

Supplemental Data

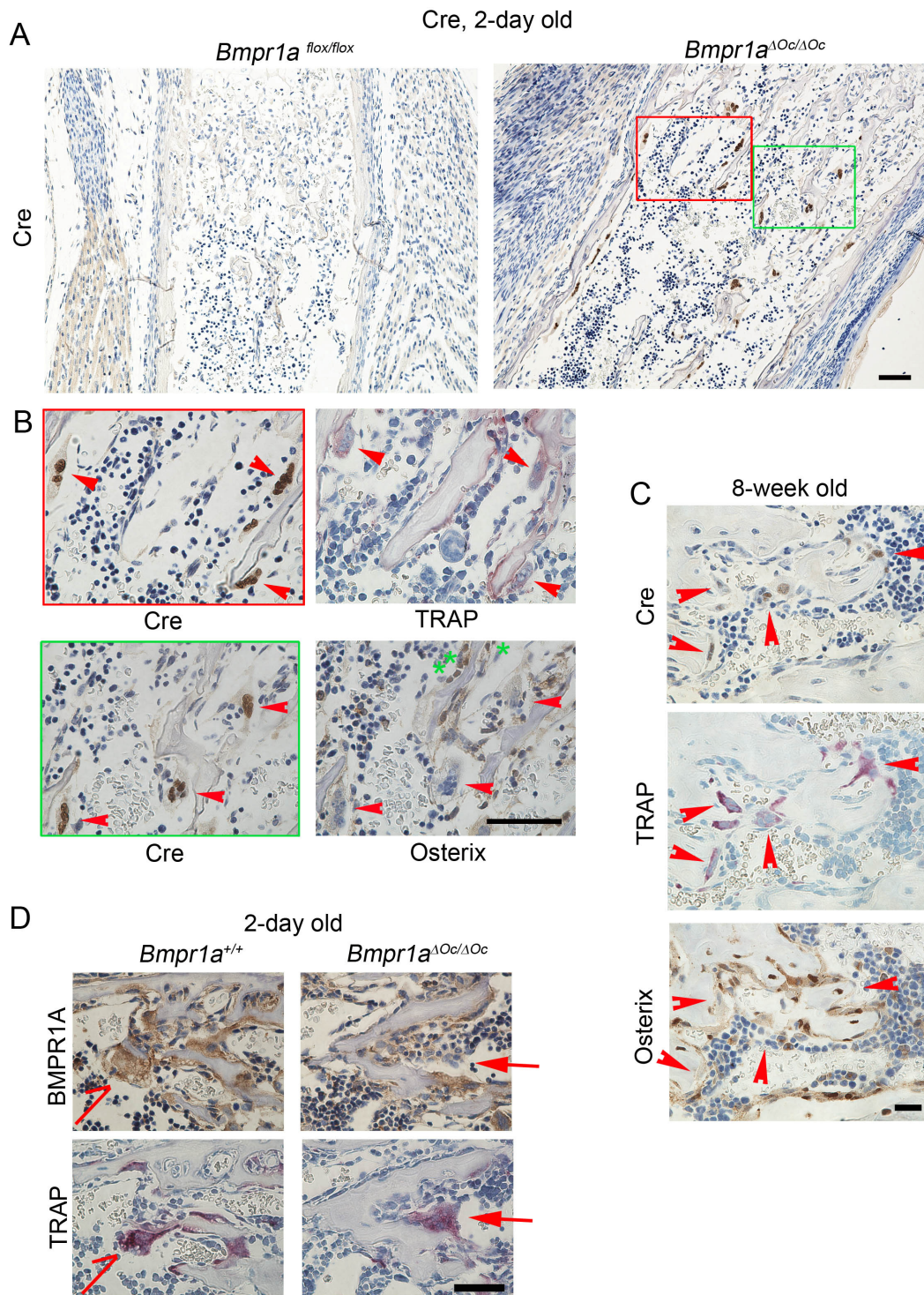
Conditional deletion of *Bmpr1a* in differentiated osteoclasts increases osteoblastic bone formation, increasing volume of remodeling bone in mice

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Supplemental Table 1. Primers used in real-time RT-PCR.

Primer	Sequence
<i>Trap</i> S	GGCCGGCCACTACCCCATCT
<i>Trap</i> AS	CACCGTAGCGACAAGCAGGACTCT
<i>Mmp9</i> S	GCCCTACAGCGCCCCCTACT
<i>Mmp9</i> AS	AGACACGCCCTTGCTGAACA
<i>Rank</i> S	TTCGGCGTTTACTACAGGA
<i>Rank</i> AS	CATTGACCCAATTCCACAAA
<i>Gapdh</i> S	GAGATGATGACCCTTTTGGCT
<i>Gapdh</i> AS	TCAAGGCCGAGAATGGGAAG
<i>Cathepsin K</i> S	AGGGCCA ACTCAAGAAGAAA ACT
<i>Cathepsin K</i> AS	TGCCATAGCCCACCACCAACT
<i>Nfatc1</i> S	CAACGCCCTGACCACCGATA
<i>Nfatc1</i> AS	GGCTGCCTTCCGTCTCATAG

Supplemental Figure 1



Supplemental Fig. 1. Expression patterns of Cre and BMPR1A in tibia.

(A) Histological sections of tibia of *Bmpr1a^{lox/lox}* and *Bmpr1a^{ΔOc/ΔOc}* mice at age 2 days were immunostained using anti-Cre antibodies.

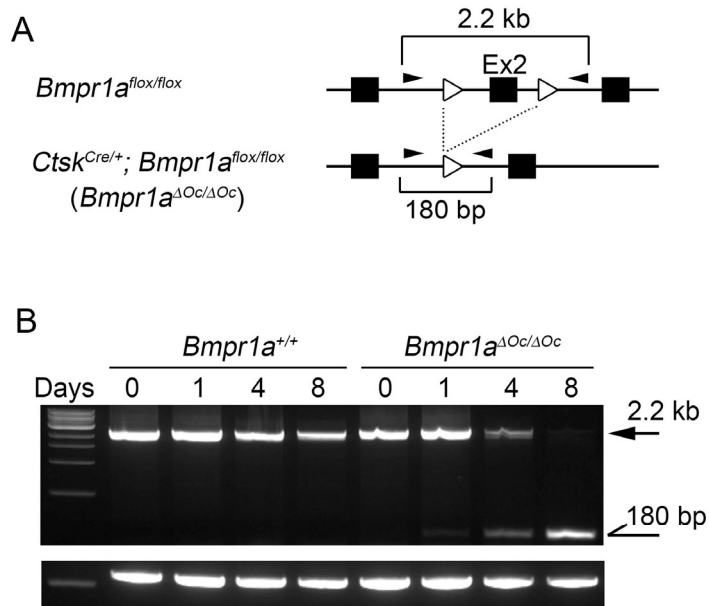
(B) *Top*, Magnification of red boxed region in (A) is shown (*left*) and serial sections were subjected to TRAP staining (*right*). *Bottom*, Magnification of green boxed region in (A) is shown (*left*) and serial sections were subjected to immunostaining with anti-Osterix antibody (*right*). Arrowheads indicate Cre-expressing TRAP-positive cells. Asterisks indicate Osterix-expressing cells.

(C) Serial sections of trabecular bone of proximal tibia of *Bmpr1a^{ΔOc/ΔOc}* mice at 8 weeks of age were immunostained with anti-Cre and anti-Osterix antibodies, and subjected to TRAP staining. Arrowheads indicate Cre-expressing TRAP-positive cells.

(D) Serial sections of trabecular bone of proximal tibia of *Bmpr1a^{+/+}* and *Bmpr1a^{ΔOc/ΔOc}* mice at age 2 days were immunostained with anti-BMPR1A antibody and subjected to TRAP staining. Half-arrows and arrows indicate multinucleated osteoclasts.

Scale bars: 50 μm

Supplemental Figure 2



Supplemental Fig. 2. Genomic PCR analysis of recombinations at loxP sites during induction of osteoclasts *in vitro*. Bone marrow macrophage cultures prepared from *Bmpr1a^{+/+}* and *Bmpr1a^{ΔOc/ΔOc}* mice with M-CSF were further cultured in the presence of M-CSF and RANKL for the indicated number of days. Genomic DNAs were extracted and subjected to PCR amplification.

(A) Primer locations for amplification of *Bmpr1a* gene fragments. *Closed boxes*, exons; *Ex2*, 2nd exon; *open triangles*, loxP sequences; *arrowheads*, primer pair fx1 and fx4.

(B) PCR products separated on the agarose gel. *Top*, PCR products with sizes around 2.2 kb (arrow) were amplified from *Bmpr1a^{+/+}* and *Bmpr1a^{flx/flx}* genomic DNA. After excision by Cre recombinase, PCR products of 180 bp (half-arrow) were amplified from *Ctsk^{Cre/+};Bmpr1a^{flx/flx}* DNA (*Bmpr1a^{ΔOc/ΔOc}*). *Bottom*, RAPSIN gene fragment (590 bp) was amplified as an internal control.