

ANNALS OF THE NEW YORK ACADEMY OF SCIENCES

Issue: *Clearance of Dying Cells in Healthy and Diseased Immune Systems***A role for calreticulin in the pathogenesis of rheumatoid arthritis**

Joseph Holoshitz, Denise E. De Almeida, and Song Ling

Department of Internal Medicine, University of Michigan Medical Center, Ann Arbor, Michigan

Address for correspondence: Joseph Holoshitz, M.D., University of Michigan School of Medicine, 5520D MSRB1, 1150 W. Medical Center Dr., Ann Arbor, MI 48109-5680. jholo@umich.edu

Calreticulin (CRT) plays a role in the clearance of dying cells and has been implicated in autoimmunity. Recent evidence indicates that cell surface CRT (csCRT) acts as a signal transducing receptor for the rheumatoid arthritis (RA) shared epitope (SE). The SE binding site on CRT has been mapped to amino acid residues 217–223 in the P-domain. Upon interaction with dendritic cells (DCs), the SE activates potent immune regulatory events. In $CD8\alpha^+$ DCs, which express higher abundance of csCRT, the SE inhibits the tolerogenic enzyme indoleamine 2,3 dioxygenase with resultant inhibition of regulatory T (Treg) cell differentiation. In $CD8\alpha^-$ DCs, the SE ligand increases secretion of IL-6 and IL-23 and facilitates generation of Th17 cells, a T cell subset known to play a role in autoimmunity. On the basis of these recent findings, we discuss the possibility that the csCRT may play a pathogenic role in RA by transducing SE-activated Th17-polarizing signals.

Keywords: calreticulin; rheumatoid arthritis; shared epitope; nitric oxide; Th17; Treg

Introduction

Calreticulin (CRT) is a ubiquitous multifunctional calcium-binding protein.¹ Although originally characterized as an endoplasmic reticulum molecular chaperone, more recently it has been shown that extracellular CRT attaches to the surface of many cells, where it is involved in signal transduction events associated with innate immunity, cell adhesion, and apoptosis.^{2–5} Because CRT lacks a trans-membrane domain, CD91 (also known as low-density lipoprotein receptor-related protein 1 or $\alpha 2$ macroglobulin receptor) has been proposed as a candidate anchoring receptor for CRT.⁶

Cell surface CRT (csCRT) is an important innate immune system receptor.^{7–12} It serves as the signal-transducing receptor for members of the collectin family, including C1q and mannose binding lectin.¹³ Collectins bind foreign organisms or apoptotic cells through their globular heads, while their collagen-like tails bind to csCRT, which leads to phagocytosis. Different from the elimination of foreign organisms, which are events associated with an intense inflammatory reaction,¹⁴ the safe clearance of apoptotic

cells critically depends on suppressing the inflammatory response.¹⁵ The decision of whether a pro- or anti-inflammatory reaction should be activated appears to depend on the presence or absence of a second set of signals that are uniquely triggered by apoptotic cells.¹⁶ Thus, csCRT plays a pivotal role in the junction between tolerance and autoimmunity because of its critical role in elimination of apoptotic cells.¹⁷ Aberrant activation of the CRT-mediated pathway can lead to autoimmunity, as exemplified by conditions that involve defective CRT-mediated clearance of apoptotic cells.¹⁸

Defective clearance of apoptotic cells has long been postulated as a cause of autoantibody production.¹⁸ Noningested apoptotic cells release danger signals and provide self-antigens for aberrant presentation by dendritic cells (DCs). Consistent with this model, both humans¹⁹ and mice²⁰ deficient in C1q develop a SLE-like disease. Donnelly *et al.*¹⁹ have reported impaired recognition of apoptotic neutrophils by the C1q/CRT pathway in patients with SLE. In this juncture, it should be mentioned that CRT itself can be targeted by autoantibodies.²⁰ It has been proposed that antigen-leak from apoptotic cells account for this phenomenon.

Little is known about the role of CRT in rheumatoid arthritis (RA). Although the protein has long been known to be targeted by autoantibodies in RA, the pathogenic significance of that immune recognition is unclear. Moreover, the possibility that the innate receptor function of CRT could play a role has not been addressed. Below we discuss recent evidence strongly implicating csCRT as an innate immune system receptor that may play an important role in the pathogenesis of RA.

The rheumatoid arthritis shared epitope activates innate signaling through cell surface calreticulin

Susceptibility to RA and the severity of the disease are both closely associated with *HLA-DRB1* alleles encoding a five amino acid sequence motif—commonly referred to as the “shared epitope” (SE)—in residues 70–74 of the DR β chain.²¹ SE-encoding *DRB1* alleles confer higher risk of developing RA in most ethnic groups.²² Interestingly, despite the strong influence of genetic factors on RA susceptibility, the concordance rate in monozygotic twins is only 12–15%,²³ suggesting that stochastic environmental or epigenetic factors are required to precipitate disease onset in genetically susceptible individuals. Consistent with this idea, epidemiological studies in Europe²⁴ and North America²⁵ have demonstrated interaction between smoking and the SE: the presence of both factors resulted in a much higher RA risk than the risk conferred by either smoking or SE alone.

In addition to being a risk factor for RA, the SE has also been shown to associate with disease severity.²⁶ The disease in SE-positive individuals is more erosive than in SE-negative individuals.²⁷ In addition, the number of SE-encoding alleles has been found to directly correlate with disease severity.²⁸

The mechanism by which the SE increases susceptibility to—and severity of—RA is unknown. The known role of MHC class II molecules in antigen presentation prompted the prevailing paradigms, which postulate that either presentation of arthritogenic self-peptides,²⁹ molecular mimicry with foreign antigens,³⁰ or T cell repertoire selection³¹ are involved. Notwithstanding their plausibility, these hypotheses are difficult to reconcile with the fact that data-supporting antigen-specific responses as the primary event in RA are inconclusive. In addition, several other human diseases have also been shown

to be associated with SE-encoding *DRB1* alleles, including polymyalgia rheumatica,³² giant cell arteritis,³² type I diabetes,³³ erosive bone changes in psoriatic arthritis,³⁴ autoimmune hepatitis,³⁵ and early-onset chronic lymphoid leukemia,³⁶ among other conditions. The SE has been also shown to be associated with spontaneous RA-like disease in dogs³⁷ and to facilitate collagen-induced arthritis,³⁸ spontaneous diabetes,³⁹ and experimental autoimmune encephalomyelitis⁴⁰ in *HLA-DRB1*0401* transgenic mice. Furthermore, the increasing incidence of RA with age⁴¹ and the positive correlations observed between the number of copies of SE alleles and RA severity²⁸ and disease penetrance⁴² are all inconsistent with an antigen presentation-based mechanism. Thus, taken together, these inconsistencies suggest that the SE may have as yet unknown antigen nonspecific effects independent of the well documented role of class II MHC molecules in antigen presentation. Our recent data reveal that the SE may have a unique ability to activate innate signaling events, which could lead to immune dysregulation.

Given the inadequacies of antigen presentation-based theories discussed earlier, over the past few years we have examined whether the SE, similar to its spatially homologous class I MHC ligands,^{43,44} can trigger innate immune signaling. In all cases, our prior studies have demonstrated that the SE could activate nitric oxide (NO) signaling in a strictly allele-specific manner. The SE was able to do this, whether expressed in its native conformation on the cell surface, as a cell-free HLA-DR tetrameric molecule, engineered into large recombinant proteins, or as a short synthetic peptide. A consensus motif comprising of the ⁷⁰Q/R-K/R-x-x-A⁷⁴ sequence was found to be necessary for triggering the signal.^{45–47}

Mapping of the shared epitope-binding site on calreticulin

We have previously identified csCRT as the cell surface receptor that transduces the SE signaling.⁴⁸ In brief, affinity chromatography purification, cell-binding assays, surface plasmon resonance (SPR), and time-resolved fluorescence resonance energy transfer techniques identified csCRT as the SE-binding molecule. SE-triggered signaling could be blocked (by anti-CRT antibodies or antibodies against CD91) and by CRT-specific

anti-sense or small interfering RNA oligonucleotides. Murine embryonic fibroblasts from *Crt*^{-/-} or *cd91*-deficient mice failed to transduce SE-activated signals. Exogenously added soluble CRT attached to the cell surface and restored signaling responsiveness in *Crt*^{-/-} cells.⁴⁸

More recently, we have mapped the SE-binding site on CRT.⁴⁹ CRT has three domains: an N-terminal domain, a C-terminal domain, and a middle domain called “P-domain.” To identify the domain that binds the SE, we first determined SE-CRT binding by SPR, using domain-selective deletion mutants. These experiments strongly suggested that the SE binding site is located in the P-domain. The role of this domain as a SE-binding region was further confirmed by a photoactive cross-linking technique.⁴⁹ To predict the interacting residues, the BioMedCache 6.1 (Fujitsu, Sunnyvale, CA) *in silico* docking software was used. The ligand and the receptor were modeled on the basis of the published crystal structure of the SE within the native HLA-DR molecule and published P-domain NMR-based structural data, respectively. Four potential docking scenarios with significant energy were identified. Intriguingly, region 217–224 of the CRT P-domain was invariably found to be the SE-binding site in all four docking models, with the most significant roles played by amino acid residues E217, D220, and E223 of the CRT P-domain.

To address the respective role of each one of these candidate SE-binding P-domain residues, point-mutants expressing single amino acid substitutions have been used. Cell-free SPR binding assays where the point-mutated CRT proteins were immobilized on a biosensor chip and the SE peptides were in the analyte, have shown a significantly diminished interaction between the SE and CRT receptors that express E217A or E223A substitutions. Experiments to determine the signal transduction efficiency of these mutant receptors in a cell culture system demonstrated that SE-expressing ligands activated significantly diminished signals when the E217A, D220A, or E223A CRT mutants were used as cell surface receptors, with the most significant role played by CRT P-domain residue E217.⁴⁹ Taken collectively, these data indicates that the SE-binding site on CRT is located in the 217–223 region in the P-domain, with a critical role played by amino acid residue E217. A proposed receptor–ligand interaction model is shown in Figure 1.



Figure 1. CRT–SE interaction. A SE-positive HLA-DR4 molecule interacts with the SE-binding site in the CRT P-domain. The HLA-DRα chain is shown in green, the HLA-DRβ chain is in yellow, the SE is in cyan, the CRT P-domain is in pink, and the groove peptide is shown in orange.

Immune polarization by the shared epitope

As mentioned earlier, one of the hallmarks of SE-induced csCRT-mediated signaling is the production of NO, a ubiquitous signaling molecule with versatile effects in the immune system. In RA, increased NO levels correlate significantly with inflammatory markers of the disease and anti-rheumatic agents have been shown to suppress NO production.^{50–52} NO has also been implicated in the pathogenesis of experimental autoimmune models in mice. For example, SJL mice are known for their NO overproduction.⁵³ These mice are susceptible to many autoimmune diseases,⁵⁴ and their autoimmune tendencies are attributed to their NO overproduction.⁵⁵ Similar to human RA, SJL mice display aging-associated increase in disease incidence,⁵⁶ excessive DNA damage,⁵⁷ higher mutation rates,⁵⁸ and a higher incidence of spontaneous lymphoma.⁵⁹ Thus, there is ample evidence that excessive NO levels may facilitate autoimmunity.

Dendritic cells are professional antigen-presenting cells strategically positioned in the interface between the innate and adaptive immune systems. Relevant to this review, csCRT is expressed on both human and murine DCs.¹² In human DCs, a maturation-dependent expression was found, with more abundant csCRT expression in immature DCs compared to mature cells.⁶⁰ In addition to their role in potentiating adaptive immune responses, DCs also induce tolerance through a variety of mechanisms, including a direct cross talk

with regulatory T (Treg) cells.⁶¹ A growing body of evidence indicates that the tolerogenic effect of DCs is mediated partly by indoleamine 2,3 dioxygenase (IDO), an enzyme that catalyzes the catabolism of tryptophan.⁶²

DCs play a key role in immune regulation and are implicated in the pathogenesis of RA. Therefore, we carried out studies to examine whether the SE can activate innate signaling in murine DCs. Our data clearly demonstrated that, similar to its effect in all other cell lines previously studied, the SE activates NO signaling in murine DCs.⁶³ Given the fact that NO has been previously found to inhibit IDO activity,^{64–66} we have examined whether SE-expressing ligands could affect IDO activity.⁶³ Our data showed that cell-free SE-positive ligands effectively and specifically blocked conversion of tryptophan to kynurenine in DCs and fibroblasts. Moreover, murine L cells expressing SE-positive DR molecules on their surface through cDNA transfection produced significantly less kynurenine in response to IFN γ , compared to transfectants expressing SE-negative DR β chains. Thus, the SE ligand effectively inhibits the activity of the tolerogenic enzyme IDO in both human and murine cells. We have confirmed published observations⁶⁷ that IDO activity is found in DCs expressing the CD8 α surface marker but not in CD8 α^- DCs and found that the SE effect on IDO activity was restricted to that DCs subset.⁶³ While the mechanism of the dichotomy of SE activity in the two subsets is presently unknown, it is worth noting that the CD8 α^+ subset showed higher abundance of the SE signal transducing receptor, csCRT.⁶³

In addition to IDO-mediated regulation, DCs can regulate immune responses by production of various cytokines that can activate or expand particular subsets of T cells, thereby polarizing the immune response.⁶⁸ To determine whether SE-mediated signaling in DCs could induce cytokine production, we have studied supernatants of SE-stimulated DCs. Our data showed that in the CD8 α^- DCs, but not in the CD8 α^+ subset, the SE ligand activated a robust production of IL-6. Although IL-23 levels did not increase following stimulation with the SE alone, in the presence of suboptimal concentrations of LPS, the SE had a prolonged synergistic effect on the production of this cytokine. Other cytokines did not show any increased production, indicating the specificity of SE effect.⁶³

Consistent with the reciprocal effects of IDO and IL-6 on Treg differentiation, we have demonstrated that the SE ligand potently inhibited Treg differentiation (Fig. 2). When DCs were pre-incubated with the SE-positive peptide 65-79*0401 Treg differentiation was significantly inhibited, while SE-negative control peptide 65-79*0402 had no effect. Similar results were obtained using SE-positive versus SE-negative HLA-DR tetramers.⁶³ Thus, the SE ligand has a potent inhibitory effect on Treg.

As mentioned earlier, in CD8 α^- DCs, the SE-activated robust production of two Th17-activating cytokines: IL-6 and IL-23. Th17 cells produce several effector cytokines, which play important roles in inflammation and autoimmunity, including, IL-17A, IL-17F, IL-21, and IL-22.⁶⁸ Over the past few years, the Th17 subset has emerged as a key player both in host defense and autoimmunity.⁶⁹ There is compelling evidence that Th17 cells may play a direct role in the pathogenesis of RA. For example, in RA, both IL-17-producing cells and IL-17 are abundantly expressed in the RA joint compartment.^{70,71} In addition, IL-17 has been shown to induce many factors known to enhance autoimmune or pro-arthritis processes, such as angiogenesis, MMPs production, osteoclastogenesis, leukocyte recruitment, angiogenesis and inflammation, among other effects.⁷² Neutralization of IL-17 prevents disease development in experimental mouse models of autoimmunity.^{73,74}

Consistent with its effect on DCs, the SE was found to have a robust effect on Th17 differentiation and expansion (Fig. 2), as well as activation of proliferative responses and secretion of IL-17.⁶³ Importantly, the SE effect on Th17 polarization could be seen both *in vitro* and *in vivo*.⁶³ Taken together, these findings indicate that the SE ligand, particularly when presented in its native conformation, has a potent Th17-polarizing effect.

A proposed model

Figure 3 shows a proposed model of CRT-mediated SE effect in RA. In physiologic condition, DCs maintain a fine balance between immune stimulation and tolerance. They are programmed to prevent autoimmune reactions by responding to tolerance-enhancing signals delivered either by Th1-derived IFN γ or by CTLA4, expressed on Treg. These signals activate IDO that increases tryptophan metabolites,

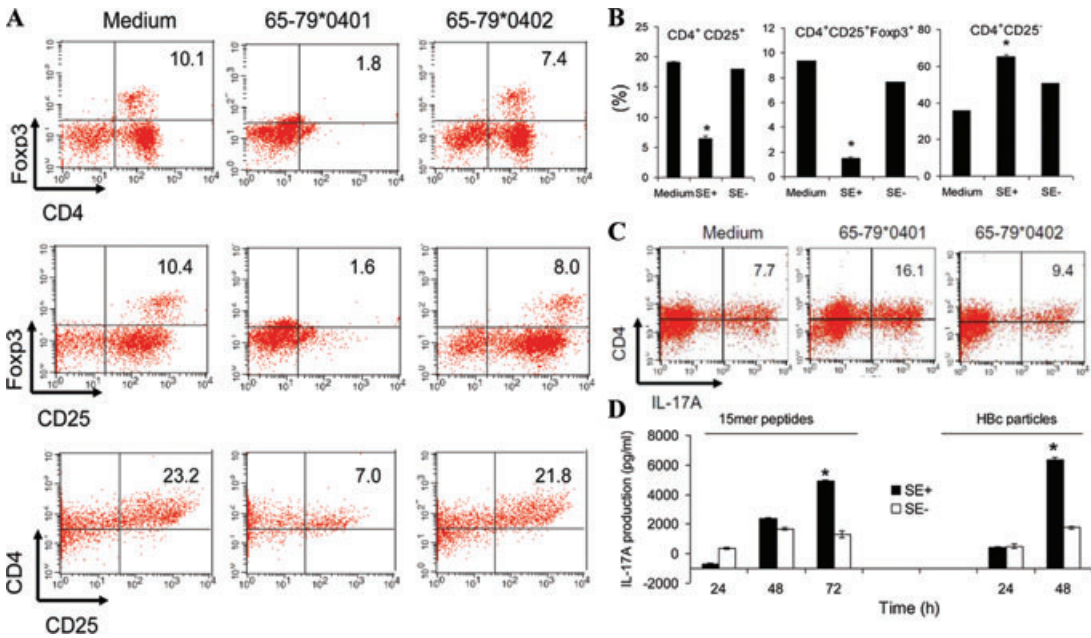


Figure 2. T cell polarization by the SE. (A) DCs were incubated overnight without (medium) or with SE-positive (65-79*0401) or SE-negative (65-79*0402) 15mer peptides. Subsequently, naive CD4 T cells were added and cocultured in the presence of TGFβ and anti-CD3 antibodies. (B) Mean ± SEM values of triplicate flow cytometry determinations of the experiment shown in A. (C) Th17 polarization by the SE. DCs were cocultured with naive CD4 T cells in Th17-differentiating medium in the absence or presence of SE-positive (65-79*0401) or SE-negative (65-79*0402) peptides. (D) DCs were incubated overnight with peptides or recombinant hepatitis B core (HBc) particles containing the SE motif QKRAA (SE+) or a non-SE sequence DERAA (SE-) and then cocultured with naive CD4 T cells as above. IL-17A concentrations in supernatants were determined by ELISA.

which preserve Treg-mediated immune tolerance. The default setting of this intricate system assures a well-balanced immune response and the prevention of autoimmune reactions.

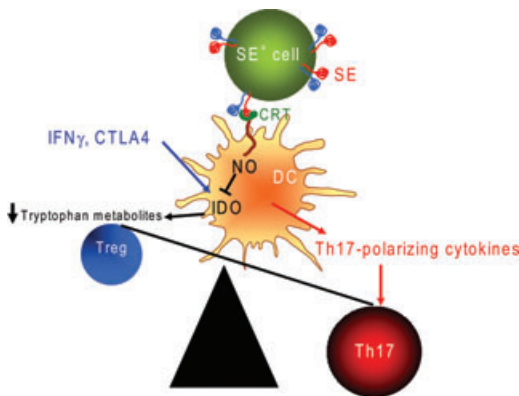


Figure 3. Proposed model. The SE is interacting with csCRT expressed on DCs. As a result of a combined effect of IDO inhibition on the one hand and production of Th17-polarizing cytokines on the other, an immune regulation shift occurs with reduced abundance of Tregs and enhanced Th17 differentiation.

In individuals carrying SE-encoding *HLA-DRB1* alleles, the SE ligand, normally expressed on APC or on lymphocytes, interacts with csCRT. Our model states that in healthy SE-positive individuals SE-csCRT interaction activates low-grade signaling with moderate Th17 polarization, which could be advantageous against pathogens.^{69,75} However, upon exposure to critical levels of environmental pollutants (e.g., cigarette smoke), this beneficial polarization effect could become excessive owing to as yet unknown stochastic events. Such aberrant interaction may lead to excessive Th17 polarization that allows the development of RA. It should be pointed out that excessive Th17 polarization has been previously shown in mice following exposure to environmental aromatic hydrocarbons⁷⁶ and the SE has been shown to synergize with cigarette smoking as a risk factor for RA.^{24,25} It is therefore tempting to entertain a scenario in which the SE and environmental pollutants may cooperatively push Th17 polarization beyond a threshold that allows RA development.

Acknowledgments

Dr. Holoshitz has been supported by the U.S. National Institutes of Health (AR55170, UL1RR02498, AI47331, GM88560, AR20557, AR48310, AR56786), the University of Michigan Global Reach Collaborative Research Fund, by a Basic Research Grant from the Arthritis Foundation, and an Innovative Basic Science Award from the American College of Rheumatology Research and Education Foundation. Special thanks to the NIH Tetramer Core Facility for providing valuable research reagents.

Conflict of interest

The authors declare no conflict of interest.

References

- Gold, L.I. *et al.* 2010. Calreticulin: non-endoplasmic reticulum functions in physiology and disease. *FASEB J.* **24**: 665–683.
- Kuroki, Y., M. Takahashi & C. Nishitani. 2007. Pulmonary collectins in innate immunity of the lung. *Cell Microbiol.* **9**: 1871–1879.
- Obeid, M. *et al.* 2007. Ecto-calreticulin in immunogenic chemotherapy. *Immunol. Rev.* **220**: 22–34.
- Tarr, J.M. *et al.* 2010. Extracellular calreticulin is present in the joints of rheumatoid arthritis patients and inhibits FasL (CD95L) mediated apoptosis of T cells. *Arthritis Rheum.* In press.
- Yan, Q., J.E. Murphy-Ullrich & Y. Song. 2010. Structural insight into the role of thrombospondin-1 binding to calreticulin in calreticulin-induced focal adhesion disassembly. *Biochemistry* **49**: 3685–3694.
- Basu, S. *et al.* 2001. CD91 is a common receptor for heat shock proteins gp96, hsp90, hsp70, and calreticulin. *Immunity* **14**: 303–313.
- Cho, J.H. *et al.* 1999. Activation of human neutrophils by a synthetic anti-microbial peptide, KLKLLLLLKLK-NH₂, via cell surface calreticulin. *Eur. J. Biochem.* **266**: 878–885.
- Gardai, S.J. *et al.* 2003. By binding SIRPalpha or calreticulin/CD91, lung collectins act as dual function surveillance molecules to suppress or enhance inflammation. *Cell* **115**: 13–23.
- Henson, P.M., D.L. Bratton & V.A. Fadok. 2001. The phosphatidylserine receptor: a crucial molecular switch? *Nat. Rev. Mol. Cell Biol.* **2**: 627–633.
- Luo, X. *et al.* 2003. C1q-calreticulin induced oxidative neurotoxicity: relevance for the neuropathogenesis of Alzheimer's disease. *J. Neuroimmunol.* **135**: 62–71.
- McGreal, E. & P. Gasque. 2002. Structure-function studies of the receptors for complement C1q. *Biochem. Soc. Trans.* **30**: 1010–1014.
- Zeng, G. *et al.* 2006. Dendritic cell surface calreticulin is a receptor for NY-ESO-1: direct interactions between tumor-associated antigen and the innate immune system. *J. Immunol.* **177**: 3582–3589.
- Ogden, C.A. *et al.* 2001. C1q and mannose binding lectin engagement of cell surface calreticulin and CD91 initiates macropinocytosis and uptake of apoptotic cells. *J. Exp. Med.* **194**: 781–795.
- Cho, J.H. *et al.* 2001. Activation of human monocyte cell line U937 via cell surface calreticulin. *Cell Stress Chaperones* **6**: 148–152.
- Chung, E.Y., S.J. Kim & X.J. Ma. 2006. Regulation of cytokine production during phagocytosis of apoptotic cells. *Cell Res.* **16**: 154–161.
- Gardai, S.J. *et al.* 2006. Recognition ligands on apoptotic cells: a perspective. *J. Leukoc. Biol.* **79**: 896–903.
- Savill, J. *et al.* 2002. A blast from the past: clearance of apoptotic cells regulates immune responses. *Nat. Rev. Immunol.* **2**: 965–975.
- Pittoni, V. & G. Valesini. 2002. The clearance of apoptotic cells: implications for autoimmunity. *Autoimmun. Rev.* **1**: 154–161.
- Donnelly, S. *et al.* 2006. Impaired recognition of apoptotic neutrophils by the C1q/calreticulin and CD91 pathway in systemic lupus erythematosus. *Arthritis Rheum.* **54**: 1543–1556.
- Eggleton, P. & D.H. Llewellyn. 1999. Pathophysiological roles of calreticulin in autoimmune disease. *Scand. J. Immunol.* **49**: 466–473.
- Gregersen, P.K., J. Silver & R.J. Winchester. 1987. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum.* **30**: 1205–1213.
- Gorman, J.D. *et al.* 2004. Impact of shared epitope genotype and ethnicity on erosive disease: a meta-analysis of 3,240 rheumatoid arthritis patients. *Arthritis Rheum.* **50**: 400–412.
- Silman, A.J. *et al.* 1993. Twin concordance rates for rheumatoid arthritis: results from a nationwide study. *Br. J. Rheumatol.* **32**: 903–907.
- Lundstrom, E. *et al.* 2009. Gene-environment interaction between the DRB1 shared epitope and smoking in the risk of anti-citrullinated protein antibody-positive rheumatoid arthritis: all alleles are important. *Arthritis Rheum.* **60**: 1597–1603.
- Karlson, E.W. *et al.* 2010. Gene-environment interaction between HLA-DRB1 shared epitope and heavy cigarette smoking in predicting incident rheumatoid arthritis. *Ann Rheum Dis.* **69**: 54–60.
- Tureson, C. *et al.* 2005. The impact of HLA-DRB1 genes on extra-articular disease manifestations in rheumatoid arthritis. *Arthritis Res. Ther.* **7**: R1386–R1393.
- Moreno, I. *et al.* 1996. Association of the shared epitope with radiological severity of rheumatoid arthritis. *J. Rheumatol.* **23**: 6–9.
- Gonzalez-Gay, M.A., C. Garcia-Porrúa & A.H. Hajeer. 2002. Influence of human leukocyte antigen-DRB1 on the susceptibility and severity of rheumatoid arthritis. *Semin. Arthritis Rheum.* **31**: 355–360.
- Wucherpfnig, K.W. & J.L. Strominger. 1995. Selective binding of self peptides to disease-associated major histocompatibility complex (MHC) molecules: a mechanism for

- MHC-linked susceptibility to human autoimmune diseases. *J. Exp. Med.* **181**: 1597–1601.
30. La Cava, A. *et al.* 1997. Genetic bias in immune responses to a cassette shared by different microorganisms in patients with rheumatoid arthritis. *J. Clin. Invest.* **100**: 658–663.
 31. Bhayani, H.R. & S.M. Hedrick. 1991. The role of polymorphic amino acids of the MHC molecule in the selection of the T cell repertoire. *J. Immunol.* **146**: 1093–1098.
 32. Weyand, C.M. *et al.* 1994. HLA-DRB1 alleles in polymyalgia rheumatica, giant cell arteritis, and rheumatoid arthritis. *Arthritis Rheum.* **37**: 514–520.
 33. Tait, B.D. *et al.* 1995. HLA-DRB1*0401 is associated with susceptibility to insulin-dependent diabetes mellitus independently of the DQB1 locus. *Eur. J. Immunogenet.* **22**: 289–297.
 34. Korendowych, E. *et al.* 2003. The influence of the HLA-DRB1 rheumatoid arthritis shared epitope on the clinical characteristics and radiological outcome of psoriatic arthritis. *J. Rheumatol.* **30**: 96–101.
 35. Doherty, D.G. *et al.* 1994. Allelic sequence variation in the HLA class II genes and proteins in patients with autoimmune hepatitis. *Hepatology* **19**: 609–615.
 36. Dorak, M.T. *et al.* 1996. Influence of the major histocompatibility complex on age at onset of chronic lymphoid leukaemia. *Int. J. Cancer* **65**: 134–139.
 37. Ollier, W.E. *et al.* 2001. Dog MHC alleles containing the human RA shared epitope confer susceptibility to canine rheumatoid arthritis. *Immunogenetics* **53**: 669–673.
 38. Rosloniec, E.F. *et al.* 1997. An HLA-DR1 transgene confers susceptibility to collagen-induced arthritis elicited with human type II collagen. *J. Exp. Med.* **185**: 1113–1122.
 39. Wen, L. *et al.* 2001. The regulatory role of DR4 in a spontaneous diabetes DQ8 transgenic model. *J. Clin. Invest.* **107**: 871–880.
 40. Forsthuber, T.G. *et al.* 2001. T cell epitopes of human myelin oligodendrocyte glycoprotein identified in HLA-DR4 (DRB1*0401) transgenic mice are encephalitogenic and are presented by human B cells. *J. Immunol.* **167**: 7119–7125.
 41. Goronzy, J.J. & C.M. Weyand. 2005. Rheumatoid arthritis. *Immunol. Rev.* **204**: 55–73.
 42. Jawaheer, D. *et al.* 1994. “Homozygosity” for the HLA-DR shared epitope contributes the highest risk for rheumatoid arthritis concordance in identical twins. *Arthritis Rheum.* **37**: 681–686.
 43. Boyington, J.C. & P.D. Sun. 2002. A structural perspective on MHC class I recognition by killer cell immunoglobulin-like receptors. *Mol. Immunol.* **38**: 1007–1021.
 44. Parham, P. 2005. MHC class I molecules and KIRs in human history, health and survival. *Nat. Rev. Immunol.* **5**: 201–214.
 45. Holoshitz, J. & S. Ling. 2007. Nitric oxide signaling triggered by the rheumatoid arthritis shared epitope: a new paradigm for MHC disease association? *Ann. N.Y. Acad. Sci.* **1110**: 73–83.
 46. Ling, S. *et al.* 2006. Activation of nitric oxide signaling by the rheumatoid arthritis shared epitope. *Arthritis Rheum.* **54**: 3423–3432.
 47. Ling, S. *et al.* 2007. The rheumatoid arthritis shared epitope increases cellular susceptibility to oxidative stress by antagonizing an adenosine-mediated anti-oxidative pathway. *Arthritis Res. Ther.* **9**: R5 1–12.
 48. Ling, S., X. Pi & J. Holoshitz. 2007. The rheumatoid arthritis shared epitope triggers innate immune signaling via cell surface calreticulin. *J. Immunol.* **179**: 6359–6367.
 49. Ling, S. *et al.* 2010. Identification of the rheumatoid arthritis shared epitope binding site on calreticulin. *PLoS ONE.* **5**: e11703.
 50. Grant, D.D. *et al.* 2001. Nitric oxide donors induce large-scale deletion mutations in human lymphoblastoid cells: implications for mutations in T-lymphocytes from arthritis patients. *Environ. Mol. Mutagen.* **38**: 261–267.
 51. Wang, B. *et al.* 2004. Triptolide, an active component of the Chinese herbal remedy Tripterygium wilfordii Hook F, inhibits production of nitric oxide by decreasing inducible nitric oxide synthase gene transcription. *Arthritis Rheum.* **50**: 2995–2303.
 52. Yki-Jarvinen, H., R. Bergholm & M. Leirisalo-Repo. 2003. Increased inflammatory activity parallels increased basal nitric oxide production and blunted response to nitric oxide in vivo in rheumatoid arthritis. *Ann. Rheum Dis.* **62**: 630–634.
 53. Ables, G.P., N. Hamashima & T. Watanabe. 2001. Analysis of genetic factors associated with nitric oxide production in mice. *Biochem. Genet.* **39**: 379–394.
 54. Chiba, A. *et al.* 2004. Suppression of collagen-induced arthritis by natural killer T cell activation with OCH, a sphingosine-truncated analog of alpha-galactosylceramide. *Arthritis Rheum.* **50**: 305–313.
 55. Ding, M. *et al.* 1998. Antisense knockdown of inducible nitric oxide synthase inhibits induction of experimental autoimmune encephalomyelitis in SJL/J mice. *J. Immunol.* **160**: 2560–2564.
 56. Weller, A.H. *et al.* 1997. Spontaneous myopathy in the SJL/J mouse: pathology and strength loss. *Muscle Nerve* **20**: 72–82.
 57. Nair, J. *et al.* 1998. Etheno adducts in spleen DNA of SJL mice stimulated to overproduce nitric oxide. *Carcinogenesis* **19**: 2081–2084.
 58. Gal, A. & G.N. Wogan. 1996. Mutagenesis associated with nitric oxide production in transgenic SJL mice. *Proc. Natl. Acad. Sci. USA* **93**: 15102–15107.
 59. Tamir, S. *et al.* 1995. Nitric oxide production in relation to spontaneous B cell lymphoma and myositis in SJL mice. *Cancer Res.* **55**: 4391–4397.
 60. Schuurhuis, D.H. *et al.* 2006. Ins and outs of dendritic cells. *Int. Arch. Allergy Immunol.* **140**: 53–72.
 61. Paust, S. & H. Cantor. 2005. Regulatory T cells and autoimmune disease. *Immunol. Rev.* **204**: 195–207.
 62. Mellor, A.L. & D.H. Munn. 2004. IDO expression by dendritic cells: tolerance and tryptophan catabolism. *Nat. Rev. Immunol.* **4**: 762–774.
 63. De Almeida, D.E. *et al.* 2010. Immune dysregulation by the rheumatoid arthritis shared epitope. *J. Immunol.* **185**: 1927–1934.
 64. Alberati-Giani, D. *et al.* 1997. Differential regulation of indoleamine 2,3-dioxygenase expression by nitric oxide and inflammatory mediators in IFN-gamma-activated murine macrophages and microglial cells. *J. Immunol.* **159**: 419–426.
 65. Hucke, C. *et al.* 2004. Nitric oxide-mediated regulation of gamma interferon-induced bacteriostasis: inhibition and

- degradation of human indoleamine 2,3-dioxygenase. *Infect. Immun.* **72**: 2723–2730.
66. Thomas, S.R., D. Mohr & R. Stocker. 1994. Nitric oxide inhibits indoleamine 2,3-dioxygenase activity in interferon-gamma primed mononuclear phagocytes. *J. Biol. Chem.* **269**: 14457–14464.
67. Fallarino, F. *et al.* 2002. Functional expression of indoleamine 2,3-dioxygenase by murine CD8 alpha(+) dendritic cells. *Int. Immunol.* **14**: 65–68.
68. McGeachy, M.J. & D.J. Cua. 2008. Th17 cell differentiation: the long and winding road. *Immunity* **28**: 445–453.
69. Awasthi, A. & V.K. Kuchroo. 2009. Th17 cells: from precursors to players in inflammation and infection. *Int. Immunol.* **21**: 489–498.
70. Chabaud, M. *et al.* 1999. Human interleukin-17: a T cell-derived proinflammatory cytokine produced by the rheumatoid synovium. *Arthritis Rheum.* **42**: 963–970.
71. Ziolkowska, M. *et al.* 2000. High levels of IL-17 in rheumatoid arthritis patients: IL-15 triggers in vitro IL-17 production via cyclosporin A-sensitive mechanism. *J. Immunol.* **164**: 2832–2838.
72. Tesmer, L.A. *et al.* 2008. Th17 cells in human disease. *Immunol. Rev.* **223**: 87–113.
73. Morgan, M.E. *et al.* 2003. CD25+ cell depletion hastens the onset of severe disease in collagen-induced arthritis. *Arthritis Rheum.* **48**: 1452–1460.
74. Nakae, S. *et al.* 2003. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. *J. Immunol.* **171**: 6173–6177.
75. Khader, S.A. & A.M. Cooper. 2008. IL-23 and IL-17 in tuberculosis. *Cytokine* **41**: 79–83.
76. Quintana, F.J. *et al.* 2008. Control of T(reg) and T(H)17 cell differentiation by the aryl hydrocarbon receptor. *Nature* **453**: 65–71.