

NSL 09014

Distribution of D₅ dopamine receptor mRNA in rat brain

James H. Meador-Woodruff^a, Alfred Mansour^a, David K. Grandy^b, Scott P. Damask^a, Olivier Civelli^b and Stanley J. Watson Jr.^a

^aMental Health Research Institute, University of Michigan Medical Center, Ann Arbor, MI 48109-0720 (USA) and ^bVollum Institute for Advanced Biomedical Research, Oregon Health Sciences University, Portland, OR 97201 (USA)

(Received 13 May 1992; Revised version received 9 July 1992; Accepted 9 July 1992)

Key words: Receptor; Dopamine; RNA; Messenger; Catecholamine

The distribution of the messenger RNA encoding the dopamine D₅ receptor was determined in the rat brain by in situ hybridization. Using [³⁵S]-labelled riboprobes to either the rat or human D₅ receptor, this mRNA was localized to the hippocampus and the parafascicular nucleus of the thalamus. This mRNA could not be visualized in the more traditional brain regions associated with dopaminergic cell bodies or projection fields. This unusual distribution suggests a novel function in the brain for this subtype of the dopamine receptor.

The recent cloning of multiple subtypes of the dopamine receptor has revealed at least five distinct receptors, which cluster into D₁-like and D₂-like families [2, 4, 7, 17, 21–25, 28, 29]. The D₂ family consists of D₂ [2], D₃ [7, 21] and D₄ [25] receptors. All three of these receptors have D₂-like pharmacology, with subtle between-receptor differences. In particular, the D₄ receptor has a 10-fold greater affinity for the atypical neuroleptic clozapine compared to the D₂ receptor, but both receptors have comparable affinities for spiperone [25]. The distributions in brain of the mRNAs encoding these receptors are also distinctive. D₂ receptor mRNA is localized in most of the traditional motor and limbic dopaminergic regions of the brain, as well in several of the dopamine-containing cell groups, reflecting autoreceptor synthesis [1, 10, 12–15, 18, 26]. D₃ receptor mRNA has a more limbic distribution, with minimal localization in the deep grey motor nuclei [1]. The D₄ receptor message appears to have a cortical, limbic and hypothalamic distribution [25].

The D₁ family of dopamine receptors at present contains only two members, the D₁ [4, 17, 23, 29] and D₅ [22, 24, 28] (also named D_{1A} and D_{1B}, resp.) subtypes. Both of these receptors have fairly similar D₁-like pharmacology, although the D₅ receptor has a higher affinity for dopa-

mine itself than the D₁ receptor [22, 24]. The mRNA encoding the D₁ receptor is widely distributed in brain, very much in parallel with the distribution of the D₂ receptor in the dopaminergic regions of the central nervous system, although not in the dopamine-containing cell regions [5, 11, 14, 16, 27]. The distribution of D₅ receptor mRNA in brain is less clear.

In the original paper describing the cloning of the human D₅ receptor [22], in situ hybridization and Northern analysis using oligonucleotide probes suggested that the mRNA for this receptor is distributed in a number of motor and limbic regions of the rat brain, indistinguishable from the previously described distribution of the mRNA encoding the D₁ receptor. Subsequently, the rat homolog of the human D₅ receptor was cloned and referred to as D_{1B} [24]. In contrast to the earlier report using oligonucleotides to the human D₅ sequence, the distribution of the D_{1B} receptor mRNA in the rat brain was reported as quite restricted, occurring only in the hippocampus, lateral mammillary nuclei, and a thalamic nucleus identified as the anterior prepectal nucleus. Based on the high degree of homology between these two sequences, it is likely that they are encoding the same functional receptor in the respective species from which they were cloned. Accordingly, it would be expected that the anatomical distributions demonstrated in the rat brain in these earlier two reports should be identical. Given the marked discrepancies in the anatomical distributions reported, the purpose of the present study was to reexamine the distribution of the mRNA encoding the D₅/D_{1B}

Correspondence: J.H. Meador-Woodruff, Mental Health Research Institute, University of Michigan Medical Center, 205 Zina Pitcher Place, Ann Arbor, MI 48109-0720, USA. Fax: (1) (313)-747-4130.

receptor in the rat brain in an attempt to resolve this inconsistency, using *in situ* hybridization with both rat and human probes.

Brains were rapidly removed from male Sprague-Dawley rats (250 g) that had been sacrificed by decapitation, and were frozen in isopentane (-30°C) for 30 s. Frozen tissue samples were cryostat-sectioned ($15\ \mu\text{m}$) and thaw-mounted onto polylysine-subbed microscope slides. Sections were obtained in a coronal plane from the frontal pole (rostral to the caudate-putamen), to just caudal to the substantia nigra. These sections were maintained at -80°C until the time of hybridization.

Riboprobes complementary to human D_5 receptor mRNA were synthesized from a partial D_5 receptor cDNA. A 1,560 bp insert corresponding to the region of the human D_5 receptor spanning transmembrane domains I–VII was synthesized using PCR and spliced into the *SaII* site of pGEM-3Z. This plasmid was linearized with *PstI*, to result in a 800 bp probe corresponding to most of the third cytoplasmic loop and transmembrane domains VI and VII. Riboprobes complementary to rat D_5 receptor mRNA were generated from a partial D_5 receptor cDNA corresponding to transmembrane domains II–VI subcloned into pGEM-3Z. This plasmid was linearized with *SaII* to generate a 650 bp probe. To generate antisense riboprobes, $1\ \mu\text{g}$ of these linearized DNAs were labelled with $250\ \mu\text{Ci}$ of $[^{35}\text{S}]\text{UTP}$ using T7 RNA polymerase. The probes were synthesized and purified as previously described for other dopamine receptor riboprobes [10–14].

For *in situ* hybridization, slides at -80°C were fixed and pre-hybridized as we have previously described [10–14]. Sections were hybridized with the $[^{35}\text{S}]$ -labelled riboprobes overnight at 55°C . Probes were diluted in hybridization buffer to a final concentration of $1\text{--}2 \times 10^6\ \text{dpm}/30\ \mu\text{l}$. The hybridization buffer was 75% (v/v) formamide for the rat probe, 50% formamide for the human probe; 10% (w/v) dextran sulfate; $3 \times \text{SSC}$; 50 mM sodium phosphate, pH 7.4; $1 \times$ Denhardt's solution (0.02% polyvinyl pyrrolidone, 0.02% Ficoll, 0.02% bovine serum albumin); 0.1 mg/ml yeast tRNA; and 10 mM dithiothreitol. Diluted probe was applied to prehybridized tissue sections, covered with a glass cover slip, and sealed with rubber cement. Following hybridization, the cover slips were removed. The sections were first rinsed in $2 \times \text{SSC}$ at room temperature for 5 min, then in RNase A ($200\ \mu\text{g}/\text{ml}$ in 10 mM Tris, 0.5 M NaCl, pH 8.0) for 60 min at 37°C , $2 \times \text{SSC}$ at room temperature for 10 min, $1 \times \text{SSC}$ at room temperature for 10 min, $0.5 \times \text{SSC}$ at 55°C for 1 h, and a final rinse in $0.5 \times \text{SSC}$ at room temperature. These slides were dehydrated in graded ethanols and air-dried. Slides were then apposed to X-ray film, or emulsion-dipped in NTB-2 emulsion.

Using both human and rat probes, only two structures were identified to contain D_5 receptor mRNA in a comprehensive survey through the rat brain, as shown in Fig. 1. The hippocampus showed low levels of D_5 receptor mRNA. In addition, the parafascicular nucleus of the thalamus was visualized. No other structures could be identified to contain D_5 receptor mRNA, including the striatum, cortex, amygdala, olfactory tubercle, nucleus accumbens and septum, or any of the dopamine-containing cell groups such as the substantia nigra, ventral tegmental area, or zona incerta. Control experiments using 'sense'-strand riboprobes or pretreatment with RNase A [10, 14] resulted in no detectable signal in these positively labelled regions.

These results are relatively similar to those reported by Tiberi et al. [24] in the rat D_5 (D_{1B}) cloning report, but are discrepant from the Sunahara et al. [22] cloning study of the human D_5 receptor. In the former paper, only the hippocampus, lateral mammillary nuclei and a thalamic nucleus which was identified as the anterior prepectal nucleus were identified as D_5 positive. Review of the relevant figure in this report indicates that these authors visualized the same thalamic nucleus that is demonstrated in the present report. Our identification of this nucleus as the parafascicular nucleus rather than the anterior prepectal nucleus is based on its histological appearance, especially its envelopment of the fasciculus retroflexus. This distinction is of significance, as these two nuclei have distinct circuitry and function.

In contrast, the Sunahara et al. study reported an anatomical distribution that suggested that this mRNA was distributed more widely, corresponding to the previously reported distribution of D_1 receptor mRNA. The difference between these reports is that the Sunahara et al. study employed short oligonucleotide probes rather than longer riboprobes, which likely hybridized with authentic D_1 receptor mRNA instead of (or in addition to) D_5 receptor mRNA, hence the apparent widespread distribution.

More recently, several pseudogenes of the D_5 receptor have been reported in the human in addition to the gene encoding the functional receptor [8, 19, 28]. It appears that these pseudogenes arose late in evolution, and are not identifiable in the rat [8, 28]. Accordingly, it is unlikely that detection of the distribution of D_5 receptor mRNA in the rat brain is confounded by possible transcription of a pseudogene. In contrast, at least one pseudogene may be transcribed in the human, thus potentially complicating detection of the distribution of D_5 receptor mRNA in the human brain [8, 28].

The restricted distribution of D_5 receptor mRNA in the rat brain suggests that this receptor may have a function distinct from the other identified dopamine recep-

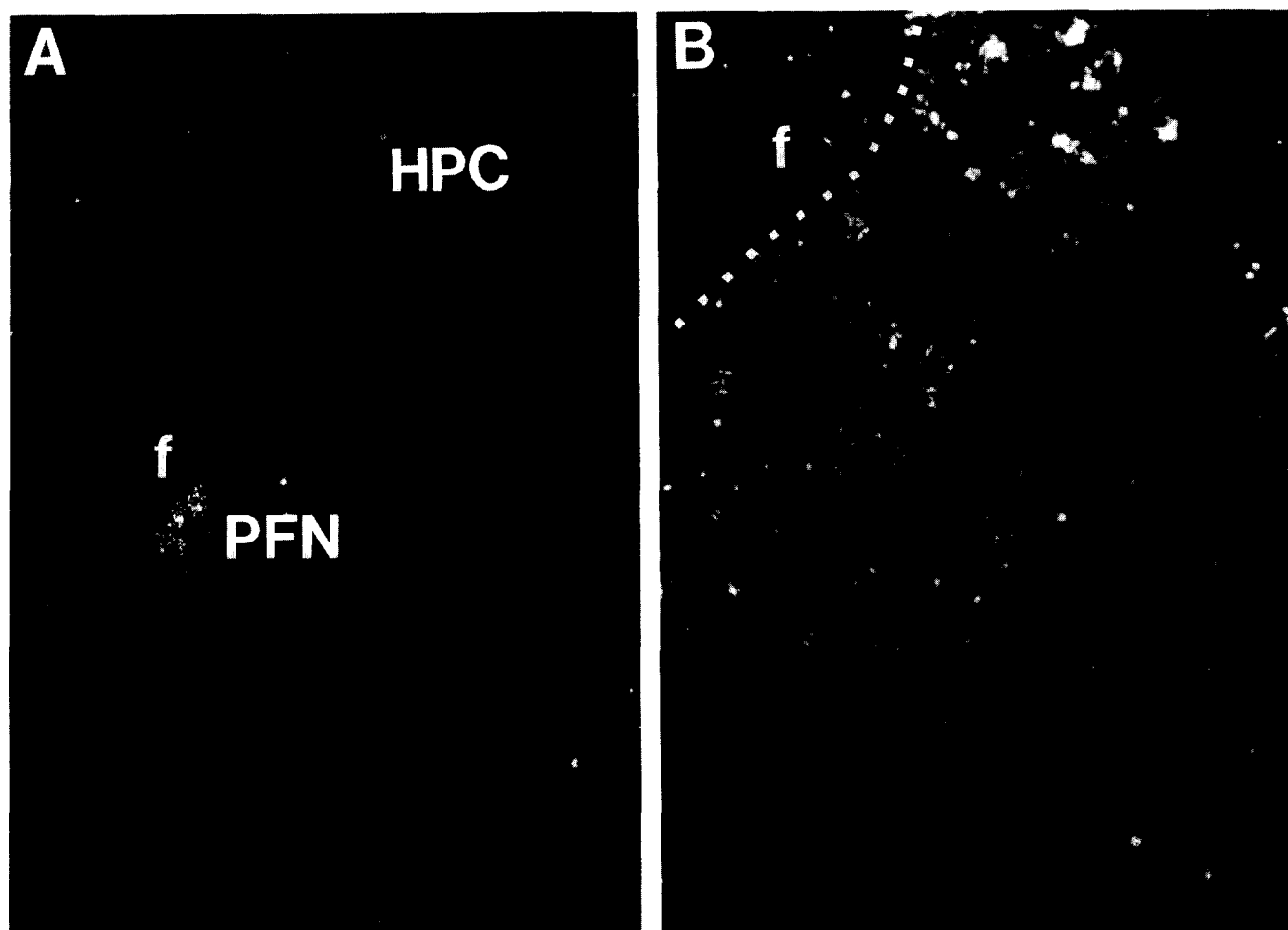


Fig. 1. Distribution of dopamine D₅ receptor mRNA in the rat brain as determined by in situ hybridization. These images were generated with a riboprobe to the rat sequence, although similar results were obtained with a human cRNA probe. A: low power view demonstrating faint labelling in the hippocampus (HPC) and parafascicular nucleus of the thalamus (PFN). The fasciculus retroflexus (f) is identified for orientation. B: high power dark field image demonstrating labelling of the parafascicular nucleus; the fasciculus (f) is dorsomedial to the PFN. Note the relatively higher density of labelling of D₅ receptor mRNA in the dorsolateral aspect of this nucleus.

tors. While all of the dopamine receptors are apparently encoded in regions of the hippocampus, this is the first dopamine receptor that has been identified as being encoded in the parafascicular nucleus. This particular nucleus receives afferents from the substantia nigra [3], making it a likely recipient of the dopamine projections arising in the midbrain. It also sends a projection to the striatum [9], thus suggesting a possible role for this structure in the integration and regulation of multiple levels of dopaminergic neurotransmission. This thalamic nucleus is involved in the perception of pain [6], and appears to be a neuroanatomical locus associated with the antinociceptive actions of opiates [20]. The localization of D₅ receptor mRNA in this nucleus suggests that this receptor subtype may be involved in the neurochemical regulation of the thalamic processing of painful stimuli, which would not be considered a traditional role for dopamine in the central nervous system.

In summary, using both rat and human riboprobes generated to the D₅ dopamine receptor, we have determined the distribution of the mRNA encoding this receptor in the rat brain, and could only identify this mRNA by in situ hybridization in the hippocampus and in the parafascicular nucleus of the thalamus. This restricted distribution suggests that the D₅ receptor may have a function quite dissimilar from other dopamine receptors.

J.H.M.-W. is the recipient of a Research Scientist Development Award from the National Institute of Mental Health (MH00818). This work was also supported by a grant from the National Institute for Mental Health (MH42251), and awards from the National Alliance for the Mentally Ill/Stanley Foundation Research Awards Program, the Scottish Rite Schizophrenia Research Program, and the National Alliance for Research on Schizophrenia and Depression (S.J.W.).

- 1 Bouthenet, M.-L., Souil, E., Martres, M.-P., Sokoloff, P., Giros, B. and Schwartz, J.-C., Localization of dopamine D₃ receptor mRNA in the rat brain using in situ hybridization histochemistry: comparison with dopamine D₂ receptor mRNA. *Brain Res.*, 564 (1991) 203–209.
- 2 Bunzow, J.R., Van Tol, H.H.M., Grandy, D.K., Albert, P., Salon, J., Christie, M., Machida, C.A., Neve, K.A. and Civelli, O., Cloning and expression of a rat D₂ dopamine receptor cDNA. *Nature*, 336 (1988) 783–787.
- 3 Clavier, R.M., Atmadja, S. and Fibiger, H.C., Nigrothalamic projections in the rat as demonstrated by orthograde and retrograde tracing techniques. *Brain Res. Bull.*, 1 (1976) 379–384.
- 4 Dearry, A., Grinrich, J.A., Falardeau, P., Fremeau, R.T., Bates, M.D. and Caron, M.G., Molecular cloning and expression of the gene for a human D₁ dopamine receptor. *Nature*, 347 (1990) 72–76.
- 5 Fremeau, R.T., Duncan, G.E., Fornaretto, M.-G., Dearry, A., Gingrich, J.A., Breese, G.R. and Caron, M.G., Localization of D₁ dopamine receptor mRNA in brain supports a role in cognitive, affective, and neuroendocrine aspects of dopaminergic neurotransmission. *Proc. Natl. Acad. Sci. USA*, 88 (1991) 3772–3776.
- 6 Giesler, G.J., Menetrey, D. and Basbaum, A.I., Differential origins of spinothalamic tract projections to medial and lateral thalamus in the rat. *J. Comp. Neurol.*, 184 (1979) 107–126.
- 7 Giros, B., Martres, M.-P., Sokoloff, P. and Schwartz, J.-C., Clonage du gène du récepteur dopaminergique D₃ humain et identification de son chromosome. *C. R. Acad. Sci., Paris, Serie III*, 311 (1990) 501–508.
- 8 Grandy, D.K., Zhang, Y., Bouvier, C., Zhou, Q.-Y., Johnson, R.A., Allen, L., Buck, K., Bunzow, J.R., Salon, J. and Civelli, O., Multiple human D₃ dopamine receptor genes: a functional receptor and two pseudogenes. *Proc. Natl. Acad. Sci. USA*, 88 (1991) 9175–9179.
- 9 Jones, E.G. and Leavitt, R.Y., Retrograde axonal transport and the demonstration of non-specific projections to the cerebral cortex and striatum from thalamic intralaminar nuclei in the rat, cat and monkey. *J. Comp. Neurol.*, 154 (1974) 349–378.
- 10 Mansour, A., Meador-Woodruff, J.H., Bunzow, J.R., Civelli, O., Akil, H. and Watson, S.J., Localization of dopamine D₂ receptor mRNA and D₁ and D₂ receptor binding in the rat brain and pituitary: an in situ hybridization-receptor autoradiographic analysis. *J. Neurosci.*, 10 (1990) 2587–2600.
- 11 Mansour, A., Meador-Woodruff, J.H., Zhou, Q.-Y., Civelli, O., Akil, H. and Watson, S.J., A comparison of D₁ receptor binding and mRNA in rat brain using receptor autoradiographic and in situ hybridization techniques. *Neuroscience*, 45 (1991) 359–371.
- 12 Meador-Woodruff, J.H. and Mansour, A., Expression of the dopamine D₂ receptor gene in brain. *Biol. Psychiatry*, 30 (1991) 985–1007.
- 13 Meador-Woodruff, J.H., Mansour, A., Bunzow, J.R., Van Tol, H.H.M., Watson, S.J. and Civelli, O., Distribution of D₂ dopamine receptor mRNA in rat brain. *Proc. Natl. Acad. Sci. USA*, 86 (1989) 7625–7628.
- 14 Meador-Woodruff, J.H., Mansour, A., Healy, D.J., Kuehn, R., Zhou, Q.-Y., Bunzow, J.R., Akil, H., Civelli, O. and Watson, S.J., Comparison of the distributions of D₁ and D₂ dopamine receptor mRNAs in rat brain. *Neuropsychopharmacology*, 5 (1991) 231–242.
- 15 Mengod, G., Martinez-Mir, M.I., Vilaro, M.T. and Palacios, J.M., Localization of the mRNA for the dopamine D₂ receptor in the rat brain by in situ hybridization histochemistry. *Proc. Natl. Acad. Sci. USA*, 86 (1989) 8560–8564.
- 16 Mengod, G., Vilaro, M.T., Niznik, H.B., Sunahara, R.K., Seeman, P., O'Dowd, B.F. and Palacios, J.M., Visualization of a dopamine D₁ receptor mRNA in human and rat brain. *Mol. Brain Res.*, 10 (1991) 185–191.
- 17 Monsma, F.J., Mahan, L.C., McVittie, L.D., Gerfen, C.R. and Sibley, D.R., Molecular cloning and expression of a D₁ dopamine receptor linked to adenylyl cyclase activation. *Proc. Natl. Acad. Sci. USA*, 87 (1990) 6723–6727.
- 18 Najlerahim, A., Barton, A.J.L., Harrison, P.J., Heffernan, J. and Pearson, R.C.A., Messenger RNA encoding the D₂ dopaminergic receptor detected by in situ hybridization histochemistry in rat brain. *FEBS Lett.*, 255 (1989) 335–339.
- 19 Nguyen, T., Sunahara, R., Marchese, A., Van Tol, H.H.M., Seeman, P. and O'Dowd, B., Transcription of a human dopamine D₃ pseudogene. *Biochem. Biophys. Res. Commun.*, 181 (1991) 16–21.
- 20 Reyes-Vazquez, C. and Dafny, N., Microiontophoretically applied morphine and naloxone on single cell activity in the parafasciculus nucleus of naive and morphine-dependent rats. *J. Pharmacol. Exp. Ther.*, 229 (1984) 583–588.
- 21 Sokoloff, P., Giros, B., Martres, M.-P., Bouthenet, M.-L. and Schwartz, J.-C., Molecular cloning and characterization of a novel dopamine receptor (D₃) as a target for neuroleptics. *Nature*, 347 (1990) 146–151.
- 22 Sunahara, R.K., Guan, H.-C., O'Dowd, B.F., Seeman, P., Laurier, L.G., Ng, G., George, S.R., Torchia, J., Van Tol, H.H.M. and Niznik, H.B., Cloning of the gene for a human dopamine D₃ receptor with higher affinity for dopamine than D₁. *Nature*, 350 (1991) 614–619.
- 23 Sunahara, R.K., Niznik, H.B., Weiner, D.M., Stormann, T.M., Brann, M.R., Kennedy, J.L., Gelernter, J.E., Rozmahel, R., Yang, Y., Israel, Y., Seeman, P. and O'Dowd, B.F., Human dopamine D₁ receptor encoded by an intronless gene on chromosome 5. *Nature*, 347 (1990) 80–83.
- 24 Tiberi, M., Jarvie, K.R., Silvia, C., Falardeau, P., Gingrich, J.A., Godinot, N., Bertrand, L., Yang-Feng, T.L., Fremeau, R.T. and Caron, M.G., Cloning, molecular characterization, and chromosomal assignment of a gene encoding a second D₁ dopamine receptor subtype: differential expression pattern in rat brain compared with the D_{1A} receptor. *Proc. Natl. Acad. Sci. USA*, 88 (1991) 7491–7495.
- 25 Van Tol, H.H.M., Bunzow, J.R., Guan, H.-C., Sunahara, R.K., Seeman, P., Niznik, H.B. and Civelli, O., Cloning of the gene for a human dopamine D₄ receptor with high affinity for the antipsychotic clozapine. *Nature*, 350 (1991) 610–614.
- 26 Weiner, D.M. and Brann, M.R., The distribution of a dopamine D₂ receptor mRNA in rat brain. *FEBS Lett.*, 253 (1989) 207–213.
- 27 Weiner, D.M., Levey, A.I., Sunahara, R.K., Niznik, H.B., O'Dowd, B.F., Seeman, P. and Brann, M.R., D₁ and D₂ dopamine receptor mRNA in rat brain. *Proc. Natl. Acad. Sci. USA*, 88 (1991) 1859–1863.
- 28 Weinshank, R.L., Adham, N., Macchi, M., Olsen, M.A., Branchek, T.A. and Hartig, P.R., Molecular cloning and characterization of a high affinity dopamine receptor (D_{1B}) and its pseudogene. *J. Biol. Chem.*, 266 (1991) 22427–22435.
- 29 Zhou, Q.-Y., Grandy, D.K., Thambi, L., Kushner, J.A., Van Tol, H.H.M., Cone, R., Pribnow, D., Salon, J., Bunzow, J.R. and Civelli, O., Cloning and expression of human and rat D₁ dopamine receptors. *Nature*, 347 (1990) 76–80.