

Possible promoter sequences for *dmmA*

Consensus sequence in parenthesis

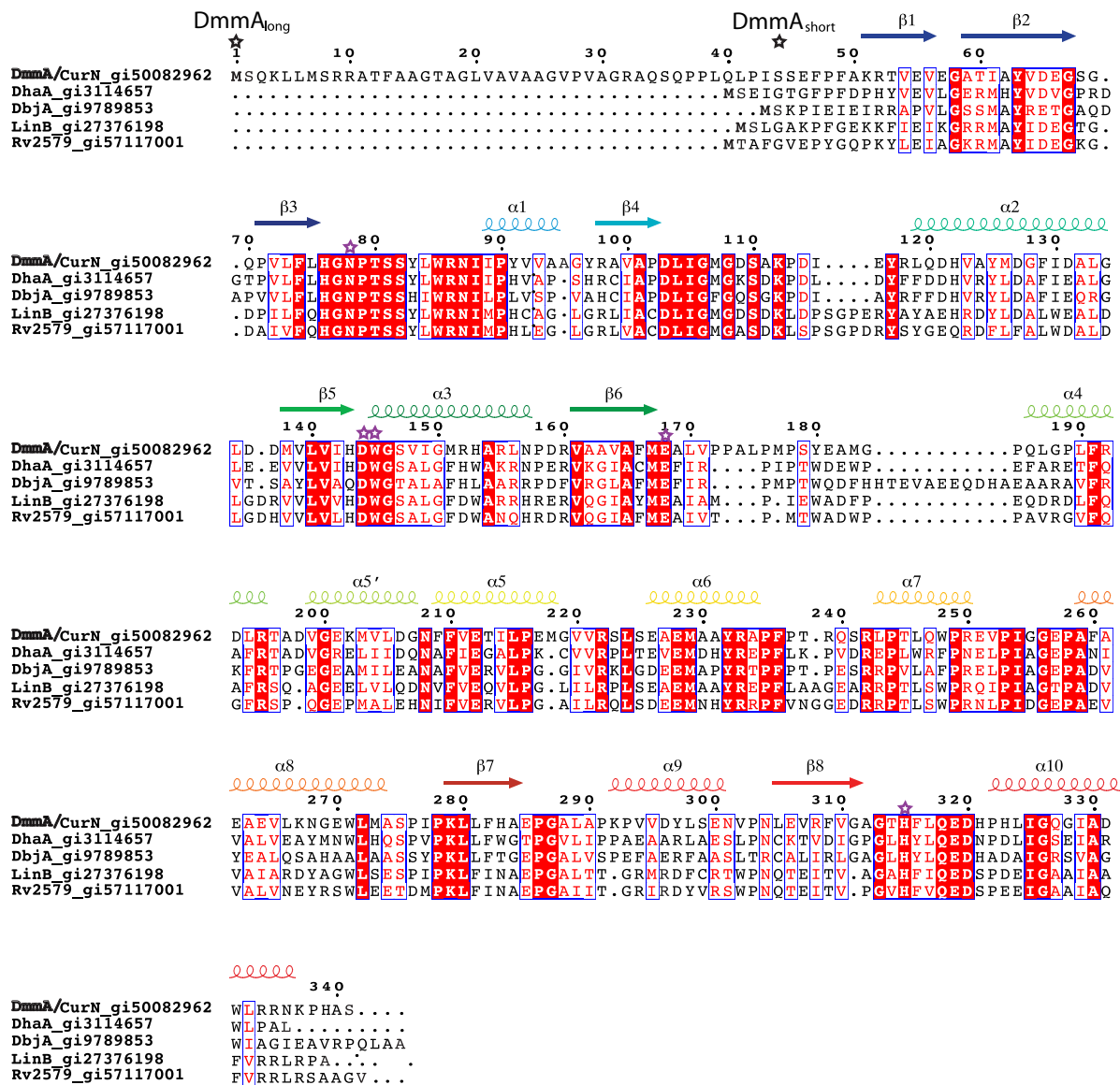
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aactgcatcatcggaacgggcctctcactggtatcgtttacgcaccgaggcggcggaatccgaccg

**-35 promoter** (TTGACA) **-10 promoter** (TATAAT) **TSS motif** (GATCGAAT)  
**TTCCGA**cagagacgtcgacttcc**TACGTAT****GATCGCGA**aggcgcgcatggtgcgcgccctgtcggg  
|← 17 bp spacer →|

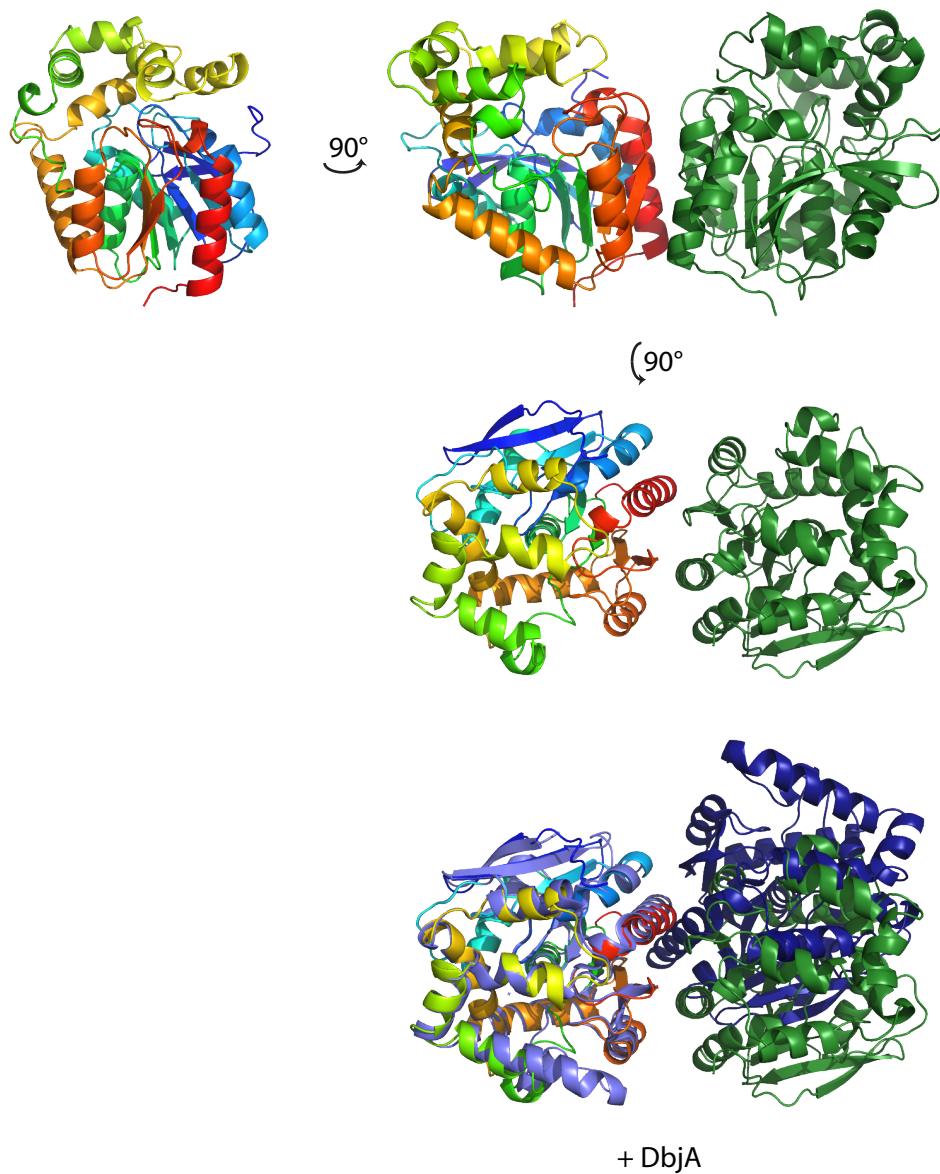
**RBS** **Translation start**  
t**GAGGA**aattca**ATG**tcgcaaaaacttctaagtgcgcgccgtgccacatttgcagctggaacggccggtct  
|← 6 bp spacer →|

ggttgccgtggcggccggtgtgcccgggttgcgggcccgggccagtcgcagccgcccgttcaactgcccgatct  
cgtcggaaatccattcgccaaaaggacggtcgaggtcgagggggcgacaatcgcctatggtgacgagggc  
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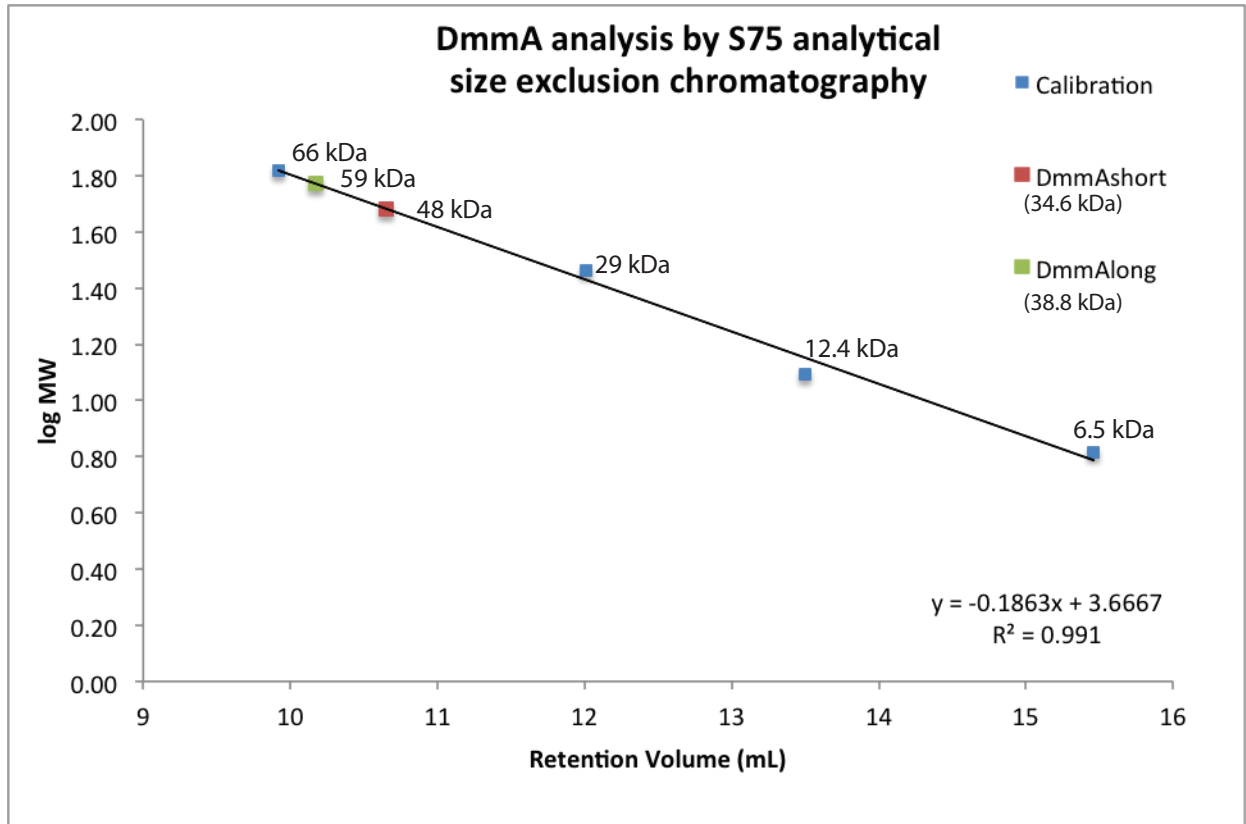
**Supplemental Figure 1:** Proposed sequence motifs and spacers similar to prokaryotic  $\sigma^{70}$  promoter and ribosome binding sequences upstream of the *DmmA* coding sequence (1). The -35 and -10 promoter regions, transcription start site (TSS), ribosome binding site (RBS) and translation start site are indicated in bold (consensus in parentheses) and labeled in red.



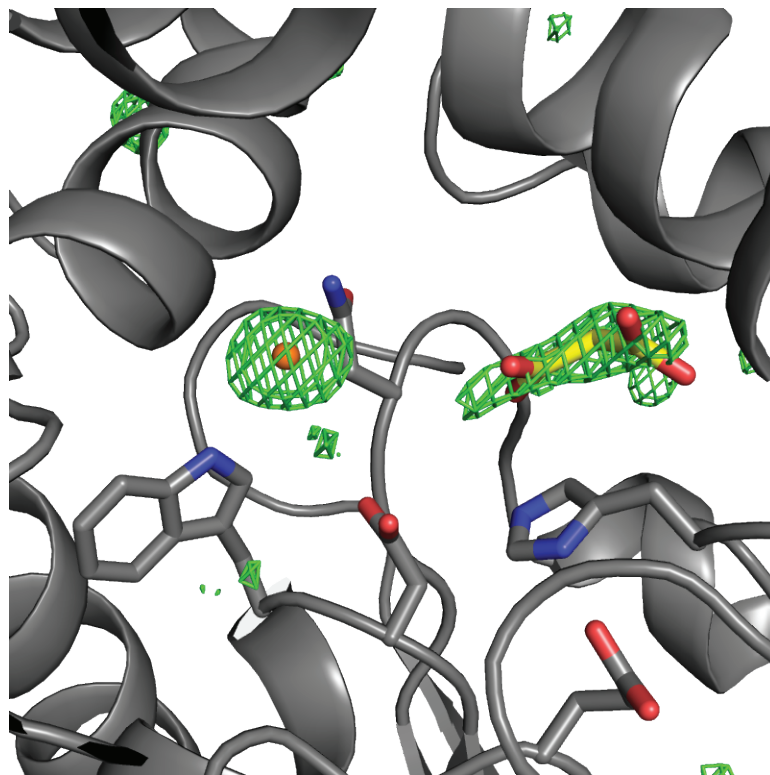
**Supplemental Figure 2:** Sequence alignment of DmmA to other subfamily II HLDs of known structure. DmmA is aligned with DhaA 1CQW (2), DbjA 3A2M (3), LinB 1IZ8 (4), and Rv2578 2O2I (5). PSI-COFFEE (6) was used to generate the alignment, ESPript (7) to prepare the figure, and the STRIDE server (8) to assign secondary structure. Black stars mark the N-termini of DmmA<sub>long</sub> and DmmA<sub>short</sub> and purple stars mark the catalytic pentad.



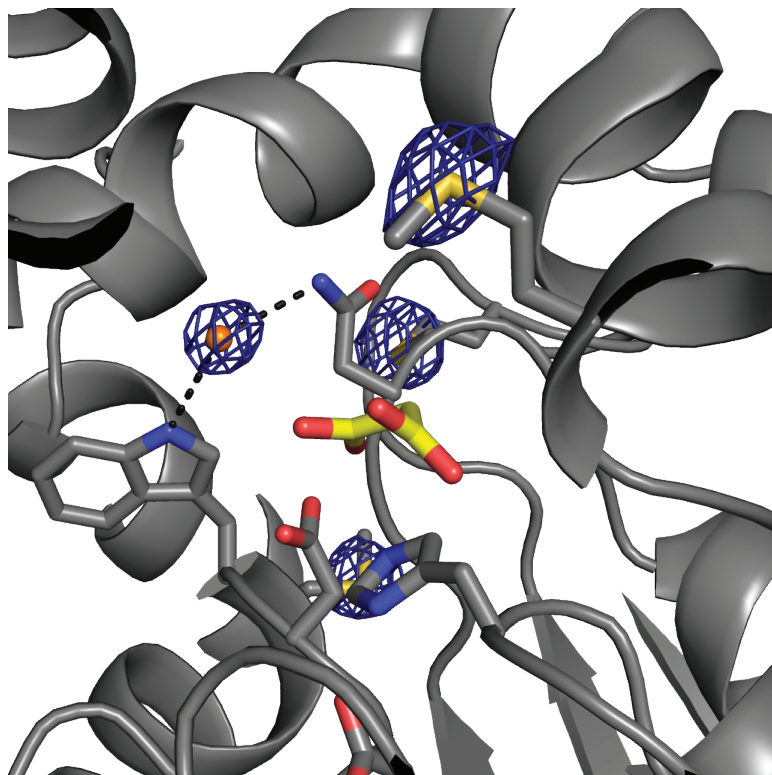
**Supplemental Figure 3:** Comparison of DbjA dimer interface and DmmA protein-protein contact. DmmA molecules are in rainbow and green and DbjA is in light and dark purple. The same side of DmmA and DbjA forms the contact, but the interactions differ substantially.



**Supplemental Figure 4:** Analysis of DmmA oligomeric state by analytical size exclusion. DmmA<sub>short</sub> and DmmA<sub>long</sub> (15 mg/mL) were eluted from a Superdex 75 10/300 GL (GE Healthcare) analytical size exclusion column, and compared against standards of known molecular weight and oligomeric state. DmmA<sub>long</sub> and DmmA<sub>short</sub> both elute with an apparent molecular weight (DmmA<sub>short</sub>: 48 kDa, DmmA<sub>long</sub>: 59 kDa) between that of the monomer (DmmA<sub>short</sub>: 34.6 kDa, DmmA<sub>long</sub>: 38.8 kDa) and the dimer (DmmA<sub>short</sub>: 69.2 kDa, DmmA<sub>long</sub>: 77.6 kDa). The elution volumes did not shift when the protein concentrations were decreased 4-fold.



**Supplemental Figure 5:** Simulated annealing omit map ( $F_o-F_c$ ) contoured at 3.5 sigma generated with  $\text{Cl}^-$  and malonate removed from the atomic model.



**Supplemental Figure 6:** Anomalous difference Fourier map using Friedel data collected at the bromine edge. A crystal of SeMet DmmA<sub>short</sub> was soaked with 1,5-dibromopentane and data were collected at 13.4761 keV. This X-ray energy is 3 eV above the theoretical Br K absorption edge and 800 eV above the Se K edge. The anomalous difference Fourier map contoured at 5.0 sigma shows density for Br<sup>-</sup> and for Se atoms in three SeMet side chains.

## References:

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