

## **Supplementary Materials**

for  
A structured brain-wide and genome-wide association study  
using ADNI PET images

by  
Yanming Li, Bin Nan, Ji Zhu, and for the Alzheimer's Disease  
Neuroimaging Initiative

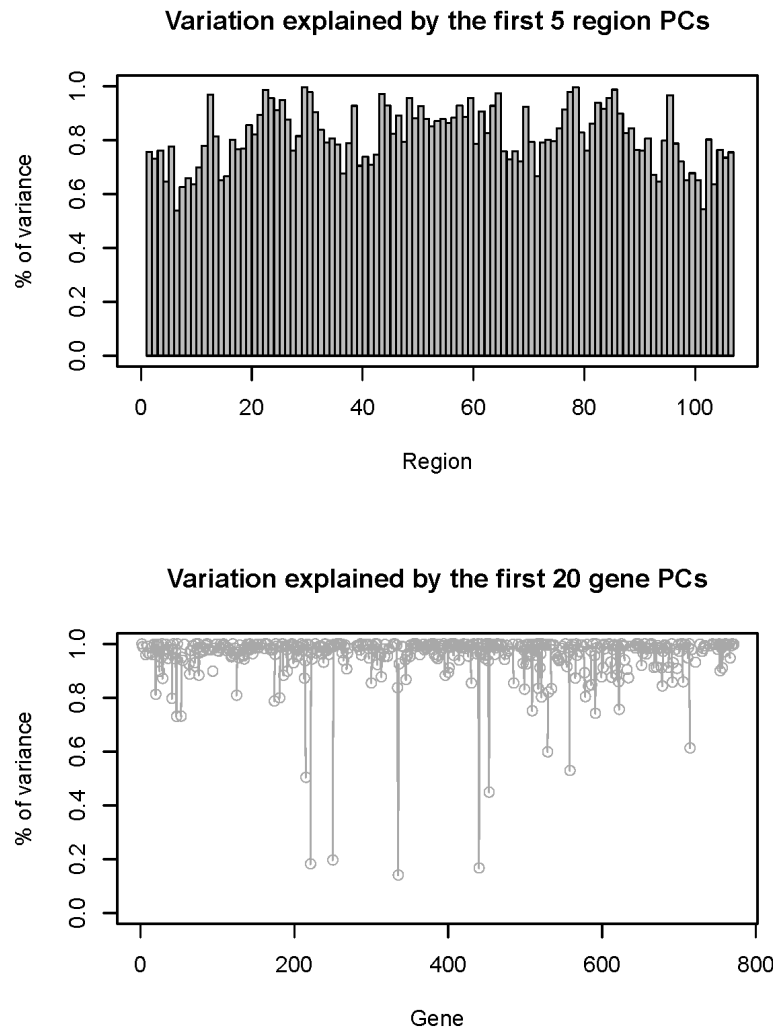
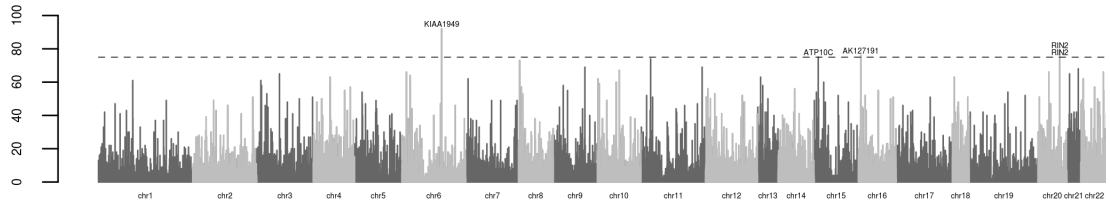
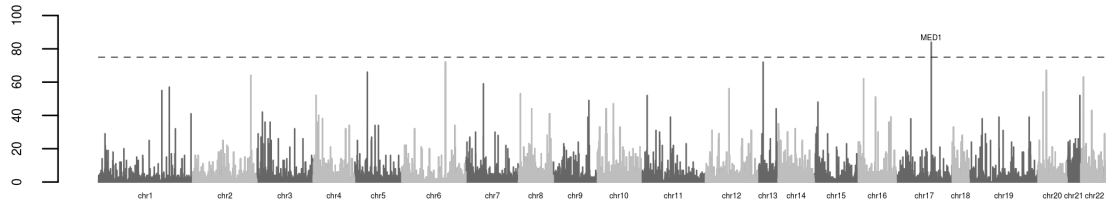


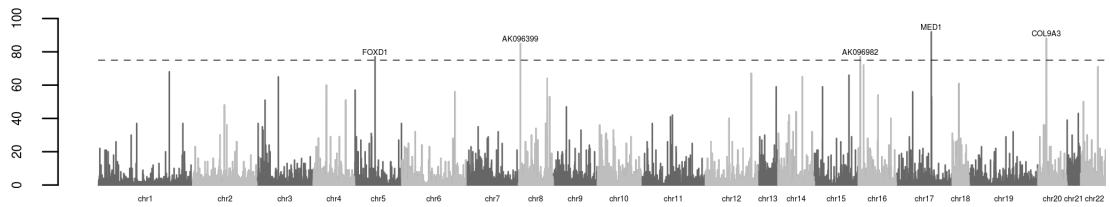
Figure S.1: (a) Percent of variation explained by the first five PCs in each of the 106 ROIs. (b) Percent of variation explained by up to the first 20 PCs for each gene on chromosome 20.



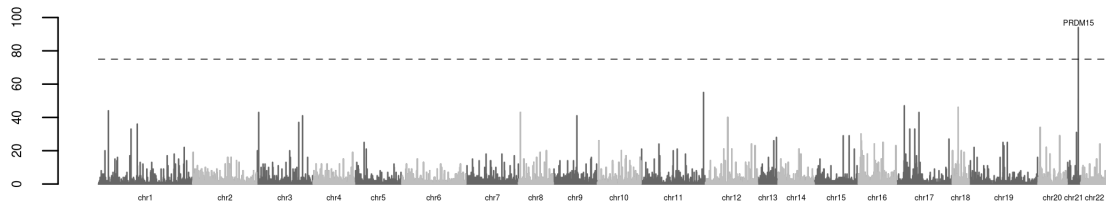
(a) Region: CERHEM(L)



(b) Region: Superior parietal cortex BA39(L)



(c) Region: Superior parietal cortex BA39(R)



(d) Region: Occipital cortex BA17(R)

Figure S.2: Stability selection frequencies. ROI versus the genome. (a) The gene PC effect selection frequency for region CERHEM(L), where the gene *RIN2* has two independent PCs with selection frequencies more than 75% and is therefore selected and passed to the second-stage selection. (b) and (c) The gene $\times$ AD interaction effect selection frequencies on regions BA39(L) and BA39, where the genes *MED1* and *COL9A3* are selected, among a few others. (d) The gene $\times$ MCI interaction effect on region BA17; the gene *PRDM15* is selected into the second stage.

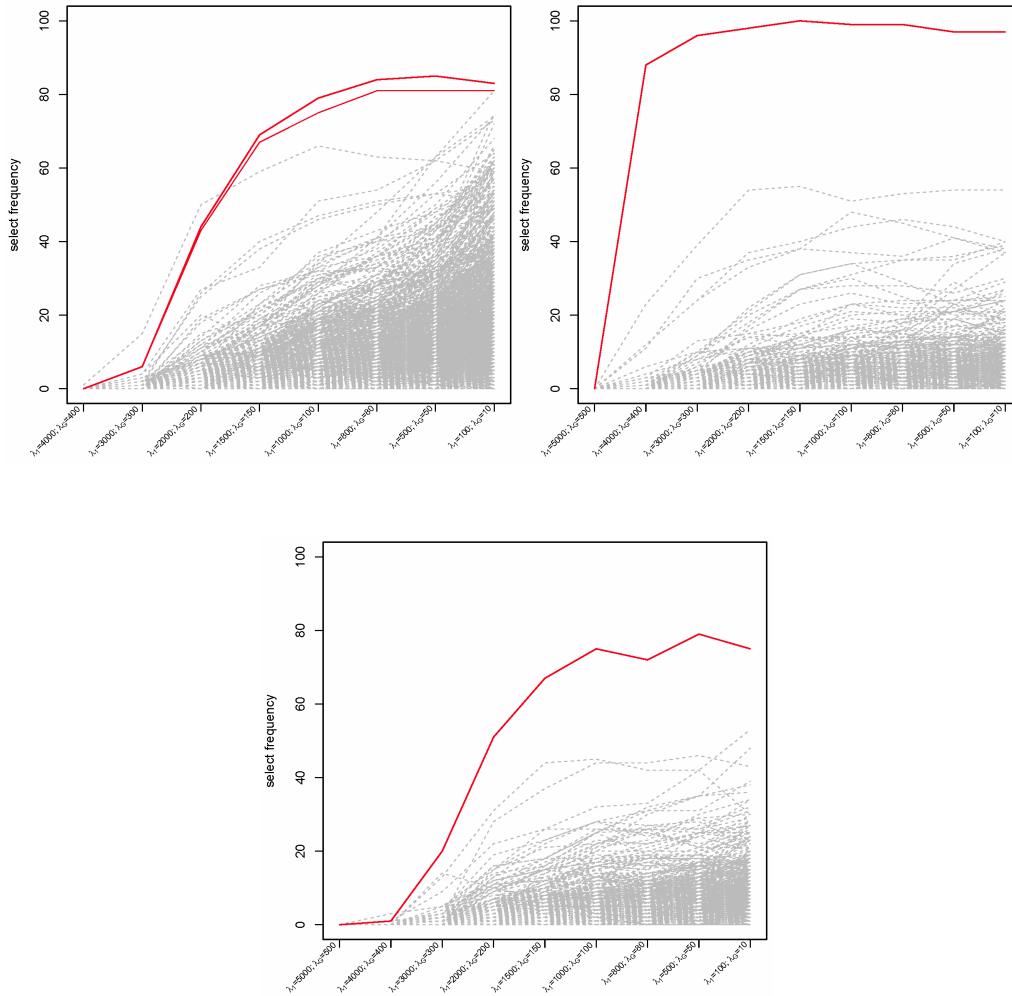


Figure S.3: Illustrating robustness to the tuning parameters of stability selection. Depicted are the selection paths of the regions CERHEM(L) and the occipital cortex BA19(R) on chromosome 20 in the first stage selection. The vertical axis is the select frequency out of 100 bootstrap datasets and the horizontal axis indicates the different settings of the tuning parameter values. We fixed the ratio of the individual level tuning parameter  $\lambda$  to the group level tuning parameter  $\lambda_1$  at 10. The highlighted selection paths are for the top selected regression coefficients. Notice that the top signals are consistently selected from the bootstrapped datasets. (a) The gene effect of the region CERHEM(L). (b) The gene AD interaction effect of the region occipital cortex BA19(R). (c) The gene MCI interaction effect of the region occipital cortex BA19(R).

Table S.1: Some other gene $\times$ AD and gene $\times$ MCI interaction effects of top SNPs that have a  $p$ -value more significant than  $10^{-5}$  and a selection frequency exceeding 80%.

name	gene information			top selective SNP in gene		associated region	effect type	reference
	chr	num. SNP in gene	% variance by 20 PCs	SNP name	most sig. p-value			
<i>HOXD4</i>	2	11	100%	rs2072590	1.6e-06	Medial frontal cortex_BA9(L)	G $\times$ AD	Nolte et al. [10],
<i>AK096399</i>	8	40	98.6%	rs6436025	1.3e-06	Primary somatosensory cortex_BA5(R)	G $\times$ MCI	Cannon et al. [2]
<i>TBC1D4</i>	13	86	87.1%	rs1864726	2.4e-06	Medial frontal cortex_BA8(L)	G $\times$ MCI	Talbot et al. [13], Yang, Li, & Liu [14], Dai et al. [4], Sakamoto & Holman [12]
<i>GPR108</i>	19	27	98%	rs2250656	6.4e-06	Occipital cortex_BA18(L)	G $\times$ MCI	Goltz, Brüggemeier, & Geerts [6]
<i>BTG3</i>	21	26	99.5%	rs2849896	6.3e-06	Pre-motor cortex_BA6(L)	G $\times$ MCI	Carson [3]

Table S.2: The top selected  $APOE-\epsilon 4$  effects for voxels that have a selection frequency exceeding 80% and a  $p$ -value not exceeding 0.1.

ROI	rank within ROI	$\hat{\beta}$	$APOE\epsilon 4$ p-value	effect type	reference
Lateral frontal BA44 (R)	1	315.3	0.10	G	Murphy et al. [9],
	2	310.4	0.07	G	Harwood et al. [7],
	3	296.1	0.09	G	Johnson et al. [8]
	4	292.8	0.08	G	
	5	291.5	0.10	G	
BA43 (R)	1	223.4	0.07	G×AD	
	2	218.8	0.10	G×AD	
	3	216.9	0.09	G×AD	
BA42 (R)	1	-303.7	0.06	G	
	2	-299.7	0.08	G	
	3	-292.8	0.07	G	
Superior parietal BA39 (R)	1	-395.8	0.09	G×MCI	Ross et al. [11],
	2	-395.2	0.09	G×MCI	Alexander et al. [1],
	3	-395.2	0.10	G×MCI	Duara et al. [5]

## References

- [1] Alexander, G. E., Furey, M. L., Grady, C. L., Pietrini, P., Brady, D. R., Mentis, M. J., & Schapiro, M. B. (1997). Association of premorbid intellectual function with cerebral metabolism in Alzheimer's disease: Implications for the cognitive reserve hypothesis. *The American Journal of Psychiatry*, 154(2), 165–172.
- [2] Cannon, D., Miller, J., Robison, R., Villalobos, M., Wahmhoff, N., Allen-Brady, K., McMahon, W., & Coon, H. (2010). Genome-wide linkage analyses of two repetitive behavior phenotypes in Utah pedigrees with autism spectrum disorders. *Molecular Autism*, 1(1), 3.
- [3] Carson, M. I. (2007). *Focus on Mental Retardation Research*. Nova Publishers.
- [4] Dai, M., Freeman, B., Shikani, H. J., Bruno, F. P., Collado, J. E., Macias, R., Reznik, S. E., Davies, P., Spray, D. C., Tanowitz, H. B., Weiss, L. M., & Desruisseaux, M. S. (2013). Altered regulation of akt signaling with murine cerebral malaria, effects on long-term neuro-cognitive function, restoration with lithium treatment. *PLoS ONE*, 7(10), e44117.
- [5] Duara R. , Grady C. , Haxby J. , Sundaram M. , Cutler N. R. , Heston L., Moore A., Schlageter N., Larson S., and Rapoport, S. I. (1986). Positron emission tomography in Alzheimer's disease. *Neurology*, 36(7), 879.
- [6] Goltz,S., Brüggemeier, U., & Geerts, A. (2004). G-protein coupled receptor lustr2 and uses thereof. EP1451324A2. European Patent Office.
- [7] Harwood, D. G., Sultzer D. L., Feil, D., Monserratt, L., Freedman, E., and Mandelkern, M. A. (2005). Frontal Lobe Hypometabolism and Impaired Insight in Alzheimer Disease. *The American Journal of Geriatric Psychiatry*, 13(11), 934–941.
- [8] Johnson, S. C., Schmitz, T. W., Moritz, C. H., Meyerand, M. E., Rowley, H. A., Alexander, A. L., et al. (2005). Activation of brain regions vulnerable to Alzheimer's disease: The effect of mild cognitive impairment. *Neurobiology of Aging*, 27(11), 1604–1612.
- [9] Murphy, J. M., Henry, R. G., Langmore, S., Kramer, J. H., Miller, B. L., & Lomen-Hoerth, C. (2007). Continuum of Frontal Lobe Impairment in Amyotrophic Lateral Sclerosis. *Arch Neurol.*, 64(4), 530–534.
- [10] Nolte, C., Rastegar, M., Amores, A., Bouchard, M., Grote, D., Maas, R., Kovacs, E., Postlethwait, J., Rambaldi, I., Rowan, S., Yan, Y., Zhang, F., &

- Featherstone, M. (2006). Stereospecificity and *PAX6* function direct *Hoxd4* neural enhancer activity along the antero-posterior axis developmental biology. *Developmental Biology*, 299(2), 582–593.
- [11] Ross, S.J., Graham, N., Stuart-Green, L., et al. (1996). Progressive biparietal atrophy: an atypical presentation of Alzheimer’s disease. *J Neurol Neurosurg Psychiatry*, 61(4), 388–395.
- [12] Sakamoto, K. & Holman, G. D. (2008). Emerging role for *AS160/TBC1D4* and *TBC1D1* in the regulation of *GLUT4* traffic. *Am J Physiol Endocrinol Metab*, 295, e29–37.
- [13] Talbot, K., Wang, H., Kazi, H., Han, L., Bakshi, K. P., Stucky, A., Fuino, R. L., Kawaguchi, K. R., Samoyedny, A. J., Wilson, R. S., Arvanitakis, Z., Schneider, J. A., Wolf, B. A., Bennett, D. A., Trojanowski, J. Q., & Arnold, S. E. (2012). Demonstrated brain insulin resistance in Alzheimer’s disease patients is associated with IGF-1 resistance, IRS-1 dysregulation, and cognitive decline. *Journal of Clinical Investigation*, 122(4), 1316–1338.
- [14] Yang, J., Li, S., & Liu, Y. (2013). Systematic analysis of diabetes- and glucose metabolism-related proteins and its application to Alzheimer’s disease. *J. Biomedical Science and Engineering*, 6, 615–644.