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Abstract

Background: Immunoglobulin E – mediated food allergy (IgE-FA) has emerged as a global public health concern. Immune dysregulation is an underlying mechanism for IgE-FA, caused by "dysbiosis" of the early intestinal microbiota. We investigated the association between infant gut bacterial composition and food-related atopy at age 3-5 years using a well-characterized birth cohort. **Methods:** The study definition of IgE-FA to egg, milk, or peanut was based on physician panel retrospective review of clinical and questionnaire data collected from birth through age 3-5 years. Using 16S rRNA sequencing, we profiled the bacterial gut microbiota present in stool specimens collected at 1 and 6 months of age. **Results:** Of 447 infants with data for analysis, 44 (9.8%) met physician panel review criteria for IgE-FA to ≥1 of the three allergens. Among children classified as IgE-FA at 3-5 years, infant stool samples showed significantly less diversity of the gut microbiota compared to the samples of children classified as no IgE-FA at age 3-5 years, especially for milk and peanut (all covariate adjusted *p*'s for alpha metrics <0.007). Testing of individual operational taxonomic units (OTUs) revealed 6-month deficiencies in 31 OTUs for IgE-FA compared to no IgE-FA, mostly in the orders *Lactobacillales, Bacteroidales*, and *Clostridiales*. **Conclusions:** Variations in gut microbial composition in infant stool were associated with a study definition of IgE-FA at 3-5 years of age. This included evidence of a lack of bacterial diversity, deficiencies in specific OTUs, and delayed microbial maturation. Results support dysbiosis in IgE-FA pathogenesis.

Key Message: Compositional differences in gut bacterial colonization were observed for children who developed IgE-mediated food allergy compared to those that did not. Results support a theory of elevated risk due to dysbiosis and delays in microbial maturation. The

importance of further research in this area lies in the potential for developing interventions that can prevent development of IgE-FA.

Key words: food allergy, IgE, microbiome

Introduction

The current 8% prevalence of food allergy for US children represents a 50% increase between 1997 and 2011[1, 2]. Food allergy can be life-threatening and undermines the quality of life of affected children and their families[1, 2].

Food allergy is a reproducible inflammatory response induced by immunoglobulin E activation of mast cells and basophils upon exposure to a given food[3]. Immunoglobulin E (IgE) - mediated food allergy (IgE-FA) occurring during infancy can be preceded by atopic eczema and increases the risk of rhinitis and asthma in later childhood[3]. Immune dysregulation is an important contributor to IgE-FA [4]. In a healthy state, oral exposure to innocuous antigens (e.g., food proteins) leads to interactions with specific antigen presenting cells followed by the induction of T regulatory cell suppression of immune responses, or oral tolerance[3]. In IgE-FA, exposure to common food proteins results in an inappropriate T helper 2 cell-mediated response to epitopes of the offending food[3]. Examining associations along the causal pathway to IgE-FA could inform processes involved in early childhood allergic disease.

Immune processes leading to the induction or failure of oral tolerance are actively influenced by the gut microbiome[5] In allergy-free individuals, commensal bacteria stimulate responses leading to immune tolerance of innocuous allergens[5]. Aberrant colonization of bacteria in the gut (dysbiosis), increases susceptibility to atopy [5]. Delayed colonization of gut bacteria leads to irregularities in the development of gut-associated lymphoid tissues, impacting downstream immune pathways[6]. Associations between gut microbiota composition and food sensitization or IgE-FA are reported in murine and human studies [7, 8].

Well-characterized birth cohorts are suited to exploring how gut bacterial colonization impacts IgE-FA risk. We use data from the Microbes, Allergy, Asthma, and Pets (MAAP) Research Program, drawn from the Wayne County Health Environment Allergy and Asthma Longitudinal Study (WHEALS) birth cohort[9] to explore early gut bacterial composition and risk of IgE-FA to egg, milk, or peanut at age 3-5 years[9].

Methods

Study population. The Institutional Review Board at Henry Ford Health System (HFHS) approved this research. The WHEALS birth cohort was established to identify environmental factors related to development of allergy and asthma[9]. Methods, eligibility and recruitment have been described previously[9]. Briefly, pregnant women ages 21 to 45 years, residing in metropolitan Detroit and receiving prenatal care at selected HFHS obstetric clinics September 2003 through November 2007 were recruited. Infant blood samples were collected at 6- and 12-month home visits. At a 24-month clinic visit, infant blood was collected to measure allergen-specific serum IgE as described previously[9], and conduct skin prick tests (SPTs) using a Duotip-test device (Lincoln Diagnostics Inc, Decatur, Illinois), and including, but not limited to, egg, milk, and peanut. A wheal diameter $\geq$ 3mm larger than saline control was considered positive for

SPTs. Infant stool samples collected at 1 and 6 month home visits, were transported to the laboratory in cryovials, stored at −80 °C, and shipped to UCSF, where they underwent sequencing of the V4 region of the 16S rRNA gene using the Illumina Nextseq (Supplement, Appendix 1)[10]. Parents were interviewed at infant ages 1, 6, 12, and 24 months for medical history, and at age 3-5 years for infant food avoidance, gastrointestinal symptoms, and reactions to food. Infant medical records (requested for those outside of HFHS), were reviewed from birth through age 5 years. The definition of IgE-FA used in this study has also been described previously[11]. Briefly, a panel of two board-certified allergists reviewed clinical and interview data from birth through age 3-5 years to classify infants as highly likely, likely, or unlikely to have IgE-FA. This was a two-step process. In Step 1, we identified infants with at least two of the following three characteristics for egg, milk, or peanut: (1) $\geq$1 specific IgE level $\geq$0.35 IU/mL, (2) a positive SPT result, or (3) parental report of infant symptoms potentially related to food allergy plus $\geq$1 specific IgE level $\geq$ of 0.10 IU/ml. Infants that did not exhibit two of these three characteristics were automatically classified as "unlikely" to have IgE-FA. Infants with at least two of the three characteristics were forwarded to the physician panel for classification (Step 2) using study protocols based on recently published guidelines[12 13]. A third allergist ruled on discordant decisions. For this analysis, "highly likely" and "likely" were collapsed into a single category and compared to "unlikely". Report of symptoms was an integral part of classification. Classification was also heavily influenced by IgE and/or SPT results meeting the 95% predictive decision points [13], and medical chart documentation of a physician diagnosis of IgE-FA or results of an oral food challenge.

Statistical Analysis. Main effects and interaction effects were considered significant at $p<0.05$; and $p<0.10$, respectively. Characteristics of children included and excluded from the analysis were compared using ANOVA and the chi-square test for numeric and categorical covariates, respectively. Children with and without IgE-FA were compared using Kruskal-Wallis for numeric covariates and the Fisher's exact test for categorical covariates. Alpha diversity measures of bacterial richness, Pielou's evenness, Faith's phylogenetic diversity (PD), and Shannon's diversity were estimated using QIIME[14] and the R vegan package.[15] Measurements used exact age at sample collection and were fit using generalized estimating equations (GEE) with a Gaussian link to account for within-subject correlations. Differences in alpha diversity by IgE-FA were examined using time interactions, followed by main effects if interactions were non-significant. Composition of the gut microbiota was defined using unweighted and weighted UniFrac (phylogenetic),[16] as well as Canberra and Bray-Curtis dissimilarity (non-phylogenetic).

Compositional differences were assessed using PERMANOVA in the R package vegan [15]. Individual operational taxonomic unit (OTU) tests were performed using zero-inflated negative binomial models (or standard negative binomial models if convergence failed), with ***Benjamini-Hochberg*** false discovery rate (FDR) adjusted p-values[17] computed to account for multiple testing (FDR adjusted $p<0.05$ significant). OTUs were only tested if they were detected in ≥25% of samples. Bacterial microbiota-for-age z-scores (BMAZ) were calculated to determine if microbial maturity (given fixed chronological age) differed in children with IgE-FA. Random forest models were fit using the randomForest package, with actual age at stool sample collection as the outcome, and the relative abundance of OTUs as predictive features. To identify a small subset of taxa that explained a large portion of the variability in age, the rfcv function of the randomForest package was applied, using 5-fold cross-validation[18]. This sparse model was then used to predict age at stool sample collection; BMAZ was calculated as described in Subramanian et al 2014,[19] with each month used as an age category, except for 2-4 and 8-10 months, which were collapsed due to data sparsity. Differences in BMAZ by IgE-FA were tested using GEE models, as described previously. The GEEmediate R package was utilized to test the mediating effect of eczema by age 2 for all microbiota metrics (alpha diversity, beta diversity, and BMAZ[20]. For beta diversity, the first principle coordinate of each metric was used in the mediation models.

Results:

Of the 590 children with sufficient data for IgE-FA classification, 447 had stool samples for microbiota analysis, (n=44 with IgE-FA and n=403 with no IgE-FA). Of the 447 children, 156 had a stool sample at 1 month only, 118 had a stool sample at 6 months only, and 173 had a stool sample for both time points. Among the 44 children with IgE-FA, 59% were allergic to one food, 30% were allergic to two, and 11% were allergic to all three foods. The most common IgE-FA in our sample was egg (73% of the IgE-FA children), followed by peanut (59%) and milk (20%).

Compared to those excluded from analysis, mothers of infants in the analytic sample were older ($p=0.020$), reported more education ($p<0.001$), higher household income ($p<0.001$), a history of atopy ($p=0.014$), greater likelihood of exclusive breastfeeding ($p=0.026$), and were less likely to be urban residents ($p=0.014$) and/or exposed to environmental tobacco smoke ($p=0.007$) (Supplement, Table 1). Children in the analytic sample meeting study definition of IgE-FA (Table 1) were more likely to have diagnosed eczema by age 2 (37.2% vs. 19.7%; $p=0.017$) compared to non-IgE-FA.

When all stool samples were modeled, all alpha diversity metrics were lower in children meeting study criteria for IgE-FA compared to those without, after covariate adjustment (Figure 1, Table 2), and this effect did not significantly differ over time (Table 2, all interaction p≥0.13). Effects were largest for milk allergic children compared to non-milk allergic children, followed by peanut and egg. Results were similar using specific IgE ≥ 0.35 IU/ml (sensitization) as an outcome (Table 2), but a significant effect was observed only for peanut (Table 2).

Bacterial compositional differences by IgE-FA were tested using PERMANOVA (Table 3). Adjusting for covariates, significant compositional differences by IgE-FA were present only at 6-months of age (Table 3). Only unweighted UniFrac and Canberra distances revealed significant differences at 6 months for all IgE-FA outcomes. Sensitization to "any food" and to peanut were associated with 1-month community composition, only.

IgE-FA was significantly associated with the abundance of 8 OTUs (Figure 2) after covariate adjustment, 5 of which were in lower abundance in IgE-FA children. At 6-months of age, 20 of 31 significant OTUs identified were deficient for those with IgE-FA, primarily *Bacteroidales*, and *Clostridiales*. OTUs at 6-months overabundant in IgE-FA children were mostly of the order *Bifidobacteriales*.

Children with IgE-FA had significantly lower adjusted bacterial microbiota-for-age z-scores BMAZ scores compared to non-IgE-FA children (Figure 3; β (95% CI)=-0.70 (-1.08, -0.33); p<0.001). This effect did not significantly differ across time (interaction p=0.97). No mediating effect was observed for the association of alpha diversity, beta diversity, and BMAZ to study definition of IgE-FA (p≥0.44) (Supplement, Table 2).

Discussion

We observed deficiencies in the alpha diversity of gut microbiota for birth cohort infants meeting study criteria for IgE-FA by age 3-5 years, primarily at 6-months of age, based on dissimilarity measures. We observed an overabundance of several *Bifidobacteriales* OTUs and a deficiency in several *Bacteroidales* and *Clostridiales* OTUs in IgE-FA children at age 6-months. Microbiota-for-age z-scores suggest delayed maturity in the infant gut microbiota of children with IgE-FA. Suggested differences in rare taxa or taxa of low abundance (UniFrac, Canberra) need further exploration. Results suggest dysbiosis is related to oral tolerance and IgE-FA.

Microbial composition can modify the risk of IgE-FA through innate and adaptive immunity[4]. Unwanted microbial antigens bound by secretory IgA transferred maternally from breastfeeding are handled by the innate immune system[4]. Oligosaccharides in human milk that induce growth of *Bifidobacterium* and *Lactobacillus* in the infant gut also induce production of

IL10 and IgA.[4] *Clostridia,* particularly clusters IV and XIVa, activate the release of TGF-β. IgA and cytokines IL10 and TGF-β are inducers of T regulatory cells that suppress undesirable immune reactions. Gut bacteria also participate in the fermentation of complex carbohydrates generating short-chain fatty acids, which contribute to regulation of inflammatory responses by influencing B-cell function and intestinal barrier function.[4, 5, 21] Thompson-Chagoyan [2011] showed that changes in infant gut composition were concomitant with decreases in levels of specific IgE against cow's milk antigens after providing hydrolyzed formula to infants with cow's milk allergy[22]. These infants had higher concentrations of butyric acid, than their non-allergic counterparts[22] providing evidence that deficiencies in taxa may represent reduced capacity to offset inflammatory processes.

Previous studies have looked at gut bacterial colonization and risk of food sensitization and IgE-FA with reports of a potential "signature" for food sensitization [23], or results suggestive of delays in gut maturity[24, 25]. In VDAART, investigators reported deficiencies in C*lostridium* that were significant for both food sensitization and food allergy[26], but unlike our analysis, showed subgroup variations for race, mode of delivery, and age at introduction of solid food[26]. Similarly, in The Consortium of Food Allergy (CoFAR) observational study of milk allergy, *Clostridia* and *Firmicutes* were enriched in the guts of children whose milk allergy resolved by 8 years[8] The CoFAR study provided additional evidence of decreased fatty acid metabolism, emphasizing potential mechanisms by which risk of IgE-FA is modified[8].

Our findings contrast with that of studies showing higher abundance of Bifidobacterium associated with reduced risk of atopic disease[27, 28]. Our results are more aligned with those reported by Stokholm et al., using the Copenhagen Prospective Studies on Asthma in Childhood (COPSAC), in which gut colonization at 1 year for infants who developed asthma at age 5 years resembled that of healthy infants at 1 month, indicating delayed gut maturation[25]. Since Bifidobacterium is 40-80% of gut colonized bacteria shortly after birth, our results may support the hypothesis that delayed gut maturity influences IgE-FA risk[25].

We did not find that atopic eczema, which typically precedes IgE-FA[3], mediates the relationship between microbiota composition and IgE-FA in our study. A systematic review (2019) found that 5 of 11 observational studies on this topic reported lower gut diversity associated with eczema[28]. While no specific bacterial species have emerged in the literature consistently, Bifidobacterium appears in several reports[27-29]. Zhang, et al. reported decreased abundance of Bifidobacterium associated with eczema only for infants >6 months [27]. Ismail et al. found modulation of eczema risk (among children at high risk of atopy) driven by certain

Bifidobacterium species[28], however, Ta, et al. found atopic eczema was associated with delayed colonization of Bacteroides, but not Bifidobacterium[29]. Evidence of immune modulatory effects that are specific to certain species highlight the need for further research.

Study variation in the orders, families, and genera of the taxa reportedly associated with sensitization or IgE-FA could be due to a myriad of factors including differences in study outcomes, age at sample collection, specific allergen under study, characteristics of the study samples, or dissimilarities in sample storage, processing, and analysis[12]. The implications of individual-level variation in the characteristics of infant intestinal microbiota continue to emerge. The substantial variation in gut colonization of healthy individuals suggests that a defined composition of microbial taxa universally present is unlikely[30]. A "healthy" gut is more likely defined by a "core" set of functions performed by a variety of colonized bacteria, as opposed to the presence of a fixed set of taxa[30].

We did not conduct oral food challenges in this birth cohort, however, documentation of an oral food challenge in the medical chart was a chief consideration in our study definition of IgE-FA[11]. We classified children from birth to 5 years as having IgE-FA using clinical and symptom data that may have been acquired at different time points, and we do not use exact age at diagnosis in our analysis. Misclassification is possible if clinical evidence of IgE-FA at 2 years resolved by age 3-5. Due to small sample size, we did not assess the impact of antibiotic exposure (reported by only 3% of cohort infants), nor the impact of environmental and sociocultural factors on the relationship between infant gut bacterial composition and IgE-FA, as done in an earlier publication[9]. We acknowledge that our observed associations are not causal.

Despite limitations, our analysis supports modulation of IgE-FA risk by colonization of infant gut bacteria. Bacterial colonization is a potentially modifiable factor along the causal pathway to IgE-FA. Continued research in this area creates potential for intervention and prevention of IgE-FA in infants and children.

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Abbreviations

BIC	Bayesian Information Criteria
BSA	Bovine serum albumin
CTAB	Cetyltrimethylammonium bromide
FDR	False Discovery Rate
HFHS	Henry Ford Health System
IgE	Immunoglobulin E
IgE-FA	Immunoglobulin E-mediated food allergy
MAAP	Microbes, Allergy, Asthma, and Pets
OTU	Operational taxonomic unit
SPT	Skin prick test
WHEALS	Wayne County Health Environment Allergy and Asthma Longitudinal Study

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Table 1. Selected cohort characteristics for food allergic and non-food allergic children included in the analyses (n=447)

Covariate	IgE-FA n=44	No IgE-FA n=403	p-value[1]
Child sex			
Male	23 (52.3)	215 (53.3)	1.000
Female	21 (47.8)	188 (46.7)	
Child race			
African American	30 (68.2)	239 (59.3)	0.071
White	6 (13.6)	100 (24.8)	
Hispanic/Latino	1 (2.3)	27 (6.7)	
Other	7 (15.9)	37 (9.2)	
Household income			
< $40K	12 (27.3)	121 (30.0)	0.460
$40K - < $80K	8 (18.2)	107 (26.6)	
$80K - >=$100K	18 (40.9)	122 (30.2)	
Refused	6 (13.6)	53 (13.2)	
Urban residence	22 (50.0)	207 (51.4)	0.875
Maternal education			
<HS diploma	2 (3.8)	15 (7.0)	0.817
HS diploma	4 (13.4)	56 (20.7)	
Some college+	38 (86.3)	332 (82.4)	
Mom age at birth, mean (sd)	30.4 (4.8)	30.0 (4.8)	0.591
Mother's marital status	29 (65.9)	260 (64.5)	1.000
Maternal atopy	13 (29.5)	165 (41.8)	0.145
Maternal history of allergies or asthma	16 (36.4)	109 (27.5)	0.220
Prenatal ETS exposure	7 (15.9)	96 (23.8)	0.514
Prenatal indoor pet(s)	14 (31.8)	151 (37.5)	0.51
Delivered by cesarean section	15 (34.1)	148 (36.7)	0.869
First born child	19 (43.2)	155 (38.5)	0.63
Breastfeeding at 1 month			
Formula feeding	7 (15.9)	80 (20.2)	0.778
Mixed feeding	29 (65.9)	254 (64)	
Exclusive breastfeeding	8 (18.2)	63 (15.9)	
Solid food introduction <4 months	16 (36.4)	168 (41.7)	0.523
Physician diagnosed eczema by age 2	16 (37.2)	72 (19.7)	0.017

[1] *Kruskal-Wallis test for numerical covariates and Fisher's exact test for categorical covariates*

Table 2. Difference in bacterial alpha diversity metrics by study definition of IgE-mediated food allergy (IgE-FA) and food sensitization[1,2]

Metric	Any IgE-FA			Milk IgE-FA			Egg IgE-FA			Peanut IgE-FA		
	Interx p^3	β (95% CI)[4]	*p*	Interx p^3	β (95% CI)[4]	*p*	Interx p^3	β (95% CI)[4]	*p*	Interx p^3	β (95% CI)[4]	*p*
Richness	0.307	-17.39 (-29.31, -5.47)	0.004	0.265	-33.3 (-49.84, -16.76)	<0.001	0.493	-20.15 (-33.66, -6.65)	0.003	0.179	-24.4 (-38.37, -10.43)	<0.001
Evenness	0.581	-0.025 (-0.05, 0)	0.047	0.224	-0.06 (-0.09, -0.03)	<0.001	0.395	-0.029 (-0.057, -0.001)	0.042	0.352	-0.04 (-0.07, -0.01)	0.005
Faith's PD[5]	0.697	-0.84 (-1.45, -0.23)	0.007	0.132	-1.76 (-2.67, -0.85)	<0.001	0.92	-1 (-1.7, -0.3)	0.005	0.274	-1.15 (-1.88, -0.42)	0.002
Shannon's[6]	0.777	-0.17 (-0.32, -0.02)	0.026	0.343	-0.39 (-0.55, -0.23)	<0.001	0.549	-0.2 (-0.36, -0.03)	0.023	0.546	-0.27 (-0.45, -0.1)	0.002
	Any Food Sensitization			**Milk Sensitized**			**Egg Sensitized**			**Peanut Sensitized**		
	Interx p^3	β (95% CI)[4]	*p*	Interx p^3	β (95% CI)[4]	*p*	Interx p^3	β (95% CI)[4]	*p*	Interx p^3	β (95% CI)[4]	*p*
Richness	0.60	-2.83 (-12.07, 6.42)	0.549	0.986	-1.67 (-11.49, 8.16)	0.74	0.99	-7.48 (-18.3, 3.33)	0.175	0.89	-17.86 (-29.62, -6.09)	0.003
Evenness	0.882	0.003 (-0.012, 0.018)	0.721	0.725	0.003 (-0.013, 0.019)	0.728	0.547	0.006 (-0.011, 0.024)	0.485	0.195	-0.02 (-0.044, 0.004)	0.109
Faith's PD	0.684	-0.19 (-0.67, 0.28)	0.426	0.925	-0.2 (-0.7, 0.31)	0.446	0.878	-0.5 (-1.05, 0.05)	0.073	0.46	-0.84 (-1.47, -0.21)	0.009
Shannon's	0.967	0.002 (-0.091, 0.094)	0.973	0.687	0.006 (-0.091, 0.103)	0.903	0.576	0.009 (-0.101, 0.118)	0.874	0.267	-0.14 (-0.285, -0.004)	0.045

[1] *Table 2 analyses based on both 1- and 6-month samples*

[2] *Sensitization based on serum specific IgE ≥ 0.35 IU/ml for egg, milk, or peanut*

[3] *Interaction p value; tests if the association between alpha diversity and outcome is time-dependent, after adjusting for exact age at stool sample collection, child race, maternal history of allergies or asthma, and breastfeeding status at 1-month.*

[4] *Interpreted as the mean difference in alpha diversity across time, comparing IgE-FA to non IgE-FA, and sensitized to non-sensitized, after adjusting for exact age at stool sample collection, child race, maternal history of allergies or asthma, and breastfeeding status at 1-month.*

[5] *Faith's phylogenic diversity*

[6] *Shannon's diversity index*

Table 3. Association between early life microbiome composition and study definition of IgE-mediated food allergy (IgE-FA) and food sensitization.[1]

Outcome	Metric	1-Month[2]			6-Months[2]		
		N	p-value	R^2	N	p-value	R^2
Any IgE-FA	Unweighted UniFrac	323	0.108	0.004	282	**0.013**	0.006
	Weighted UniFrac	323	0.674	0.002	282	**0.009**	0.012
	Canberra	323	0.196	0.003	282	**0.008**	0.005
	Bray-Curtis	323	0.285	0.003	282	**0.018**	0.006
Milk IgE-FA	Unweighted UniFrac	323	0.148	0.004	282	**0.008**	0.006
	Weighted UniFrac	323	0.313	0.004	282	**0.008**	0.014
	Canberra	323	0.075	0.004	282	**0.008**	0.005
	Bray-Curtis	323	0.145	0.004	282	0.071	0.005
Egg IgE-FA	Unweighted UniFrac	323	0.089	0.004	282	**0.032**	0.005
	Weighted UniFrac	323	0.603	0.002	282	**0.040**	0.009
	Canberra	323	0.123	0.003	282	**0.020**	0.004

	Bray-Curtis	323	0.370	0.003	282	**0.021**	0.006
Peanut IgE-FA	Unweighted UniFrac	323	0.182	0.004	282	**0.031**	0.005
	Weighted UniFrac	323	0.628	0.002	282	0.079	0.007
	Canberra	323	0.172	0.003	282	**0.029**	0.004
	Bray-Curtis	323	0.182	0.004	282	0.158	0.004
Any Food Sensitization	Unweighted UniFrac	308	0.259	0.003	275	0.591	0.003
	Weighted UniFrac	308	0.366	0.003	275	0.737	0.002
	Canberra	308	0.263	0.003	275	0.696	0.003
	Bray-Curtis	308	**0.039**	0.006	275	0.294	0.004
Milk Sensitized	Unweighted UniFrac	317	0.412	0.003	280	0.485	0.003
	Weighted UniFrac	317	0.489	0.003	280	0.651	0.002
	Canberra	317	0.470	0.003	280	0.590	0.003
	Bray-Curtis	317	0.148	0.004	280	0.556	0.003
Egg Sensitized	Unweighted UniFrac	315	0.175	0.004	280	0.299	0.004
	Weighted UniFrac	315	0.720	0.002	280	0.931	0.001
	Canberra	315	0.334	0.003	280	0.613	0.003
	Bray-Curtis	315	0.299	0.003	280	0.339	0.004
Peanut Sensitized	Unweighted UniFrac	305	**0.041**	0.005	271	0.163	0.004
	Weighted UniFrac	305	0.451	0.003	271	0.813	0.002
	Canberra	305	0.058	0.004	271	0.375	0.004
	Bray-Curtis	305	0.110	0.005	271	0.428	0.004

[1] *Serum specific IgE ≥ 0.35 IU/ml for egg, milk, or peanut*

[2] *After adjusting for exact age at stool sample collection, child race, maternal history of allergies or asthma, and breastfeeding status at 1-month.*