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Design of a Three-Helix Bundle Capable of Binding Heavy Metals in a Triscysteine Environment**

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Supporting Information

α ₃ DIV (μM)	Hg ["] (μM)	рН	ν _Q (GHz)	η	Δω₀/ω₀×100	1/τ _c (μs ⁻¹)	A×100	χ^{2}
200	80	5.8	1.48(2)	0.15(5)	2(2)	18(21)	12(1)	0.63
200	80	7.5	1.48 ^b	0.15 ^b	2 ^b	10(18)	4(1)	0.67
			1.11 ^b	0.40 ^b	6 ^b		11(1)	
200	80	8.6	1.11(2)	0.40(3)	6(2)	0(20)	10(1)	0.62

Table S1. Parameters fitted to the ^{199m}Hg PAC data.^a

^a Numbers in the parenthesis are the standard deviations of the fitted parameters. ^b Fixed in the fit.

Table S2. Parameters fitted to the ¹¹¹	^m Cd PAC data. ^a
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Peptide (C _{Cdll} /C _{peptide})	ω_0 (rad/ns)	η	$\Delta \omega_0 / \omega_0 \times 100$	1/τ _c (μs ⁻¹)	A ×100	χ^2_r
α ₃ D IV (1/12)	0.350(6)	0.0(1)	14(3)	4.5(6)	5(1)	1.36
	0.268(4)	0.18(7)	1(2)	4.5(6)	0.3(3)	
	0.17(2)	0.5(2)	24(3)	4.5(6)	5.8(8)	

^a Numbers in the parenthesis are the standard deviations of the fitted parameters.



Figure S1. A) CD spectrum of α_3 DIV at pH 8.0 and 25°C. The double minima at 222 and 208 nm are representative of α helical structure. Molar ellipticity of –34726 deg cm² dmol⁻¹ res⁻¹ at 222 nm corresponds to a 97% folded structure. **B)** ¹H-¹H NOESY spectrum of α_3 DIV showing chemical shift dispersion characteristic of a well folded α -helical structure.



Figure S2. GuHCl denaturation titration curve of α_3 DIV plotted as a function of concentration of folded protein vs. concentration of GuHCl. The solid line represents fit to the experimental data.



Figure S3. UV/Vis spectra of solutions containing 20 μ M α_3 DIV, 40 μ M TCEP, and 1 equivalent of CdCl₂ at pH 8.0 (dashed line); 30 μ M α_3 DIV; 60 μ M TCEP, and 1 equivalent of HgCl₂ at pH 8.6 (solid line), and 20 μ M α_3 DIV, 40 μ M TCEP, and 1.0 equivalent of PbCl₂ at pH 8 (dotted line). Shown is a plot of normalized absorbance vs. wavelength.



Figure S4. pH dependence of the binding of 1 equivalent of Cd^{II} (open circles), Hg^{II} (squares), and Pb^{II} (filled circles) to 20 μ M, 30 μ M, and 20 μ M α_3 DIV, respectively, along with the fits of the experimental data. Experiments were followed by monitoring UV/Vis absorbance due to LMCT bands at 232, 247, and 236 nm for Cd^{II}, Hg^{II}, and Pb^{II}, respectively. Cd^{II} and Pb^{II} titration data were fit to the model simultaneous dissociation of two Cys thiol protons; Hg^{II} was fit to a single 1-H step corresponding to the formation of HgS₃ from HgS₂(SH).



Figure S5. Fourier transformed ^{199m}Hg PAC spectra of α_3 DIV (faint lines) under different conditions along with the Fourier transformed fits (dark lines). Sample conditions were A) 200 μ M α_3 DIV, 80 μ M HgCl₂, 100 mM phosphate buffer at pH 5.8, B) 200 μ M α_3 DIV, 80 μ M HgCl₂, 100 mM phosphate buffer at pH 7.4, and C) 200 μ M α_3 DIV, 80 μ M HgCl₂, 100 mM CHES buffer at pH 8.6.



Figure S6. ¹¹³Cd NMR spectrum of 3 mM α_3 DIV with 0.8 equivalents of ¹¹³Cd(NO₃)₂ recorded at pH 8.



Figure S7. Fourier transformed ^{111m}Cd PAC spectrum of 300 μ M α_3 DIV loaded with 1/12 equivalents of Cd^{II} (faint line) along with the Fourier transformed fits (dark line). Sample was prepared in 20 mM TRIS buffer at pH 8.1. The data analysis of this spectrum relied on identification of typical signals from CdS₃O species reported in the literature.^[37]