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## Severe autosomal recessive retinitis pigmentosa maps to chromosome 1p13.3–p21.2 between D1S2896 and D1S457 but outside ABCA4

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**Abstract** A severe form of autosomal recessive retinitis pigmentosa (arRP) was identified in a large Pakistani family ascertained in the Punjab province of Pakistan. All affected individuals in the family had night blindness in early childhood, early complete loss of useful vision, and typical RP fundus changes plus macular degeneration. After exclusion of known arRP loci, a genome-wide scan was performed using microsatellite markers at about 10 cM intervals and calculating two-point lod scores. PCR cycle dideoxynucleotide sequencing was used to sequence candidate genes inside the linked region for mutations. RP in this family shows linkage to markers in a 10.5 cM (8.9 Mbp) region of chromosome 1p13.3–p21.2 between D1S2896 and D1S457. D1S485 yields the highest lod score of 6.54 at  $\theta=0$ . Sequencing the exons and intron–exon boundaries of five candidate genes and six ESTs in this region, OLFM3, GNAI3, LOC126987, FLJ25070, DKFZp586G0123, AV729694,

BU662869, BU656110, BU171991, BQ953690, and CA397743, did not identify any causative mutations. This novel locus lies approximately 4.9 cM (7.1 Mbp) from ABCA4, which is excluded from the linked region. Identification and study of this gene may help to elucidate the phenotypic diversity of arRP mapping to this region.

### Introduction

Retinitis pigmentosa (RP; OMIN268000) is a clinically and genetically heterogeneous group of progressive retinal degenerations primarily affecting rod photoreceptors. Clinically, they share common features including night blindness, gradual constriction of visual fields, typical fundus changes and loss of central vision (Heckenlively et al. 1988). RP is the most common hereditary retinal dystrophy causing irreversible blindness and is most frequently inherited as an autosomal recessive trait (Boughman et al. 1980; Buch et al. 2004). A number of loci or genes responsible for RP have been reported (RetNet: <http://www.sph.uth.tmc.edu/Retnet/disease.htm>). arRP has been associated with 21 loci (Bareil et al. 2001; den Hollander et al. 1999; Dryja et al. 1995; Finckh et al. 1998; Gal et al. 2000; Gerber et al. 2000; Gu et al. 1999; Hagstrom et al. 1998; Hameed et al. 2001; Huang et al. 1995; Martinez-Mir et al. 1998; Maw et al. 1997; McLaughlin et al. 1993; Morimura et al. 1998, 1999; Nakazawa et al. 1998; Rivolta et al. 2000; Rosenfeld et al. 1992; Ruiz et al. 1998; Thompson et al. 2001; Tuson et al. 2004), and of these, the mutations responsible for arRP have been identified in ABCA4 (Martinez-Mir et al. 1998), CERKL (Tuson et al. 2004), CNGA1 (Dryja et al. 1995), CNGB1 (Bareil et al. 2001), CRB1 (den Hollander et al. 1999), LRAT (Thompson et al. 2001), MERTK (Gal et al. 2000), NR2E3 (Gerber et al. 2000), PDE6A (Huang et al. 1995), PDE6B (McLaughlin et al. 1993),

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RGR (Morimura et al. 1999), RHO (Rosenfeld et al. 1992), RLBP1 (Maw et al. 1997), RPE65 (Morimura et al. 1998), SAG (Nakazawa et al. 1998), TULP1 (Hagstrom et al. 1998), and USH2A (Rivolta et al. 2000).

The genes causing arRP in most families have yet to be identified. With the exception of USH2A, mutations in each individual gene identified so far have been responsible for arRP in only about 2–5% of families. Overall, causative mutations have been identified in a small fraction of arRP families, possibly as low as 30%. The limited number of large families in which arRP is segregating has hindered the search for additional loci. Searching for additional loci and genes associated with arRP not only provides valuable information for the diagnosis and differentiation of retinal degenerations but also forms a scientific basis for their prevention and treatment. Identification of disease genes is also a valuable way to ascertain the function and physiological role of genes in the retina, especially for recessive diseases which generally are caused by the loss of gene function. Identification of genes and their physiological role in the retina is valuable especially for recessive diseases caused by loss of gene function.

In this study, we describe a large consanguineous family with severe form of arRP and map the disease to a novel locus on chromosome 1p between DIS2896 and DIS457. This region does not include the ABCA4 gene, which is proximal on chromosome 1. In addition to the typical symptoms and signs of RP, affected individuals in this family showed relatively severe loss of central visual acuity in many affected individuals over 20 years old absent or limited only to hand movement. Consistent with this clinical finding, macular degeneration is present in all patients in whom the macular region is visible.

## Methods

### Family and clinical data

Family 61030 with arRP described in this study was ascertained from Punjab province of Pakistan as part of a collaborative project between the CEMB, Lahore, Pakistan and the NEI to identify genetic cause of eye diseases. It contains three consanguineous marriages. Nine affected and seven unaffected persons spanning three generations participated in this study. The diagnosis of RP was based on night blindness beginning in early childhood, progressive loss of peripheral vision, attenuation of retinal vessels, pigment disturbance on fundus examination, and decreasing visual acuity with age. Electroretinogram (ERG) responses were recorded consistent with ISCEV standards (Anonymous 1989) in selected cases, using commercial ERG equipment (LKC, Gaithersburg, MD, USA). Under scotopic (dark-adap-

ted) conditions, the single bright-flash stimulus elicits a response dominated by rod activity but that also contains a cone component. The photopic (light-adapted) 30 Hz flicker stimulus elicits activity exclusively from the cone system. Ophthalmological examination was performed by Dr. Z.A. Qazi, Dr. M. Amer and Ms Fareeha Zulfiqar at the LRBT/CEMB, Lahore, Pakistan. Informed consent was obtained from all participating individuals, conforming to the tenets of the Declaration of Helsinki. This project was approved by the IRBs of the National Eye Institute, Bethesda, MD and the Centre of Excellence in Molecular Biology, Lahore, Pakistan.

### Genotyping and linkage analysis

Genomic DNA was prepared from white blood cells as previously described (Smith et al. 1989). Genotyping for all participating family members was performed using 5'-fluorescently labeled microsatellite markers. A genome-wide scan was carried out using panels 1–27 of the ABI PRISM linkage Mapping Set Version 2, which includes 382 markers spaced at intervals of about 10 cM. PCR was conducted at 94°C for 8 min, followed by 10 cycles of amplification at 94°C 15 s, 55°C 15 s, and 72°C 30 s; then 20 cycles at 89°C 15 s, 55°C 15 s, 72°C 30 s; finally at 72°C for 10 min. After mixing with GENE-SCAN™ 400 HD [ROX™] standard (ABI) and deionized formamide, PCR products were denatured at 95°C for 5 min and then immediately placed on ice for 5 min. The amplicons were separated on Long Ranger sequencing gels (Cambrex) on an ABI 377 DNA sequencer. Genotyping data were collected by using GeneScan Analysis 3.0 and analyzed using the Genotyper 2.5 software package from ABI. Two-point linkage analysis was performed by using the MLINK program of the FASTLINK implementation of the LINKAGE program package (Lathrop and Lalouel 1984; Schaffer et al. 1994). RP in this family was analyzed as autosomal recessive trait with full penetrance and with a disease-gene allele frequency of 0.0001. For fine mapping, the markers in the candidate region were arranged according to the National Center for Biotechnology Information (NCBI) map. Linkage analysis of family 61030 was carried out with three loops broken at individuals 5, 15, and 25. Haplotypes were generated using the Cyrillic 2.1 program and confirmed by inspection. Equal marker allele frequencies were arbitrarily assumed for the genome wide scan and were calculated from 24 unrelated, unaffected individuals for fine mapping.

### Mutation screening of candidate genes

Genes located in the linked region were selected for mutation screening based on (1) expression in the retina and/or RPE; (2) encoding a functional domain seen in

known causative genes for RP; (3) belonging to a gene family known to cause RP; or (4) having a biochemical function related to a clinical phenotype that was observed in a syndrome with RP. Primers situated in the flanking region were designed to amplify each exon including exon-intron boundaries (primer sequences available on request). The amplicon was sequenced with the ABI BigDye Terminator cycle sequencing kit v3.1, according to the manufacturer's recommendations, using an ABI 3100 Genetic Analyzer. Sequencing results from 2 affected and 2 unaffected individuals (individual 12 (affected), individual 13 (unaffected), individual 16 (unaffected carrier), and individual 27 (affected)) in family 61030 as well as consensus sequences from the NCBI human genome database were imported to SeqMan (DNASTAR) and aligned to identify the variations.

Analysis of the ABCA4 gene for known sequence changes was carried out using the ABCA4 gene chip as described by Jaakson et al. (2003). Haplotypes were constructed with the sequence changes observed in the ABCA4 gene so as to minimize recombination events.

## Results

The presenting symptom experienced by all nine affected individuals is night blindness, which appears at 3–10 years of age. Visual acuity gradually decreased accompanied by a progressive loss of peripheral vision. The visual acuity in eight of the nine affected individuals is severely limited, to hand movement in three cases, light perception in one case, and no light perception in four cases, even though they were only 22–47 years old (Table 1). The youngest affected boy (individual 17, 11 years old), had a visual acuity of 6/36 (right) and 6/48 (left).

Photophobia and nystagmus are reported in every patient recently examined. There is no evidence of keratoconus. Recent funduscopy reveals waxy-pale discs, obvious attenuation of the retinal arteries, a generalized grayish carpet-like retinal degeneration, and typical bone-spicule pigmentation in the mid-peripheral retina. The range and amount of pigmentation increased with age. Macular degeneration is

seen in all affected individuals in whom the macular region is visible (Fig. 1). A bull's eye like appearance in the macular region surrounded by a grayish carpet-like retina in the posterior fundus was observed in younger patients as shown in Fig. 1: individuals 10, 11, 12, and 17. In older patients, the macular degeneration advanced to show atrophic changes accompanied by irregular pigment clumping as shown in Fig. 1: individuals 9, 14, and 30. ERG recordings document extensive loss of rod and cone function in two affected family members. Individual 10 at age 22 shows no appreciable rod or cone responses for either eye. Even individual 17 at age 11 has no identifiable cone responses to 30 Hz flicker and at best only slight residual rod function in only one eye. This indicates severe photoreceptor dysfunction and vision loss at young ages and indicates a severe form of rod-cone degeneration which has been termed "retinitis pigmentosa" (Fig. 2).

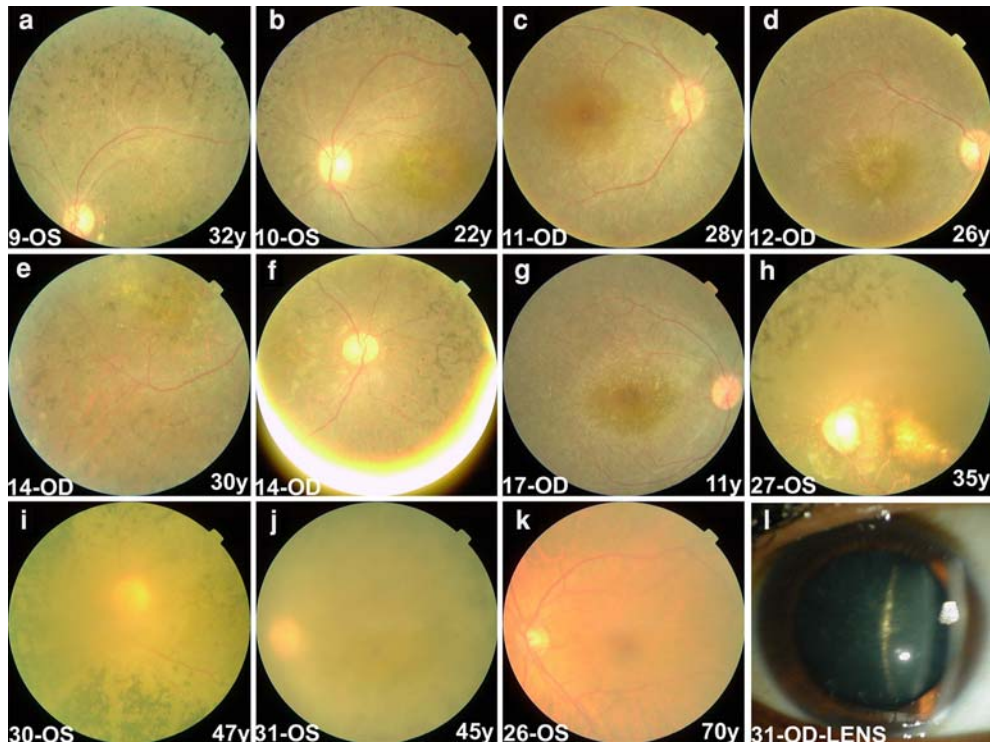
Initially, examining markers near known candidate genes excluded the RP locus in this family from all known arRP loci [data not shown]. Upon a complete genome-wide scan of chromosomes 1–22, two-point linkage analysis yields lod scores greater than 1.5 with markers corresponding to two possible loci, on chromosomes 1 and 13. The locus on chromosome 13 was excluded by lod scores of minus infinity of the nearby markers and by non-segregation of supposed disease alleles (haplotypes) in affected family members (data not shown). Fine mapping and haplotype analysis confirms the locus on chromosome 1p13.2–p21.2 (Fig. 3, Table 2). This locus maps to a 10.5 cM or 8.9 Mbp region between D1S2896 and D1S457. All eight microsatellite markers examined inside this region generated positive lod scores, with D1S485 showing the highest lod score, 6.54 at  $\theta=0$ .

Haplotypes of family members in this region support the linkage results (Fig. 3). Recombination at D1S2896 in individual 27 with further recombination at D1S236 in individual 25 confirmed in individuals 30 and 31 set the telomeric boundary for the linked region at D1S2896. Recombination at D1S457 for individuals 12, 14, and 15, confirmed in individual 17 as well as lack of homozygosity for this marker in individual 17 sets the proximal boundary for the linked region. Of the infor-

**Table 1** Clinical information for family members affected with arRP

ID #	Gender	Age (years)	First symptom	Age at onset (years)	Visual acuity	Fundus changes			
						Macular degeneration	Pale optic disc	Artery attenuation	Pigment deposit
9	M	32	Night blindness	8	NPL	Yes	Yes	Yes	Yes
10	F	22	Night blindness	10	HM	Yes	Yes	Yes	Yes
11	F	28	Night blindness	7	HM	Yes	Yes	Yes	Yes
12	F	26	Night blindness	9	HM	Yes	Yes	Yes	Yes
14	M	30	Night blindness	8	NPL	Yes	Yes	Yes	Yes
17	M	11	Night blindness	3	3/36,6/48	Yes	Yes	Yes	Yes
27	M	35	Night blindness	8	NPL	Yes	Yes	Yes	Yes
30	F	47	Night blindness	6	NPL	Yes	Yes	Yes	Yes
31	F	45	Night blindness	5	PL	UN	UN	UN	Yes

UN unknown. Posterior fundus could not be seen clearly because of posterior subcapsular cataract



**Fig. 1** Fundus photographs of all nine affected individuals and of a normal individual (26-OS) in the family as well as a photo showing posterior subcapsular cataract (31-lens). *OD* right eye. *OS* left eye. Grayish carpet-like retinal degeneration is shown in individuals 9, 10, 11, 12, 14, 17, 27, and 30 as compared to a normal fundus color in 26. Waxy-pale disc and arteriolar attenuation are shown in individuals 9, 10, 11, 12, 14, 17, 27, and 30. Bone-spicule pigment deposit in mid-peripheral retina is shown in individuals 9, 10, 12,

14, 27, 30, and 31. Such pigment deposit was recorded in 11 and 17 but is not shown in the photos because of further peripheral location. Bull's eye macular change is shown in individuals 10, 11, 12, and 17. Macular atrophy with pigment clumping is shown in individuals 9, 14, 27, and 30. The macular region is not visible for individual 31. The haze appearance of fundus photos for individuals 26, 27, 30, and 31 results from various degrees of posterior subcapsular cataract as shown in 31-lens

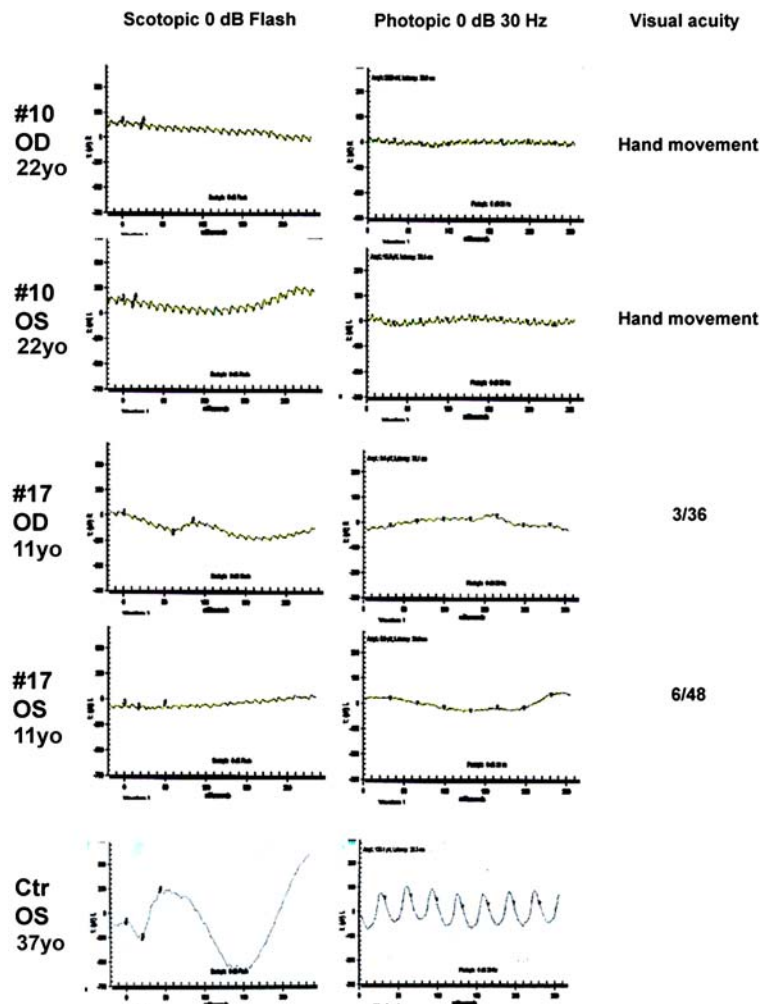
mative markers in the linked region, only D1S485 is homozygous in all affected individuals in both the left (offspring of individuals 5 and 6) and right (offspring of individuals 25 and 26) branches of the family. The variant affected haplotype inherited through individual 26 suggests an independent origin for the disease allele in that individual. Alternatively, if this disease allele in individual 26 is inherited from individual 40 or 41, or in the more likely event that it represents inheritance of the same disease allele from an even more distant progenitor, the candidate locus might lie in a very small region (1 Mbp) around D1S485 between D1S2699 and D1S429. Unfortunately, only six ESTs and no identified genes reside in the 1 Mbp region flanked by D1S2699 and D1S429.

The new *arRP* locus is close to *ABCA4*, which lies between D1S2868 and D1S236. *ABCA4* is about 7.1 Mbp proximal to the new locus and is not linked with *arRP* in this family (Fig. 3 and Table 2). Recombination at D1S2896 in individual 27 with further recombination at D1S236 in individual 25 confirmed in individuals 30 and 31 exclude *ABCA4* as a candidate locus. The linkage results are supported by haplotype analysis of SNPs within *ABCA4* (Table 3). Individuals 9 and 17, from the left-hand branch of the family, are

homozygous for one *ABCA4* haplotype. In contrast, siblings 27 and 30 have inherited at least 3 distinct haplotypes, the data being consistent with these two siblings sharing one haplotype (shown on the right for these individuals in table 3), and no haplotype being identical to those in individuals 9 and 17. Similarly, recombination with D1S457, which is given as the proximal flanking marker for *CORD8* by Khaliq et al. (2000), excludes *CORD8* as a candidate locus, although *CORD8* is also described as having a distinct phenotype.

There are over 73 genes listed in the linked region. Of these, 5 genes were selected for initial mutational screening based on their expression pattern and presumed functional properties. Genomic DNA encompassing the six ESTs between D1S2699 and D1S429 was also screened for mutation. Sequence analyses of LOC126987 (3 exons), FLJ25070 (14 exons), OLFM3 (6 exons), *GNAI3* (8 exons), and DKFZp586G0123 (10 exons) as well as genomic DNA flanking six ESTs (AV729694, BU662869, BU656110, BU171991, BQ953690, and CA397743) did not reveal any potentially causative mutations. Both affected and unaffected family members are heterozygous for two SNPs in LOC126987, dbSNP2060700 and dbSNP6582952. No variation was detected in FLJ25070. Three individuals

**Fig. 2** ERG responses from two affected family members. For comparison, the normal control (*ctr*) subject to the scotopic single flash stimulus shows a rod-dominated initial negative-going a-wave and positive b-wave of 500  $\mu$ V, and has a 120  $\mu$ V flicker response of the cone system to photopic 30 Hz stimulus



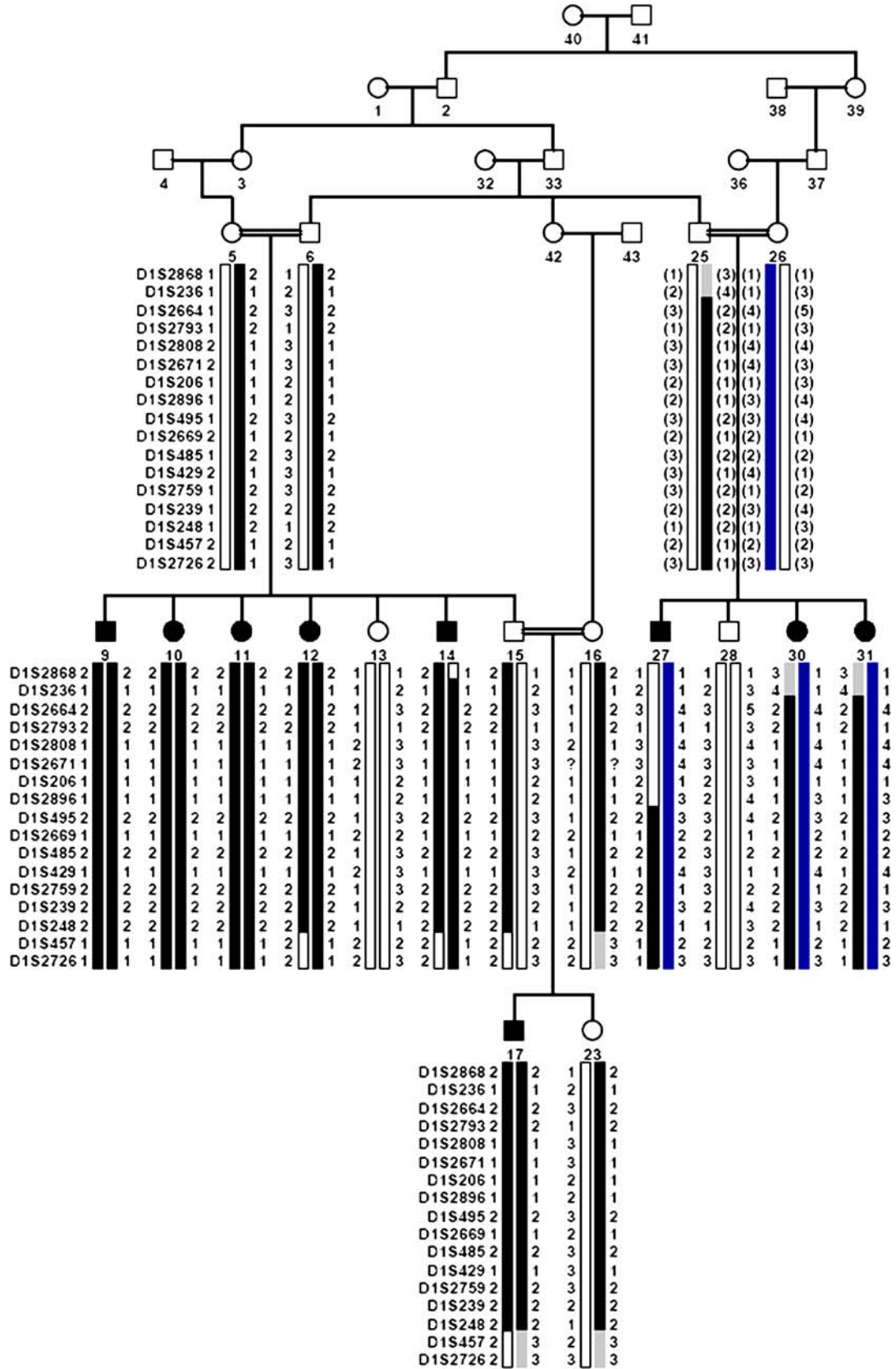
were each heterozygous for a SNP, dbSNP11276972, dbSNP10493973, and dbSNP4907955, in OLFM3. Both affected and unaffected individuals were homozygotes for three variations, c.311-11C>T, c.311-43C>T, and C.398+45T>C, in SLC25A24 (NM\_013386).

## Discussion

In this study, a severe type of arRP in a large Pakistani family with consanguinity was assigned to a new locus on 1p13.3–p21.2 between D1S2896 and D1S457. This interval does not include ABCA4, which was excluded by haplotype and linkage analysis producing lod scores below  $-2$  (Table 2). Similarly, the CORD8 locus described by Khaliq and coworkers is distinct from this locus by linkage analysis (Khaliq et al. 2000). Five putative candidate genes and six ESTs in this interval have been screened by sequencing without identification of any potentially causative mutations. Nearly half of the 73 genes in the linked region are expressed in the retina and/or retinal pigment epithelium (NCBI human genome database) and the function for many of them is

currently unknown. Additional genes in the linked region include HEJ1, AMY2B, COL11A1, CESLR2, SORT1, GPR61, and GNAT2. Mutations in COL11A1 cause Stickler syndrome (Vikkula et al. 1995) and mutations in GNAT2 cause achromatopsia (Kohl et al. 2002). Identification of new gene structure of those retinally expressed EST sequences in the linked region and analysis of additional genes may finally lead to the target causative gene.

The RP disease phenotype in this family is quite severe for both rod and cone vision. All affected family members reported night blindness by age 10, and all have severe loss of central vision with reduction of visual acuity to “hand motion” or less at age 22–47, and even the 11-year-old has severe acuity reduction. All including the 11-year-old had typical and advanced RP fundus changes accompanied by macular changes. The disease in this family is differentiated from the congenital blindness of Leber congenital amaurosis (OMIM 204000) in which patients generally are born blind or lose vision within a few months after birth. It is distinguished from most forms of cone–rod dystrophy in which visual function at night is spared initially and for



**Fig. 3** Pedigree and haplotypes on 1p13–p21. *Blackened bars* indicate disease allele. *Filled squares or filled circles* represent individuals affected with RP. The *individual number* in the pedigree is the same as the number used in Fig. 1 and Table 3

**Table 2** Two-point linkage results for markers in the ARRP region at 1p13.2–p21.2

Markers	Position		Lod score at $\theta$ value							$Z_{\max}$	$\theta_{\max}$
	cM	Mbp <sup>a</sup>	0.00	0.01	0.05	0.10	0.20	0.30	0.40		
D1S2868	129.90	93.05	–∞	–1.96	0.40	1.07	1.19	0.81	0.29	1.24	0.16
D1S236	132.40	93.82	–∞	1.03	2.07	2.22	1.88	1.27	0.56	2.22	0.10
D1S2664	133.00	95.66	–∞	2.91	3.81	3.78	3.06	2.01	0.86	3.85	0.07
D1S2793	133.00	96.81	–∞	3.26	4.14	4.08	3.29	2.17	0.96	4.17	0.07
D1S2808	135.20	98.97	–∞	2.33	3.25	3.25	2.62	1.70	0.69	3.30	0.07
D1S2671	137.40	100.98	–∞	3.47	4.34	4.29	3.48	2.34	1.07	4.38	0.07
D1S206	137.60	101.40	–∞	2.16	2.54	2.44	1.90	1.23	0.52	2.54	0.05
D1S2896	137.30	101.68	–∞	1.05	2.07	2.21	1.86	1.23	0.53	2.21	0.09
D1S495	140.80	102.27	3.35	4.57	4.76	4.42	3.39	2.17	0.93	4.80	0.03
D1S2699	140.70	104.49	1.52	2.77	3.06	2.86	2.14	1.29	0.46	3.06	0.04
D1S485	140.60	104.98	6.54	6.41	5.89	5.23	3.86	2.46	1.07	6.54	0.00
D1S429	140.50	105.41	3.70	4.90	5.06	4.68	3.56	2.25	0.94	5.11	0.03
D1S2759	140.30	105.56	3.70	5.25	5.41	5.01	3.85	2.49	1.09	5.46	0.03
D1S239	143.10	106.55	0.04	1.27	1.70	1.67	1.31	0.82	0.31	1.71	0.07
D1S248	143.30	106.87	3.41	4.63	4.82	4.48	3.45	2.23	0.97	4.85	0.04
D1S457	147.80	110.59	–∞	–4.63	–1.44	–0.30	0.42	0.45	0.20	0.74	0.80
D1S2726	149.00	110.90	–∞	–3.52	–0.47	0.51	0.97	0.81	0.41	0.97	0.21

<sup>a</sup>Build 35.1 (9-15-04)

which some vision typically persists beyond the third to fourth decade (Evans et al. 1995; Gregory-Evans et al. 2000; Klevering et al. 1999; Small and Gehrs 1996). Like this family, macular degeneration in association with RP has been reported in arRP caused by mutations in such genes as MERTK (McHenry et al. 2004), RLBP1 (Burstedt et al. 2001) and ABCA4 (Cremers et al. 1998; Martinez-Mir et al. 1997, 1998).

This new locus is of particular interest since it is close to but distinct from ABCA4, lying about 7.1 Mbp telomeric. Mutations in the ABCA4 gene can cause autosomal recessive Stargardt disease (STGD)/fundus flavimaculatus (FFM), cone-rod dystrophy (CRD), and arRP (Allikmets et al. 1997; Briggs et al. 2001; Cremers et al. 1998; Fukui et al. 2002; Gerber et al. 1998; Klevering et al. 1999, 2004; Lewis et al. 1999; Martinez-Mir et al. 1998; Maugeri et al. 1999, 2000; Nasonkin et al. 1998; Paloma et al. 2002; Rozet et al. 1999; Rudolph et al. 2002; Shroyer et al. 2001; Webster et al.

2001). Two arRP families with macular degeneration and ABCA4 mutations were independently assigned to ABCA4 region by linkage studies for candidate loci (Cremers et al. 1998; Klevering et al. 1999; Martinez-Mir et al. 1997, 1998). In one family, five of the six affected individuals were homozygous for markers extending more centromerically without a determined border (Martinez-Mir et al. 1997). The second patients were classified as either RP- or CRD-like but were typical for neither disease. (Cremers et al. 1998; Klevering et al. 1999) The RP locus was found to lie between D1S198 and D1S239, containing ABCA4 and overlapping part of the new arRP locus determined in this study. The RP in the family reported here shares some common features with the two families linked to ABCA4: waxy-pale disc, attenuated retinal vessels, pigment deposits in the mid-peripheral retina, and degeneration or atrophy involving the macula. However, choroidal atrophy in the posterior pole is much more

**Table 3** Haplotypes of four affected individuals in the pedigree using the sequence changes in the ABCA4 gene

Exon	Nucleotide change	Amino acid change	Individual number			
			9	17	27	30
10	1268 A > G	H423R	<i>A/A</i>	<i>A/A</i>	<i>A/A</i>	<i>G/A</i>
	1269 C > T	H423H	<u>T/T</u>	<u>T/T</u>	<u>C/C</u>	<u>C/C</u>
	delG IVS+5	Splice	<u>G/G</u>	<u>G/G</u>	<u>G/T</u>	<u>G/T</u>
19	2828 G > A	R943Q	<u>A/A</u>	<u>A/A</u>	<u>G/G</u>	<u>G/G</u>
33	IVS+48 C > T	Splice	<u>C/C</u>	<u>C/C</u>	<u>T/T</u>	<u>C/T</u>
45	6249 C > T	I2083I	<u>C/C</u>	<u>C/C</u>	<u>T/C</u>	<u>C/C</u>
46	6285 T > C	D2095D	<u>T/T</u>	<u>T/T</u>	<u>C/T</u>	<u>T/T</u>
48	6529 G > A	D2177N	<u>G/G</u>	<u>G/G</u>	<u>G/G</u>	<u>A/G</u>
49	6764 G > T	S2255I	<u>G/G</u>	<u>G/G</u>	<u>T/G</u>	<u>G/G</u>

Italic—wild-type alleles; underlined—nucleotide substitutions which do not lead to the amino acid substitution and/or common polymorphisms; bold—nucleotide substitution which results in an amino acid substitution; the individual numbers in Table 3 are consistent with those in Figs. 1 and 2 and Table 1

obvious in the two families with ABCA4 mutations (Cremers et al. 1998; Klevering et al. 1999; Martinez-Mir et al. 1997, 1998). The generalized grayish carpet-like retinal change seen in this Pakistani family has also been reported previously in a family (Cremers et al. 1998; Klevering et al. 1999) in which the linked region also overlaps with the novel locus described here and which shows mutations in ABCA4.

It is not impossible that other factors may contribute to the phenotypic diversity of RP associated with ABCA4 mutations, and much evidence supports a complex relationship of arRP with mutations in ABCA4 (Cremers et al. 1998; Fukui et al. 2002; Klevering et al. 1999, 2004; Martinez-Mir et al. 1998; Paloma et al. 2002; Rozet et al. 1999; Rudolph et al. 2002; Shroyer et al. 2001). It has been hypothesized that two null mutations in ABCA4 result in arRP while a combination of a null mutation with a second mutation preserving residual ABCA4 function cause STGD or CRD (Cremers et al. 1998). However, homozygous null mutations and two null alleles have also been identified in patients with STGD and CRD (Briggs et al. 2001; Gerber et al. 1998; Lewis et al. 1999; Simonelli et al. 2004). Macular degeneration has been described in most RP patients with ABCA4 mutations (Cremers et al. 1998; Fukui et al. 2002; Klevering et al. 2004; Martinez-Mir et al. 1997, 1998). Some patients diagnosed as RP or RP-like had somewhat uncertain phenotypes (Klevering et al. 1999, 2004), or could not be excluded as having CRD (Klevering et al. 2004), or were actually diagnosed as having STGD (Fukui et al. 2002) or CRD (Paloma et al. 2002) in earlier examinations. The retinal degeneration seen in STGD and CRD could progress to show RP-like fundus changes (Fukui et al. 2002; Paloma et al. 2002). RP and STGD or CRD may present together in one family just by chance (Paloma et al. 2002). The high variability of ABCA4 is noteworthy, with an average of 3.8 variations per individual seen in all participants including controls and 76% of controls harboring at least one variation (Webster et al. 2001). Homozygous ABCA4 knock out mice still have ERG findings comparable to wild-type mice which is consistent with STGD but not RP (Radu et al. 2003; Weng et al. 1999). Therefore, it remains at least a theoretical possibility that another gene close to ABCA4 may cause RP independently or may act with ABCA4, contributing to the phenotypic diversity of ABCA4. The possibility is further suggested by the results presented in this study. Cloning of this new arRP gene linked to 1p13.3–p21.2 may elucidate this possibility.

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