017). **Figure 1**
69 shows the potential key microbial groups and their interactions during complete nitrogen
70 removal via nitrite in a single stage GSR downstream of an AnMBR A-stage. The schematic
71 emphasizes that competition for nitrite will occur among different anaerobic organisms, and
72 suggests that successful N removal requires sustained partial nitrification while suppressing
73 nitrite oxidation by NOB. Therefore, the success of an energy efficient aerobic GSR as a B-
74 stage N removal system requires a robust operating strategy that favors simultaneous partial
75 nitrification and denitrification by suppressing NOB.

76 One of the main challenges for N-removal in mainstream wastewater via nitrite is
77 suppression of NOB. In particular, the suppression of NOB in PN/A process has been
78 demonstrated at full-scale for concentrated sidestream applications, which have favorable
79 conditions such as: low C/N ratio (<2 g COD/g N); high temperature (20 to 30 °C); high free
80 ammonia concentrations (> 0.1 mg N/L) (Philips et al., 2002); and high (> 0.2 mg N/L) free
81 nitrous acid (Kornaros, Dokianakis, & Lyberatos, 2010). Unfortunately, most of these
82 conditions are atypical for dilute mainstream systems (Cao, van Loosdrecht, & Daigger,
83 2017). However, if a sequencing batch reactor (SBR) is used and a minimum residual
84 ammonium concentration (RAC) is maintained throughout the reaction time, NOB
85 suppression can still be achieved via free ammonia that is present in sufficient concentration
86 during most of the reaction cycle. Nonetheless, since this alone may not be enough to
87 effectively suppress NOB, new strategies are needed for NOB suppression. For this reason,
88 we propose the use of real-time sensor-mediated control (SMC) for robust aeration control to
89 suppress NOB. Online sensors have been used to suppress NOB in sidestream applications
90 by manipulating the DO setpoint, or by using intermittent aeration and ammonium-based
91 aeration control (ABAC). For example, Regmi et al. (2014) used real-time ABAC with
92 intermittent aeration to suppress NOB in a suspended culture for mainstream nitrification.
93 Lemaire et al. (2008) used DO and pH sensors to control aerobic duration for shortcut N
94 removal by suppressing NOB. Both studies demonstrate that SMC for mainstream NOB
95 suppression is a viable option.

96 A-stage effluent has a low organic load in concert with a much lower N load than side-
97 stream granular systems, which can tolerate low organic loads given the high N loading (Wett

98 et al., 2015). The combination of low organic and N loading with a low C/N ratio in dilute
99 mainstream A-B applications makes it challenging to develop and sustain granules. Tay et al.
100 (2004) reported that they were not able to produce granules when the organic loading was
101 below $2 \text{ kg COD m}^{-3} \text{ d}^{-1}$. The lowest organic loading rate that we found reported to date for
102 successful granule formation came from Ni et al. (2008) and Zhang et al. (2015) who used
103 0.6-1 and 0.37-0.56 $\text{kg COD m}^{-3} \text{ d}^{-1}$, respectively. Hence, developing stable granules in low
104 loaded circumstances is another challenge that must be addressed to advance the GSR
105 technology.

106 In this study, we focused on developing and operating an aerobic GSR as an exemplary
107 B-stage N removal system for an A-stage AnMBR effluent. We use this reactor
108 configuration to develop and demonstrate an SMC strategy that supports NOB suppression
109 and reduces aeration energy. We evaluated the degree to which the strategy supports PN/A
110 for N removal, and highlight the conditions needed to support stable granule formation.

111 2. METHODS

112 2.1. Reactor setup.

113 A glass bubble column reactor with 76.2 mm (3 inches) diameter and 711.2 mm (28
114 inches) height with a working volume of 4.5 L was operated for 474 days. The reactor was
115 initially inoculated by mixing a nitrifying activated sludge from the Ann Arbor Wastewater
116 Treatment Plant (Michigan, USA) and biomass from a full-scale deammonification
117 (DEMON) unit (Hampton Roads Sanitation District, Virginia, USA). The reactor was fed
118 with a simulated mainstream anaerobic digester effluent containing ammonium ($48 \pm 6 \text{ mg/L-N}$),
119 VFAs (acetate and propionate, 100 mg/L-COD), dissolved methane at saturation (~ 22
120 mg/L-CH_4) and other trace elements. Details of the media preparation procedure can be found
121 in the SI Section 12. The C/N ratio, considering VFA and ammonium, was from 1.85 to 2.5.
122 The reactor was monitored and controlled using online optical DO (WTW, FDO 925, Xylem
123 Inc.), pH (accumet[®] Electrode, Fisher Scientific), and $\text{NH}_4^+/\text{NO}_3^-$ (IQ SensorNet VARiON[®]
124 Plus Sensors, Xylem Inc.) probes to suppress NOB and favor the growth of AOB and
125 anammox species. A bulk DO concentration was held at specific DO levels between 0.2 and
126 $1.5 (\pm 0.1 \text{ mg/L O}_2)$. The pH was monitored using an online probe and was maintained
127 between 7.3 ± 0.2 and 8.0 ± 0.2 by dosing NaHCO_3 . The entire experiment was conducted at
128 an ambient temperature between 20 and 23°C.

129 2.2. Cycle operation.

130 The reactor was operated in a sequencing batch mode with a 40 min anoxic slow feed
131 from the bottom of the reactor, followed by intermittent aeration with anoxic and aerobic
132 cycles for 220-300 min, a settling time of 4 to 10 min, and 4 min of decanting with a
133 volumetric exchange ratio of 50%. Air was supplied using a glass diffuser with a superficial
134 upflow velocity of 1.6 cm/sec. During the anoxic phase, gas from the head space of the
135 reactor was pressurized by diaphragm pump, blended with dinitrogen gas and recirculated
136 through the reactor. This supported the development of anoxic conditions and resupplied
137 stripped methane gas to enhance the chance for its dissolution and metabolism. During the
138 aerobic phase a mixture of dry air, dinitrogen gas and head space gas were pumped into the
139 reactor. The dry air flow rate was controlled with a mass flow controller (MFC) device to
140 maintain the desired setpoint using in-house developed partial differential and integral (PID)
141 controller (developed in LabView® software).

142 2.3. Sensor-mediated control development.

143 Across the aerobic phases of a single SBR cycle, DO was controlled in the first two
144 phases based on a specific setpoint, and ammonia-based aeration control was used for the last
145 phase of operation. Both aeration schemes were implemented by developing a PID controller
146 in LabView (Figure SI-5). The developed LabView SMC program was designed to allow
147 time-based aeration for DO setpoint only and ABAC with a DO setpoint. The program also
148 controls all the pumps and sensor-based devices, thus automating the entire operation.
149 Information reported as sensor-derived concentration was adjusted based on correction
150 factors determined by using simple linear regression between sensor output and analytically-
151 determined concentrations (i.e., for ammonium and nitrate). Effluent nitrite corrections
152 reported in **Figure 5** only were derived from its correlations with effluent ammonium and
153 nitrate.

154 2.4. Long-term reactor operation

155 The reactor was operated for 474 days, which can be broken down into four phases (Table
156 SI-2). During the first phase (days 1-60), granule development occurred and the reactor was
157 operated at a DO setpoint of 1.5 mg/L. A lower DO setpoint (0.5 mg/L) without ABAC was
158 used during the second phase (days 61-200). The third phase (days 201 - 410) was operated at
159 a DO setpoint of 0.75 mg/L without ABAC. The final and fourth phase (days 411-474) was
160 operated with low DO setpoint (0.5 mg/L) and ABAC. Ammonium-based aeration was

161 implemented to maintain residual ammonium at or above 5 mg/L as N, both to promote
162 anammox activity and to suppress NOB as indicated by Cao et al. (2017). During all phases,
163 samples were usually collected three times a week from the reactor after settling using a
164 sampling port, and analyzed for ammonium (Standard Method (SM) Sec. 4500-NH₃F),
165 nitrite (SM Sec. 4500-NO₂⁻ B), nitrate (SM Sec. 4110 B) and methane (SM 2720, 6211, and
166 6010). Biweekly cross-cycle sampling was done over a single operating cycle to obtain
167 profiles of soluble N species, also volatile fatty acids (VFAs) using ion chromatography as
168 described in Smith et al. (Smith, Skerlos, & Raskin, 2013). Additionally, in-situ batch activity
169 tests were conducted for Phases 2, 3 and 4 to determine nitrification and anammox activities.
170 The detailed procedure for this can be found in SI section 13. Physical characteristics of
171 granules, such as size, sludge volume index (SVI), and solids retention time (SRT), were also
172 monitored as described in the SI. All sensors data (i.e., NH₄⁺, NO₃⁻, DO and pH) and other
173 operation information were logged every minute. All values errors are given at a 95%
174 confidence interval from a t-test distribution.

175 2.5. Microbial community analysis

176 Biomass samples were taken during the first two months of the reactor granulation phase
177 and also at later stages of operation to monitor the microbial composition using 16S rRNA
178 gene amplicon sequencing. The inoculum used to obtain an anammox organism was from a
179 full-scale deammonification unit at Hampton Roads Sanitation District (HRSD), Virginia.
180 DNA was extracted using three bead-beating steps followed by extraction with a Maxwell 16
181 LEV automated nucleic acid extractor (Promega, Madison WI) using DNA blood kits.
182 Amplicon sequencing of the V4 region of the 16S rRNA gene was performed on the Illumina
183 MiSeq (MiSeq Reagent Kit V2 500 cycles, Illumina Inc., San Diego, CA) platform using the
184 previously developed dual-indexing sequencing strategy (Kozich, Westcott, Baxter,
185 Highlander, & Schloss, 2013). Additional details of the procedure can be found in Delgado
186 Vela et al. (2018). Post-processing of the Illumina MiSeq data was done using the Mothur
187 MiSeq SOP (Schloss et al., 2009) without rarefaction and including archaea in the analysis.
188 Data from the MiSeq analysis have been uploaded to NCBI and is openly available under
189 accession number PRJNA549919.

190 3. RESULTS

191 3.1. Phase 1: Granule formation

192 The initial phase of granule development took about 60 days (**Figure 2**), which is
193 relatively rapid compared to other granular systems (Ni et al., 2009). For granule
194 development and selection, the reactor was operated in SBR mode with a short (5 min)
195 settling time and a superficial upflow velocity of 1.6 cm/sec. The granules went through
196 different morphological transformations as they were formed (**Figure 2**). First, micro-
197 granules began to form after two weeks of operation. Then, large and fluffy granules formed
198 after a month. Finally, the granules developed into mature granules after two months of
199 operation. The mature granules had an average diameter of 0.97 ± 0.06 mm (Figure SI-2) and
200 an average settling velocity of 12 m/hr. The steady state mean VSS concentration was $1,520$
201 ± 176 mg/L with a mean 5 min SVI of 70 mg/L. We calculated an average SRT of 4.8 ± 0.6
202 days, 12.2 ± 2.9 days, 11.9 ± 2.1 days, and 17.5 ± 3.5 days for Phase 1, Phase 2, Phase 3, and
203 Phase 4, respectively (Figure SI-1).

204 During granulation, the biomass color changed from dark red (inoculum, not shown) to
205 pale yellow (**Figure 2**), indicating a shift in microbial composition. Whole community
206 analysis based on the 16S rRNA gene was used to characterize how the microbial community
207 shifted over the course of reactor operation, including during the granulation period (**Figure**
208 **3**). Community analysis showed that only anammox taxa (AMX) was detected in the
209 inoculum of the order “Candidatus Brocadiales,” and comprised approximately 13% of the
210 community. Subsequently, the relative abundance of AMX decreased to 2.2% by the end of
211 granulation. During this same period, nitrite and ammonium accumulated while nitrate was
212 mostly absent. Although substrates required for anammox metabolism were present, the loss
213 of AMX suggests that this metabolism was not occurring to a significant degree.
214 Concurrently, the inoculum contained relatively equal fractions of AOB (genus
215 *Nitrosomonas*, 3.7%) and NOB (genus *Nitrospira*, 3.4%); however, by the end of the
216 granulation period AOB had a higher relative abundance (5.4%) than NOB (0.6%), consistent
217 with NOB out-selection. The nitrite accumulation rate ($NAR = 0.67 \pm 0.24$) was consistent
218 with this result (see **Figure 4**). Furthermore, OHOs increased from 20% to 50% over this
219 same period; however, we presume that insufficient organic carbon was available as an
220 electron donor to fully consume the accumulated nitrite as it was observed in Phase 2 and 3
221 (Figure SI-6). These results suggest that a rapid, or aggressive, granulation period makes it
222 difficult to establish stable redox niches that are needed to support slow growing
223 microorganisms, such as AMX and NOB. Furthermore, the high DO of 1.5 mg/L used during
224 the aerobic period might have reduced the size of the anoxic zone in the granules, which may

225 have limited AMX activity. Collectively, these factors all likely contributed to the reduction
226 in N removal during the granulation phase.

227 3.2. Phase 2: Low (0.5 mg/L) dissolved oxygen operational phase

228 In Phase 2 operation when the bulk DO setpoint was 0.5 mg-O₂/L, nitrite was routinely
229 present in the effluent at an average concentration of 13.9 ± 3.4 mg N/L while nitrate
230 averaged 0.4 ± 0.2 mg N/L with high NAR of 0.93 ± 0.06 (**Figure 4**). The effluent ammonium
231 concentration was 8.4 ± 3.5 mg N/L and quite variable (Figure SI-4), ranging between 25 and
232 below detection (seven of 25 measurements during this phase were at or below 0.1 mg/L as
233 N). The overall TIN removal during this phase was $49.6 \pm 5.6\%$ and the overall ammonia
234 conversion was $80.7 \pm 8.1\%$ (**Figure 4**). This broad range of performance during Phase 2 may
235 have made it difficult to sustain significant anammox growth in the system.

236 To estimate of the relative contributions of AMX and OHOs to TIN removal, we used
237 cross-cycle data (i.e., one batch cycle N and VFA profile data) with theoretical stoichiometric
238 equation (See SI section 10, from Tables SI-9 to 16). The VFA data show that at least one-
239 third was utilized by the end of the first anoxic period and was consumed at the same time as
240 residual nitrite held over from the prior cycle was consumed, implying that nitrite served as
241 the electron acceptor during that period. The remaining VFA was rapidly oxidized during the
242 first aerobic period. After accounting for nitrogen consumption for cell growth, our
243 calculations suggest that TIN removal via OHOs occurred up to the end of the first anoxic
244 period, and for the rest of the cycle via anammox. Our stoichiometric predictive analysis
245 shows that VFA removed up to 72% of the oxidized inorganic nitrogen on average during
246 Phase 2. We did not detect residual methane at the end of the anoxic feed period (Figure SI-
247 7) nor did we detect DAMO through our microbial community analysis; therefore, it is
248 reasonable to assume that methane was stripped out as soon as mixing started in the first
249 anoxic zone. Considering all these factors leaves an unaccounted source of TIN removal of at
250 least 28%, which we assume is attributed to anammox during this Phase. Consequently,
251 despite its low relative abundance, AMX may have contributed to TIN removal.

252 In-situ nitrification and anammox activity results from tests conducted during Phase 2
253 suggest that NOB were suppressed and AMX were active, corroborating the stoichiometric
254 predictive analysis. AOB (0.32 g N-NO₂⁻ formed/g VSS/day) were 4 times more active than
255 NOB (0.08 g N-NO₃⁻ formed/g VSS/day), even though DO exceeded 1.0 mg/L (Table SI-6).
256 The Phase 2 in-situ anammox activity test yielded a specific total inorganic nitrogen
257 (ammonium + nitrite) utilization rate of 0.104 mg-N/mg-VSS/day, supporting our prior

258 conclusion that anammox was likely actively involved in TIN removal (See Table SI-3).

259 3.3. Phase 3: high (0.75 mg/L) dissolved oxygen operational phase

260 Phase 3 was operated at a higher bulk DO setpoint of 0.75-1 mg-O₂/L, and resulted in
261 both a sustained loss of nitrite and increase in nitrate. During this phase, effluent nitrite was
262 present at an average concentration of 4.4 ± 1.1 mg N/L while the effluent nitrate increased to
263 an average concentration of 14.7 ± 2.6 mg N/L, which resulted in low NAR of 0.33 ± 0.08
264 (**Figure 4**). The effluent ammonium concentration was 4.0 ± 1.5 mg N/L with an overall
265 conversion rate of $92 \pm 3\%$, which is higher than in Phase 2 ($p < 0.001$). The overall TIN
266 removal for this phase was $53.2 \pm 4.1\%$, which is similar to the performance of Phase 2 ($p >$
267 0.05).

268 Cross-cycle analysis showed that AMX and OHO both continued to contribute to TIN
269 removal. VFA was predominantly consumed during the anoxic feed period via heterotrophic
270 denitrification that used the residual nitrate from the prior cycle (see Figure SI-6). The
271 remaining VFA was oxidized rapidly during the first aerobic period. Consequently, any TIN
272 removal observed after the first aerobic period was assumed to be due to AMX. Based on
273 cross-cycle data (Tables SI-17 through 24) and after accounting for N loss for growth, we
274 estimate that VFA could, at best, remove 66% of the oxidized inorganic nitrogen present in
275 the influent. Since AMX are present in the system and exposed to intermittent anaerobic
276 periods, and a persistent anaerobic inner core exists in the granules despite a measureable
277 bulk DO, we conclude that AMX removed up to 34% of TIN.

278 In-situ nitrification and anammox activity tests indicated that NOB were more active than
279 AOB, and anammox activity was detected but lower than what was measured during Phase 2
280 (See Table SI-3 & SI-6). The in-situ nitrification test conducted on day 376 showed that the
281 NOB activity rate (0.32 g N-NO₃⁻ formed/g VSS/day) was at least 3 times more active than
282 AOB (0.099 g N-NO₂⁻ formed/g VSS/day), indicating that the NOB suppression observed
283 during Phase 2 was reversed. In addition, the in-situ anammox activity test conducted on day
284 362 showed that AMX had a specific total inorganic nitrogen utilization rate of 0.08 mg-
285 N/mg-VSS/ day, indicating that AMX were active but at a lower rate than what was measured
286 during Phase 2. As a confirmation, we estimated the average rate of the net TIN oxidized by
287 AMX only during Phase 3 to be 0.06 mg-N/mg-VSS/day from the cross-cycles data. Since
288 the in-situ activity tests show the highest rate achievable under ideal conditions, this
289 comparison shows that the anammox activity measured can explain the loss of oxidized TIN
290 residual during a single cycle.

291 Microbial community analysis produced results consistent with the in-situ activity
292 experiments. Illumina Miseq results from samples collected on day 396 (toward the end of
293 Phase 3) indicate that the granules continued to contain AMX, AOB and NOB but with a
294 lower relative abundance compared to the granulation phase. In addition, granule size did not
295 significantly change during Phase 2 (0.97 ± 0.05 mm) and Phase 3 (0.92 ± 0.02 mm) (two-tail t-
296 Test $p=0.1$). This suggests that the shift in performance and establishment of nitrite oxidation
297 was motivated primarily by the small change in bulk liquid DO, which could have supported
298 higher NOB activity and caused loss of NOB suppression.

299 3.4. Phase 4: Ammonium-based aeration control (ABAC)

300 Phase 2 performance showed that a low bulk DO concentration could maintain a higher
301 nitrite concentration and support anammox metabolism; however, nitrite was highly variable
302 and made anammox-based total N removal vulnerable to instability. The variability observed
303 likely occurred because the available ammonium was periodically used up before the end of a
304 cycle, which would have reduced AMX growth and supported NOB growth. Hence, ABAC
305 was implemented during Phase 4 to maintain a minimum residual ammonium concentration
306 throughout each reaction cycle to create a condition that suppressed NOB and supported
307 AOB and AMX activity (Lotti et al., 2014; Pérez, Lotti, Kleerebezem, Picioreanu, & van
308 Loosdrecht, 2014). For an SBR, the suppression of NOB by residual ammonium is much
309 more pronounced at the beginning of the reaction phase than the end given the concentration
310 gradient in the reactor. This also allows for sufficient free ammonia to be present that can
311 inhibit NOB, despite the eventual decrease in total ammonia-N by the end of each reaction
312 cycle. The ABAC strategy used an $\text{NH}_4^+/\text{NO}_3^-$ sensor to maintain a residual ammonium
313 concentration around 5 mg/L as N, and a DO sensor to maintain a low bulk DO setpoint (0.5
314 mg O_2/L). This strategy reduced variability in bulk liquid nutrient concentrations and
315 increase aeration efficiency.

316 With ABAC, we saw a significant shift in both performance and microbial community
317 population composition. Detectable effluent nitrite (16.0 ± 1.6 mg-N/L) and nitrate
318 (2.26 ± 0.72 mg-N/L) resulted in a high NAR of 0.88 ± 0.04 . Also, a lower residual ammonium
319 concentration of 3.5 ± 1.3 mg/L was maintained. The average overall inorganic nitrogen
320 removal efficiency improved slightly to 54.8 ± 3.3 % compared to the other phases (**Figure**
321 **5**); however, the improvement was not statistically significant compared to both Phase 2
322 ($p=0.12$) and Phase 3 ($p=0.75$). Nevertheless, we saw evidence of improved anammox

323 activity. From the in-situ nitrification test on day 459 (Table SI-8), AOB (0.31 g N-NO_2^-
324 formed/g VSS/day) were 3.5 times more active than NOB (0.09 g N-NO_3^- formed/g
325 VSS/day), while the in-situ anammox activity test performed on day 463 resulted in an
326 anammox specific activity of $0.153 \text{ mg-N/mg-VSS day}$, which is about 1.5 times faster than
327 Phase 2 and almost 2 times faster than Phase 3 (See Table SI-4). From the cross-cycle data
328 (Tables SI25-26) analysis, we estimated AMX contributed at least 40% of TIN removed.
329 Illumina Miseq analysis (Day 460, Figure 3) provided additional evidence that the anammox
330 population had recovered significantly during Phase 4 to a relative abundance of 13% while
331 OHOs declined. Together, the performance and microbial community data demonstrate that
332 coupling three aspects of SMC (low DO setpoint, intermittent aeration, and ABAC) created a
333 favorable condition for partial nitrification/anammox in a granular system that received a
334 simulated dilute mainstream anaerobic effluent and reduced the variability in TIN removal
335 (**Figure 4**).

336 Our data also suggests that implementing ABAC improved aeration efficiency, as shown
337 in **Figure 5**. To take a closer look at the improvement made by ABAC in our system, we
338 show sensor-based performance measurements from the reactor for 50 consecutive cycles as
339 we transitioned from Phase 3 to Phase 4. ABAC determined the aerobic duration by limiting
340 aeration up until the residual ammonium dropped below the setpoint of. This caused the
341 overall aerobic duration to be shorter as compared to using a DO-based setpoint only. In this
342 case, the overall aerobic duration was reduced by up to 25% and on average by 15% with the
343 use of ABAC versus DO setpoint operation with fixed aerobic duration (**Figure 5b**).

344 4. DISCUSSION

345 4.1. Stable granulation is possible in a B-stage nitrogen removing GSR system

346 Our results show that it is possible to produce stable granules in an aerobic granular
347 sludge system receiving a dilute wastewater containing residual organic carbon and
348 ammonium. Mature granules with a mean diameter of 1 mm were developed within two
349 months of operation on a synthetic feed with a COD loading of $0.2 \text{ kg m}^{-3} \text{ d}^{-1}$ and nitrogen
350 loading of $0.1 \text{ kg N m}^{-3} \text{ d}^{-1}$. The larger granules are likely to have a larger intra-granular
351 anaerobic zone that can support anaerobic metabolisms. Generally, granules are much easier
352 to develop when the organic loading rate is higher than $1 \text{ kg m}^{-3} \text{ d}^{-1}$ (Jafari Kang & Yuan,
353 2017; Tay, Pan, He, & Tay, 2004). A few studies have reported successful granulation at low
354 organic loading rates (between 0.4 and $1 \text{ kg m}^{-3} \text{ d}^{-1}$) but with a longer time for stable

355 granulation (65 to 120 days) than what was observed in this study (60 days) (Ni et al., 2009;
356 C. Zhang, Zhang, & Yang, 2015). Therefore, with this study we demonstrated that it is
357 possible to produce granules at low organic loading conditions pertinent to mainstream
358 applications.

359 The organic carbon load to the GSR played two major roles. First, it was a major
360 contributor for N removal in all phases. Using stoichiometry, we estimate that it functioned
361 as an electron donor and contributed around 60%, 64% and 60% of the oxidized TIN removal
362 observed during Phases 2, 3 and 4, respectively. The loss of organic carbon due to its
363 reaction with oxidized TIN was limited to the first anoxic period and the availability of
364 residual NO_x . Any residual organic carbon present in the first aerobic zone was oxidized by
365 O_2 . Second, the organic carbon load to the GSR provided the minimum loading needed to
366 develop and sustain granules in our dilute system. Peyong (2012) observed that reducing the
367 organic loading rate below $0.54 \text{ kg COD m}^{-3}\text{day}^{-1}$ resulted in disintegration of granules and
368 the subsequent loss of biomass. Hence, our study showed that it is indeed possible to
369 maintain a stable granular system with organic loading as low as $0.2 \text{ kg COD m}^{-3}\text{day}^{-1}$.

370 4.2. Low bulk DO with intermittent aeration supported NOB suppression

371 Operating the reactor at a bulk DO setpoint of 0.5 mg/L combined with intermittent
372 aeration effectively suppressed NOB activity. The NAR observed during Phases 2 and 4 was
373 significantly higher than Phase 3 ($p < 0.001$) and indicates that operating at low bulk DO was
374 key to effective NOB suppression (Figure 4). Suppression of NOB by low DO is also known
375 to occur in both activated sludge (Peng & Zhu, 2006) and biofilm systems (Brockmann &
376 Morgenroth, 2010; Ma et al., 2015). Rapid intermittent aeration also suppressed NOB
377 because they are known to adapt slowly under transient conditions when shifting from an
378 anaerobic to an aerobic environment, and leads to an accumulated growth disadvantage
379 (Gilbert et al., 2014; Kornaros et al., 2010; Regmi et al., 2014). Concurrently, specific to our
380 system, the quick loss of both VFA and methane means the residual ammonia creates a
381 condition that supports the growth of AOB and ANX more so than NOB for most of the
382 reaction cycle. On top of this, the presence of residual ammonium between 2 and 5 mg NH_4^+
383 N/L has been reported to differentially limit the activity of NOB relative to that of AOB
384 (Pérez et al., 2014; Poot, Hoekstra, Geleijnse, van Loosdrecht, & Pérez, 2016). Therefore,
385 our SMC operation strategy with low and intermittent DO mainly favored AOB while
386 suppressing NOB.

387 The success of using low bulk DO to achieve NOB suppression can be attributed in part
388 to the known differences in growth rate between the AOB and NOB genera present in our
389 system. A 16S rRNA gene-based community analysis showed that the only NOB types
390 detected in our reactor are from the genus *Nitrospira*, which are typically found to have a
391 lower maximum specific growth rate and lower oxygen affinity (Blackburne, Vadivelu,
392 Yuan, & Keller, 2007) than the AOB detected in our system, the genus *Nitrosomonas*, which
393 are typically found to have a higher maximum specific growth rate and oxygen affinity
394 (Blackburne, Yuan, & Keller, 2008). These two genera of bacteria have been found to coexist
395 in many partial nitrification systems, as summarized by Cao et al. (2017) and reported by other
396 authors (Sinha, Ajit, & Annachhatre, 2006; Wett et al., 2013). Hence, the AOB in our system
397 are likely to have a higher oxygen affinity under low DO conditions than the NOB, resulting
398 in suppression of the latter.

399 4.3. Coupling ABAC with low DO setpoint enhanced energy efficient, anammox-supported 400 N removal

401 The performance of the GSR was stable and aeration energy demand was reduced with
402 the addition of ABAC. When the reactor was operated without residual ammonium control
403 during Phase 3, it was not possible to consistently maintain residual ammonium through the
404 end of the reaction cycle; hence, the residual ammonium concentration tended to vary
405 substantially (**Figure 5a**). This is undesirable, since a minimum residual ammonium
406 concentration is needed throughout the reaction zones for successful partial nitrification. Thus,
407 this underscores the benefit brought by ABAC to ensure that an ammonium residue is
408 maintained throughout the reaction time. When we added ABAC, aeration duration was
409 reduced by up to 25% relative to what occurred when we used a DO setpoint only (**Figure**
410 **5b**). This reduction in aerobic duration translates into a reduction in aeration energy cost.
411 Consequently, the use of ABAC resulted in tighter aeration control, which yielded more
412 stable residual ammonium and overall TIN removal performance for the system. Translated
413 to full-scale treatment systems that often have a dynamic influent composition, these results
414 imply that the use of ABAC will be critical to the cost-effective deployment of mainstream
415 B-stage GSR applications that must achieve stable nitrogen reduction.

416 The use of ABAC with low DO setpoint and intermittent aeration also improved the
417 retention of AMX in our system. Whole community (16S rRNA gene) sequencing data
418 showed that coupling ABAC with low intermittent DO setpoint control corresponded with the
419 recovery of AMX to around 13% relative abundance, four times higher than was seen without

420 ABAC (**Figure 3**). The increase in AMX relative abundance corresponded with a 1.5-fold
421 increase in the in-situ rate of anammox specific activity relative to what was observed during
422 Phase 2 (low-DO without ABAC). Furthermore, the specific anammox rate measured during
423 Phase 4 (low DO with ABAC, 0.153 mg-N/mg-VSS-day) is similar the rate reported by Lotti
424 et al. (2015b) for a partial nitrification/anammox SBR controlled with low DO and ABAC at
425 20°C (0.11 mg-N/mg-VSS day) and at 25°C (0.14 mg-N/mg-VSS day). To achieve N-
426 removal via nitrite by suppressing NOB, other studies have used DO, pH and ORP in aerobic
427 granular sludge reactors (Lochmatter, Gonzalez-Gil, & Holliger, 2013; Tao, Gao, Fu, Wu, &
428 Ren, 2012) while DO and ABAC have been used in conventional activated sludge systems
429 (Regmi et al., 2015, 2014) to control DO setpoint and aerobic duration. All of these studies
430 had higher organic and/or nitrogen volumetric loading than this study. Here, we developed
431 and demonstrated a SMC strategy that integrated a DO setpoint, intermittent aeration and
432 residual ammonia control to promote partial nitrification/anammox in a mainstream GSR fed
433 with dilute wastewater to achieve N-removal with reduced aeration expense.

434 4.4. Less aggressive start-up is required for better nitrogen removal

435 The manner with which the GSR was operated during granulation influenced the system's
436 ability to retain anammox activity. As the 16S rRNA sequencing results show, AMX was
437 substantially reduced in abundance during the granulation period, consistent with the
438 corresponding higher nitrite accumulation and lower levels of nitrate. We believe the
439 observed reduction in AMX relative abundance has to do with two unfavorable start-up
440 conditions. First, during the start-up period granules were developed with a short settling time
441 to select against flocculent sludge. This created a short residence time of 4.8 ± 0.6 days
442 (Figure SI-1), which was less than the minimum reported doubling time of 11 days for AMX
443 (Strous et al., 1998) in an SBR, and this possibly caused washout within the first few days
444 before granules started developing. While others have predicted that SRTs as short as three
445 days are possible (Lotti et al., 2015a; Zhang et al., 2017), we did not observe that with our
446 data. Second, since the influent contained only ammonium and not nitrite, AMX growth had
447 to rely on AOB activity and achieving nitrite accumulation, neither of which was stable
448 during the start-up period.

449 Collectively, these results suggest that a less aggressive start up condition is required to
450 retain a higher percentage AMX population. A less aggressive start-up condition could be
451 implemented to include: (i) gradually decreasing the settling time to maintain an adequate

452 SRT until granules start to appear; (ii) supplementing the feed with nitrite during start-up
453 until the initial development of granules is observed; and (iii) incorporating an anaerobic
454 phase at the beginning of the run. The last two actions were demonstrated by Winkler et al.
455 (2012) for integration of anammox to out-compete acetate in a granular system that received
456 a higher organic ($0.6 \text{ kg-COD/m}^3/\text{d}$) and N ($1.14 \text{ kg-N/m}^3/\text{d}$) loading than this study. In
457 addition to these actions, start-up can be further improved by incorporating intermittent
458 aeration to match the rate of nitrification with the rate of anammox activity in the system. This
459 can be achieved by using a SMC to dynamically adjust the aerobic and anaerobic durations to
460 promote partial nitrification/anammox while suppressing NOB. Further studies are needed to
461 demonstrate the viability of these ideas.

462 **5. Conclusions**

463 We demonstrated that successful N-removal from a dilute mainstream wastewater requires a
464 robust real-time control strategy for effective utilization of resources and reduced energy
465 expense. We showed that it is possible to develop a granular sludge in a low carbon-loaded
466 system that can effectively suppress NOB activity so that N-removal can be achieved via
467 partial nitrification/anammox. Key operational strategies were identified and include using low
468 DO intermittent aeration with ABAC (i.e. to maintain minimum residual ammonium). The
469 findings of this research indicate that it is possible to remove nitrogen in a single compact
470 system and with less aeration energy expended if simultaneous nitrification, anammox and
471 heterotrophic denitrification are enabled with the assistance of SMC.

472 **Practitioner points**

- 473 • Tight sensor-mediated aeration control is need for better PN/A
- 474 • Low DO intermittent aeration with minimum ammonium residual results in a stable N
475 removal
- 476 • Low DO aeration results in a stable NOB suppression
- 477 • Using sensor-mediated aeration control in a granular sludge reactor reduces aeration
478 cost

479 **Acknowledgements**

480 This work was supported through NSF GOALI grant number CBET1438560. Any opinions,
481 findings and conclusions or recommendations expressed in this material are those of the
482 authors and do not necessarily reflect those of the National Science Foundation. Zerihun

483 Bekele was also supported by Hampton Roads Sanitation District and the Department of
484 Civil and Environmental Engineering. Jeseth Delgado was supported through an NSF
485 Graduate Research Fellowship, a Rackham Engineering Merit Fellowship, and a Ford
486 Foundation Fellowship. The authors would like to thank Robert Smith (Xylem Inc.) for
487 providing the ammonium/nitrate sensors used in this work. The authors also thank Adriana
488 Arcelay and Yan Du for their assistance in the laboratory. The authors declare no conflict of
489 interest.

490 REFERENCES

- 491 Barr, J. J., Cook, A. E., & Bond, P. L. (2010). Granule formation mechanisms within an
492 aerobic wastewater system for phosphorus removal. *Applied and Environmental*
493 *Microbiology*, 76(22), 7588–7597. <https://doi.org/10.1128/AEM.00864-10>
- 494 Blackburne, R., Vadivelu, V. M., Yuan, Z., & Keller, J. (2007). Kinetic characterisation of an
495 enriched *Nitrospira* culture with comparison to *Nitrobacter*. *Water Research*, 41(14),
496 3033–3042. <https://doi.org/10.1016/J.WATRES.2007.01.043>
- 497 Blackburne, R., Yuan, Z., & Keller, J. (2008). Demonstration of nitrogen removal via nitrite
498 in a sequencing batch reactor treating domestic wastewater. *Water Research*, 42(8–9),
499 2166–2176. <https://doi.org/10.1016/J.WATRES.2007.11.029>
- 500 Brockmann, D., & Morgenroth, E. (2010). Evaluating operating conditions for outcompeting
501 nitrite oxidizers and maintaining partial nitrification in biofilm systems using biofilm
502 modeling and Monte Carlo filtering. *Water Research*, 44, 1995–2009.
503 <https://doi.org/10.1016/j.watres.2009.12.010>
- 504 Cao, Y., van Loosdrecht, M. C. M., & Daigger, G. T. (2017). Mainstream partial nitrification–
505 anammox in municipal wastewater treatment: status, bottlenecks, and further studies.
506 *Applied Microbiology and Biotechnology*, 101(4), 1365–1383.
507 <https://doi.org/10.1007/s00253-016-8058-7>
- 508 Daigger, G. T. (2014). Oxygen and Carbon Requirements for Biological Nitrogen Removal
509 Processes Accomplishing Nitrification, Nitritation, and Anammox. *Water Environment*
510 *Research*, 86(3), 204–209. <https://doi.org/10.2175/106143013X13807328849459>
- 511 de Graaff, M. S., van den Brand, T. P. H., Roest, K., Zandvoort, M. H., Duin, O., & van
512 Loosdrecht, M. C. M. (2016). Full-Scale Highly-Loaded Wastewater Treatment
513 Processes (A-Stage) to Increase Energy Production from Wastewater: Performance and
514 Design Guidelines. *Environmental Engineering Science*, 33(8), 571–577.
515 <https://doi.org/10.1089/ees.2016.0022>
- 516 de Kreuk, M. K., Heijnen, J. J., & van Loosdrecht, M. C. M. (2005). Simultaneous COD,

517 nitrogen, and phosphate removal by aerobic granular sludge. *Biotechnology and*
518 *Bioengineering*, 90(6), 761–769. <https://doi.org/10.1002/bit.20470>

519 Delgado Vela, J., Dick, G. J., & Love, N. G. (2018). Sulfide inhibition of nitrite oxidation in
520 activated sludge depends on microbial community composition. *Water Research*, 138,
521 241–249. <https://doi.org/10.1016/j.watres.2018.03.047>

522 Delgado Vela, J., Stadler, L. B., Martin, K. J., Raskin, L., Bott, C. B., & Love, N. G. (2015).
523 Prospects for Biological Nitrogen Removal from Anaerobic Effluents during
524 Mainstream Wastewater Treatment. *Environmental Science & Technology Letters*, 2(9),
525 234–244. <https://doi.org/10.1021/acs.estlett.5b00191>

526 Diamantis, V., Verstraete, W., Eftaxias, A., Bundervoet, B., Vlaeminck, S. E., Melidis, P., &
527 Aivasidis, A. (2013). Sewage pre-concentration for maximum recovery and reuse at
528 decentralized level. *Water Science and Technology*, 67(6), 1188–1193.
529 <https://doi.org/10.2166/wst.2013.639>

530 Gilbert, E. M., Agrawal, S., Brunner, F., Schwartz, T., Horn, H., & Lackner, S. (2014).
531 Response of different *Nitrospira* Species to anoxic periods depends on operational DO.
532 *Environmental Science and Technology*, 48(5). <https://doi.org/10.1021/es404992g>

533 Jafari Kang, A., & Yuan, Q. (2017). Long-term stability and nutrient removal efficiency of
534 aerobic granules at low organic loads. *Bioresource Technology*, 234, 336–342.
535 <https://doi.org/10.1016/J.BIORTECH.2017.03.057>

536 Jetten, M. S. M., Horn, S. J., & van Loosdrecht, M. C. M. (1997). Towards a more
537 sustainable municipal wastewater treatment system. *Water Science and Technology*,
538 35(9). Retrieved from <http://wst.iwaponline.com/content/35/9/171>

539 Jimenez, J., Miller, M., Bott, C., Murthy, S., De Clippeleir, H., & Wett, B. (2015). High-rate
540 activated sludge system for carbon management – Evaluation of crucial process
541 mechanisms and design parameters. *Water Research*, 87, 476–482.
542 <https://doi.org/10.1016/J.WATRES.2015.07.032>

543 Kornaros, M., Dokianakis, S. N., & Lyberatos, G. (2010). Partial nitrification/denitrification
544 can be attributed to the slow response of nitrite oxidizing bacteria to periodic anoxic
545 disturbances. *Environmental Science & Technology*, 44(19), 7245–7253.
546 <https://doi.org/10.1021/es100564j>

547 Kozich, J. J., Westcott, S. L., Baxter, N. T., Highlander, S. K., & Schloss, P. D. (2013).
548 Development of a Dual-Index Sequencing Strategy and Curation Pipeline for Analyzing
549 Amplicon Sequence Data on the MiSeq Illumina Sequencing Platform. *Applied and*
550 *Environmental Microbiology*, 79(17), 5112–5120. <https://doi.org/10.1128/AEM.01043->

- 552 Lemaire, R., Marcelino, M., & Yuan, Z. (2008). Achieving the nitrite pathway using aeration
553 phase length control and step-feed in an SBR removing nutrients from abattoir
554 wastewater. *Biotechnology and Bioengineering*, 100(6), 1228–1236.
555 <https://doi.org/10.1002/bit.21844>
- 556 Liu, Y., Yang, S.-F., & Tay, J.-H. (2003). Elemental compositions and characteristics of
557 aerobic granules cultivated at different substrate N/C ratios. *Applied Microbiology and*
558 *Biotechnology*, 61(5–6), 556–561. <https://doi.org/10.1007/s00253-003-1246-2>
- 559 Lochmatter, S., Gonzalez-Gil, G., & Holliger, C. (2013). Optimized aeration strategies for
560 nitrogen and phosphorus removal with aerobic granular sludge. *Water Research*, 47(16),
561 6187–6197. <https://doi.org/10.1016/j.watres.2013.07.030>
- 562 Lotti, T., Kleerebezem, R., Abelleira-Pereira, J. M., Abbas, B., & van Loosdrecht, M. C. M.
563 (2015). Faster through training: The anammox case. *Water Research*, 81, 261–268.
564 <https://doi.org/10.1016/j.watres.2015.06.001>
- 565 Lotti, T., Kleerebezem, R., Hu, Z., Kartal, B., Jetten, M., & Van Loosdrecht, M. C. M.
566 (2014). Simultaneous partial nitrification and anammox at low temperature with granular
567 sludge. *Water Research*, 66, 111–121. <https://doi.org/10.1016/j.watres.2014.07.047>
- 568 Lotti, T., Kleerebezem, R., & Loosdrecht, M. C. M. Van. (2015). Effect of Temperature
569 Change on Anammox Activity. *Biotechnology and Bioengineering*, 112(1), 98–103.
570 <https://doi.org/10.1002/bit.25333>
- 571 Lotti, T., Kleerebezem, R., & van Loosdrecht, M. C. M. (2015). Effect of temperature change
572 on anammox activity. *Biotechnology and Bioengineering*, 112(1), 98–103.
573 <https://doi.org/10.1002/bit.25333>
- 574 Ma, B., Bao, P., Wei, Y., Zhu, G., Yuan, Z., & Peng, Y. (2015). Suppressing Nitrite-
575 oxidizing Bacteria Growth to Achieve Nitrogen Removal from Domestic Wastewater
576 via Anammox Using Intermittent Aeration with Low Dissolved Oxygen. *Nature*
577 *Publishing Group*. <https://doi.org/10.1038/srep13048>
- 578 Miller, M. W., DeArmond, J., Elliott, M., Kinyua, M., Kinnear, D., Wett, B., ... Bott, C. B.
579 (2016). Settling and dewatering characteristics of an A-stage activated sludge process
580 proceeded by shortcut biological nitrogen removal. *International Journal of Water and*
581 *Wastewater Treatment*, 2(5), 1–8. [https://doi.org/http://dx.doi.org/10.16966/2381-](https://doi.org/http://dx.doi.org/10.16966/2381-5299.133)
582 [5299.133](https://doi.org/http://dx.doi.org/10.16966/2381-5299.133)
- 583 Ni, B.-J., Xie, W.-M., Liu, S.-G., Yu, H.-Q., Wang, Y.-Z., Wang, G., & Dai, X.-L. (2009).
584 Granulation of activated sludge in a pilot-scale sequencing batch reactor for the

585 treatment of low-strength municipal wastewater. *Water Research*, 43(3), 751–761.
586 <https://doi.org/10.1016/j.watres.2008.11.009>

587 Peng, Y., & Zhu, G. (2006). Biological nitrogen removal with nitrification and denitrification
588 via nitrite pathway. *Applied Microbiology and Biotechnology*, 15–26.
589 <https://doi.org/10.1007/s00253-006-0534-z>

590 Pérez, J., Lotti, T., Kleerebezem, R., Picioreanu, C., & van Loosdrecht, M. C. M. (2014).
591 Outcompeting nitrite-oxidizing bacteria in single-stage nitrogen removal in sewage
592 treatment plants: A model-based study. *Water Research*, 66, 208–218.
593 <https://doi.org/10.1016/J.WATRES.2014.08.028>

594 Peyong, Y. N., Zhou, Y., Abdullah, A. Z., & Vadivelu, V. (2012). The effect of organic
595 loading rates and nitrogenous compounds on the aerobic granules developed using low
596 strength wastewater. *Biochemical Engineering Journal*, 67, 52–59.
597 <https://doi.org/10.1016/J.BEJ.2012.05.009>

598 Philips, S., Laanbroek, H. J., & Verstraete, W. (2002). Origin, causes and effects of increased
599 nitrite concentrations in aquatic environments. *Reviews in Environmental Science and*
600 *Biotechnology*. <https://doi.org/10.1023/A:1020892826575>

601 Poot, V., Hoekstra, M., Geleijnse, M. A. A., van Loosdrecht, M. C. M., & Pérez, J. (2016).
602 Effects of the residual ammonium concentration on NOB repression during partial
603 nitrification with granular sludge. *Water Research*, 106, 518–530.
604 <https://doi.org/10.1016/j.watres.2016.10.028>

605 Raghoebarsing, A. A., Pol, A., Van De Pas-Schoonen, K. T., Smolders, A. J. P., Ettwig, K.
606 F., Rijpstra, W. I. C., ... Strous, M. (2006). A microbial consortium couples anaerobic
607 methane oxidation to denitrification. *Nature*, 440(7086), 918–921.
608 <https://doi.org/10.1038/nature04617>

609 Regmi, P., Bunce, R., Miller, M. W., Park, H., Chandran, K., Wett, B., ... Bott, C. B. (2015).
610 Ammonia-based intermittent aeration control optimized for efficient nitrogen removal.
611 *Biotechnology and Bioengineering*. <https://doi.org/10.1002/bit.25611>

612 Regmi, P., Miller, M. W., Holgate, B., Bunce, R., Park, H., Chandran, K., ... Bott, C. B.
613 (2014). Control of aeration, aerobic SRT and COD input for mainstream
614 nitrification/denitrification. *Water Research*, 57, 162–171.
615 <https://doi.org/10.1016/j.watres.2014.03.035>

616 Sarma, S. J., Tay, J. H., & Chu, A. (2017). Finding Knowledge Gaps in Aerobic Granulation
617 Technology. *Trends in Biotechnology*. <https://doi.org/10.1016/j.tibtech.2016.07.003>

618 Schloss, P. D., Westcott, S. L., Ryabin, T., Hall, J. R., Hartmann, M., Hollister, E. B., ...

- 619 Weber, C. F. (2009). Introducing mothur: Open-source, platform-independent,
620 community-supported software for describing and comparing microbial communities.
621 *Applied and Environmental Microbiology*, 75(23), 7537–7541.
622 <https://doi.org/10.1128/AEM.01541-09>
- 623 Sinha, B., Ajit, A., & Annachhatre, P. (2006). Partial nitrification—operational parameters
624 and microorganisms involved. <https://doi.org/10.1007/s11157-006-9116-x>
- 625 Smith, A. L., Skerlos, S. J., & Raskin, L. (2013). Psychrophilic anaerobic membrane
626 bioreactor treatment of domestic wastewater. *Water Research*, 47, 1655–1665.
627 <https://doi.org/10.1016/j.watres.2012.12.028>
- 628 Smith, A. L., Stadler, L. B., Cao, L., Love, N. G., Raskin, L., & Skerlos, S. J. (2014).
629 Navigating Wastewater Energy Recovery Strategies: A Life Cycle Comparison of
630 Anaerobic Membrane Bioreactor and Conventional Treatment Systems with Anaerobic
631 Digestion. *Environmental Science & Technology*, 48, 5972–5981.
632 <https://doi.org/10.1021/es5006169>
- 633 Souza, T. S. O., & Foresti, E. (2013). Sulfide-Oxidizing Autotrophic Denitrification: an
634 Evaluation for Nitrogen Removal from Anaerobically Pretreated Domestic Sewage.
635 *Applied Biochemistry and Biotechnology*, 170(5), 1094–1103.
636 <https://doi.org/10.1007/s12010-013-0261-8>
- 637 Strous, M., Jetten, M. S. M., Heijnen, J. J., & Kuenen, J. G. (1998). The sequencing batch
638 reactor as a powerful tool for the study of slowly growing anaerobic ammonium-
639 oxidizing microorganisms. *Applied Microbiology and Biotechnology*, 50(1998), 589–
640 596. <https://doi.org/10.1007/s002530051340>
- 641 Szabó, E., Liébana, R., Hermansson, M., Modin, O., Persson, F., & Wilén, B. M. (2017).
642 Microbial population dynamics and ecosystem functions of anoxic/aerobic granular
643 sludge in sequencing batch reactors operated at different organic loading rates. *Frontiers*
644 *in Microbiology*, 8(MAY). <https://doi.org/10.3389/fmicb.2017.00770>
- 645 Tao, Y., Gao, D.-W., Fu, Y., Wu, W.-M., & Ren, N.-Q. (2012). Impact of reactor
646 configuration on anammox process start-up: MBR versus SBR. *Bioresource*
647 *Technology*, 104, 73–80. <https://doi.org/10.1016/j.biortech.2011.10.052>
- 648 Tay, J.-H., Pan, S., He, Y., & Tay, S. T. L. (2004). Effect of Organic Loading Rate on
649 Aerobic Granulation. I: Reactor Performance. *Journal of Environmental Engineering*,
650 130(10), 1094–1101. [https://doi.org/10.1061/\(ASCE\)0733-9372\(2004\)130:10\(1094\)](https://doi.org/10.1061/(ASCE)0733-9372(2004)130:10(1094))
- 651 Wan, J., Gu, J., Zhao, Q., & Liu, Y. (2016). COD capture: a feasible option towards energy
652 self-sufficient domestic wastewater treatment. <https://doi.org/10.1038/srep25054>

- 653 Wett, B., Omari, A., Podmirseg, S. M., Han, M., Akintayo, O., Gómez Brandón, M., ...
654 O'Shaughnessy, M. (2013). Going for mainstream deammonification from bench to full
655 scale for maximized resource efficiency. *Water Science and Technology*, 68(2), 283–
656 289. <https://doi.org/10.2166/wst.2013.150>
- 657 Wett, B., Podmirseg, S. M., Gómez-Brandón, M., Hell, M., Nyhuis, G., Bott, C., & Murthy,
658 S. (2015). Expanding DEMON Sidestream Deammonification Technology Towards
659 Mainstream Application. *Water Environment Research*, 87(12), 2084–2089.
660 <https://doi.org/10.2175/106143015X14362865227319>
- 661 Winkler, M.-K. H., Kleerebezem, R., & van Loosdrecht, M. C. M. (2012). Integration of
662 anammox into the aerobic granular sludge process for main stream wastewater treatment
663 at ambient temperatures. *Water Research*, 46(1), 136–144.
664 <https://doi.org/10.1016/j.watres.2011.10.034>
- 665 Zhang, C., Zhang, H., & Yang, F. (2015). Diameter control and stability maintenance of
666 aerobic granular sludge in an A/O/A SBR. *Separation and Purification Technology*, 149,
667 362–369. <https://doi.org/10.1016/j.seppur.2015.06.010>
- 668 Zhang, L., Narita, Y., Gao, L., Ali, M., Oshiki, M., & Okabe, S. (2017). Maximum specific
669 growth rate of anammox bacteria revisited. *Water Research*, 116, 296–303.
670 <https://doi.org/10.1016/J.WATRES.2017.03.027>
- 671 Zhou, Y., Zhang, D. Q., Le, M. T., Pua, A. N., Ng, W. J., Qing, D., ... Le, T. (2013). Energy
672 utilization in sewage treatment-a review with comparisons.
673 <https://doi.org/10.2166/wcc.2013.117>
- 674
- 675 [dataset] Zerihun A. Bekele; 2019; Granular activated sludge Raw sequence reads; NCBI
676 SRA; Accession: PRJNA549919

677 **Figure Captions**

678 **Figure 1.** Potential metabolic pathways in a B-stage GSR for removal of ammonia, VFA,
679 methane and sulfide present in an anaerobically-treated A-stage. SBD: sulfur based
680 denitrification, MOB: methane oxidizing bacteria, and SOB: sulfur oxidizing bacteria.

681 **Figure 2.** (a) Granules change over time during start-up Phase 1. They began as micro-
682 granules, then became large and fluffy, and finally developed into mature granules. (b)
683 Mature granules with an average size of 1 mm. (c) Granules in the reactor after 5 minutes of
684 settling.

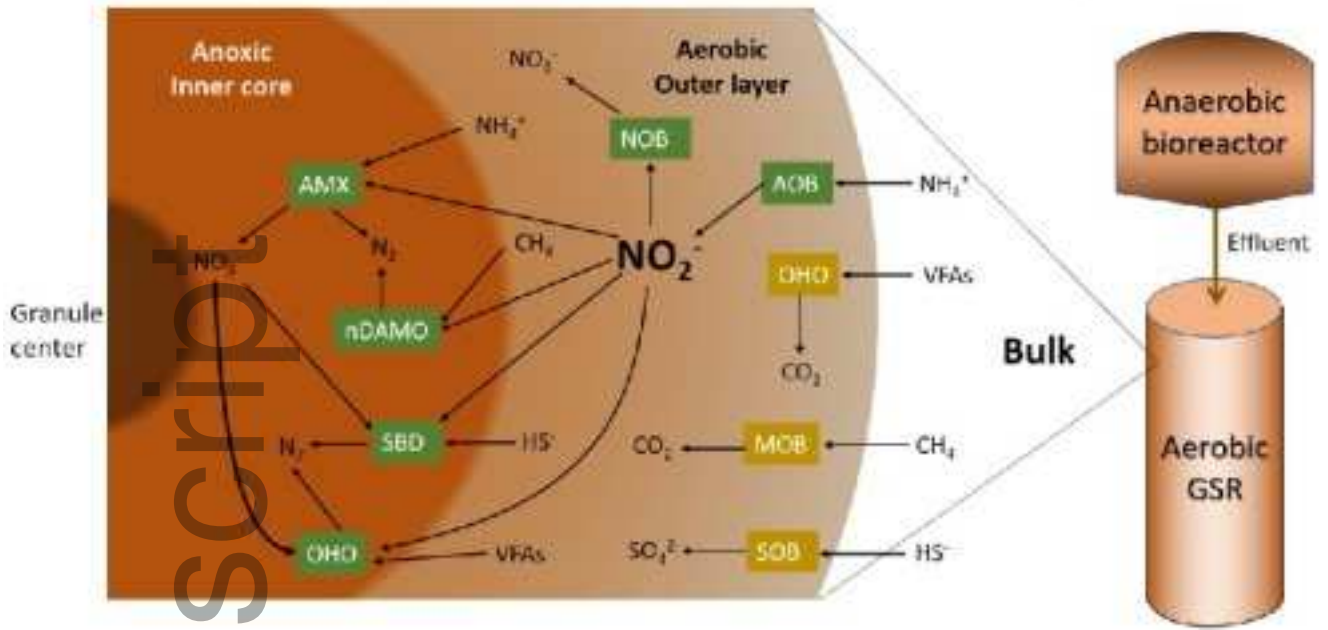
685 **Figure 3.** Microbial composition dynamics at order level OTUs for the period of granule

686 development (through day 50), day 396 which is in phase 3, and day 460 which is in phase 4.
687 Solid greytone colors are OHOs, hatched greytones are other bacteria with either known or
688 unknown functions such as EPS production, hydrolysis and filament formation, and solid
689 non-greytone colors are AOB, NOB, anammox and the order plactomycetales.

690 **Figure 4.** Boxplots showing comparisons across the four operation stages for (a) percent
691 nitrification, (b) percent total inorganic nitrogen removal, and (c) nitrite accumulation ratio
692 (NAR = effluent nitrite-N:effluent [nitrite-N + nitrate-N]). Note: 'x' indicates the mean, the
693 inside horizontal line indicates the median.

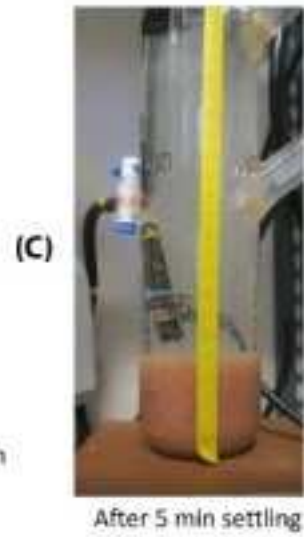
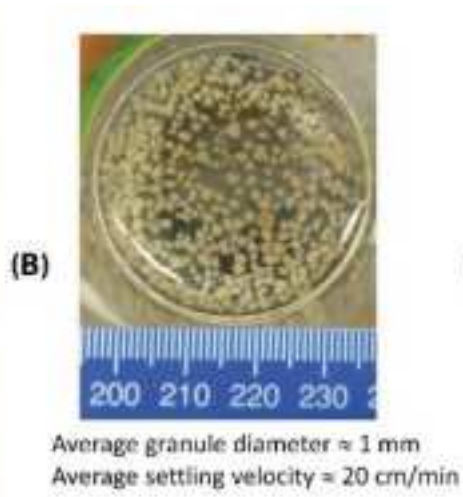
694 **Figure 5.** Reactor operation with and without ABAC (Days from 396 to 417). (a) Effluent
695 nitrogen species concentration profile under both scenarios (measurements determined using
696 sensors and corrected with analytically determined values). (b) The corresponding total
697 aerobic/anoxic duration fraction.

Author Manuscript

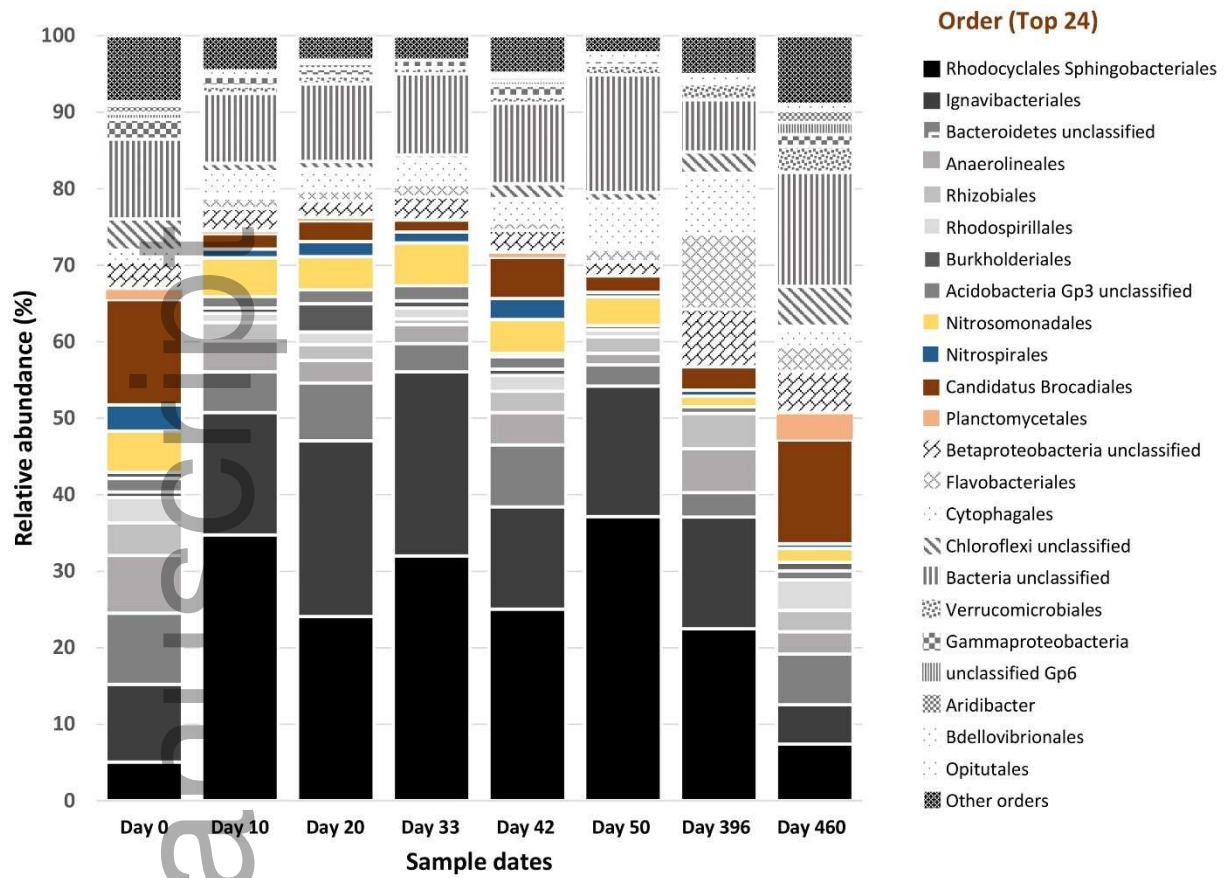


wer_1296_f1.jpg

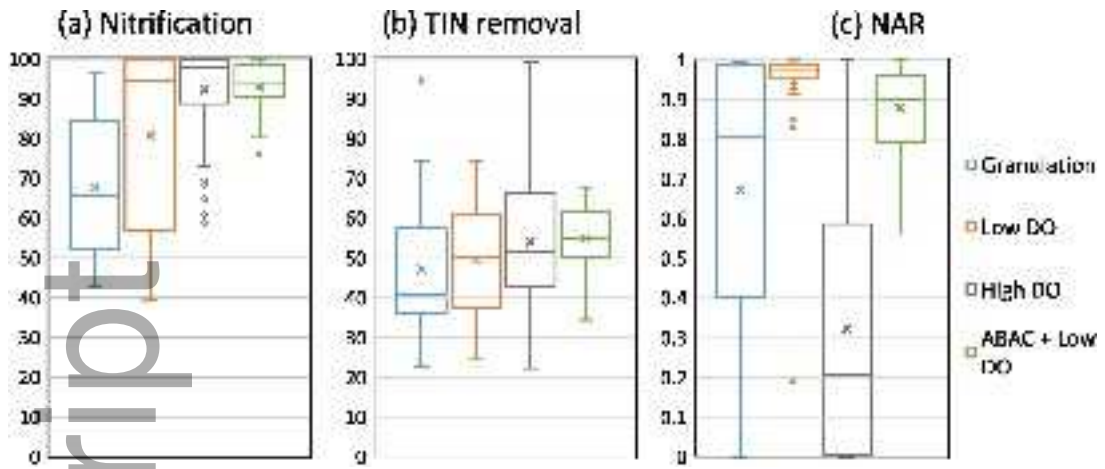
Author Manuscript



wer_1296_f2.jpg

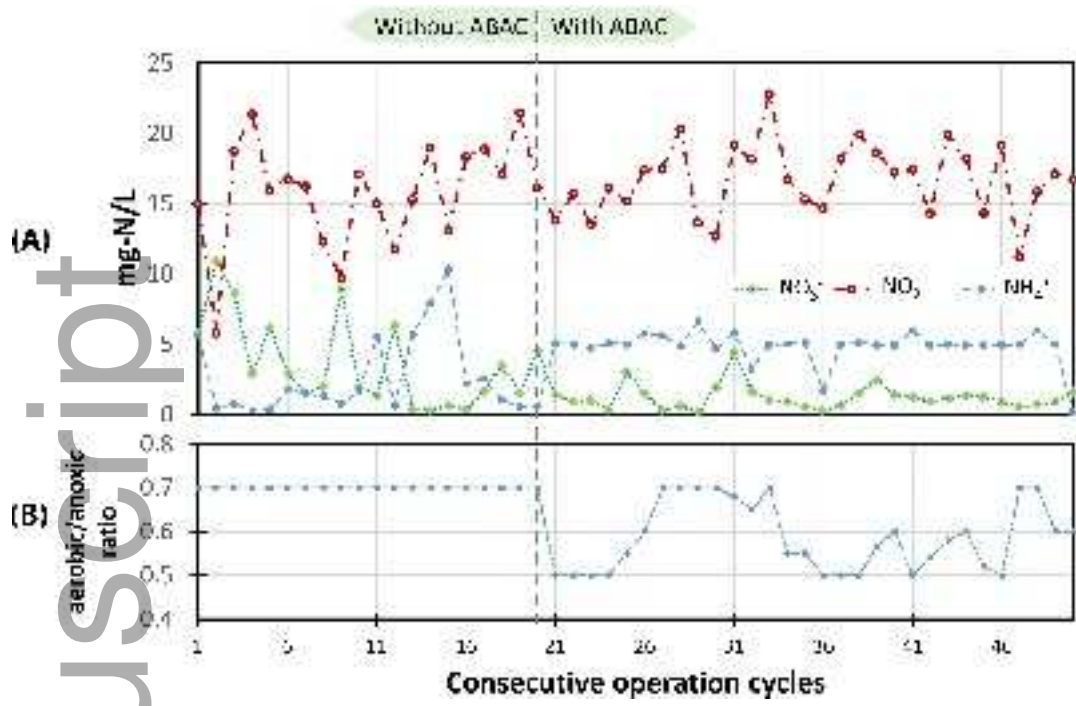


wer_1296_f3.jpg



wer_1296_f4.jpg

Author Manuscript



wer_1296_f5.jpg