

# Supernumerary chromosome variation and heterochromatin distribution in the endemic New Zealand frog *Leiopelma hochstetteri*

David M. Green<sup>1</sup>, James Kezer<sup>2</sup>, and Ronald A. Nussbaum<sup>3</sup>

<sup>1</sup> Redpath Museum, McGill University, Montreal, PQ, H3A 2K6 Canada

<sup>2</sup> Department of Biology, University of Oregon, Eugene, OR 97403 USA

<sup>3</sup> Museum of Zoology, University of Michigan, Ann Arbor, MI 48109 USA

**Abstract.** Specimens of the endemic New Zealand frog *Leiopelma hochstetteri* from Tapu on North Island were found to have six, nine or ten supernumerary chromosomes in their karyotypes. In comparison with previously published data, these results further indicate probable geographic variation in supernumerary chromosome number between populations. Increased numbers of supernumeraries in these frogs is correlated with apparent decrease of centromeric heterochromatin in the five large metacentric chromosomes of the karyotype, as detected by C-banding. Meiosis was abnormal in a male with a high number of supernumeraries. In lampbrush preparations from a single female with one supernumerary univalent, the supernumerary often had a denser, beaded appearance in comparison with the regular bivalents. Evidence is consistent with the notion that these supernumerary chromosomes may have arisen from centromeric fragments.

## Introduction

The three species comprising the endemic New Zealand genus *Leiopelma* are often considered to be among the most primitive of living frogs (Bell 1982; Duellman and Trueb 1986). The most widespread of these species, *L. hochstetteri*, is found in damp seepages and stream banks in scattered localities on the North Island of New Zealand (Fig. 1). *L. hamiltoni* and *L. archeyi* are extremely restricted in range. The three species are considered to be threatened in New Zealand (Bell 1986), especially *L. hamiltoni*. While *L. hamiltoni* and *L. archeyi* have both been shown to have  $2n = 18$  chromosomes, *L. hochstetteri* has  $2n = 22$  chromosomes plus varying numbers of very small supernumerary, or B-chromosomes (Morescalchi 1967; Stephenson et al. 1972, 1974; Green et al. 1984a). The regular karyotype, or A-set of chromosomes, of *L. hochstetteri* consists of five pairs of large metacentric chromosomes and six pairs of smaller telocentric chromosomes, one of which has a prominent secondary constriction.

Supernumerary chromosomes, always considered to be unusual in animals, are rare in anurans; only four species other than *L. hochstetteri* are reported in the literature to possess them (Jones and Rees 1982) although there exist additional, unpublished examples (J.P. Bogart, personal communication). The origin of supernumerary chromosomes in animal karyotypes has not been convincingly explained (Jones and Rees 1982). Considering the highly het-

erochromatic nature of supernumerary chromosomes in many species, it is possible that they may have arisen, in some cases, as free centromeres or centromeric fragments (see Jones and Rees 1982), but there is little corroborating evidence. Supernumerary chromosomes have been associated, in various instances, with particular kinds of meiotic chromosomal behaviour, decreased developmental rate, reduced fitness or viability and/or increased C-values (Harvey and Hewitt 1979; Jones and Rees 1982).

Earlier, we reported on the supernumerary chromosomes and heterochromatin of specimens of *L. hochstetteri* obtained from Dome Valley, near Warkworth north of Auckland, New Zealand (Fig. 1), including comments on a triploid female (Green et al. 1984a). The number of supernumerary chromosomes per individual in that sample was low. All females except the triploid had one supernumerary while the males had none. The triploid had two supernumeraries. The supernumeraries were not unusually heterochromatic; C-bands were confined to the centromeres, as is usual among the regular set of chromosomes. All other chromosomes were similarly C-band positive at the centromeres with an additional C-band observed in association with the prominent secondary constriction on chromosome 7. Stephenson et al. (1972) presented karyotype data from frogs taken from Tokatea Ridge on the Coromandel Peninsula of the North Island of New Zealand (Fig. 1). These specimens, all males, had either two or four supernumeraries. Another male from Dome Valley was without supernumeraries. Two females from Dome Valley were examined by Morescalchi (1967). They had 1 and 12 supernumeraries, respectively.

We are now able to present karyotypic data, including C-banding information on heterochromatin distribution, from additional specimens of *L. hochstetteri* from a third locality, Tapu, on the Coromandel Range (Fig. 1). In addition, we present further information from two more specimens of this rare frog from Dome Valley, including data from lampbrush chromosomes obtained from the oöcytes of a Dome Valley female. This information gives additional insight into variation in heterochromatin distribution and supernumerary chromosome numbers within the karyotype of this species.

## Materials and methods

Four frogs collected on the west side of the Tapu-Coroglen Saddle of the Coromandel Range on the North Island of New Zealand by Dr. Joan Robb in September, 1983 were



**Fig. 1.** Map of North Island of New Zealand showing the range of *Leiolopma hochstetteri* (cross-hatching) and localities mentioned in the text where chromosome information has been obtained. 1 Dome Valley, Warkworth; 2 Tapu; 3 Tokatca Ridge

examined. Chromosomes from one male and one female, deposited at the California Academy of Sciences (CAS Nos. 156252-3), were obtained using Bogart's (1981) corneal epithelium squash method while chromosomes from two males, deposited at the Museum of Zoology, University of Michigan (UMMZ Nos. 180360-1) were obtained with Kezer and Sessions' (1979) gut epithelium and testis squash methods. Lampbrush chromosome preparations, using methods described by Kezer et al. (1960), were made from the oocytes of two females from Dome Valley, Warkworth,

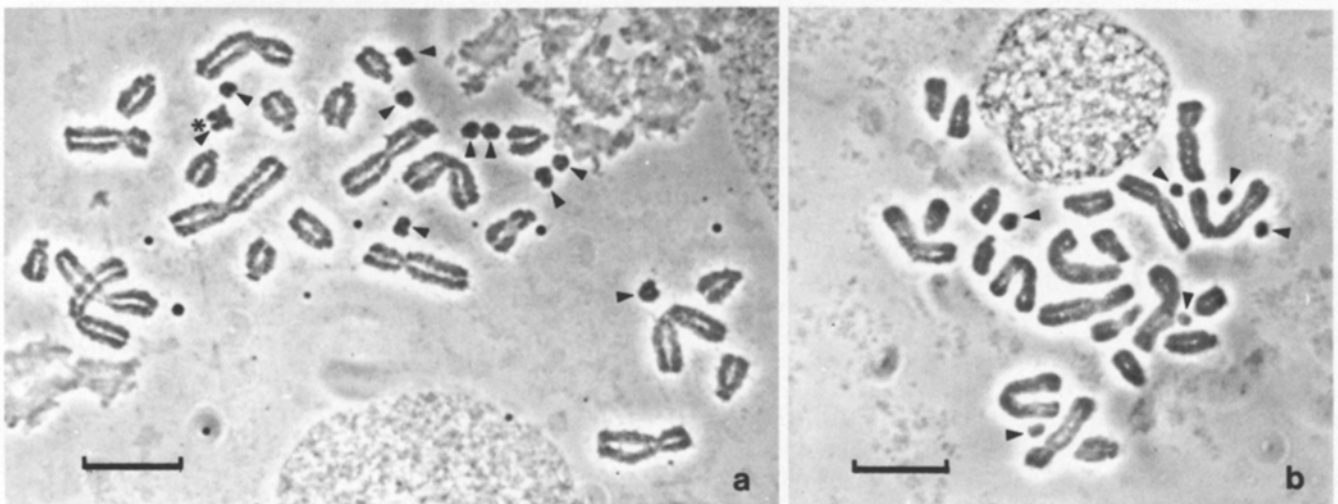
New Zealand collected in March, 1976, by Joan Robb (UMMZ Nos. 146848 and 146850).

At least 20 metaphase chromosome spreads per specimen were examined under phase contrast optics. Photomicrographs of good spreads were numerically analyzed with the CHROMPAC III computer-assisted analysis system (Green et al. 1984b). The nomenclature used for chromosomes is that suggested by Levan et al. (1964) and modified by Green et al. (1980). Schmid's (1978) C-banding method was used to demonstrate the presence of the heterochromatic regions.

## Results

When examined unstained with phase contrast (Fig. 2a, b), the 11 pairs of chromosomes of the A-set of the Tapu specimens did not differ in any marked way from results obtained in previous studies of the karyotype of *L. hochstetteri*. Neither were significant differences found upon measurement and numerical analysis of the A-chromosomes when compared with published data (Green et al. 1984a). All Tapu specimens had five pairs of large metacentric chromosomes and six pairs of smaller telocentric chromosomes, one of which, previously identified as the seventh chromosome (Green et al. 1984a), had a conspicuous median secondary constriction. Some of the telocentric chromosomes possessed very small, but definable, short arms. However, the Tapu specimens had different numbers of supernumerary chromosomes present than had previously been observed in *L. hochstetteri* (Table 1). One male and one female each had six supernumerary chromosomes, one male had nine supernumerary chromosomes and one male had ten supernumerary chromosomes. All supernumerary chromosomes observed were strictly telocentric with one exception: one of the supernumerary chromosomes of the male with ten was metacentric (Fig. 2a).

The supernumerary chromosomes seen in *L. hochstetteri* from Tapu were not all equal in size. If the combined length of the A-chromosomes is set to equal 100%, the largest supernumeraries observed were about 1.4% of the length of the A-set while the smallest supernumeraries were about



**Fig. 2a, b.** Unstained mitotic chromosome spreads obtained from corneal epithelium of *Leiolopma hochstetteri* from Tapu, New Zealand, viewed with phase contrast. Supernumerary chromosomes (arrows) are clearly visible. **a** Male with ten supernumerary chromosomes, one of which is metacentric (asterisk); **b** female with six supernumeraries. Note that supernumerary chromosomes can be of different sizes. Bars represent 10  $\mu$ m

0.87% of the A-set. The supernumerary chromosomes seen in females from Dome Valley examined by Green et al. (1984a) were measured at about 0.98% of the A-set. The metacentric supernumerary observed in one male from Tapu was among the largest of the supernumeraries observed; each of its two arms was smaller (about 0.72% of the A-set) than the smallest telocentric supernumerary.

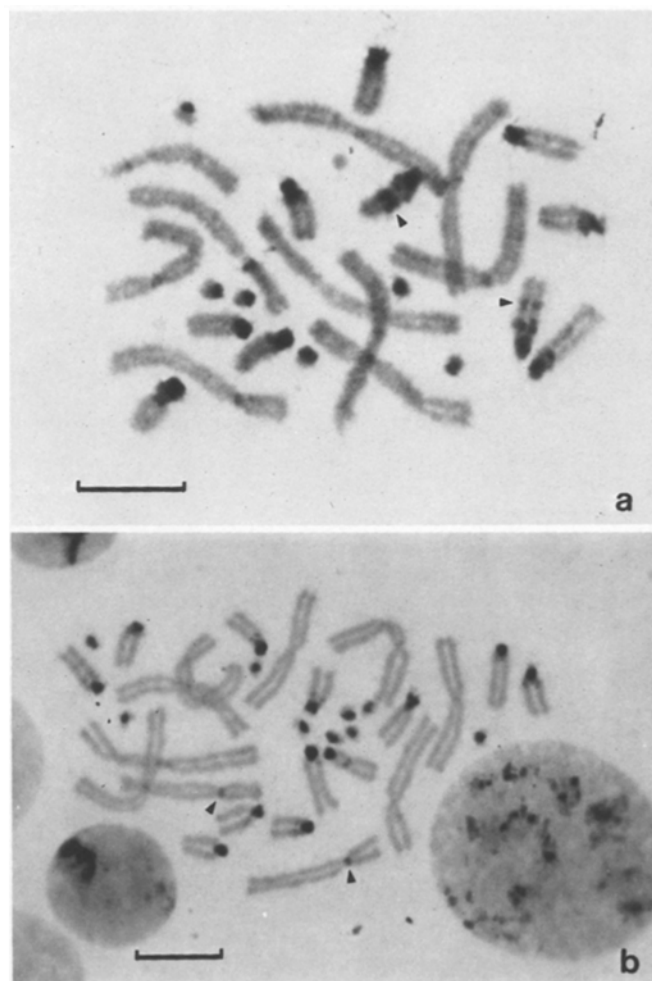
The possibility of mitotic instability in inheritance of one or more of the B-chromosomes was investigated by counting the numbers of supernumeraries in many different chromosome spreads of the same individual. In one male, 23 cells were examined. Of these, equal numbers of cells were observed to have 5 supernumeraries visible as were observed to have 6 supernumeraries (6 being the maximum number in this individual). Five of the 23 cells were observed to have fewer than 5 supernumeraries visible. In the male with 10 supernumeraries, many cells were observed where only 9 such chromosomes could be counted. However, the modal number of supernumeraries per cell was the same as the maximum number of supernumeraries per cell in every individual, indicating that the observed variation in number was probably artifactual.

C-banding revealed the presence of strongly staining heterochromatic regions at the centromeres of all the telocentric chromosomes and the supernumeraries of the Tapu specimens. However, the centromeres of the large metacentric chromosomes of these individuals stained only weakly, or not at all (Fig. 3a, b). By contrast, results obtained from C-banding of chromosomes from Dome Valley frogs (Green et al. 1984a) showed that all their chromosomes, including the metacentrics, stained darkly at the centromeres. Only one of the metacentric chromosomes of the Tapu frogs, chromosome 3, revealed dense C-band heterochromatin at its centromere. This band stained much less strongly than did bands on the telocentrics and supernumeraries.

C-bands in addition to those associated with the centromeres of the chromosomes were noted in the Tapu frogs. The seventh chromosome had a prominent C-band associated with its secondary constriction (Fig. 3a, b), as was observed in Dome Valley frogs (Green et al. 1984a). In the male with six supernumeraries, however, this band appeared to be heteromorphic in that one of the pair of seventh chromosomes had a much more intensely stained band than the other (Fig. 3a). This frog also featured numerous weak C-bands on the "shoulders" of the long arms of the telocentric chromosomes, proximal to the centromeres. The male with nine supernumeraries did not have the heteromorphism in C-banding on chromosome 7.

C-banding of the supernumerary chromosomes of the Tapu frogs was as observed before in Dome Valley specimens (Green et al. 1984a). Only the centromeres were heterochromatic (Fig. 3a, b). The one small metacentric supernumerary observed in one of the males likewise stained only at its centromere.

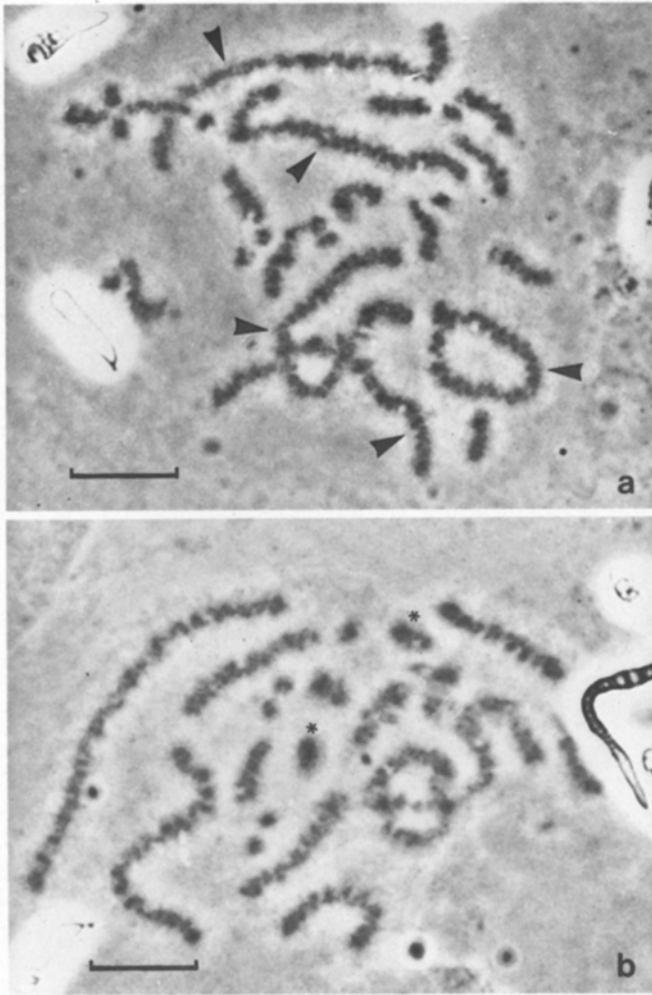
Meiotic chromosome preparations were obtained from the spermatocytes of one of the Tapu males. Meiosis in this frog, which had nine supernumeraries and had been inoculated with colchicine, was highly abnormal (Fig. 4a, b). In some spreads, only the metacentric chromosomes appeared to be paired. Five large bivalents were observed among numerous small univalent elements (Fig. 4a). One of the metacentric pairs formed a conspicuous ring bivalent, indicating the presence of two terminal, or near-terminal,



**Fig. 3a, b.** C-banded metaphase chromosome preparations from gut epithelium of *Leiopelma hochstetteri* from Tapu, New Zealand, illustrate differential intensity of staining of centromeric heterochromatin between metacentric and telocentric chromosomes. **a** Male with six supernumerary chromosomes. Interstitial C-bands associated with the secondary constriction on chromosome 7 (arrows) are heteromorphic. **b** Male with nine supernumeraries. Only one metacentric chromosome pair, 3, has appreciable centromeric heterochromatin (arrows). Bars represent 10  $\mu$ m

chiasmata. All other bivalents had a single, terminal chiasma. A ring bivalent was not observed in meiotic preparations of *L. hochstetteri* as illustrated by Stephenson et al. (1972). Similar ring bivalents, though, are frequently observed in meiotic preparations from more "advanced" frogs such as *Bufo* or *Rana* (Morescalchi 1973). In other meiotic spreads from the same frog, however, differing numbers of telocentric A-chromosomes appeared to be paired. A C-banded preparation (Fig. 5) shows all of the metacentrics to be paired while some of the telocentrics are paired, but not others. In another figure, all of the A-set chromosomes were paired. The supernumerary chromosomes always appeared as univalents.

Lampbrush chromosomes were prepared from oocytes of two females from Dome Valley. Since these individuals had not been treated with colchicine, we had not been able to secure mitotic chromosomes from them. As in other females from this locality that we had examined earlier (Green et al. 1984a), each of these two frogs had one super-

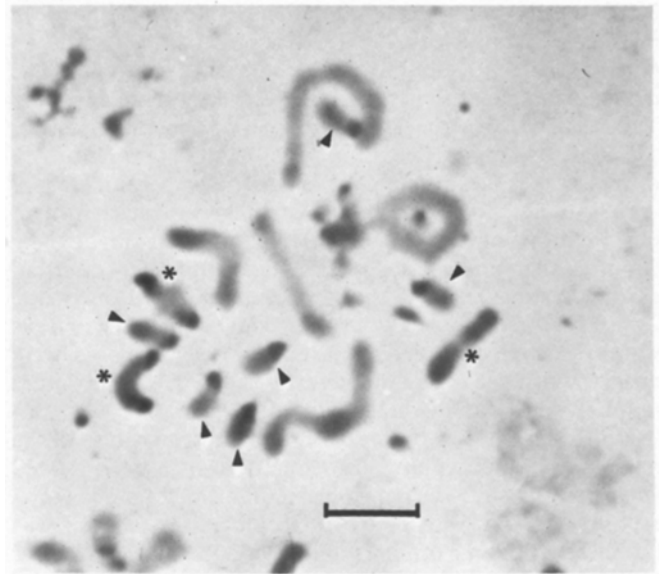


**Fig. 4a, b.** Meiotic chromosomes at prometaphase from the colchicine-treated testis of a male *Leiopelma hochstetteri* with nine supernumeraries. **a** Only the five pairs of metacentric chromosomes (arrows) appear as bivalents, one as a ring bivalent. The six pairs of telocentric chromosomes are unpaired, as are the small supernumeraries. **b** All chromosomes are paired except the supernumeraries and one pair of telocentrics (asterisks). Bars represent 10  $\mu\text{m}$

numerary chromosome per cell, clearly discernible in the lampbrush preparations (Fig. 6a, b). While the A-set lampbrush chromosomes were paired as normal meiotic bivalents, displaying chiasmata and lateral loops, the supernumerary chromosome appeared as a more condensed univalent, with a dense, beaded axis and relatively few lateral loops (Fig. 6a, b).

### Discussion

As of this study, 9 different cytotypes are known from 18 specimens of *L. hochstetteri*. Diploid karyotypes with 0, 1, 2, 4, 6, 9, 10 and 12 supernumeraries have been found. Despite the small samples studied, there is an indication that the average number of supernumerary chromosomes per individual differs between populations (Table 1). Average supernumerary number appears to increase with increasing latitude from Dome Valley to populations at Tokatea and Tapu along the Coromandel Peninsula (Fig. 1). One observation is at odds with this pattern: a 12 supernumer-



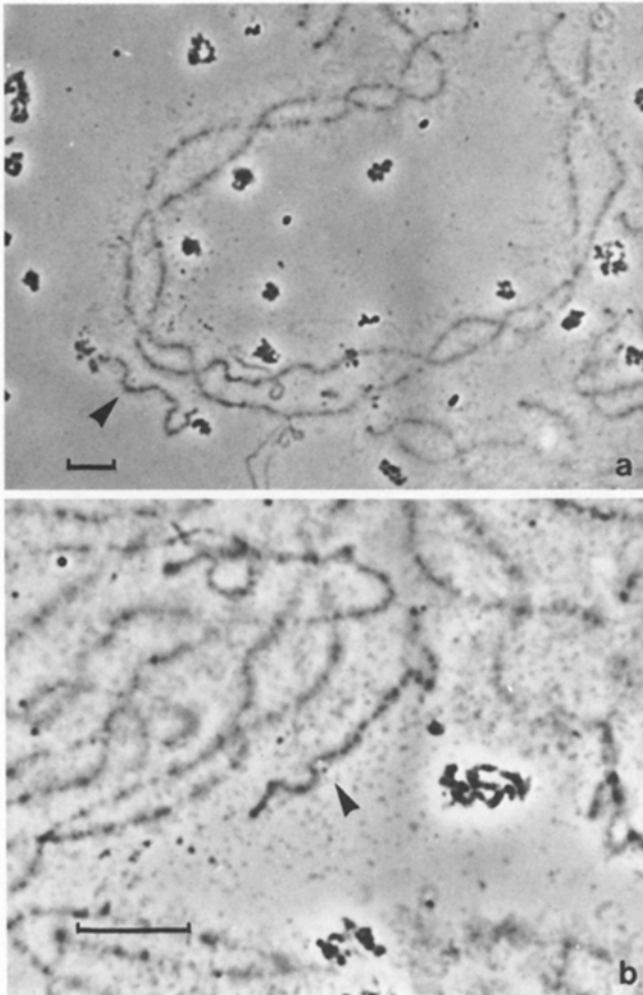
**Fig. 5.** C-banded meiotic preparation from the testis of the same specimen as in Figure 4 showing paired (asterisks) and unpaired (arrows) telocentric chromosomes. Bar represents 10  $\mu\text{m}$

ary chromosomal karyotype found by Morescalchi (1967) in a frog reported to be from Dome Valley. Further investigations will reveal if a geographic trend is real and whether it may continue to outlying parts of the species' range.

While it has been suggested that supernumerary chromosomes are usually transcriptionally inactive (Jones and Rees 1982), visual evidence from lampbrush chromosomes has been lacking. Our results indicate that the supernumerary chromosome in the lampbrush state (Fig. 6a, b) does have short lateral loops indicative of transcriptional activity, though at a lower level than the A-chromosomes.

The variation in the staining intensity of centromeric heterochromatin in the large metacentric chromosomes of *L. hochstetteri* is unusual. Variation in C-band staining has been seen in association with heteromorphic nucleolar organizer regions (NORs) in various frogs (King 1980; Schmid 1983). In one of the specimens of *L. hochstetteri* from Tapu, a C-band heteromorphism can be seen in the NOR-bearing chromosome 7 (Fig. 3a). However, the striking variation observed in *L. hochstetteri* involves only the metacentric chromosomes in apparent correlation with supernumerary chromosome number and/or with geographic distribution. The Dome Valley frogs had strongly C-band positive centromeric regions on all chromosomes while the Tapu frogs had little or no C-banding at the centromeres of the metacentrics.

Variation in heterochromatin banding has been extensively studied in many rodents. Subspecies of the house mouse, *Mus musculus*, have differing amounts of heterochromatin in certain chromosomes (Dev et al. 1976). A similar sort of variation occurs in subspecies of the rat, *Rattus rattus* (Yosida and Sagai 1975). Patton (1977) found evidence of three types of supernumerary chromosomes in the pocket mouse, *Perognathus baileyi*, and slight associated variation in C-banding in the chromosomes of the A-set. Patton, however, did not see any correlation between heterochromatin variation and supernumerary number. The supernumeraries of *P. baileyi* include both totally heterochromatic and, rarely, partially heterochromatic types.



**Fig. 6a, b.** Lampbrush chromosomes from oocytes of a female *Leiopelma hochstetteri* with one supernumerary chromosome (arrows). **a** Size and configuration of lampbrush supernumerary relative to a metacentric A-chromosome bivalent in lampbrush condition. **b** A lampbrush supernumerary showing its relatively dense, beaded condition with few loops. Bars represent 10  $\mu$ m

Among other amphibians, differences in amount of centromeric heterochromatin have been noted in chromosomes of other species, particularly subspecies of the red-legged frog, *Rana aurora* (Green 1985), and of the crested newt, *Triturus cristatus* (Sessions 1984). In these species, all chromosomes were affected and no supernumerary chromosomes were present although the additional heterochromatin differs markedly in its precise location in *R. aurora* and *T. cristatus*.

An example perhaps more like that seen in *L. hochstetteri* with regard to heterochromatin and supernumerary chromosomes was studied in the Australian rodent *Uromys caudimaculatus* (Baverstock et al. 1976). Animals from two disjunct parts of this species' range had great differences in heterochromatin and supernumeraries. Northern animals had many C-band positive regions in the A-set, mostly as telomeric or interstitial bands, and had no supernumeraries. Southern animals had few heterochromatic regions in the A-set yet had six to nine totally heterochromatic supernumerary chromosomes. Unlike the chromosomes of *L. hochstetteri*, however, the centromeres of the A-chromosomes

**Table 1.** Numbers of supernumerary chromosomes known to be found in karyotypes of *Leiopelma hochstetteri* with the numbers of individuals, separated by sex, that have been found to possess those cytotypes

Number of supernumerary chromosomes per karyotype	Number of individuals and population	
	Males	Females
0	2, Dome Valley <sup>b,c</sup>	
1		7, Dome Valley <sup>a-d</sup>
2	2, Tokatea <sup>b</sup>	1, Dome Valley <sup>c</sup> (triploid)
4	1, Tokatea <sup>b</sup>	
6	1, Tapu <sup>d</sup>	1, Tapu <sup>d</sup>
9	1, Tapu <sup>d</sup>	
10	1, Tapu <sup>d</sup>	
12		1, Dome Valley <sup>a</sup>

Data computed from all known sources

<sup>a</sup> Morescalchi (1967)

<sup>b</sup> Stephenson et al. (1972)

<sup>c</sup> Green et al. (1985)

<sup>d</sup> Present study

of *U. caudimaculatus* were relatively unaffected, and the supernumeraries had no euchromatic regions.

The pattern of heterochromatin variation in *L. hochstetteri* is interesting in that only centromeric heterochromatin seems to be involved and only certain morphologically distinct chromosomes seem to be affected. The presence of increasing numbers of supernumerary chromosomes appears to relate to meiotic behaviour of the A-set as well. It has been argued that in certain plants, for instance, supernumerary chromosomes can affect pairing of the A-set in meiosis. Parker et al. (1981) suggested that supernumerary chromosomes affect chiasma frequency in the A-set in *Hypochaeris maculata* (Family Compositae) and produce other effects which disrupt normal meiosis when the number of supernumeraries exceeds one. Patton (1977) also found effects on meiotic chiasma formation among the A-chromosomes in the presence of supernumeraries in the pocket mouse *P. baileyi*. Limited evidence from the present investigations of the karyology of *L. hochstetteri* seems to indicate that supernumeraries may influence synapsis of bivalents in meiosis in *L. hochstetteri*. A single male without supernumeraries from Dome Valley from whom meiotic chromosome preparations were made by Stephenson et al. (1972) showed no deviations from normally expected pairing of all normal chromosome homologs. However, our results from a male frog from Tapu with nine supernumeraries showed a case in which different elements exhibited varying degrees of asynapsis or chiasma failure. One pair of large metacentrics also formed a circular bivalent with terminal chiasmata. The individual we examined may have been aberrant but the colchicine used is unlikely to have been responsible for the effects upon meiosis that we observed.

The origins of supernumerary chromosomes are as yet without clear explanation. Two phenomena must be considered in this context: the origination of supernumeraries and the accumulation of pre-existing supernumeraries. Certainly supernumeraries have accumulated in certain, but not all, populations of *L. hochstetteri*. Among Dome Valley specimens, supernumeraries were found only in females (Table 1). Meiotic accumulation mechanisms have been pro-

posed, pertaining in particular to certain plants (Jones and Rees 1982). Progressive origination of supernumeraries cannot be ruled out in *L. hochstetteri*, however. The different sizes of supernumeraries indicate that they are not all identical and thus probably do not represent proliferation of a single chromosome. The metacentric supernumerary chromosome found in one male has no counterpart in other individuals examined. It may have arisen separately, or be the product of fusion of two very small telocentric supernumeraries, or be an iso-chromosome.

It is possible that at least some of the supernumeraries in the Tapu frogs may have arisen as centromeric fragments, especially as fragments from the large metacentrics. This sort of mechanism has been discussed by Patton (1977) and Jones and Rees (1982). The loss of centromeric fragments preferentially from the large metacentrics, with their accompanying heterochromatin, is an explanation consistent for both the possible origination of the supernumerary chromosomes of *L. hochstetteri* and the observed patterns of heterochromatin distribution seen in this species. However, the apparent increase in functional kinetochores represented by the supernumerary chromosomes awaits explanation.

We have made a number of assertions concerning the distribution, effect and origin of the supernumerary chromosomes of *L. hochstetteri* based on the information at hand. Obviously, however, the volume of that information is limited by the relatively small number of specimens available of this rare frog. Our hypotheses concerning apparent geographical differentiation in numbers of supernumerary chromosomes present per individual could be proven false if additional animals with other numbers of supernumeraries are found: higher numbers from Dome Valley and lower numbers from Tapu, for example. This would also provide information on the possible occurrence of karyotypes with very high numbers of supernumerary chromosomes, such as described by Morescalchi (1967). If these other karyotypes are present, the apparent correlation of supernumeraries with heterochromatin distribution could be tested. Is the observed variation related to geographic divergence or to gain of supernumeraries? At present, these cannot be separated. Other populations of *L. hochstetteri*, on Great Barrier Island or in the East Cape region, for example, have not been examined karyotypically. The lampbrush behavior of the supernumerary chromosomes where more than one is present is also unknown. We hope that further studies may be possible to address these questions.

*Acknowledgements.* We are grateful to Dr. Joan Robb for providing the specimens used in this study and to Dr. R.C. Drewes for making available two of those sent to the California Academy of Sciences. Dr. W.F. Grant kindly provided use of his computer and Drs. S.K. Sessions and C.H. Daugherty offered valuable comments upon the manuscript. This study was supported in part by NSERC Canada grants A2198 and UO-526 to DMG.

## References

Baverstock PR, Watts CHS, Hogarth JT (1976) Heterochromatin variation in the Australian rodent *Uromys caudimaculatus*. *Chromosoma* 57:397-403  
 Bell BD (1982) The amphibian fauna of New Zealand. In: Newman DG (ed) *New Zealand herpetology*. New Zealand Wildlife Service, Wellington, pp 27-89  
 Bell BD (1986) The conservation status of New Zealand wildlife. *Occ Publ N-Z Wildlife Serv* 12:1-103

Bogart JP (1981) Chromosome studies in *Sminthillus* from Cuba and *Eleutherodactylus* from Cuba and Puerto Rico (Anura, Leptodactylidae). *Life Sci Contrib R Ontario Museum* 129:1-22  
 Dev VG, Miller DA, Tantravani RR, Schreck RR, Roderick TH, Erlanger BF, Miller OJ (1976) Chromosome markers in *Mus musculus*: differences in C-banding between the subspecies *M. m. musculus* and *M. m. molossinus*. *Chromosoma* 53:335-344  
 Duellman WE, Trueb L (1986) *The biology of amphibians*. McGraw-Hill Book Co. New York, p 670  
 Green DM (1985) Differentiation in amount of centromeric heterochromatin between subspecies of the red-legged frog, *Rana aurora*. *Copeia* 1985:1071-1074  
 Green DM, Bogart JP, Anthony EH, Genner DL (1980) An interactive, microcomputer based karyotype analysis system for phylogenetic cytogenetics. *Comput Biol Med* 10:219-227  
 Green DM, Kezer J, Nussbaum RA (1984a) Triploidy in *Hochstetter's* frog *Leiopelma hochstetteri* from New Zealand. *NZ J Zool* 11:457-461  
 Green DM, Myers PZ, Reyna DL (1984b) CHROMPAC III: an improved package for micro-computer assisted analysis of karyotypes. *J Hered* 75:143  
 Harvey AW, Hewitt GM (1979) B-chromosomes slow development in a grasshopper. *Heredity* 42:397-401  
 Jones RN, Rees H (1982) *B-Chromosomes*. Academic Press, New York, p 266  
 Kezer J, Sessions SK (1979) Chromosome variation in the plethodontid salamander *Aneides ferreus*. *Chromosoma* 71:65-80  
 Kezer J, Léon PE, Sessions SK (1980) Structural differentiation of the meiotic and mitotic chromosomes of the salamander *Ambystoma macrodactylum*. *Chromosoma* 81:177-197  
 King M (1980) C-banding studies on Australian hylid frogs: Secondary constriction structure and the concept of euchromatin transformation. *Chromosoma* 80:191-217  
 Leván A, Fredga D, Sandberg AA (1964) Nomenclature for centromeric position on chromosomes. *Hereditas* 52:201-220  
 Morescalchi A (1967) The karyotype of two specimens of *Leiopelma hochstetteri* Fitz. (Amphibia Salientia). *Caryologia* 21:37-46  
 Morescalchi A (1973) Amphibia. In: Chiarelli A, Capanna E (eds) *Cytotaxonomy and vertebrate evolution*. Academic Press, New York, pp 233-283  
 Patton JL (1977) B-chromosome systems in the pocket mouse, *Perognathus baileyi*: Meiosis and C-band studies. *Chromosoma* 60:1-14  
 Parker JS, Ainsworth CC, Taylor S (1981) The B-chromosome system of *Hypochoeris maculata* II. B-effects on meiotic A-chromosome behaviour. *Chromosoma* 67:123-143  
 Schmid M (1978) Chromosome banding in amphibia I. Constitutive heterochromatin and nucleolus organizer regions in *Bufo* and *Hyla*. *Chromosoma* 66:361-388  
 Schmid M (1983) Chromosome banding in Amphibia. VII. Analysis of the structure and variability of NORs in Anura. *Chromosoma* 87:327-344  
 Sessions SK (1984) *Cytogenetics and evolution in salamanders*. Ph.D. dissertation. University of California, Berkeley  
 Stephenson EM, Robinson ES, Stephenson NG (1972) Karyotype variation within the genus *Leiopelma* (Amphibia: Anura). *Can J Genet Cytol* 14:691-702  
 Stephenson EM, Robinson ES, Stephenson NG (1974) Inter-specific relationships of *Leiopelma* (Amphibia: Anura). Further karyological evidence. *Experientia* 30:1248-1250  
 Yosida T, Sagai T (1975) Variation of C-bands in the chromosomes of several subspecies of *Rattus rattus*. *Chromosoma* 50:283-300

Received February 18, 1987; in revised form May 19, 1987  
 Accepted by H. Macgregor