

Resonant energy transfer in electron-driven proton pumps

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We examine proton and electron transport in living cell membranes and show that the Coulomb interaction leads to a resonant energy transfer, from the electrons to protons during their simultaneous tunnelings, which allows the proton pump to work. The proton pump efficiency is controlled by the

electron voltage build-up, external temperature, and the molecular electrostatics driving the electron and proton energies to their resonant conditions. We also show that physiological (e.g., human body) temperatures are optimal for the operation of this proton pump.

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1 Introduction Charge and energy transfers in living cells require nanoscale machinery working with high efficiency and reliability. An example of such nanomachine is the *cytochrome c oxidase* (*CcO*) which converts atmospheric oxygen into water, reducing the energy of electrons (reduction/oxidation, or redox, process) and simultaneously creating a proton gradient across the inner mitochondrial membrane (proton pump) [1]. This membrane separates regions with different proton concentrations, and the proton gradient is used for driving the biological motor of adenosine triphosphate (ATP) synthase, which converts mechanical motion into the chemical energy stored in ATP molecules. In recent years, the structure of *CcO* has been determined for various organisms, but the specific mechanism of the proton pump remains elusive [2].

Recent progress in nanotechnology brings the parameters of modern electronic devices closer to the scales of living cell machinery. This makes it possible both to mimic biological systems for device purposes and to analyze the electron and proton transport mechanisms in living cells using methods of nanoelectronics and nanomechanics. Here, we apply such methods to examine the electroni-

cally-controlled proton transfer through an inner mitochondrial membrane and to try to understand the elusive mechanism of proton pumps. To create and maintain the proton gradient, it is necessary to transfer energy from the intra-membrane electron transport chain to the proton subsystem. We propose a model for this energy transduction based on the Förster-type energy exchange [3], i.e. when the simultaneous electron and proton tunnelings are accompanied by a transfer of energy under resonant conditions between the electrons and protons. It should be emphasized that such quantum-mechanical processes [4] (i.e. electron and proton tunnelings) are very important for living cell dynamics, despite temperatures much larger than that of conventional nanoelectronics. We show below that our proton pump mechanism operates optimally at human body temperatures (about 36 °C).

2 Formulation We examine the situation where the electron transport between the left, *L*, and right, *R*, sites embedded into the inner membrane of the mitochondrion is accompanied by the proton transfer from the “lower” (*D*) protonable site that is close to the negative (inner) side of

the membrane to the “upper” (U) protonable site located nearby the positive (outer) surface of the membrane (Fig. 1).

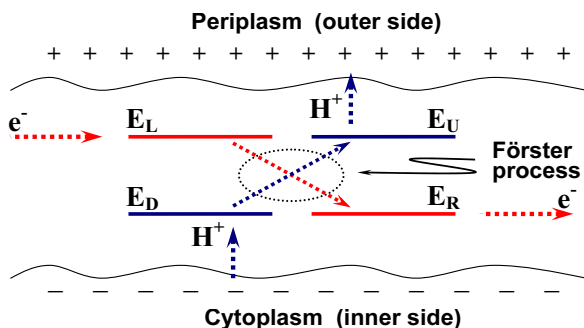


Figure 1 Schematic diagram of the Förster-type energy exchange between electron and proton subsystems. The electron transfer from left to right is accompanied by the proton transfer from the site D with lower energy to the site U with higher energy.

In the case of CcO , a specific site (known as *heme a*) and a binuclear site can be considered [5, 6] as a double-dot system for electrons with two dots (sites) having different energies E_L and E_R [6]. For the proton component, the two proton dots (sites) having different energies, E_D and E_U , correspond to [6] a carboxylic residue E278 from the negative (inner) side of the membrane and a protonable site from the positive (outer) side of the membrane. It is assumed [6] that the left (right) electron site is coupled only to the left (right) electronic reservoir and the proton D and U sites are also coupled only to the corresponding proton reservoirs. The electron and proton tunnelings between their two sites are described by the tunneling splittings, Δ_a and Δ_b , respectively. In addition, the Coulomb interaction between electrons and protons localized on the corresponding sites leads to the Förster transfer [3], i.e., to the process when the energy is transferred from the electrons to protons (or vice versa) during their simultaneous tunnelings. The same mechanism is responsible for the fluorescence resonant energy transfer (FRET) in biological systems [7] as well as for exciton transfer in condensed matter [8]. To be effective, the Förster transfer requires a resonant condition: when the energy difference between the initial (L) and final (R) electron states is equal to the energy separation of the final (U) and the initial (D) proton states: $E_L - E_R \approx E_U - E_D$. We show that, in the presence of the electron-proton Förster interaction, the voltage applied between the left and the right *electron* reservoirs (redox potential) can be used for generating a *proton* current in the direction of *higher* proton concentration, thus realizing a proton pump.

To analyze this process quantitatively, we examine an electron double-dot connected to the left and right electron reservoirs and a proton double-dot connected to the “up” and “down” proton reservoirs. The Hamiltonian describing

double-dots is given by

$$H = \sum E_\alpha a_\alpha^\dagger a_\alpha + \sum E_\beta b_\beta^\dagger b_\beta - \Delta_a a_L^\dagger a_R - \Delta_a^* a_R^\dagger a_L - \Delta_b b_D^\dagger b_U - \Delta_b^* b_U^\dagger b_D - \lambda a_L^\dagger a_R b_D^\dagger b_U - \lambda^* a_R^\dagger a_L b_U^\dagger b_D \quad (1)$$

and the coupling of the double dots to the leads has the form

$$H_T = -\sum t_{k\alpha} c_{k\alpha}^\dagger a_\alpha - \sum T_{k\beta} d_{k\beta}^\dagger b_\beta + H.c. \quad (2)$$

Here, a_α^\dagger (a_α) and b_β^\dagger (b_β) are the creation (annihilation) operators in the α -sites for the electrons and in the β -sites for the protons, respectively ($\alpha = L, R$; $\beta = D, U$); E_α and E_β are the energies of electrons and protons in the corresponding α - and β -dot, respectively. Terms with tunneling amplitudes Δ_a and Δ_b describe the *conventional* electron and proton tunnelings, whereas the *resonant Förster double-tunneling* is described by the terms with the coupling constant. The direct Coulomb coupling of electrons and protons only leads to the renormalization of their energies; however, the overlap of their wave function produces the Coulomb terms responsible for the Förster-type interaction [3, 9].

Evaluating the equations of motion for electron and proton operators, we obtain self-consistent set of equations for electron and proton *populations* in the form:

$$\langle n_L \rangle = \int \frac{d\omega}{2\pi} \left(\gamma_L f_L(\omega) |F_L(\omega)|^2 + |\Delta_a|^2 \gamma_R |G_L(\omega)|^2 |G_R(\omega)|^2 \right) + |\lambda|^2 \langle N_U \rangle (1 - N_D) \gamma_R \int \frac{d\omega}{2\pi} f_R \left(\omega - \frac{E_U - E_D}{2} \right) \times \left| G_L \left(\omega - \frac{E_U - E_D}{2} \right) \right|^2 \left| G_R \left(\omega - \frac{E_U - E_D}{2} \right) \right|^2 \quad (3)$$

$\langle n_R \rangle$ is obtained from Eq. (3) by replacing L by R , D by U , and vice versa. Essentially identical equations hold for the proton populations, $\langle N_U \rangle$ and $\langle N_D \rangle$. The Green's functions involved in Eq. (3) (as well as in the equations for the proton populations, with replacement of α by β and γ by Γ) are given by

$$G_\alpha(\omega) = \frac{1}{\omega - E_\alpha + i\gamma_\alpha/2} \quad (4)$$

and

$$F_\alpha(\omega) = \frac{1}{\omega - E_\alpha + i\gamma_\alpha/2 - \Sigma_\alpha(\omega)} \quad (5)$$

where γ_α (Γ_β) are the electron (proton) level broadenings due to the couplings to their respective leads, f_α (f_β) are the Fermi distribution functions in the corresponding electron (proton) reservoirs with the chemical potential $\mu_L - \mu_R = V_e$ ($\mu_U - \mu_D = V_p$), and V_e (V_p) are the voltages build-up in the electron (proton) subsystems, respectively. The self-energy

$\Sigma_L(\omega)$ is given by

$$\Sigma_L(\omega) = |\Delta_a|^2 G_R(\omega) + |\lambda|^2 \langle N_D \rangle (1 - \langle N_U \rangle) G_R(\omega + E_D - E_U) \quad (6)$$

with $\Sigma_R(\omega)$, $\Sigma_U(\omega)$, and $\Sigma_D(\omega)$ having similar forms.

Using the equations of motion for electron and proton operators of the leads, we obtain expressions for the electron and proton currents in terms of the steady-state populations, Fermi distributions, and Green's functions, as

$$I_e = |\Delta_a|^2 \gamma_L \gamma_R \int \frac{d\omega}{2\pi} |G_L(\omega)|^2 |G_R(\omega)|^2 (f_L(\omega) - f_R(\omega)) + |\lambda|^2 \gamma_L \gamma_R \int \frac{d\omega}{2\pi} \left| G_L\left(\omega + \frac{E_U - E_D}{2}\right) \right|^2 \left| G_R\left(\omega - \frac{E_U - E_D}{2}\right) \right|^2 \times \left[\langle N_D \rangle (1 - \langle N_U \rangle) f_L\left(\omega + \frac{E_U - E_D}{2}\right) - \langle N_U \rangle (1 - \langle N_D \rangle) f_R\left(\omega - \frac{E_U - E_D}{2}\right) \right] \quad (7)$$

and

$$I_p = |\Delta_b|^2 \Gamma_D \Gamma_U \int \frac{d\omega}{2\pi} |G_D(\omega)|^2 |G_U(\omega)|^2 (f_U(\omega) - f_D(\omega)) + |\lambda|^2 \Gamma_D \Gamma_U \int \frac{d\omega}{2\pi} \left| G_D\left(\omega + \frac{E_R - E_L}{2}\right) \right|^2 \left| G_U\left(\omega - \frac{E_R - E_L}{2}\right) \right|^2 \times \left[\langle n_R \rangle (1 - \langle n_L \rangle) f_U\left(\omega - \frac{E_R - E_L}{2}\right) - \langle n_L \rangle (1 - \langle n_R \rangle) f_D\left(\omega + \frac{E_R - E_L}{2}\right) \right] \quad (8)$$

The positive direction is chosen to be from the higher chemical potential to the lower chemical potential, both for the electrons and protons.

3 Results and discussion

The self-consistent equations for the electron and proton populations (e.g., as in Eq. (3)) were solved numerically and their solutions

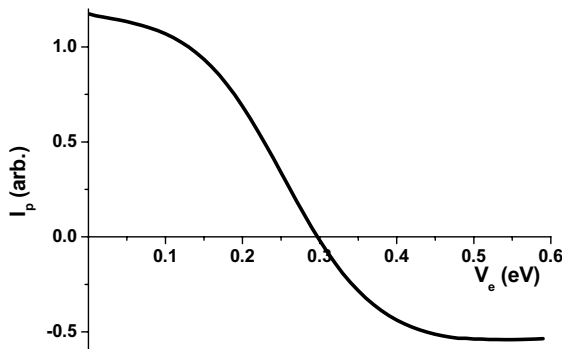


Figure 2 Proton current, I_p , as a function of electron voltage build-up.

were substituted into Eqs. (7),(8) for I_e and I_p . The electron current-voltage characteristics (not shown) exhibit the usual Ohmic region at small voltages, followed by a saturation regime.

The dependence of the proton current, I_p , on the *electron* voltage is shown in Fig. 2 for the electron energies $E_R = 100$ meV and $E_L = 200$ meV, proton energies $E_U = 200$ meV and $E_D = 100$ meV (the equilibrium chemical potentials are taken as the reference energies), tunnel splittings $\Delta_a = \Delta_b = 10$ meV, Förster constant, = 40 meV, and the temperature of the human body, 36.6 °C. The proton voltage build-up is taken to be $V_p = 200$ meV, which is typical for a living cell [1]. It is evident from Fig. 2 that at small electron voltages the proton current is positive and it becomes negative with increasing electron voltage. It should be noted that the negative proton current saturates at electron voltages larger than 450 meV, which is close to the maximal redox potential change along the *CcO* [1].

The temperature dependence of the proton current is of special interest. It is presented in Fig. 3 for the parameters of Fig. 2 and for the electron voltage $V_e = 350$ meV corresponding to the linear regime of Fig. 2.

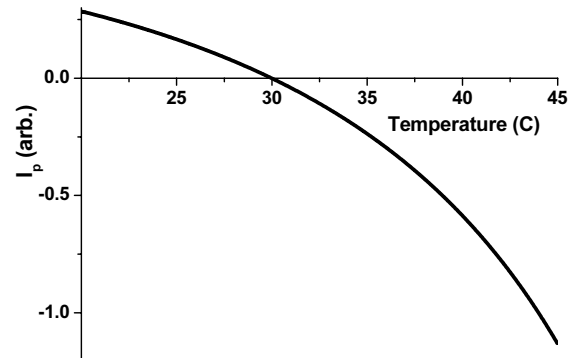


Figure 3 Temperature dependence of the proton current.

One can see that at temperatures below 30 °C, the proton current is positive and the proton pump cannot function. The optimal regime occurs at temperatures about that of the human body (36 °C), and with further temperature increase the negative proton current becomes so strong that it could break the electrostatic balance within the cell.

Förster-type processes require a resonant condition and are thus sensitive to detuning from the resonance. To illustrate this, we plot the proton (Fig. 4(a)) and electron (Fig. 4(b)) currents as functions of the detuning for varying E_U .

These figures show that the proton pump can only work if such detuning is sufficiently small. It should be noted that the electron current is strongly enhanced at the resonant condition because the energy drain provided by

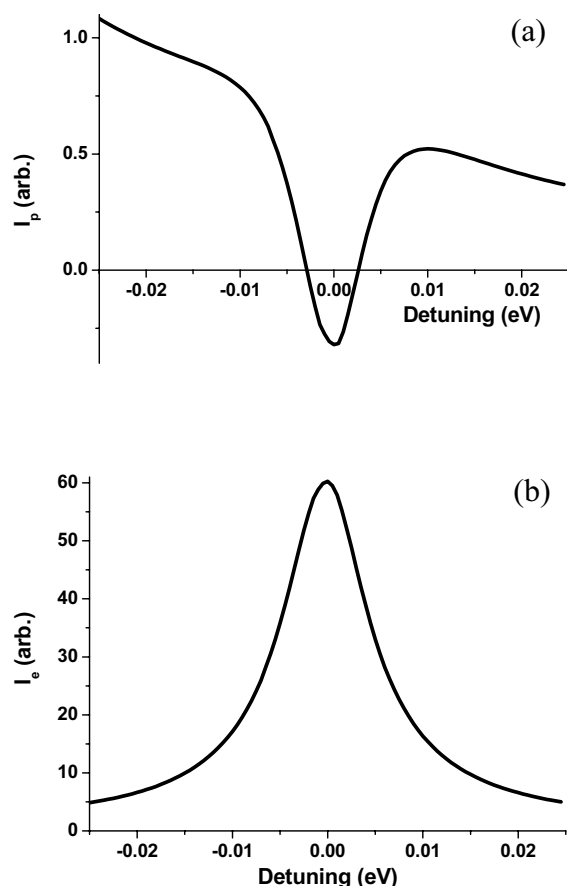


Figure 4 The proton, I_p , (a) and electron, I_e , (b) currents as functions of the detuning of the proton and electron energies from the resonant conditions.

the proton subsystem compensates the energy difference between electron levels in the two dots. The slope in Fig. 4(a) around the resonant current dip is produced by the direct proton current (the first term in Eq. (8)), which is maximal when $E_U = E_D$ and, therefore, increases when EU decreases.

4 Conclusion In conclusion, we have proposed a model to describe the electronically-driven proton pump using a Förster-type resonant process for the energy transfer from the electron to proton subsystems. Our calculations, based on reasonable parameters, demonstrate that the proton pump works optimally at physiological temperatures. We have examined the effects of the electron voltage build-up, external temperature, and molecular electrostatics driving the electron and proton energies to the resonant conditions, and have shown that these parameters can control the proton pump operation.

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References

- [1] B. Alberts et al., *Molecular Biology of the Cell* (Garland Science, New York, 2002), Ch. 14.
- [2] For recent reviews on proton pumps, see, e.g., J. P. Hosler, S. Ferguson-Miller, and D. A. Mills, *Annu. Rev. Biochem.* **75**, 165 (2006); J. Quenneville, D. M. Popovic, and A. A. Stuchebrukhov, *Biochem. Biophys. Acta* **1757**, 1035 (2006); M. Wikstrom and M. I. Verkhovsky, *Biochem. Biophys. Acta* **1757**, 1047 (2006); S. Papa, G. Capitanio, and P. L. Martino, *Biochem. Biophys. Acta* **1757**, 1133 (2006).
- [3] T. Förster, in: *Modern Quantum Chemistry*, edited by O. Sinanogly (Academic, New York, 1965).
- [4] See, e.g., A. Garg, J. N. Onuchic, and V. Ambegaokar, *J. Chem. Phys.* **83**, 4491 (1985).
- [5] M.-L. Tan, I. Balabin, and J. N. Onuchic, *Biophys. J.* **86**, 1813 (2004).
- [6] K. Faxen et al., *Nature* **437**, 286 (2005).
- [7] I. L. Medintz et al., *Nature Mater.* **2**, 630 (2003).
- [8] M. Achermann et al., *Nature* **429**, 642 (2004).
- [9] A. O. Govorov, *Phys. Rev. B* **71**, 15532