# Nucleotide sequences of the PA and PB1 genes of B/Ann Arbor/1/66 virus: comparison with genes of B/Lee/40 and type A influenza viruses 

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#### Abstract

Summary The complete sequences of the PA and PB1 genome RNA segments of B/Ann Arbor/1/66 virus have been determined. The PA vRNA is 2308 bases long. Its complementary RNA has a single open reading frame of 2187 bases, capable of encoding a PA protein of 726 amino acids with a molecular weight of $83,175 \mathrm{Da}$. The predicted PA polypeptide has an overall net charge of -7.5 at pH 7.0 . The PB1 vRNA is 2369 bases long. Its complementary RNA has a single open reading frame of 2277 bases, capable of encoding a PB1 protein of 752 amino acids with a molecular weight of $84,332 \mathrm{Da}$. The predicted PB1 polypeptide has an overall net charge of +18.5 at pH 7.0 . Sequence homology comparisons of the PA and PB1 polypeptides from B/Ann Arbor/1/66 virus to the PA and PB1 polypeptides of type A influenza virus reveal respective homologies of approximately 38 and $60 \%$. This high cross-type homology ( $61 \%$ ) was previously reported for the PB1 protein of B/Lee/40 virus (Kemdirim et al., 1986). The cross-type homology for the PA protein is similar to that of other non-polymerase proteins, but is substantially lower than that seen for the PB1 protein. Thus, the high cross-type homology that exists for the PB1 gene does not appear to be a characteristic of all polymerase genes.


PA; PB1; Polymerase; Dideoxynucleotide sequencing; Influenza type A and B virus

[^0]Until 1986, complete sequence information for the RNA of type B influenza virus existed only for the non-polymerase genes of $B /$ Lee/40 virus; then the B/Lee/40 PB1 gene sequence was published (Kemdirim et al., 1986). The polypeptide predicted from this PB1 gene showed a significantly higher level of sequence homology with the PB1 polypeptides of influenza type A viruses than that occurring for any of the other non-polymerase proteins, suggesting that a functional constraint might be operating as a significant selection mechanism, restricting sequence variation in the PB1 gene. In the absence of sequence information for the PB2 and PA genes of influenza type B virus, it was not known if this high level of cross-type homology would be typical of all three polymerase genes.

The PB1 and PA genes of influenza B/Ann Arbor/1/66 (B/AA/1/66) wild-type (wt) virus were sequenced as part of a project in which all six non-glycoprotein genes of both the cold-adapted (ca) and wt B/AA/ $1 / 66$ viruses are being compared in order to catalogue the changes that occur during the process of cold-adaptation. With the sequencing of both wt and ca $\mathrm{B} / \mathrm{AA} / 1 / 66$ viruses, we confirm the high cross-type homology for the PB1 polypeptide, but show that the PA polypeptide exhibits a level of cross-type homology similar to that of the NP (37\%), HA 2 (39\%), and NA (35\%) proteins (Kemdirim et al., 1986).

The complete sequences of the PA and PB1 genes were determined by a combination of two RNA sequencing techniques. The first $60-70$ nucleotides at the $3^{\prime}$-termini of both the PA and PB1 vRNA segments were sequenced by a direct chemical method described in Peattie (1979) using vRNA segments isolated on, and subsequently eluted from, a $3 \%$ polyacrylamide gel. The remaining sequences were determined by dideoxynucleotide chain termination sequencing procedures described previously (DeBorde et al., 1986). All ambiguities in these sequences were resolved using terminal deoxynucleotidyl transferase enzyme (DeBorde et al., 1986). The sequence that extended from each primer overlapped the position of the next primer by at least 20 nucleotides in every case. The complete nucleotide and predicted amino acid sequences for the PA and PB1 genes are presented in Figs. 1 and 4 , respectively. The oligodeoxynucleotide primers were all 15 bases long, except for a 12 -nucleotide-long primer beginning at residue 9 in PB1, and their positions are underlined in Figs. 1 and 4.

All sequence comparisons, manipulations, and calculations were performed using the programs developed by Queen and Korn (1984) and distributed by Beckman Instruments, Inc., Palo Alto, CA, as the Microgenie Sequence Software package.

The PA vRNA segment is 2308 nucleotides long (Fig. 1). The first protein initiation codon in the complementary RNA (cRNA) starts 30 nucleotides in from its $5^{\prime}$-end, and is encompassed by an open reading frame of 2187 nucleotides extending to the first termination codon beginning at nucleotide 2208. The PA polypeptide encoded by this open reading frame is 726 amino acids in length with a calculated molecular weight of $83,175 \mathrm{Da}$. A potential polyadenylation site composed of five consecutive adenine residues is present at nucleotides 2288-2292. The predicted mRNA would be 2292 nucleotides long, prior to capping and the addition of poly (A). No other major open reading frame exists. The next longest unterminated coding sequence in any of the reading frames would encode only 44 amino acids.

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 $2^{211}$ an cal ace nug aca cca caa lay gaa at moto























1740 1800 1770



 1950
${ }^{1950}$ गTIT



2160 , 2290




Fig. 1. Nucleotide and predicted polypeptide sequence of the PA gene of B/AA/1/66 virus. The sequence is presented $5^{\prime} \rightarrow 3^{\prime}$ in the $(+)$ messenger strand sense. The underlined areas indicate the position and sequence of the oligodeoxynucleotide primers that were used. The numbers in () are the lengths of the nucleic acid and polypeptide at that point.

A comparison of the PA protein sequences of $\mathrm{B} / \mathrm{AA} / 1 / 66$ and $\mathrm{A} / \mathrm{NT} / 60 / 68$ using the Microgenie alignment program is shown in Fig. 2. Homologous amino acids are underlined. The overall charge of the $\mathrm{B} / \mathrm{AA} / 1 / 66$ PA protein is predicted to be -7.5 , based on $a+1$ charge for each arginine and lysine residue, $a+0.5$ charge for each histidine residue, and a -1.0 charge for each aspartic acid and glutamic acid residue calculated at pH 7.0 . However, the net charge is asymmetrically distributed over the length of the protein as follows: the $\mathrm{NH}_{2}$-terminal half of
 A/NT/60/68 PA N-terminus Met Glu Asp Phe val ArgGln Cys Phe Asn Pro Met Ile Val Glu Leu Ala Glu Lys Ala Met (21) Ala Glu Phe Ser Glu Asp Pro Glu Leu Gin Pro Ala Met leu Phe Asn le Cys Va His Leu Glu Val Gys Tyr Val le Ser Asp (50) Lys Glu Tyr Gly Glu Asp Leu Lys lie Glu Thr Asn Lys Phe Ala Ala lle Cys Thr His Leu Glu Val Cys Phe het Tyr Ser Asp (50)
 Phe His Phe Jle Asn glu Gingly Glu Ser lle val Val glu leu Asp Asp Pro Asn Ala Leu Leu Lys His Arg Phe glu Ile (78)
Ile Glu Gly Met Pro Arg Asn lle Ala Tro Met Val Gia Arg Ser Lev Ala Ging gu His Gly lle GTu Thr Pro Arg Tyr Leu Ala fiog lle GluGly Arg Asp Arg Thr Het Ala Trp Thr Val Val Asn Ser lie Gys Asn Thr Thr Giy Ala Glu Lys Pro lys phe Leu Pro (iof)

Asp Leu Phe Asp Tyr Lys Thr Lys Arg Phe lle glu va? Giy lle Thr Lys Gyy Leu Ala Asp Asp lyr Phe Trp Lys Lys (i35) Asp Leu Tyr Asp Tyr Lys Glu Asn Arg Phe lle glu lie Gly val Thr Arg Arg Glu val His lle Tyr Tyr Leu Glu Lys Ala Asn (i3g) Lys Giu Lys Leu Giy Asn Ser Met Glu Leu Met lle Phe Ser $\quad$ Tyr Asn Gln Asp Tyr Ser leu Ser Asn Glu His Ser Leu Asp (l63)
Lys lle Lys Ser Glu Asn $\quad$ Fhr His lle His lle Phe Ser Phe Thr Gly Glu Glu Met Ala Thm Lys Ala Asp Tyr The Leu Asp (lgu) Lys lle Lys Ser Glu Asn Thr His lle His lle Phe Ser Phe Thr Gly Glu Glu Met Ata Thr Lys Ala Asp Tyr Thr Leu Asp (igu)
 Glu Glu Ser Arg Ala Arg lie Lys Thr Arg Leu Phe Jor Ile Arg Gin Glu Met Ala Ser Arg Gly Leu frp Asp Ser Phe Arg gla ligh)
Val Leu lle Gly Glu Glu Asp lle Glu Lys Gly lle Asp phe lys Lew Gly Gin Thr lle Ser Lys Leu Arg Asp lle Ser val pro gith Ser Glu Arg Gly glu Glu Thr lle Glu Giu Arg Phe Glu Me Thr Giy Thr Met Arg Argleu Ala Asp Gin Ser leu Pro (z2ol

Ala Gily Phe Ser Asn Phe Glu Gly Met Arg Ser Tyr lle Asp Asn lle Asp Pro lys Gly Ala lle Glu Arg Asn Leu Ala Arg Met (24b) Pro Asn Phe Ser Cys lea Glu Asn Phe ArgAla Tyr val Asp Gly Phe Glu Pro Asn Gly Tyr Ile Glu Gly Lys Leu Ser Gln Met (z4g)

Ser Pro Leu Val Ser Val Thr Pro Lys lys Leu Lys Trp Glu Asp Leu Arg Pro lle Giy Pro His fle Tyr Ser His Glu Leu (274) Ser Lys Giu Yal Asf Ala Lys Ihe Glu Pro Phe lew lys Thr Thr Pro Arg Pro Ile Arg Leu Pro Asp Gly Pro (274) Pro Giu Val Pro Tyr Asn Ala Phe Leu Leu Met Ser Asp Glu leu Gly beu Ala Asn Met Thr Giu Gly Lys Ser lys Lys Pro Lys (303) Pro Cys Phe Gin Arg Ser Lys Phe Leu Leu Met Asp Ala Leu lys Leu Ger Ile Glu Asp Pro Ser His Giu Gly Glu (300)

Thr beu Ala lys glu Cys leu Glu Lys Jyr Ser Thr Leu Arg Asp Gln The Asp Pro ITe Leu lle Met Lys Ser Gif lys (330) Gly lle Pro Leu fyr Asp Ala lle Lys Cys Met Arg for Phe Phe Gly Trplys Glu Pro Tyr Ile Val Lys Pro His Glu Lys (328)

Ala Asn Glu Asn Phe Leu Trp Lys Leu Trp Arg Asp Cys Va? Asn Thr Ile Ser Asm Glu Glu Gly lle Asn Pro Asn Tyr Leu Leu Ser Trp Lys Glo Val Leu Ala Glu Lew Gln Asp Ile Glu Asn Glu Glu lys lle Pro Arg Thr (357)

Ser Asn Giu Leu Gln Lys Thr Asn Tyr Ala Lys Trp Ala Thr Gly Asp Gly Lew Thr Tyr Gln Lys lle Met Lys Glu Val Ala lle (381) Lys Asm Met Lys Lys Thr Ser Gin Leu Lys Trp Ala Leu Gly Glu Asn Met Ala Pro Glu Lys Vat Asp Phe Asp Asn Cys Arg (385)
Asp Asp Giu Thr Met Tyr Gin Gluglu Pro lys lle Pro Asn lys Cys Arg Val Ala Ala Trp Val Gin thr Giu Met Asn beu leu (4io\} Asp Val Ser Asp Leu Lys Gin Tyr Asp Ser Asp Glu Pro Glu Lew Arg Ser lew Ser Ser Trp Ile Gin Asn Glu Phe Asn hys Ala (4la)

Ser Thr Leu Thr Ser lys Arg Ala Leu Asp Lew Pro Glu lle Gly Pro Asp Val Ala Pro Val glu His val Gly Ser Glu Arg Arg (abg) Cys Glu Leu Thr Asp Sef Thr Tro lle Glu lew Asp Glu lle Gly Gla Asp Val Ala Pro lle Glu Tyr lle Ala Ser Met Arg Arg (443)
lys Tyr Phe Val Asn Glu Lle Asn Tyr Cys Lys ATa Ser Thr Val Met Met Lys Jyr Val Leu phe His Thr Ser Leu Leu Asn Glu fag) Asn Iyr Phe Thr Ala Glu Val Ser his Cys Arg Ala Thr Gu Tyr lle Met lys Gly Val Iyr ile Asn Thr Ala leuleu Asn Ala (472)
Ser Asn Ala Ser Met Gly lys Tyr Lys Val lle Pro fle Thr Ash Arg Val Val Asn Glu Lys Gly Ghu Ser Phe Asp dle Leu (4g6) Ser Gys Ala Ala Met Asp Asp Phe Gln Lee lle Pro Met lle Ser lys Gys Arg Thr Lys Glu Gly Arg Arg Lys Ihr Asn Leu Soob

Tyr Gly Leu Ala Val Lys Gly Gin Ser His leu Arg Giy Asp Thr Asp Val Val Thr Vał Val Thr Phe Glu Phe Ser Ser Thr Asp (525) Tyr Gly Phe Ile Ile Lys Gly Arg Ser His Leu Arg Asn Asp Thr Asp Val Val Asn Phe Val Ser Met Glu Phe Ser Leu Thr Asp. (52g)
Pro Arg Val Asp Ser Gly Lys Trp Pro Lys Tym Thr Val Phe Arg lle Gly Ser leu Phe val Ser Giy Arg Glu Lys Ser (55z) Pre Aro Leu Glu Pro His Lys Trp Giu Lys Tyr Gys Val Leu Glu jle Giy Asp Met Leu Leu Arg Ser Ala Ile Gly Gin Met Ser (558)

Val Yyr Leu Tyr Gys Arg Val Asn Gly Thr Asn bys lle gin Met lys Trp Gly Met Giu Ala Arg Arg Cys Len Leu Gln (57g) Arg Pro Met Phe Leu Tyr Val Arg Thr Asn Gly Thr Ser Lys lle lys Het ys Irp Gly Met Giu Met Arg Arg Cys leuleu Gin fol)
Ser Met Gln GIn Met Glu Ala Ile Val Asp, Gln Giu Ser Ser Ile Gin Giy Tyr Asp Met Thr Lys Ala Cys Phe Lys Giy Asp Arg (608) Ser Leu Gln Gln lle Glu Ser Met lle Glu Ala Glu Ser Ser val tys Glu Lys Asp Met inr Lys Glu Phe Phe
 Glu Asn Lys Ser Glu Thr Trp Pro Ile Gly Glu Ser Pro bys Gly Val Glu Asp Gly Ser Ile Gly lys Val Cys Arg Thr Leu (640)
Phe Thr Lys Cys Leu Met His Tyr Val Phe Gly Asn Ala Gin Leu Glu Gly Phe Ser Ala Glu Ser Arg Arg Leu Leu Leu Leu Ile (665) Leu Ala Lys Ser Val Phe Asn Ser Leu Tyr Ala Ser Pro Gln Leu Glu Gly Phe Ser Ala Giu Ser Arg Lys Leu leu Leu Val Val (6gg)

Gin Ala Leu lys Asp Arg lys Gly Pro irp Val Phe Asp Leu Glu Gly Met Tyr Ser Gly lle Giu Glu Gys lle Ser Asn Asn Pro foat GIn Ala Leu Arg Asp Asn Leu Glu Pro Gly Thr Phe Asp Leu Glu Gly Leu Tyr Glu Ala lie Glu Glu Gys Leu lle Asn Asp Pro figol
Trp Val Ile Gin Ser Ala Tyr Trp Phe Asn Glu Trp Leu Gly Phe Glu lys Glu Gly Ser Lys Val Leu Glu Ser lle Aso Glu lle (723) Trp Val Leu leu Asn Ala Ser Irp Phe Asn Ser Phe leu Thr His Ala Leu Arg C-terminus (716)
Met Asp Glu C-terminus (726)
Fig. 2. Predicted polypeptide sequences of the PA genes of B/AA/1/66 and A/NT/60/68 viruses. The sequence is presented in the $\mathrm{NH}_{2}$-terminal $\rightarrow \mathrm{COOH}$-terminal direction. The underlined areas indicate the amino acids that are in common for the two polypeptides. The numbers in () are the lengths of the polypeptides at that point. Gaps were inserted in either sequence as necessary to provide the optimal alignment.
the $\mathrm{B} / \mathrm{AA} / 1 / 66 \mathrm{PA}$ protein has a net charge of -8.5 , while its COOH -terminal half has a net charge of +1.0 . This unequal distribution of charge is not seen in the $\mathrm{A} / \mathrm{NT} / 60 / 68 \mathrm{PA}$ protein. Its net charge is divided between its $\mathrm{NH}_{2}-$ and $\mathrm{COOH}-$ terminal ends as -8.0 and -11.0 , respectively. Thus the overall net drop in negative charge between the influenza A and influenza B PA genes occurs in the COOH-terminal half of the protein.

While the two PA proteins exhibit a $38 \%$ overall level of homology, the amino acids range from a low of $22 \%$ conservation (histidine) to a high of $57 \%$ conservation (tryptophan). The charged amino acids are conserved as follows: histidine, $22 \%$; arginine, $47 \%$; lysine, $37 \%$; aspartic acid, $43 \%$; and glutamic acid, $48 \%$. If the acidic or basic residues are considered as equivalent amino acids then the conservation for the acidic sites is $58 \%$ and for the basic sites, $47 \%$. While these values are higher than the $38 \%$ overall homology, they do not imply rigid conservation of charged sites. Indeed, no amino acid is dramatically conserved between the sequences of these two viruses. The conserved regions are scattered throughout the protein, but the COOH-terminal half of the PA protein is slightly more conserved than the $\mathrm{NH}_{2}$-terminal half ( 46 to $33 \%$ ). Fig. 3 illustrates this homology difference over the length of the protein by a matrix comparison using a segment length of 40 amino acids in which 20 amino acids must match to give a positive result. It is obvious that the COOH -terminal end contains the region of highest homology overall, with a small region of high homology near the $\mathrm{NH}_{2}$-terminal end. Thus, the COOH-terminal half of the polypeptide may be more important to the PA protein's function than the $\mathrm{NH}_{2}$-terminal half. The importance of the difference in net charge between the $\mathrm{B} / \mathrm{AA} / 1 / 66$ and $\mathrm{A} / \mathrm{NT} / 60 / 68$ PA proteins in this region is not clear. Essentially the same asymmetric homology and net charge patterns were obtained with the PA gene of B/Singapore/222/79 (Dr. Debi Nayak, pers. commun.).


Fig. 3. Matrix comparison plot of B/Ann Arbor/1/66 and A/NT/60/68 PA polypeptides. Each symbol represents a site where at least 20 out of 40 amino acids werc conserved. The numbers on the $x$ and $y$ axes represent the amino acid position in from the $\mathrm{NH}_{2}$ terminus of each protein.

The PB1 vRNA segment is 2369 nucleotides long (Fig. 4). The first protein initiation codon in the cRNA starts 22 nucleotides in from the $5^{\prime}$-end, and an open reading frame of 2277 nucleotides extends from the first nucleotide at the $5^{\prime}$-end to the first termination codon beginning at nucleotide 2278. The PB1 polypeptide encoded in this open reading frame is 752 amino acids in length with a molecular weight of $84,332 \mathrm{Da}$. A polyadenylation site composed of six consecutive adenine residues is present at nucleotides 2348 -2353. The predicted mRNA prior to capping and poly (A) addition would be 2353 nucleotides long. The PB1 RNA of $\mathrm{B} / \mathrm{AA} / 1 / 66$ virus is one nucleotide longer than the B/Lee/40 PB1 RNA (Kemdirim et al., 1986). The difference in length is due to an addition of one cytidine residue in the B/AA/1/66 PB1 cRNA sequence at nucleotide 15 prior to the start codon for the PB1 polypeptide (see Fig. 4). The next longest stretch of nucleotides without a termination codon in either of the other reading frames can only code for 55 amino acids. The sequence predicts a protein with a net charge of +18.5 at pH 7.0 , similar to the $\mathrm{B} / \mathrm{Lee} / 40$ PA protein.

In addition to the one nucleotide insertion at position 15, there were 109 nucleotide mismatches resulting in 11 amino acid changes between the B/AA/1/66 and B/Lee/40 PB1 RNA and protein, respectively. Thus, the variation between these two viruses' predicted polypeptides is only $1.5 \%$. Table 1 shows a compilation of the variation between five polypeptides from B/AA/1/66 and B/Lee/40 predicted by their respective nucleotide sequences. B/AA/1/66 sequences have been determined in this laboratory (manuscript in preparation) while sequence data for B/Lee/40 virus were derived from Kemdirim et al. (1986), Briedis and Tobin (1984), Briedis et al. (1982) and Briedis and Lamb (1982). The overall nucleotide variation was similar for all genes compared, but the amino acid variation ranged from a low of $1.5 \%$ (PB1) to a high of $7.8 \%\left(\mathrm{NS}_{1}\right)$. These results are echoed in the percentage of possible non-silent and silent changes observed. The NS RNA was interesting in that the $\mathrm{NS}_{1}$ gene had the highest percentage of non-silent changes, while $\mathrm{NS}_{2}$ had the third lowest percentage of non-silent changes. $\mathrm{NS}_{2}$ protein also had higher cross-type homology than did $\mathrm{NS}_{1}$ protein ( $16.2 \%$ vs $9.7 \%$ ) (Kemdirim et al., 1986) although these values are among the lowest for any of the viral proteins. Only the $\mathrm{M}_{1}$ polypeptide shows as little variation as the PB1 polypeptide between these two viruses. Interestingly, the $\mathrm{M}_{1}$ polypeptide does not show a correspondingly high cross-type homology with $\mathrm{M}_{1}$ polypeptides of influenza $A$ viruses as does the PB1 polypeptide. Its cross-type homology is only $25 \%$ (Kemdirim et al., 1986). This data may reflect a type-specific functional constraint existing for the $\mathrm{M}_{1}$ gene of influenza B viruses as opposed to a cross-type constraint for the PB1 gene.

Because the PB1 gene of B/Lee/40 virus showed an extremely high cross-type homology with the PB1 genes of influenza A virus, approximately $60 \%$ (Kemdirim et al., 1986), we were interested in determining whether this relatedness would hold for polymerase genes in general. The $\mathrm{B} / \mathrm{AA} / 1 / 66 \mathrm{PA}$ and PB 1 polypeptides were compared to the PA and PB1 polypeptides of $\mathrm{A} / \mathrm{PR} / 8 / 34$ and $\mathrm{A} / \mathrm{NT} / 60 / 68$ viruses. The average cross-type homologies were 38 and $60 \%$, respectively. Fig. 2 shows an optimal alignment of the predicted PA protein sequences of B/AA/1/66 and $\mathrm{A} / \mathrm{NT} / 60 / 68$ viruses. Comparison with the $\mathrm{A} / \mathrm{PR} / 8 / 34$ PA gene gave almost

 3/AMm Arberfi/65 PQ1 Protein



 ya)



 Gin Asp lle ile Asp Ser leu Asp lys pro glu Met Thr Phe Phe ser










 Lys














Fig. 4. Nucleotide and predicted polypeptide sequences of the PB1 genes of B/AA/1/66 and B/Lee/40 viruses. The sequence is presented $5^{\prime} \rightarrow 3^{\prime}$ in the $(+)$ messenger strand sense. The underlined areas indicate the position and sequence of the oligodeoxynucleotide primers that were used. The numbers in () are the lengths of the nucleic acids and polypeptides at those points. Only sites of change are shown in the $B /$ Lee/ 40 nucleotide and polypeptide sequences. Missing nucleotides are represented by - .

TABLE 1
VARIATION BETWEEN B/LEE/40 AND B/AA/1/66 VIRUSES.

| Gene | No. nucleotide ${ }^{\text {a }}$ <br> mismatchcs/ <br> total length | No. amino acid <br> mismatchcs/ <br> total length | No. non-silent <br> mismatches/No. <br> possible | No. silent <br> mismatches/No. <br> possible |
| :--- | :--- | :---: | :--- | :--- |
| PB1 | $109 / 2368(4.6)$ | $11 / 752(1.5)$ | $11 / 1922(0.57)$ | $93 / 800(11.6)$ |
| NP | $113 / 1841(6.1)$ | $26 / 560(4.6)$ | $28 / 1407(1.9)$ | $77 / 590(13.0)$ |
| $\mathrm{M}_{1}$ | $65 / 1191(5.5)$ | $4 / 248(1.6)$ | $4 / 624(0.64)$ | $31 / 271(11.4)$ |
| $\mathrm{N}_{1}$ | $61 / 1096(5.6)$ | $22 / 281(7.8)$ | $22 / 706(3.1)$ | $26 / 302(8.6)$ |
| $\mathrm{NS}_{2}$ |  | $3 / 122(2.5)$ | $3 / 315(0.95)$ | $9 / 131(6.9)$ |

${ }^{a}$ Values in this column are based on total RNA length, not on polypeptide encoding regions.
The total number of non-silent and silent positions possible in each coding region was calculated as previously described (Bishop et al., 1982).
identical homology results (data not shown). A similar figure for the PB1 gene is also not included, because we demonstrate that there is very little change between the PB1 proteins of $B /$ Lee $/ 40$ and $B / A A / 1 / 66$ viruses (sce Fig. 4), and a comparison of $\mathrm{B} /$ Lee/ 40 and $\mathrm{A} / \mathrm{WSN} / 33$ virus was previously published (Kemdirim et al., 1986). The cross-type homology data shows that the high level of relatedness seen for the $\mathrm{B} /$ Lee/40 PB1 gene extends to the PB1 gene of B/AA/1/66 virus, but that this high level of relatedness is not applicable to the PA gene, and hence to polymerase genes in general. In fact, this high cross-type homology appears to be unique for the PB1 gene since preliminary data comparing the first 435 amino acids of the $\mathrm{B} / \mathrm{AA} / 1 / 66 \mathrm{~PB} 2$ polypeptide with PB 2 polypeptides of the same influenza $A$ viruses used above, yields a homology of $38-40 \%$ (sequence not shown) similar to the PA proteins.

PB1 has been identified as the most likely polymerase protein to catalyze each successive nucleotide addition to the influenza RNA growing chains (Braam et al., 1983). This function should be essentially the same whether influenza A or B virus is involved, and thus little or no variation due to virus type would be expected. No divergence due to influenza virus type, coupled to a strong functional constraint, may explain why the PB1 protein, alone, has retained such a high sequence homology across influenza type $A$ and $B$ viruses.

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