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80 Abstract

81 Three decades of research have demonstrated that biodiversity can promote the 82 functioning of ecosystems. Yet, it is unclear whether the positive effects of 83 biodiversity on ecosystem functioning will persist under various types of global 84 environmental change drivers. We conducted a meta-analysis of 46 factorial 85 experiments manipulating both species richness and the environment to test how 86 global change drivers (i.e. warming, drought, nutrient addition or CO₂ enrichment) 87 modulated the effect of biodiversity on multiple ecosystem functions across three 88 taxonomic groups (microbes, phytoplankton and plants). We found that biodiversity 89 increased ecosystem functioning in both ambient and manipulated environments, but 90 often not to the same degree. In particular, biodiversity effects on ecosystem 91 functioning were larger in stressful environments induced by global change drivers, 92 indicating that high-diversity communities were more resistant to environmental 93 change. Using a subset of studies, we also found that the positive effects of 94 biodiversity were mainly driven by interspecific complementarity, and that these This article is protected by copyright. All rights reserved

95 effects increased over time in both ambient and manipulated environments. Our
96 findings support biodiversity conservation as a key strategy for sustainable ecosystem
97 management in the face of global environmental change.

98

99 Introduction

100 Global environmental changes are reshaping the structure and processes of 101 ecosystems, potentially threatening the functioning and services that ecosystems 102 provide to human society (Zhou et al. 2012; Song et al. 2019; Jiang et al. 2020). In 103 the face of environmental change, biodiversity is a key factor maintaining (Naeem et 104 al. 2009; Cardinale et al. 2012; Hooper et al. 2012) and buffering ecosystem 105 functioning (Yachi & Loreau 1999; Hisano et al. 2018; Pires et al. 2018). However, 106 environmental change also alters species dynamics and interactions, causing rapid 107 changes in biodiversity at various spatial scales (Vellend et al. 2013; Dornelas et al. 2014; Urban 2015; Blowes et al. 2019; IPBES 2019). Therefore, changing 108 109 environments can affect ecosystem functioning both directly, via altering rates of 110 ecosystem processes (Spaak et al. 2017), and indirectly, via changing biodiversity 111 (Hautier et al. 2015; Giling et al. 2019). Furthermore, environmental changes can 112 interact with biodiversity in regulating ecosystem functioning, such that the effects of 113 biodiversity on ecosystem functioning may be enhanced or weakened under altered 114 environmental conditions, even if biodiversity itself does not change (Eisenhauer et al. 115 2019; Benkwitt et al. 2020). However, the potential interactions between 116 environmental changes and biodiversity are much less understood compared with the 117 direct and indirect effects of environmental change on ecosystems, leaving it unclear 118 whether the positive effects of biodiversity will persist in future environments 119 impacted by global change drivers (De Laender et al. 2016).

120 While existing biodiversity experiments have mostly manipulated species 121 richness under ambient environmental conditions, a growing number of experiments 122 examine how environmental change (or global change drivers) may alter the effects of 123 biodiversity on ecosystem functioning across different taxonomic groups, such as 124 terrestrial plants, microbes, or phytoplankton (Hautier et al. 2014; Craven et al. 2016; 125 García et al. 2018; Bestion et al. 2020). Using factorial designs in which biodiversity 126 and environmental manipulations were combined, these studies revealed strengthened 127 (Reich et al. 2001; Steudel et al. 2011; García et al. 2018), constant (Thakur et al. 128 2015; Eisenhauer et al. 2018), or weakened (De Boeck et al. 2008) biodiversity This article is protected by copyright. All rights reserved

129 effects impacted by various types of global change drivers. Such mixed results may be 130 explained by the context dependency of biodiversity effects, which vary in strength 131 across different types and magnitudes of environmental manipulations, different 132 taxonomic groups, or different aspects of ecosystem functions being measured. For 133 instance, experimental warming in microbial and phytoplankton communities has 134 been found to either dampen or enhance ecosystem productivity (Yvon-Durocher et al. 135 2015; García et al. 2018; Tabi et al. 2019; Bestion et al. 2020). Currently, we lack a 136 synthetic understanding of both how different types of environmental change may 137 influence biodiversity effects on ecosystem functioning, and how these interactive 138 effects depend on the experimental context (e.g. taxonomic group, experimental time, 139 etc.) (Eisenhauer et al. 2019).

140 One hypothesis about the interactive effects of biodiversity and environmental 141 change is that global change drivers alter the strength and even the type of 142 interspecific interactions (He et al. 2013; Hoek et al. 2016; Baert et al. 2018), which 143 underlie the effects of biodiversity on ecosystem functioning. Two classes of 144 processes have been proposed to explain biodiversity effects, namely 145 complementarity (CE) and selection (SE) effects (Loreau & Hector 2001; Tilman et al. 146 2014). CE arises from interspecific niche partitioning or facilitation, and SE arises 147 from a disproportionally high contribution of one or few species to ecosystem 148 functioning. Many studies suggest that the strength and type of species interactions 149 may shift as the environment changes (Maestre et al. 2009; He et al. 2013; Hoek et al. 1502016; Olsen et al. 2016). In particular, the stress-gradient hypothesis predicts that 151 species interactions can switch from higher competition in favorable environments to 152lower competition or even facilitation in stressful environments (Bertness & Callaway 1531994; He et al. 2013; Hoek et al. 2016; but see Metz & Tielbörger 2016). Such a 154switch may enhance CE and hence overall biodiversity effects in stressful 155environments. On the other hand, SE may also increase with environmental stress, 156 because more diverse communities may have a higher probability of including stress-157 tolerant species and such species may be expected to outcompete stress-intolerant 158 ones and dominate in stressful environments (Maestre et al. 2009; Baert et al. 2018; 159 Bestion et al. 2020). Thus, stress intensity induced by environmental changes may 160 provide a useful indicator to predict the influence of environmental changes on the 161 magnitude of biodiversity effects on ecosystem functioning (Baert et al. 2018).

162 Previous experiments also suggested that biodiversity effects, particularly CE, 163 increased over time (Cardinale et al. 2007; Reich et al. 2012; Huang et al. 2018). 164 Such enhanced biodiversity effects were at least in part due to character displacement 165 during the succession of experimental communities (Zuppinger-Dingley et al. 2014; 166 van Moorsel et al. 2018). In stressful environments, interspecific facilitation and the 167 compensation of more resistant species may allow high-diversity communities to cope 168 better with stress through time and maintain ecosystem functions at higher levels than 169 less diverse communities (Hisano et al. 2018; Wright et al. 2021). In such cases, the 170 influence of environmental changes on biodiversity effects may strengthen over time, 171but the magnitude of these effects is uncertain.

172 In this study, we performed a meta-analysis of 46 factorial experiments in which 173 species richness was manipulated together with at least one of four types of global 174change drivers (namely warming, drought, nutrient addition or CO2 enrichment) to 175 systematically assess how environmental change may modify biodiversity effects on 176 ecosystem functioning across three taxonomic groups (namely microbes, 177phytoplankton, and terrestrial plants [plants for short]). These four global change 178drivers capture common anthropogenic impacts on ecosystems and are most 179 frequently investigated in the literature (e.g. Hooper et al. 2012; Song et al. 2019). 180 That said, only warming could be assessed in all three taxonomic groups (i.e. 181 microbes, phytoplankton, and plants, with 4, 3, and 6 studies, respectively), and 182 drought, nutrient and CO₂ enrichment were only assessed in plants (with 14, 11, and 8 183 studies, respectively). Using these data, we asked two questions: (i) does biodiversity 184 promote ecosystem functioning under both ambient and manipulated environmental 185 conditions? (ii) do global change drivers influence the magnitude of biodiversity 186 effects, and do such influences vary through time and depend on the stress intensity 187 induced by environmental changes? Our hypothesis is that biodiversity promotes 188 ecosystem functioning across a range of environmental conditions, but its effect size 189 can be modulated by global change drivers. In stressful environments, mixtures may 190 be less influenced than monocultures if CE increases with stress intensity as predicted 191 by the stress gradient hypothesis (Bertness & Callaway 1994), or if SE increases with 192 stress intensity due to the presence of stress-tolerant species (Baert et al. 2018; 193 Bestion et al. 2020). In such cases, biodiversity effects increase with stress intensity 194 (H₁ in Fig. 1). Otherwise, biodiversity effects can decrease with stress intensity if 195 mixtures are more sensitive to environmental stress (H₂ in Fig. 1). Finally, This article is protected by copyright. All rights reserved

biodiversity effects will not change with stress intensity if mixtures and monocultures are similarly sensitive to environmental stress (H_0 in Fig. 1).

198

199 Methods

200 Data collection

201 We searched the literature and extracted all factorial experiments that 202 manipulated species richness and at least one of the four focal types of global change 203 drivers, i.e. warming, drought, nutrient addition or CO₂ enrichment (see Appendix B: 204 Table B1 for search terms). We performed the search on February 17th 2020 in the 205 Web of Science and retrieved 2386 papers in total. We then selected studies using the 206 following criteria (See PRISMA diagram, Appendix B: Fig. B1): (i) the experiment 207 includes monocultures and mixtures under both ambient and manipulated 208 environmental conditions; (ii) the study provides measures of at least one ecosystem 209 function under different treatments. In total, 35 experimental studies were selected 210 based on our criteria, and we added 11 experimental studies from cross-referencing 211 that met our criteria (Appendix B: Table B7). This led to a total of 46 experimental 212 studies, including 7,582 experimental units (ranging from a culture dish to a field plot) 213 covering different taxonomic groups and types of global change drivers. Note that the 214 46 experimental studies were treated as independent in our meta-analysis, although 215 some of them came from the same site but involved different types of environmental 216 manipulations (e.g., BioCON experiment with nutrient addition or CO₂ enrichment; 217 Reich et al. 2001). This simplification was justified by a likelihood ratio test which 218 reported no significant difference between models including and excluding the non-219 independence between studies (see Appendix A).

220 We categorized our dataset by taxonomic groups (microbes, phytoplankton, and 221 plants), types of global change drivers (warming, drought, nutrient addition, and CO₂ 222 enrichment) and two categories of ecosystem functions (biomass production and 223 biogeochemical process) (Appendix B: Table B2; Chapin et al. 2011). In our study, 224 microbes referred to non-photosynthetic microorganisms, such as bacteria, fungi and 225 protozoans. Because only the warming treatment was found for microbes and 226 phytoplankton, we have in total six combinations of taxonomic groups and global 227 change drivers, i.e. warming in all three taxonomic groups, as well as drought, 228 nutrient and CO₂ enrichment in plants. The 46 studies included 3, 4, and 39 229 experimental studies on microbes, phytoplankton, and plants, respectively (Appendix

230 B: Table B7). For studies on plants, 10 studies were from pots in the greenhouse, and 231 29 studies were from field experiments. Experimental duration ranged from 6 to 40 232 days in microbial studies, from 7 to 19 days in phytoplankton studies, and from 1 to 233 15 years in plant studies. The highest species richness varied among studies on 234 microbes (6 ~ 24), phytoplankton (12 ~ 64), and plants (2 ~ 60). For each type of 235 global change drivers, we recorded the level(s) of experimental manipulation, e.g. the 236 degree of temperature increase, the proportion of water reduction, the type and 237 amount of nutrient added (e.g. N and NPK enrichment), and the amount of CO₂ 238 enrichment. For warming treatments in microbes and phytoplankton, which usually 239 contained several temperature levels in addition to the ambient or long term-culture 240 temperature (as the control, e.g. 15°C in Pennekamp et al. 2018 and 20°C in García et 241 al. 2018), we only used temperature levels above this control as warming treatments 242 and excluded data with cooling treatments.

243 Regarding the two categories of ecosystem functions, biomass production 244 captured the total biomass produced during a given period (e.g. grassland biomass 245 production within a year), which has been the most commonly used metric of 246 ecosystem functioning; biogeochemical processes captured soil characteristics (e.g. 247 the concentration of key elements or organic materials in the soil), soil microbial 248 functions (e.g. microbial biomass, microbial growth and abundance) and nutrient 249 cycling (e.g. ectoenzyme activity and organic phosphorus). We considered soil 250 microbial growth and abundance as functions, because soil microbial community 251composition and activity play key roles in mediating biogeochemical cycling 252 (Cavicchioli et al. 2019) and have been used to indicate belowground secondary 253 productivity (Eisenhauer et al. 2018).

254

255 Biodiversity effects under ambient and manipulated environments

We first quantified the net biodiversity effect (NBE) on ecosystem functioning by calculating the bias-corrected estimation of the log response ratio (Koricheva *et al.* 2013; Lajeunesse 2015):

259
$$NBE = ln\left(\frac{\overline{X}_{mix}}{\overline{X}_{mono}}\right) + \frac{1}{2}\left(\frac{SD_{mix}^2}{n_{mix}\overline{X}_{mix}^2} - \frac{SD_{mono}^2}{n_{mono}\overline{X}_{mono}^2}\right)$$
(1)

where \overline{X}_{mix} , SD_{mix} , and n_{mix} represent the average, standard deviation, and sample size (i.e. number of replicates) of ecosystem function, respectively, in mixtures at a given biodiversity level. \overline{X}_{mono} , SD_{mono} , and n_{mono} represent similar metrics in

263 monocultures. A positive value of NBE indicates a higher ecosystem functioning in 264 mixtures compared with that in monocultures, and vice versa. We calculated NBE 265 under both ambient (NBE_A) and manipulated (NBE_M) environments, for each type 266 and level of global change drivers, each taxonomic group, each ecosystem function, 267 each level of species richness in mixtures, and each time point (if the experiment 268 contained multiple observations). Overall, we obtained 1997 NBE values, including 269 508, 74 and 146 pairs of NBE_A and NBE_M for the warming treatment in microbes, 270 phytoplankton, and plants, respectively, and 170 for drought, 201 for nutrient addition 271and 136 for CO₂ enrichment in plants (see Appendix B: Table B1).

To quantify the influence of environmental manipulations (i.e. global change drivers) on biodiversity effects, we calculated the difference in biodiversity effects between ambient and manipulated environments (Lajeunesse 2011, 2015):

275

$$\Delta NBE = NBE_M - NBE_A \tag{2}$$

where the subscripts A and M indicate ambient and manipulated environments, respectively. A positive value of Δ NBE indicates a higher biodiversity effect under the manipulated environment, and vice versa. The relative change in biodiversity effects induced by environmental manipulations can be derived by rescaling Δ NBE (Hooper *et al.* 2012): $e^{\Delta NBE} - 1 = \frac{e^{NBE_M}}{e^{NBE_A}} - 1$.

281 To understand the mechanisms underlying biodiversity effects, we used the 282 additive partition by Loreau & Hector (2001) to derive the complementarity (CE) and 283 selection effect (SE) on biomass production, based on the 12 studies with available 284 raw data of monocultures (Appendix B: Table B7). We first calculated CE and SE for 285 each mixture and then obtained the mean and standard deviation for each species 286 richness level, each level of environmental manipulation and each experimental time 287 point. For comparison among studies, we standardized CE and SE by the average 288 value of ecosystem function in monocultures (Loreau & Hector 2001; Craven et al. 289 2016). In doing so, we excluded monocultures with too low biomass (i.e. less than 2.5 g m⁻² in grassland and 0.005 mg ml⁻¹ in microbes) following Reich et al. (2012), and 290 291 omitted 100 (out of 1974) CE or SE values accordingly. Specifically, CE and SE were 292 calculated as:

293
$$CE = T_{sqrt} \left(\frac{N \cdot \overline{\Delta RY} \cdot \overline{X}_{mono}}{\overline{X}_{mono}} \right)$$
(3)

294
$$SE = T_{sqrt} \left(\frac{N \cdot cov(\overline{\Delta RY}, \overline{X}_{mono})}{\overline{X}_{mono}} \right)$$
(4)

295where N and ΔRY are the number of species in the mixture and difference between 296 species' observed relative yield in the mixture and expected relative yield, respectively (Loreau & Hector 2001). $T_{sqrt}(x) = sign(x) * \sqrt{|x|}$ represented a 297 298 square-rooted transformation while keeping the original sign, which has commonly 299 been used in previous studies to reduce the influence of skewed distributions of CE 300 and SE (e.g. Isbell et al. 2009). By definition, we can derive the link between different metrics: $e^{NBE} = \frac{N \cdot \overline{\Delta RY} \cdot \overline{X}_{mono}}{\overline{X}_{mono}} + \frac{N \cdot cov(\overline{\Delta RY}, \overline{X}_{mono})}{\overline{X}_{mono}} + 1$. We calculated CE and SE in both 301 302 ambient (CE_A and SE_A) and manipulated environment (CE_M and SE_M). We then 303 calculated the difference in CE or SE between the manipulated and ambient 304 environments:

$$\Delta CE = CE_M - CE_A \tag{5}$$

305

$$\Delta SE = SE_M - SE_A \tag{6}$$

307 In addition to NBE, we also quantified biodiversity effects using the regression 308 slope (BE_{Slope}) between ecosystem functioning and species richness (Baert et al. 309 2018). We calculated the log-log regression slope between species richness and 310 ecosystem functioning in both ambient ($BE_{Slope A}$) and manipulated ($BE_{Slope M}$) 311 environments, for each type and level of environmental manipulations, each 312 taxonomic group, each ecosystem function, and each time point. We then derived 313 their difference ($\Delta BE_{Slope} = BE_{Slope_M} - BE_{Slope_A}$) to characterize the response of 314 biodiversity effects to global change drivers.

315 Both NBE and BE_{Slope} have been widely used for quantifying biodiversity effects 316 in experimental studies and meta-analyses (e.g. Balvanera et al. 2006; Hooper et al. 317 2012; Hautier et al. 2015; Duffy et al. 2017; Baert et al. 2018). NBE is more 318 appropriate if ecosystem functioning saturates rapidly, e.g. from monocultures to two-319 species mixtures. In comparison, BE_{Slope} is more appropriate if ecosystem functioning 320 exhibits a (quasi-) linear increase with species richness on log-log scales (Appendix 321 A). As the realistic scenarios should occur along the continuum between a rapid 322 saturating curve and a (quasi-) linear trend (Cardinale et al. 2012), our usage of both 323 NBE and BE_{slope} facilitates a comprehensive picture of biodiversity effects (see 324 Appendix A for more detailed discussion). For brevity, we presented the results of 325 NBE (and its additive partitions into CE and SE) in the main text and those of BE_{Slope} 326 in the Appendix B.

327

328 Environmental stress intensity and biodiversity effects

For a given taxonomic group and type of global change drivers, experimental communities might respond to environmental manipulations differently across studies. Following Steudel *et al.* (2012), we defined stress intensity as the response of monoculture functions to environmental manipulations within each study:

333
$$Stress intensity = 1 - \begin{pmatrix} X_{mono_M} \\ \overline{X}_{mono_A} \end{pmatrix}$$
(7)

where \overline{X}_{mono_M} and \overline{X}_{mono_A} were average functioning of monocultures in manipulated 334 335 and ambient environments, respectively. A positive (negative) value of stress intensity 336 indicated that global change drivers decreased (increased) monoculture functions and 337 thus provided a stressful (favorable) condition. We calculated stress intensity for each 338 ecosystem function, level of environmental manipulation, and experimental time in 339 each study. We then derived average stress intensity through time over the 340 experimental duration, and we also tested the robustness of our results using the value 341 of stress intensity at the end of the experiment (Steudel et al. 2012).

We note that in our definition of stress intensity, the responses of mixtures were not taken into account because they involved both the direct effects of global change drivers and the potential buffering effects of biodiversity (Hisano *et al.* 2018), which would have confounded our analysis on the relationship between stress intensity and ANBE. However, in the appendix, we also quantified the responses of mixtures to global change drivers, in order to examine the overall effects of environmental changes.

349

350 Statistical analyses

351 To derive pooled effect sizes for biodiversity effects (NBE, Δ NBE, BE_{Slope}, and 352 ΔBE_{Slope}) and their 95% confidence intervals (CIs), we used three-level, hierarchical 353 mixed-effects models to account for the non-independence between biodiversity 354 effect sizes (Nakagawa & Santos 2012; see Appendix A for details). In these models, 355 samples were weighted by the inverse of their variance (see Appendix A for 356 derivation). Fixed-effects terms are type of global change drivers (α), ambient or 357 manipulated environmental conditions (τ), taxonomic group (β) and category of 358 ecosystem functions (γ). Random-effects terms are study (θ) and the combination of 359 species richness (n), level of environmental manipulation (λ), specific ecosystem

function (ψ) and experimental time (φ) nested within study (i.e. as a sub-study ID). The models thus read:

362
$$NBE \sim (\alpha \tau \beta \gamma)_{ijkl} + (\theta / \eta \lambda \psi \varphi)_{mnrst} + \epsilon_{ijklmnrst}$$
(8)

363
$$\Delta NBE \sim (\alpha \beta \gamma)_{ijk} + (\theta / \eta \lambda \psi \phi)_{mnrst} + \epsilon_{ijkmnrst}$$
(9)

364
$$BE_{Slope} \sim (\alpha \tau \beta \gamma)_{ijkl} + (\theta / \lambda \psi \varphi)_{mnrst} + \epsilon_{ijklmnrst}$$
(10)

365
$$\Delta BE_{Slope} \sim (\alpha \beta \gamma)_{ijk} + (\theta / \lambda \psi \varphi)_{mnrst} + \epsilon_{ijkmnrst}$$
(11)

Similarly, we estimated the average and 95% CIs of CE and SE and the differences between ambient and manipulated environment (i.e. ΔCE and ΔSE) using the following models:

369

$$CE \text{ or } SE \sim (\alpha \beta \tau)_{ijk} + (\theta / \eta \lambda \varphi)_{mnrs} + \epsilon_{ijkmnrs}$$
(12)

370
$$\Delta CE \text{ or } \Delta SE \sim (\alpha\beta)_{ij} + (\theta/\eta\lambda\phi)_{mnrs} + \epsilon_{ijmnrs}$$
(13)

We used the function "rma.mv()" of the *metafor* package in R to estimate these effect
sizes (Viechtbauer 2010).

Between-study heterogeneity of effect sizes was evaluated using the extended I^2 373 374 and Cochran's Q statistic (Nakagawa et al. 2012; see Appendix A for details). The extended I^2 quantified the fraction of variation in effect size attributed to between-375 study variance in multi-level models, and the Cochran's Q statistic evaluated the 376 377 significance of between-study variance (Appendix A). Our analyses found high between-study heterogeneity of effect sizes ($I^2 > 60\%$ and p < 0.01 for NBE, ΔNBE , 378 379 BE_{Slope} and ΔBE_{Slope} ; see Appendix B: Table B3). We thus conducted multi-level 380 meta-analyses for both NBE and BE_{Slope} to explore how between-study heterogeneity 381 may be explained by potential moderators. Our analyses showed significant 382 differences in NBE and BE_{Slope} across different combinations of taxonomic groups, 383 type of global change drivers and category of ecosystem functions (Appendix B: 384 Table B4 & B5). Therefore, we did not estimate overall effect sizes across all 46 385 studies. We also tested the effects of other potential moderators (type of nutrient 386 addition, vegetation type, and experimental unit), which were found to have non-387 significant effects on NBE or BE_{Slope} and thus omitted in the main text (see Appendix 388 A).

Using five long-term studies in our dataset (one on microbes and four on grasslands; see Appendix B: Table B7), we tested whether the effects of global change drivers on NBE and BE_{Slope} for biomass production varied through time. For each study, we used mixed-effects models with experiment duration as a fixed effect,

and the combination of species richness level and level of environmental manipulation as the random effect. Similarly, we further tested the relationship between $\Delta NBE/\Delta BE_{Slope}$ and experimental duration. We fitted each model using linear, quadratic, and logarithmic forms of environment duration and calculated the Akaike information criterion (AIC) to determine the best model. We used the function "Imer()" of the *nlme* package in R to fit these models.

399 We also used mixed-effects models to test the relationships between ΔNBE , 400 ΔBE_{Slope} , ΔCE or ΔSE and stress intensity (SI):

401 $\Delta NBE, \Delta BE_{Slope}, \Delta CE \text{ or } \Delta SE \sim SI + (\alpha \beta \gamma / \theta / \eta \lambda \psi)_{ijkmnrs} + \epsilon_{ijkmnrs}$ (14)

402 where *SI* was the fixed effect, and other variables $(\alpha, \beta, \gamma, \theta, \eta, \lambda, \psi)$ were random effects. 403 We also tested the two-way interactions of stress intensity with taxonomic group, type 404 of global change drivers, and category of ecosystem function (Appendix B: Table B6). 405 Again we used the function "lmer()"to fit this model.

406 Publication bias was analyzed using the function "funnel()" from the metafor 407 package in R (Viechtbauer 2010) and Egger's test was conducted to test the 408 asymmetry of the funnel plot of our model (with only random effect) in NBE, Δ NBE, 409 CE, Δ CE, SE, and Δ SE, as well as BE_{Slope} and Δ BE_{Slope} (Nakagawa *et al.* 2017). We 410 found overall low publication biases in our dataset; a significant bias occurred only 411 for ΔSE (Egger's p = 0.0110, Appendix B: Fig. B2). But our further sensitivity 412 analysis showed that the publication bias in ΔSE did not influence the robustness of 413 our results (see Appendix A).

414

415 **Results**

416 Biodiversity had generally positive effects (NBE and BE_{Slope}) on biomass 417 production in both ambient and manipulated environments, regardless of the type of global change drivers and taxonomic group (p < 0.05; Fig. 2 & Appendix B: Fig. B3). 418 419 Biodiversity also tended to enhance biogeochemical processes, but these effects were 420 weaker and non-significant for certain combinations of global change drivers and 421 taxonomic groups (p > 0.05; Fig. 2 & Appendix B: Fig. B3). NBEs under ambient and 422 manipulated environmental conditions were positively correlated in each of the six 423 combinations of global change drivers and taxonomic groups ($r = 0.42 \sim 0.82$; 424 Appendix B: Fig. B4).

425 Although warming did not change the sign of NBEs, it modulated their 426 magnitude (as measured by ΔNBE) in microbes and phytoplankton (Fig. 2; $Q_M =$ 427 24.76, p < 0.0001, Appendix B: Table B4). In microbial communities, warming 428 decreased NBEs on biomass production on average by 20% (e^{-0.222}, number of effect sizes (N) = 506, number of studies (n) = 3, p = 0.0726). In phytoplankton 429 430 communities, however, warming increased NBEs on biomass production on average 431 by 61% (e^{0.360}, N = 64, n = 3, p = 0.002; Fig. 2), and this increase was stronger as the 432 degree of warming increased (Appendix B: Fig. B5). For biodiversity effects 433 measured by BE_{Slope}, warming also increased BE_{Slope} on biomass production in 434 phytoplankton ($\Delta BE_{Slope} = 0.109$, N = 12, n = 3, p = 0.019; Appendix B: Fig. B3), but 435 it had a non-significant effect in microbes (p > 0.05; Appendix B: Fig. B3).

436 In plants, global change drivers had overall non-significant effects on NBE, 437 regardless of the type of global change drivers and ecosystem functions measured (p > 438 0.05; Fig. 2). Global change drivers also did not influence BE_{Slope} for biogeochemical 439 processes, but BE_{Slope} for biomass production responded significantly to global 440 change drivers, except for CO₂ enrichment. Specifically, BE_{Slope} was decreased by 441 warming ($\Delta BE_{Slope} = -0.103$, N = 28, n = 4, p = 0.034; Appendix B: Fig. B3) and nutrient addition ($\Delta BE_{Slope} = -0.085$, N = 49, n = 7, p = 0.011; Appendix B: Fig. B3), 442 whereas it was increased by drought ($\Delta BE_{Slope} = 0.099$, N = 21, n = 10, p = 0.026; 443 444 Appendix B: Fig. B3).

445 Based on the 12 studies with monocultures where raw data were available, we 446 found that complementarity effects (CE) were generally positive (p < 0.05) and 447 selection effects (SE) were generally not different from 0 in both ambient and 448 manipulated environments, regardless of the type of global change drivers or 449 taxonomic group (Fig. 3). Warming decreased CE for microbial biomass production by 24.3% (N = 470, n = 2, p < 0.0001) and increased CE for phytoplankton 450 451 production by 82.6% (N = 10, n = 1, p = 0.008) (Fig. 3a). But warming did not alter 452 SE in either microbes or phytoplankton communities (Fig. 3b). In plant communities, 453 global change drivers had no significant effects on either CE or SE on biomass 454 production (p > 0.05; Fig. 3).

Using the five long-term studies in our dataset, we found that biodiversity effects
(i.e. NBE and BE_{Slope}) generally increased with experimental duration in both ambient
and manipulated environments (Fig. 4; Appendix B: Fig. B6). The difference in
biodiversity effects between ambient and manipulated environments, measured by
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459 either ΔNBE or ΔBE_{Slope} , decreased with experimental duration in microbial 460 experiments (p < 0.05; Appendix B: Fig. B7 & B8). Yet, such differences did not vary 461 with experimental duration in the four grassland studies (p > 0.1; Fig. 4; Appendix B: 462 Fig. B7 & B8).

463 Warming in phytoplankton communities and drought in plant communities on 464 average decreased monoculture functions (positive value of stress intensity), and 465 nutrient addition in plant communities on average increased monoculture functions 466 (negative value of stress intensity) (p < 0.05; Fig. 5a). However, the effects of drought 467 and nutrient addition on ecosystem functions became non-significant in more diverse 468 plant communities (p > 0.10; Appendix B: Fig. B9). We found that both ΔNBE and 469 ΔBE_{Slope} increased as stress intensity increased (p < 0.001 for both; Fig. 5b,c), 470 indicating stronger biodiversity effects in more stressful environments. These results 471 were robust whether we used biodiversity effects (i.e. ΔNBE and ΔBE_{Slope}) averaged 472 across time or at the end of the experiment (Appendix B: Fig. B10). Moreover, the 473 positive relationship between stress intensity and ΔNBE or ΔBE_{Slope} also held in 474 models that incorporated the effects of taxonomic groups, type of global change 475 drivers, category of ecosystem function, and their interaction with stress intensity 476 (Appendix B: Table B6).

477

478 Discussion

479 The past decades have seen major progress in understanding the responses of 480 ecosystems to environmental and biodiversity changes. Recent meta-analyses have 481 documented the individual effects of global environmental change (Yuan et al. 2017; 482 Song et al. 2019) or biodiversity (Balvanera et al. 2006; Cardinale et al. 2012; Hooper 483 et al. 2012) on ecosystem functioning. Our synthesis of 46 factorial experiments 484 advances current understanding by demonstrating the interactive effects between 485 biodiversity and environmental changes on ecosystem functions. We found that 486 biodiversity consistently enhanced ecosystem functioning in both ambient and 487 manipulated environments, but global change drivers could modulate the strength of 488 biodiversity effects. Specifically, the effects of biodiversity on ecosystem functioning 489 were stronger in stressful environments than in more favorable environments. 490 Therefore, biodiversity contributes to buffering ecosystem functions across a range of 491 stressful conditions associated with current and future global change scenarios.

492

493 Biodiversity effects in ambient and manipulated environments

494 Biodiversity generally promoted ecosystem functioning in both ambient and 495 changing environments (Fig. 2; Appendix B: Fig. B3), although its positive effects on 496 biogeochemical processes were relatively weak and often non-significant, possibly 497 due to small sample sizes. Based on a subset of our dataset (12 studies where the 498 necessary data were available), we found that these positive biodiversity effects were 499 mainly due to complementarity effects (Fig. 3), which was consistent with earlier 500 findings from plant diversity experiments under ambient environmental conditions 501 (Loreau & Hector 2001; Tilman et al. 2014; Huang et al. 2018) as well as under 502 altered conditions (Craven et al. 2016).

503 While global change drivers did not change the directions of biodiversity effects 504 (both NBE and BE_{Slope}), they mediated the magnitude of biodiversity effects (Fig. 2, 505 Appendix B: Fig. B3). Warming increased biodiversity effects on biomass production 506 in phytoplankton, but decreased it in microbes. The contrasting responses of 507 biodiversity effects to warming in microbes versus phytoplankton may be explained 508 by differential effects of warming on interspecific interactions. In microbial 509 communities, warming has been shown to enhance interspecific competition between 510 culturable microbial species, even leading to competitive exclusion (Jiang & Morin 511 2004). Such intensified competition by warming should lead to a reduction in CE in 512 microbes (Fig. 3; Parain et al. 2019). In phytoplankton communities, however, 513 warming might promote interspecific facilitation, for instance by stimulating certain 514 groups or species such as N₂-fixing cyanobacteria (Brauer et al. 2015; Striebel et al. 515 2016), which would lead to an increased CE (Fig. 3).

516 In plant communities, warming and nutrient addition decreased, and drought 517 increased biodiversity effects quantified by the regression slope (BE_{slope}) between 518 species richness and biomass production (Appendix B: Fig. B3). These global change 519 drivers also similarly influenced NBEs (as well as CE), but the effects were weak and 520 often non-significant (Figs. 2 & 3). These apparently inconsistent results for ΔNBE 521 and ΔBE_{slope} may be due to the fact that BE_{slope} usually have smaller variances 522 compared with NBE, which makes it easier to detect statistically significant effects 523 for ΔBE_{slope} (see Appendix A). Furthermore, the responses of NBE and BE_{slope} to 524 global change drivers are reconciled from the perspective of stress intensity (see 525 below).

526 Several recent studies showed that biodiversity effects increased with time 527 (Reich et al. 2012; Guerrero-Ramírez et al. 2017; Huang et al. 2018). In line with this 528 finding, our analysis based on five long-term studies (one on microbes and four on 529 plants) showed that biodiversity effects generally increased with time under both 530 ambient and altered environmental conditions (Fig. 4, Appendix B: Fig. B6). 531 Moreover, we found a negative interaction between warming and experimental 532 duration on biodiversity effects in the microbial experiment (Appendix B: Fig. B7 & 533 B8), suggesting that the negative effect of warming on biodiversity effects increased 534 over time. In the four plant experiments, interactions between global change drivers 535 and time were all non-significant, possibly explained by their longer generation times 536 and less intense manipulations of global change drivers compared with microbes. In 537 particular, the longest plant experiment in our data lasted 15 years, which may not be 538 sufficient for the plants to acclimate to environmental changes and for novel 539 interspecific interactions to manifest or evolve (Jessup et al. 2004; but see Zuppinger-540 Dingley et al. 2014).

- 541
- 542

Biodiversity buffers environmental stress

543 By quantifying the responses of monocultures in each study, we found that the 544 stress induced by the altered environment differed markedly among studies, even for 545 the same global change driver and taxonomic group (Fig. 5). In other words, the same 546 environmental manipulation can be perceived as either more or less stressful by 547 different experimental communities (e.g. different plant species pools used in 548 different experiments). Interestingly, we found that stress intensity was positively 549 related to both ΔNBE and ΔBE_{Slope} . Thus, stress intensity provides a useful indicator 550 to predict how global change drivers alter biodiversity effects. The positive 551 relationship between ΔNBE or ΔBE_{Slope} and stress intensity could be due to higher 552 resistance to environmental stress of diverse mixtures compared with monocultures 553 (H1 in Fig. 1; Appendix B: Fig. B9). When global change drivers induced high stress 554 intensity (e.g. warming in phytoplankton communities or drought in plant 555 communities), diverse mixtures exhibited higher resistance to environmental stress 556 and better maintained their functions (in line with Isbell et al. 2015), resulting in an 557 enhanced biodiversity effect. Conversely, when global change drivers reduced 558 environmental stress compared with ambient conditions (e.g. nutrient addition on 559 plants), diverse mixtures benefitted less than monocultures, resulting in a weakened This article is protected by copyright. All rights reserved

560 biodiversity effect. Overall, communities with higher species diversity were less 561 influenced by environmental changes, because biodiversity could buffer the negative 562 or positive effects of global change drivers on ecosystem functioning (Fig. 5, 563 Appendix B: Fig. B9). This can occur directly through niche complementarity, via 564 buffering of microenvironmental stress (Wright et al. 2014), or both. Such buffering 565 effects can increase not only ecosystem resistance to directional changes in the 566 environment, but also the temporal and spatial stability of ecosystems in fluctuating 567 environments (Yachi & Loreau 1999; Wang et al. 2019).

568 Our results are consistent with recent findings that biodiversity can increase the 569 resistance and resilience of ecosystems (Mori et al. 2013; Isbell et al. 2015; Hisano et 570 al. 2018; but see Baert et al. 2016; De Boeck et al. 2018; Pennekamp et al. 2018). The 571results are also in line with the stress gradient hypothesis (Bertness & Callaway 1994), 572 which predicts increased biodiversity effects (particularly CE) with increasing stress 573 intensity due to reduced interspecific competition or enhanced facilitation (He et al. 574 2013; Wright et al. 2017). That said, our analyses based on a small sample size 575 showed that neither CE nor SE exhibited a significant relationship with stress 576 intensity, although both showed positive trends (Appendix B: Fig. B11).

577 Recent studies hypothesized that biodiversity effects could first increase but then 578 decrease with stress intensity, because high stress intensity may restrict population 579 growth to such a level that ecosystem functions are extremely low regardless of the 580 level of biodiversity (Baert et al. 2018). Our results did not support this hypothesis. 581 This could be due to the fact that our data included few experiments with extreme 582 environmental conditions, hence most values of stress intensity in our data fell into a 583 relatively narrow interval (e.g. the 5% and 95% quantiles of stress intensity were [-584 0.54, 0.60], see Fig. 5). Alternatively, it may be intrinsically difficult to detect the 585 threshold level of environmental conditions based on empirical data, even if such a 586 threshold exists (Hillebrand et al. 2020). Disentangling these possibilities will require 587 future experiments that cover more extreme values of stress intensity (De Boeck et al. 588 2018).

589

590 Future directions

591 Several future directions emerge from our study. On the experimental side, more 592 efforts are needed to better understand the interactions between biodiversity and the 593 environment. First, most studies in our datasets (34 out of 46) are from grasslands; so, This article is protected by copyright. All rights reserved 594 research efforts should be extended to other ecosystems (e.g. forests) and taxonomic 595 groups (e.g. microbes and phytoplankton). In particular, we did not find any study that 596 tested the interaction between biodiversity and eutrophication in microbes or 597 phytoplankton. Recent studies showed that nutrient addition in microbial communities 598 could shift species interactions from facilitation to competition (Hoek et al. 2016; 599 Piccardi et al. 2019), whereas nutrient addition in phytoplankton communities shifted 600 competition from nutrients to light and thus altered species interactions (Burson et al. 601 2018). Thus, biodiversity effects might be altered by eutrophication in microbes and 602 phytoplankton. Second, all studies in our dataset manipulated species richness as the 603 main facet of biodiversity. However, recent studies highlighted effects of other facets 604 of biodiversity (e.g. functional or phylogenetic diversity) on ecosystem functioning (Steudel et al. 2016; Craven et al. 2018; Huang et al. 2018). Whether environmental 605 606 change alters the effects of different facets of biodiversity in different ways remains 607 unclear and should be explored further. Third, very few experiments (one in our 608 dataset) tested the interaction between biodiversity and multiple environmental 609 change drivers (Reich et al. 2001). Although a recent meta-analysis suggested that 610 different global change drivers might have weak interactive effects (Song et al. 2019), 611 there are case studies showing significant interactions (Rillig et al. 2019; Reich et al. 612 2020), and whether they exhibit higher-order interactions with biodiversity remains 613 unknown. Future experiments should address the interactions between different global 614 change drivers, including additional ones to those considered here (e.g. climatic 615 variability). On the other hand, new theory is required for clarifying the ecological 616 mechanisms underlying interactions between biodiversity and the environment. 617 Classic theories on biodiversity-ecosystem functioning were built upon competition 618 models that assumed constant environmental conditions (Tilman et al. 1997; Loreau 619 1998). A constant environment can be readily achieved in experimental settings but 620 rarely in natural ecosystems. New theoretical studies should allow for different 621 environmental conditions under which biodiversity-ecosystem functioning 622 relationships are predicted, to explore possible mechanisms by which the physical and 623 biological environments might modify biodiversity effects (e.g. De Laender 2018). 624

Lastly, experimental and theoretical findings need to be reconciled with the increasing number of observational studies along natural gradients of biodiversity and environmental conditions, for both terrestrial (Fei *et al.* 2018; Guo *et al.* 2019) and marine (Benkwitt *et al.* 2020) ecosystems. Recent observational studies revealed This article is protected by copyright. All rights reserved 628 either enhanced (Fei et al. 2018; Hisano & Chen 2020), similar (Liang et al. 2016) or 629 weakened (Mori 2018) effects of tree diversity on ecosystem productivity in warmer 630 regions. In comparison, our meta-analysis revealed an overall weak, negative effect of 631 warming on plant biodiversity effects. Moreover, several continental-scale studies 632 showed that plant diversity increased ecosystem productivity or biomass in dry 633 regions, but such effects were weakened or even reversed in moist regions (Ratcliffe 634 et al. 2017; Fei et al. 2018; Guo et al. 2019). These results are consistent with the 635 overall positive effects of drought on plant biodiversity effects in our analyses (Fig. 2 636 & Appendix B: B3). Syntheses across different approaches (e.g. experimental, 637 theoretical, and observational) will help clarify which patterns and mechanisms are 638 general and which are context dependent.

639

640 Conclusions

641 By synthesizing experiments across various taxonomic groups and types of 642 global change drivers, our study demonstrates persistent positive effects of 643 biodiversity on ecosystem functions across taxa despite environmental change. The 644 positive effects of biodiversity increased over time under both ambient and 645 manipulated environmental conditions and were stronger when environmental change 646 increased stress intensity. Independent of the level of environmental stress, 647 biodiversity helps to maintain ecosystem functioning at relatively stable levels, 648 whereas stability is lowered in species-poor communities or monocultures. Moreover, 649 our synthesis contributes to exposing knowledge gaps and informing future research 650 to advance our understanding of the interaction between biodiversity and environment. 651 Recent studies showed that the magnitude of biodiversity effects on ecosystem 652 functioning was comparable to that of effects of environmental changes in both 653 experimental and natural communities (Hooper et al. 2012; Tilman et al. 2012; Duffy 654 et al. 2017). Taken together, biodiversity provides an important biological buffer to 655 maintain ecosystem functioning in the face of environmental changes.

656

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669

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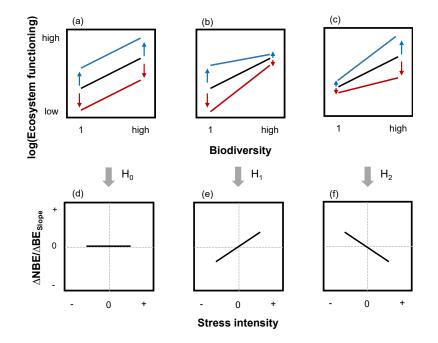
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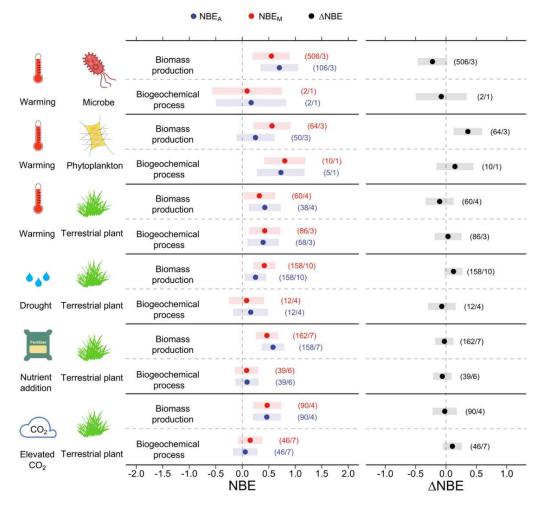
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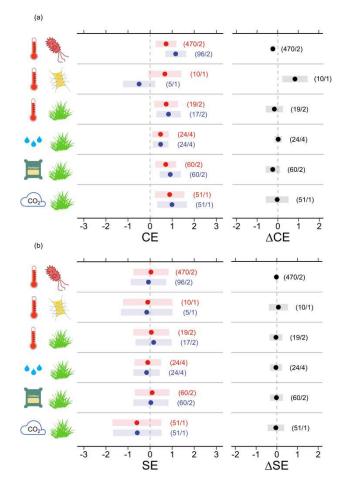
945 Figure 1 Scenarios underlying the responses of biodiversity-ecosystem functioning 946 relationships to stress intensity (a-c) and the resulting relationship between changes in 947 biodiversity effects (ΔNBE , ΔBE_{Slope}) and stress intensity (e-f). Global change drivers 948 can either increase or decrease ecosystem functioning and thus result in a more 949 favorable or stressful environment for ecological communities. Under the null 950 scenario (H_0) where mixtures are similarly influenced by global change driver as 951 monocultures, biodiversity effects are not altered by global change drivers and we 952 will observe no relationship between stress intensity and ΔNBE or ΔBE_{Slope} . Under 953 the scenario (H_1) where mixtures are less influenced by global change drivers 954 compared with monocultures, biodiversity effects are enhanced in more stressful 955 environments but weakened in more favorable environments, and we will observe a 956 positive relationship between ΔNBE or ΔBE_{Slope} and stress intensity. Under the 957 scenario (H₂) where mixtures are more influenced by global change drivers compared 958 with monocultures, biodiversity effects are weakened in more stressful environments

but enhanced in more favorable environments, and we will observe a negative relationship between ΔNBE or ΔBE_{Slope} and stress intensity.



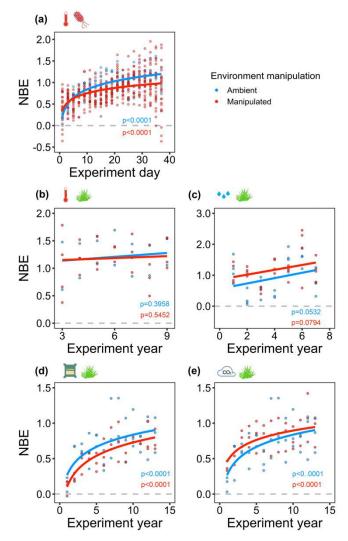
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962 Figure 2 Net biodiversity effects (NBE) on ecosystem functioning under ambient 963 (NBE_A, blue) and manipulated environmental conditions (NBE_M, red), and the 964 difference between them ($\Delta NBE = NBE_M - NBE_A$, black) across different 965 combinations of types of global change drivers (warming, drought, nutrient addition 966 or elevated CO₂), taxonomic groups (microbes, phytoplankton or plants) and 967 ecosystem functions (biomass production or biogeochemical process). The numbers in 968 brackets show the number of effect sizes and studies. The points and shades represent 969 the estimated mean and confidence interval, respectively, from linear mixed-effects 970 models (see Methods). Confidence intervals (95%) not overlapping with the dashed 971 line (i.e. 0) indicate statistical significance (p < 0.05). 972



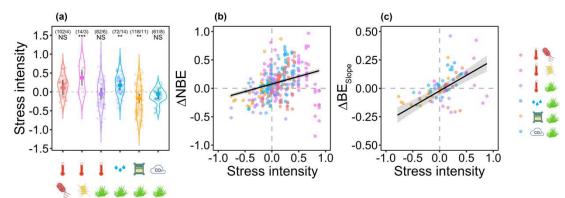
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974 Figure 3 Complementarity (CE) (a) and selection (SE) (b) effects on biomass 975 production under ambient (CE_A and SE_A, respectively, blue) and manipulated 976 environmental conditions (CE_M and SE_M, respectively, red) and the difference between them ($\Delta CE = CE_M - CE_A$, $\Delta SE = SE_M - SE_A$, black), across different 977 978 combinations of types of global change drivers (warming, drought, nutrient addition 979 or elevated CO_2) and taxonomic groups (microbes, phytoplankton and plants). The 980 numbers in brackets show the number of effect sizes and studies. Analyses were based 981 on 12 experiments with raw data of monocultures, and CE and SE were square-root 982 transformed (restoring the sign after transformation; see Methods). The points and 983 shades represent the estimated means and confidence intervals, respectively, from 984 linear mixed-effects models (see Methods). Confidence intervals (95%) not 985 overlapping with the dashed line (i.e. 0) indicate statistical significance (p < 0.05).



986

Figure 4 Net biodiversity effect (NBE) as a function of experimental duration across
different types of environmental manipulations and taxonomic groups: (a) warming
on microbes (Pennekamp *et al.* 2018); (b) warming on plants (Cowles *et al.* 2016); (c)
drought on plants (Wagg *et al.* 2017); (d) nutrient addition on plants (Reich *et al.*2001); (e) CO₂ enrichment on plants (Reich *et al.* 2001).



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Figure 5 Distribution of environmental stress intensity for each combination of types
of environmental manipulations and taxonomic group (a), and relationship between
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995 changes in biodiversity effects (ΔNBE , ΔBE_{Slope}) and stress intensity (b, c). In (a), 996 light-colored points represent the distribution of stress intensity, which was calculated 997 as the relative difference under ambient vs. manipulated environmental conditions in 998 each study. Positive (negative) values mean lower (higher) monoculture functions in 999 manipulated environments. Colored points represent their averages with 95% 1000 confidence intervals (black bars), and numbers on the top represent sample sizes. The 1001 numbers in brackets show the number of effect sizes and studies. In (b) and (c), 1002 changes in biodiversity effects between manipulated and ambient environmental 1003 conditions were calculated in two ways: the difference in net biodiversity effect (Δ NBE; N = 327, n = 46, p < 0.0001) (b) and the difference in the log-log slope 1004 1005 between ecosystem functioning and species richness (ΔBE_{Slope} ; N = 76, n = 46, p = 1006 0.0003) (c). Trendlines and 95% confidence intervals are given. NS, not significant 1007 (p>0.1); *, p<0.05; **, p<0.01; ***, p<0.001.