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# For Pony

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#### **List of Abbreviations**

2D Two dimensional

3D Three-dimensional

αΜΕΜ Alpha Minimum Essential Medium

BMP-2 Bone morphogenic protein-2

DMEM Dubecco's Modified Eagle Media

D-PBS Dulbecco's Phosphate Buffered Saline

EBS Embryoid bodies

ECM Extracellular matrix

ESC Embryonic stem cells

FBS Fetal bovine serum

FGF2 Fibroblast growth factor 2

HBSS Hank's buffered salt solution

hESC Human embryonic stem cells

IGF Insulin-like growth factor I

LIF Leukemia inhibitory factor

NF Nanofibrous

PCL Poly(caprolactone)

PDAC Poly(diallyldimethylammonium chloride)

PEO Poly(ethylene oxide)

PET Polyethylene terephthalate

PHB Poly(3-hydroxybutyrate)

PLGA Poly(lactic-co-glycolic acid)

PLLA Poly(L-lactic acid)

PVA Poly(vinyl alcohol)

SW Solid-walled

TGF-β1 Human transforming growth factor-beta1

TIPS Thermally induced phase separation

TUJ1 Neuronal Class III β-Tubulin

**ABSTRACT** 

Effects of Nanofibrous Scaffolding Architecture on Bone Tissue Development

from Embryonic Stem Cells

by

Laura Ann Smith

Chair: Peter X. Ma

Embryonic stem cells, typically isolated from the inner cell mass of blastocysts,

represent a potentially unlimited cell source for tissue engineering. However, the

potential tumorgencity of the undifferentiated cells and the heterogeneous cell population

generated by current differentiation protocols impede the use of embryonic stem cells as

a clinical cell source for tissue engineering applications. This thesis examines the effects

of emulating the differentiation signals provided by the extracellular matrix during

development with synthetic poly (L-lactic acid) nanofibers on the differentiation of the

embryonic stem cells to osteoblasts.

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First, undifferentiated mouse embryonic stem cells were seeded onto two dimensional nanofibrous thin matrices or flat (solid) films. With osteogenic supplementation the nanofibrous architecture was found to promote the osteogenic differentiation and mineralization of the mouse embryonic stem cells.  $\alpha$ 2 and  $\alpha$ 5 integrin appear to contribute to this osteogenic differentiation.

Next, the effects of biologically active factors and three dimensional culture were examined on mouse embryonic stem cells which were partially differentiated via embryoid body formation prior to seeding on the materials. The nanofibrous architecture was found to facilitate further differentiation of the cells in the absence of osteogenic stimulation, while the cells cultured on solid film required osteogenic supplements and growth factors to support osteogenic differentiation. Three dimensional culture on nanofibrous scaffolding was found to further enhance the osteogenic differentiation and mineralization more than two dimensional culture on either the nano-fibrous or solid architecture and three dimensional culture on the solid-walled scaffolding.

The osteogenic differentiation of human embryonic stem cells was examined next. In both two and three dimensional culture, the nanofibrous architecture enhanced the osteogenic differentiation and mineralization of the human embryonic stem cells compared to the solid architecture.

In summary, the nanofibrous architecture enhances the osteogenic differentiation of mouse and human embryonic stem cells compared to the more traditional solid-walled tissue engineering scaffolding architecture. This indicates that emulating size scale of the

extracellular matrix with synthetic nanofibers is advantageous in promoting osteogenic differentiation of embryonic stem cells.