### **REVIEW ARTICLE**

# The B7 Homologues and their Receptors in Hematologic Malignancies

## Ryan A. Wilcox<sup>1</sup>, Stephen M. Ansell<sup>2</sup>, Megan S. Lim<sup>3</sup>, Weiping Zou<sup>4</sup>, Lieping Chen<sup>5</sup>

<sup>1</sup>Department of Internal Medicine, Division of Hematology/Oncology, University of Michigan, Ann Arbor, MI; <sup>2</sup>Department of Internal Medicine, Division of Hematology, Mayo Clinic, Rochester, MN; <sup>3</sup>Department of Pathology, University of Michigan, Ann Arbor, MI; <sup>4</sup>Department of Surgery, University of Michigan, Ann Arbor, MI; <sup>5</sup>Department of Immunobiology and Yale Comprehensive Cancer Center, Yale University, New Haven, CT, USA

#### Abstract

The B7 homologues and their receptors regulate both peripheral tolerance and adaptive immunity. This field is rapidly evolving as new ligands and receptors are being identified. Much of the work supporting their role in the regulation of host anti-tumor immunity has been derived from experimental models and clinical trials in solid malignancies. However, a growing body of evidence demonstrates that the B7-H family has important immunologic and non-immunologic functions in a variety of hematologic malignancies. Herein, we will review recent evidence that supports the therapeutic targeting of the B7 homologues in hematologic malignancies.

Key words leukemia; lymphoma; B7-H1; PD-1; T cell; costimulation

**Correspondence** Ryan Wilcox, MD, PhD, Division of Hematology/Oncology, University of Michigan, 1500 E. Medical Center Drive, Room 4310 CC, Ann Arbor, MI 48109-5948, USA. Tel: +1 734 764 8100; Fax: +1 734 936 7376; e-mail: rywilcox@med.umich.edu

Accepted for publication 31 January 2012

doi:10.1111/j.1600-0609.2012.01766.x

The 'classical' B7 family members, B7-1 (CD80) and B7-2 (CD86), provide T-cell costimulatory or coinhibitory signals upon binding their receptors CD28 or CTLA-4, respectively [reviewed in (1)]. Over the past decade, a number of ligands/counter-receptors with homology to B7-1/B7-2 have been identified. These B7 homologues ('B7-H'), including B7-H1 (CD274 or PD-L1), B7-H2 (CD275 or ICOS-L), B7-H3 (CD276), B7-H4 (B7S1 or B7x), B7-H6, B7-DC (CD273 or PD-L2), and BTLN2, play a critically important role in the maintenance of self-tolerance and the regulation of innate and adaptive immunity in the tumor-bearing host (Fig. 1). Many of these B7-H family members are exploited by tumor cells to escape and suppress host immunity and are being actively investigated as therapeutic targets in solid tumors. While a comprehensive discussion of the B7-H family is beyond the scope of this review, recently published reviews provide an excellent overview of the B7-H family, emphasizing their role in solid tumor immunity (2–6). Instead, we hope to highlight areas that are of particular relevance in hematologic malignancies.

#### The B7-H family: a primer

The rapid expansion of the B7-H family coupled with improved understanding of their immunologic functions has changed the view of T-cell co-stimulation within the context of the 'two-signal hypothesis' (7, 8). Simply stated, the 'two-signal hypothesis' postulates that combined antigen-dependent signaling via the T-cell receptor and a second 'costimulatory' signal are required for optimal T-cell activation. In this view, costimulatory ligands function as molecular 'toggle' switches. It is now apparent that B7-H family members not only influence the activation of naïve T cells following antigen presentation, but also have broad roles in the control of T-cell differentiation, effector functions, deactivation, and survival. Therefore, members of the B7-H family are molecular 'dimmer' switches that fine-tune adaptive (and innate) immunity.

B7-H family members with either stimulatory or inhibitory functions are widely expressed by both tumor cells and constituents of the tumor microenvironment in hematologic malignancies (Table 1)(9–24). B7-H2 provides a T-cell costimulatory signal (25–29) upon engaging

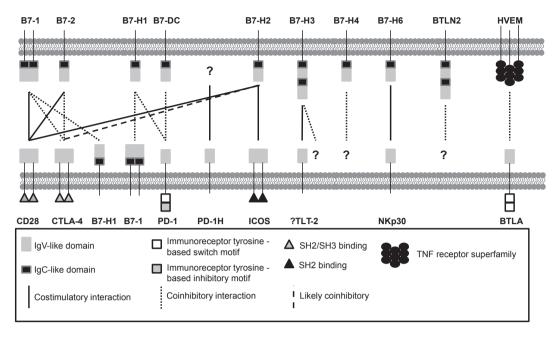


Figure 1. The B7-H family and their receptors.

either of its receptors, CD28 and ICOS (25, 30). In contrast, B7-H1 (2) and B7-H4 (31–33) both inhibit T-cell immunity. The available evidence suggests both inhibitory and stimulatory functions for B7-H3 (6). Whether the triggering receptor expressed on myeloid cells family member TLT-2 is a bona fide costimulatory receptor for B7-H3 is also controversial (34–36). The identification of B7-H3 receptors using novel techniques should help clarify its role in immunity (30). Ultimately, the B7 ligands exert disparate effects on T-cell immunity depending upon the particular receptor/counter-receptor engaged (Fig. 1; B7-1/CD28 cf. B7-1/CTLA-4/B7-H1).

With the exception of CTLA-4, the B7-H1/PD-1/B7-1 axis is best understood among the inhibitory B7-H family members. In contrast to CTLA-4-deficient mice that develop a profound lymphoproliferative disorder, PD-1deficient mice, depending upon their genetic background, develop a variety of more subtle autoimmune manifestations (37-39). This work clearly supports the role of PD-1 as an important inhibitory receptor. The striking difference in phenotypes between CTLA-4- and PD-1-deficient mice suggests that targeting PD-1 may be associated with fewer immune-related adverse events compared with CTLA-4 blockade. While it may be premature to draw any definitive conclusions, the available evidence supports this conclusion (40-42). Recruitment of SH2-domain containing protein tyrosine phosphatases (SHP-1 and/or SHP-2) to the immunoreceptor tyrosinebased switch motif (ITSM) within the PD-1 cytoplasmic tail inhibits signaling events, particularly PI3K/AKT activation, downstream of the T-cell receptor (43). PD-1

demonstrate that B7-1 engagement by B7-H1 downregulates T-cell immunity (44). Engagement of B7-H1 on tumor cells confers resistance to cytotoxic T-cell (CTL)mediated killing, thus functioning as a 'molecular shield' (43). In the setting of persistent antigenic exposure, PD-1 engagement contributes to the maintenance of functionally incompetent antigen-specific T cells (45). This has been nicely demonstrated in chronic viral infections, like HIV. In most patients with HIV, viral-specific CTL highly express PD-1 and are functionally suppressed or 'exhausted' (46, 47). However, T-cell effector functions and proliferation are restored upon B7-H1 blockade. In contrast, a low viral load and maintenance of peripheral blood CD4<sup>+</sup> T-cells are observed in the absence of antiviral therapies in a subset of HIV patients. These longterm non-progressors have a pool of functional HIV-specific T cells that express little PD-1. These observations have significant implications for virally associated lymphoproliferative disorders. Adult T-cell leukemia/lymphoma (ATLL) is associated with human T-cell leukemia virus-1 (HTLV-1) infection, but only develops in a minority of infected patients after a long latency period. In contrast to HTLV-1 carriers, antigen-specific T cells in ATLL patients highly expressed PD-1 and are suppressed by B7-H1-expressing lymphoma cells (20). The contribution of the B7-H1/PD-1 axis in the maintenance of T-cell anergy or exhaustion is unlikely to be limited to virally associated lymphoproliferative disorders (48).

also promotes tumor cell evasion of host immunity as a

major counter-receptor for B7-H1. Recent studies also

	Tumor Cell B7-H Expression (%)						
	B7-H1	B7-H2	B7-H3	B7-H4			
MDS	Present (<25%) (9)						
AML	≈25% (10–13)	$pprox$ 15% $^{\dagger}$ (21)					
ALL			Absent (23)				
DLBCL	≈25% (14)		≈5% (24)				
PMBCL	>50% (16)						
BL			Rare (23, 24)				
FL	Absent (14)		Rare (24)				
CLL/SLL			Rare (24)				
MCL			Absent (24)				
CBCL				Present (131			
cHL	≈90% (14, 15)		≈5% (24)				
NLPHL			Absent (24)				
MM	>50% (17, 49)	Rare (22)					
PTCL-U	≈20% (18)						
AITL	Rare (18)						
ALCL, ALK+	33% (18)		pprox20% (ALK status unknown) (24)				
ALCL, ALK-	15% (18)						
ATLL	≈20% (20)						
CTCL	≈25% (18, 19)						

Table 1 Expression of selected B7 homologues in hematologic malignancies	Table 1	Expression	of	selected	B7	homologues	in	hematologic	malignancies
--------------------------------------------------------------------------	---------	------------	----	----------	----	------------	----	-------------	--------------

MDS, myelodysplastic syndrome; AML, acute myelogenous leukemia; ALL, acute lymphoblastic leukemia; DLBCL, diffuse large B-cell lymphoma; PMBCL, primary mediastinal large B-cell lymphoma; BL, Burkitt lymphoma; FL, follicular lymphoma; CLL/SLL, chronic lymphocytic leukemia/small lymphocytic lymphoma; MCL, mantle cell lymphoma; CBCL, cutaneous B-cell lymphoma; cHL, classical Hodgkin lymphoma; NLPHL, nodular lymphocyte-predominant Hodgkin lymphoma; MM, multiple myeloma; PTCL-U, peripheral T-cell lymphoma, unspecified; AITL, angioimmunoblastic lymphoma; ALCL, anaplastic large cell lymphoma; ALK, anaplastic lymphoma kinase; ATLL, adult T-cell leukemia/lymphoma; CTCL, cutaneous Tcell lymphoma.

<sup>†</sup>Associated with survival.

In addition to regulating T-cell immunity, B7-H family members have recently been shown to influence NK-cell activation. This observation is particularly relevant in hematologic malignancies given the widespread use of monoclonal antibodies that mediate antibody-dependent, NK-cell-mediated cytotoxicity (ADCC). NK cells derived from multiple myeloma patients, in contrast to healthy donors, highly express PD-1. As myeloma cells highly expressed B7-H1, PD-1 blockade enhanced conjugate formation between NK cells and myeloma cells, increasing NK-cell-mediated cytotoxicity (49). Interestingly, lenalidomide decreased myeloma cell B7-H1 expression. Therefore, lenalidomide may increase ADCC when used in conjunction with monoclonal antibody-based therapies by directly stimulating NK-cell cytotoxicity and by impairing B7-H1 expression on target cells (50, 51). Analogous to its role in maintaining a state of T-cell exhaustion, PD-1 was found to mediate NK-cell 'exhaustion' in EBV-associated post-transplant lymphoproliferative disorders (52). In contrast to B7-H1, B7-H2 and B7-H6 promote NK-cell cytotoxicity (53-56).

The role of the B7-H family is not limited to direct interactions between malignant cells and effector cells of the adaptive immune response. B7-H family members expressed on non-malignant cells within the tumor

© 2012 John Wiley & Sons A/S

microenvironment play a critically important role in providing a niche favorable for tumor growth and survival in hematologic malignancies. For example, in various T-cell lymphoproliferative disorders B7-H1 expression was far more common within the tumor microenvironment and was less commonly expressed by clonal T cells (18). A considerable body of work now supports the important role of both myeloid-derived cells and regulatory T cells (Treg) in many hematologic malignancies. Monocytes and their progeny within the tumor microenvironment (e.g., lymphoma-associated macrophages) provide trophic factors that support the growth of malignant lymphocytes, promote angiogenesis, and regulate host immunity (57). Not surprisingly then, increased frequencies of peripheral blood monocytes and lymphoma-associated macrophages are associated with inferior outcomes in both Hodgkin and non-Hodgkin lymphomas (58-61). Dendritic cells (DC) and other myeloid cells, including subsets of monocytic cells (18, 62, 63), determine the fate of activated T cells, at least in part, by the provision of costimulatory/coinhibitory ligands. For example, T-cell lymphoma cells were found to stimulate B7-H1 expression on both monocytes and DC (18). In vitro co-culture experiments demonstrated that DC promote Treg generation in a B7-H1-dependent

manner (18, 64). This was further supported by immunohistochemical analyses demonstrating the co-localization of B7-H1<sup>+</sup> DC and Treg in T-cell lymphoma specimens (18). While these studies support the role of B7-H1 in Treg induction, B7-H1 may also promote the expansion of Treg (65). Regulatory T cells have also been implicated in the pathogenesis of MDS/AML (66, 67) and B-cell malignancies (68, 69). B7-H1, expressed by either malignant cells or tumor-infiltrating DC, appears to similarly regulate the induction and suppressive functions of Treg in these diverse hematologic malignancies (69–72).

Given their inhibitory role, expression of many B7-H family members by tumor cells is an adverse prognostic factor in most solid tumors that have been examined (4-6). This is usually attributed to their immunologic affects. However, recent evidence demonstrates that 'reverse signaling' mediated by B7-H1, B7-H3, and B7-H4 may regulate tumor cell survival. Using tumor cells transfected with B7-H1 constructs lacking the cytoplasmic domain, Azuma et al. demonstrated that B7-H1 signaling following engagement by T-cell PD-1 led to resistance to CTL-mediated killing (73). A more generalized role for B7-H1 in regulating tumor cell survival is demonstrated by the observation that these cells were similarly resistant to Fas- or chemotherapy-mediated apoptosis. In a murine model, B7-H1 increased the survival of CD8<sup>+</sup> T cells during the contraction phase of the immune response by upregulating Bcl-x<sub>1</sub> expression (74). Silencing of B7-H3 or B7-H4 expression have similarly been shown to increase tumor cell apoptosis (75, 76). Therefore, these ligands promote tumorigenesis by both immunologic and non-immunologic affects. Not surprisingly then, malignant cells inducibly express members of the B7-H family by a variety of different mechanisms.

#### **B7-H expression in hematologic malignancies**

Given their important immunoregulatory roles, B7-H expression is strictly regulated. For example, B7-H1 is post-transcriptionally regulated by type I and II interferons and by MyD88, JAK/STAT, TRAF6, MEK, and PI3K/AKT signaling in solid tumors (2). While B7-H1 may be similarly regulated in hematologic malignancies (9, 10, 17), both anaplastic large cell lymphoma (ALCL) and primary mediastinal large B-cell lymphoma (PMBCL) regulate the expression of B7-H family members by novel mechanisms, including chromosomal translocations and gene amplification.

A subset of ALCL harbor a novel nucleophosmin (NPM) – anaplastic lymphoma kinase (ALK) fusion protein resulting from a chromosomal translocation [most commonly t(2;5)(p23;q35)]. Constitutively active NPM-ALK is oncogenic and culminates in the activation of

multiple signaling pathways, including STAT3. In ALK<sup>+</sup> ALCL, B7-H1 expression was strictly dependent upon NPM-ALK expression and activity and was attributed to NPM-ALK-dependent activation of STAT3 and its subsequent binding to the B7-H1 promoter (77). In similarly performed experiments, STAT3 was shown to bind the ICOS promoter and inhibit the expression of miR-219, culminating in ICOS expression (78). Whether NPM-ALK further regulates B7-H1 expression in a miRNA-dependent manner is unknown (79). In addition, exposure to a hypomethylating agent further increased ICOS expression owing to its effects on the putative ICOS enhancer, which is methylated in ALK<sup>+</sup> ALCL cell lines (78).

Genomic amplifications involving chromosome 9 (9p24.1) are a characteristic finding observed in over 50% of PMBCL (80). This subtelomeric region of chromosome 9 includes the genes for B7-H1 and B7-DC. Integrative analyses have clearly demonstrated that amplification at these loci results in increased expression of B7-H1 and B7-DC in PMBCL (16). Furthermore, JAK2 is also involved in this amplification and further enhances B7-H1 gene transcription in PMBCL (16). It remains to be determined whether JAK2 amplifications or mutations observed in other malignancies are associated with the induction of B7-H1 expression (81). Alternatively, recurrent translocations involving the major histocompatibility complex class II transactivator (CIIT-A) and the B7-H1 and B7-H2 loci are associated with overexpression of these ligands in PMBCL and classical Hodgkin lymphoma (82). These observations raise the possibility that the B7-H family may bridge oncogenic events driving tumor proliferation with the suppression of host immunity.

In contrast to ALCL and PMBCL, B7-H expression in other hematologic malignancies may be explained by derivation from a cell subset that normally expresses specific B7-H family members. Therefore, B7-H expression provides important clues about the 'cell of origin' in these hematologic malignancies.

#### Defining the 'cell of origin': follicular helper T-cell lymphomas

Germinal center formation is regulated by follicular helper T cells ( $T_{FH}$ ), a subset of differentiated CD4<sup>+</sup> T cells regulated by the 'master' transcriptional repressor Bcl-6 [reviewed in (83)].  $T_{FH}$  cells express chemokines (CXCL13), chemokine receptors (CXCR5), and adhesion molecules (SLAM family receptors) that permit co-localization with germinal center B cells and follicular dendritic cells (FDC). Specific cytokines (IL-4, IL-21) and cell-surface ligands (CD40L) expressed by  $T_{FH}$  promote somatic hypermutation, class-switch recombination, and B-cell proliferation/survival leading to memory B cell and plasma cell generation. Angioimmunoblastic T-cell lymphoma (AITL) is an aggressive peripheral T-cell lymphoma (PTCL) that originates from clonally expanded  $T_{FH}$  cells [reviewed in (84)]. This is supported by gene expression profiling similarities between AITL and  $T_{FH}$  cells (85). The ontogeny of malignant T cells in AITL explains the histologic findings (varying expansion of germinal center B cells and an expanded meshwork of FDC) and clinical manifestations (polyclonal hypergammaglobulinemia, autoimmunity) characteristic of this PTCL. Malignant T cells in AITL share a common immunophenotype with  $T_{FH}$  cells, including the expression of B7-H family members PD-1(86–89) and ICOS (90–93).

ICOS and PD-1 play a critical role in T<sub>FH</sub> regulation. Soon after antigen presentation by dendritic cells, ICOS costimulation induces Bcl-6, leading to T<sub>FH</sub> differentiation (94). Continued ICOS costimulation, provided by germinal center B cells, is required for T<sub>FH</sub> maintenance (83). Not surprisingly then, a subset of patients with common variable immunodeficiency owing to the genomic loss of ICOS have a severe loss of memory B cells and IgG (95, 96). Conversely, Sanroque mice harbor a missense mutation in Roguin, a negative regulator of ICOS expression, and thus develop T<sub>FH</sub> hyperplasia, a robust germinal center reaction, and spontaneous autoimmunity (97). PD-1 is highly expressed on  $T_{FH}$  and may provide inhibitory signals upon engagement by B7-H1-expressing germinal center B cells. The observation that PD-1- and B7-H1-deficient mice develop increased frequencies of T<sub>FH</sub> cells following vaccination is consistent with this notion. Perhaps unexpectedly, increased B-cell apoptosis and impaired generation of long-lived plasma cells was observed in PD-1- and B7-H1-deficient mice (98). This phenotype was attributed to decreased cytokine production by PD-1-deficient T<sub>FH</sub>. However, reverse signaling by B7-H1 was recently demonstrated to promote T-cell survival (74), thus raising the possibility that reverse signaling by B7-H1 may similarly regulate the survival of germinal center B cells. The extent to which malignant T cells in AITL are regulated by ICOS or PD-1 signaling remains to be defined, although it was recently demonstrated that ICOS provides growth and survival signals to malignant T cells in ALK<sup>+</sup> ALCL (78). In addition to classic AITL, both a follicular variant of peripheral T-cell lymphoma, unspecified (PTCL-U) and cutaneous T-cell lymphoma subtypes express ICOS and PD-1 and may be derived from T<sub>FH</sub> cells (91, 93, 99, 100). Furthermore, non-malignant T<sub>FH</sub> cells represent an important constituent of the tumor microenvironment in some B-cell lymphomas [e.g., nodular lymphocyte-predominant Hodgkin lymphoma (101, 102) and follicular lymphoma (103-105)]. Therefore,

PD-1 and ICOS are rational therapeutic targets in these lymphomas.

#### **B7-H4 and neutropenia**

While their role in the regulation of adaptive immunity is increasingly appreciated, emerging evidence suggests that B7-H family members also regulate the function of the innate immune response (106). This was illustrated by the phenotype observed in B7-H4-deficient mice. Surprisingly, these mice were resistant to infection with Listeria monocytogenes (107). This was observed within a few days of inoculation, suggesting a role for the innate immune response. Therefore, investigators examined the neutrophil response in these mice. B7-H4-deficient mice were found to mount a more robust neutrophil response following infection when compared with their normal littermates. This was explained by the inhibition of neutrophil progenitors by B7-H4. A constellation of hematologic malignancies, most obviously T-cell large granular lymphocytic leukemia (T-LGL), are associated with an expansion of cytotoxic T-lymphocytes and significant neutropenia. The association between severe neutropenia and T-LGL is well described, but its pathogenesis is poorly understood, with increased peripheral destruction as well as impaired neutrophil production being described (108-110). For example, FasL is expressed by T-LGL cells and may be cleaved from the cell surface, thus explaining the elevated levels of soluble FasL observed in the sera of patients (110). Serum from these patients may trigger neutrophil apoptosis in a Fas-dependent manner (110). While reverse signaling via B7-H4 may upregulate FasL in EBV-transformed B cells, we are unaware of any data to suggest that a similar mechanism may exist in T cells (111). T-LGL cells have also been demonstrated to suppress neutrophil colony growth in a manner that is independent of Fas/FasL interactions (109). The observation that T-LGL cells highly express B7-H4 (R.A.W. unpublished observation), in conjunction with the recently described phenotype of B7-H4-deficient mice, implicates B7-H4 in the pathogenesis of disorders associated with immune-mediated neutropenia (112-114).

# Targeting B7-H: novel therapeutic approaches in hematologic malignancies

The B7-H family members are important regulators of adaptive and innate immunity in the tumor-bearing host and have non-immunologic effects that promote the survival of malignant cells. Therefore, targeting B7-H ligands with antagonistic monoclonal antibodies is rational and supported by preclinical studies performed in animal models. A phase I study of CT-011, a humanized IgG1 monoclonal antibody (mAb) that

blocks PD-1, included 17 patients with various hematologic malignancies, including nine with MDS/AML and 7 with non-Hodgkin lymphoma (41). A maximum tolerated dose was not reached in this study. Diarrhea was the most common adverse event, being observed in two patients. Grade 4 graft-versus-host disease (GVHD) developed in one of these patients who had undergone prior allogeneic transplantation for AML. Whether antibody administration may have exacerbated GVHD in this patient is unclear. Otherwise, no serious immunerelated adverse events were observed in this study. A single complete remission was observed in a previously untreated patient with bulky stage III follicular lymphoma. A minimal response was observed in a single patient with AML and stable disease reported in four patients (2 CLL, 1 cHL, and 1 MM). Peripheral blood CD4<sup>+</sup> and CD8<sup>+</sup> T cells were monitored during the course of the study. A statistically significant increase in the CD4<sup>+</sup> T-cell count was observed in patients treated at higher dose levels. Lymphopenia is an adverse prognostic factor at diagnosis and is associated with disease relapse in many lymphoproliferative disorders (59, 115-121). For example, in a cohort of 149 consecutive DLBCL patients treated with R-CHOP at a single institution, the cumulative incidence of relapse was 79% for patients with an absolute lymphocyte count (ALC)  $<0.96 \times 10^{9}$ /L at the time of follow-up (121). In contrast, the relapse rate was 6% among those with a higher ALC. In a similarly performed study including DLBCL patients following autologous stem cell transplantation, the development of new-onset lymphopenia during follow-up was associated with a cumulative incidence of relapse of 92%, compared with a cumulative incidence of relapse of only 19% for those with a higher ALC (122). PD-1 blockade may represent a novel therapeutic strategy to reverse lymphopenia and decrease the incidence of disease relapse in lymphopenic, high-risk patients. Two observations may further support this approach. First, elevated serum levels of a soluble form of B7-H1 are associated with lymphopenia in lymphoma patients (123). Finally, soluble B7-H1 was shown to promote T-cell apoptosis (124). In a phase I study conducted in patients with refractory solid tumors, tumor cell expression of B7-H1 appeared to predict the likelihood of response following PD-1 blockade with MDX-1106 (42). Theoretically, antibodies targeting B7-H1 interactions with both B7-1 and PD-1 may be superior to antibodies blocking either receptor/counter-receptor alone. A clinical trial in hematologic malignancies using a B7-H1 targeting antibody (BMS-936559) is planned (http:// www.clinicaltrials.gov). The identification of predictive biomarkers will be important in the selection of patients for therapeutic strategies targeting the B7-H1/PD-1/B7-1 axis.

Allogeneic transplantation is frequently considered for patients with various high-risk or relapsed hematologic malignancies. The goal of allogeneic stem cell transplantation is the generation of a robust graft-versus-leukemia (GVL) response. Unfortunately, concomitant graftversus-host disease (GVHD) is a frequent complication that contributes to significant morbidity and mortality. Recent work demonstrates that therapeutic manipulation of B7-H family members may promote the GVL response (10, 125-127), leading to the eradication of minimal residual disease, and prevent GVHD (128, 129). For example, B7-H1 expression by residual leukemic blasts confers resistance to T-cell mediated eradication and may thus promote immune evasion and disease recurrence following allogeneic stem cell transplantation (126, 127). In a murine model, B7-H1 blockade restored the GVL effect following the adoptive transfer of leukemiaspecific T cells without exacerbating GVHD (128). This data should be interpreted cautiously, however, as PD-1 was shown to inhibit GVHD in another model (130). These studies raise the possibility that manipulation of B7-H family members may augment the GVL response without exacerbating GVHD and may thus increase the therapeutic index associated with allogeneic stem cell transplantation.

The significant survival advantage observed in many lymphoproliferative disorders following the introduction of rituximab highlights the important role of mAb-mediated targeting in these malignancies. These antibodies mediate ADCC, complement-dependent cytotoxicity, or have direct apoptotic affects. As B7-H1 impairs NK-cellmediated ADCC, therapeutic strategies targeting B7-H1 may be rationally combined with targeting mAb, like rituximab (49–51).

#### Conclusions

Members of the B7-H family are widely expressed by malignant cells, and within the tumor microenvironment, in many hematologic malignancies. Inhibitory B7-H ligands promote the suppression of host anti-tumor immunity while those with stimulatory functions may directly stimulate the growth and survival of malignant cells. It is anticipated that further clarification of their pathogenic role in hematologic malignancies will have significant implications for the classification and risk stratification of these disorders. The development of novel therapeutic strategies targeting the B7-H family in hematologic malignancies is warranted.

#### References

1. Sharpe AH, Freeman GJ. The B7-CD28 superfamily. *Nat Rev Immunol* 2002;**2**:116–26.

- 2. Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. *Annu Rev Immunol* 2008;**26**:677–704.
- 3. Greenwald RJ, Freeman GJ, Sharpe AH. The B7 family revisited. *Annu Rev Immunol* 2005;**23**:515–48.
- Flies DB, Chen L. The new B7s: playing a pivotal role in tumor immunity. *J Immunother* 2007;30:251–60.
- 5. Zou W, Chen L. Inhibitory B7-family molecules in the tumour microenvironment. *Nat Rev Immunol* 2008;**8**:467–77.
- Yi KH, Chen L. Fine tuning the immune response through B7-H3 and B7-H4. *Immunol Rev* 2009;229:145– 51.
- Bretscher P, Cohn M. A theory of self-nonself discrimination. Science 1970;169:1042–9.
- Lafferty KJ, Cunningham AJ. A new analysis of allogeneic interactions. *Aust J Exp Biol Med Sci* 1975;53:27–42.
- Kondo A, Yamashita T, Tamura H, *et al.* Interferongamma and tumor necrosis factor-alpha induce an immunoinhibitory molecule, B7-H1, via nuclear factor-kappaB activation in blasts in myelodysplastic syndromes. *Blood* 2010;**116**:1124–31.
- Berthon C, Driss V, Liu J, Kuranda K, Leleu X, Jouy N, Hetuin D, Quesnel B. In acute myeloid leukemia, B7-H1 (PD-L1) protection of blasts from cytotoxic T cells is induced by TLR ligands and interferon-gamma and can be reversed using MEK inhibitors. *Cancer Immunol Immunother* 2010;**59**:1839–49.
- Salih HR, Wintterle S, Krusch M, Kroner A, Huang YH, Chen L, Wiendl H. The role of leukemia-derived B7-H1 (PD-L1) in tumor-T-cell interactions in humans. *Exp Hematol* 2006;**34**:888–94.
- Chen X, Liu S, Wang L, Zhang W, Ji Y, Ma X. Clinical significance of B7-H1 (PD-L1) expression in human acute leukemia. *Cancer Biol Ther* 2008;7:622–7.
- Ge W, Ma X, Li X, Wang Y, Li C, Meng H, Liu X, Yu Z, You S, Qiu L. B7-H1 up-regulation on dendritic-like leukemia cells suppresses T cell immune function through modulation of IL-10/IL-12 production and generation of Treg cells. *Leuk Res* 2009;33:948–57.
- 14. Andorsky DJ, Yamada RE, Said J, Pinkus GS, Betting DJ, Timmerman JM. Programmed death ligand 1 is expressed by non-hodgkin lymphomas and inhibits the activity of tumor-associated T cells. *Clin Cancer Res* 2011;**17**:4232–44.
- Yamamoto R, Nishikori M, Kitawaki T, *et al.* PD-1-PD-1 ligand interaction contributes to immunosuppressive microenvironment of Hodgkin lymphoma. *Blood* 2008;111:3220–4.
- 16. Green MR, Monti S, Rodig SJ, *et al.* Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. *Blood* 2010;**116**:3268–77.
- Liu J, Hamrouni A, Wolowiec D, Coiteux V, Kuliczkowski K, Hetuin D, Saudemont A, Quesnel B. Plasma cells from multiple myeloma patients express B7-H1

(PD-L1) and increase expression after stimulation with IFN-{gamma} and TLR ligands via a MyD88-, TRAF6-, and MEK-dependent pathway. *Blood* 2007;**110**:296–304.

- Wilcox RA, Feldman AL, Wada DA, *et al.* B7-H1 (PD-L1, CD274) suppresses host immunity in T-cell lymphoproliferative disorders. *Blood* 2009;114:2149–58.
- Kantekure K, Yang Y, Raghunath P, Schaffer A, Woetmann A, Zhang Q, Odum N, Wasik M. Expression patterns of the immunosuppressive proteins PD-1/CD279 and PD-L1/CD274 at different stages of cutaneous T-cell lymphoma/mycosis fungoides. *Am J Dermatopathol* 2011;**34**:126–8.
- Kozako T, Yoshimitsu M, Fujiwara H, et al. PD-1/PD-L1 expression in human T-cell leukemia virus type 1 carriers and adult T-cell leukemia/lymphoma patients. *Leukemia* 2009;23:375–82.
- Tamura H, Dan K, Tamada K, Nakamura K, Shioi Y, Hyodo H, Wang SD, Dong H, Chen L, Ogata K. Expression of functional B7-H2 and B7.2 costimulatory molecules and their prognostic implications in de novo acute myeloid leukemia. *Clin Cancer Res* 2005;**11**:5708–17.
- 22. Yamashita T, Tamura H, Satoh C, Shinya E, Takahashi H, Chen L, Kondo A, Tsuji T, Dan K, Ogata K. Functional B7.2 and B7-H2 molecules on myeloma cells are associated with a growth advantage. *Clin Cancer Res* 2009;15:770–7.
- Gregorio A, Corrias MV, Castriconi R, Dondero A, Mosconi M, Gambini C, Moretta A, Moretta L, Bottino C. Small round blue cell tumours: diagnostic and prognostic usefulness of the expression of B7-H3 surface molecule. *Histopathology* 2008;53:73–80.
- 24. Wey EA, Laliberta J, McNeil C, Basrur V, Keyoumarsi F, Thomas D, Suh C, Huh J, Elenitoba-Johnson K, Lim MS. Expression of B7-H3 (CD276) an Immunoregulatory Cell Surface Molecule in Non-Hodgkin Lymphomas Identified by Mass Spectrometry. Presented at: Annual Meeting of the United States and Canadian Academy of Pathology. 2011.
- 25. Wang S, Zhu G, Chapoval AI, Dong H, Tamada K, Ni J, Chen L. Costimulation of T cells by B7-H2, a B7-like molecule that binds ICOS. *Blood* 2000;**96**:2808–13.
- 26. Swallow MM, Wallin JJ, Sha WC. B7h, a novel costimulatory homolog of B7.1 and B7.2, is induced by TNFalpha. *Immunity* 1999;**11**:423–32.
- 27. Yoshinaga SK, Whoriskey JS, Khare SD, *et al.* T-cell costimulation through B7RP-1 and ICOS. *Nature* 1999;**402**:827–32.
- 28. Brodie D, Collins AV, Iaboni A, Fennelly JA, Sparks LM, Xu XN, van der Merwe PA, Davis SJ. LICOS, a primordial costimulatory ligand? *Curr Biol* 2000;**10**:333–6.
- Mages HW, Hutloff A, Heuck C, Buchner K, Himmelbauer H, Oliveri F, Kroczek RA. Molecular cloning and characterization of murine ICOS and identification of B7h as ICOS ligand. *Eur J Immunol* 2000;**30**:1040–7.

- Yao S, Zhu Y, Zhu G, et al. B7-h2 is a costimulatory ligand for CD28 in human. *Immunity* 2011;34:729– 40.
- Zang X, Loke P, Kim J, Murphy K, Waitz R, Allison JP. B7x: a widely expressed B7 family member that inhibits T cell activation. *Proc Natl Acad Sci USA* 2003;100:10388–92.
- Prasad DV, Richards S, Mai XM, Dong C. B7S1, a novel B7 family member that negatively regulates T cell activation. *Immunity* 2003;18:863–73.
- 33. Sica GL, Choi IH, Zhu G, Tamada K, Wang SD, Tamura H, Chapoval AI, Flies DB, Bajorath J, Chen L. B7-H4, a molecule of the B7 family, negatively regulates T cell immunity. *Immunity* 2003;18:849–61.
- Hashiguchi M, Kobori H, Ritprajak P, Kamimura Y, Kozono H, Azuma M. Triggering receptor expressed on myeloid cell-like transcript 2 (TLT-2) is a counter-receptor for B7-H3 and enhances T cell responses. *Proc Natl Acad Sci USA* 2008;105:10495–500.
- 35. Kobori H, Hashiguchi M, Piao J, Kato M, Ritprajak P, Azuma M. Enhancement of effector CD8 + T-cell function by tumour-associated B7-H3 and modulation of its counter-receptor triggering receptor expressed on myeloid cell-like transcript 2 at tumour sites. *Immunology* 2010;**130**:363–73.
- Leitner J, Klauser C, Pickl WF, *et al.* B7-H3 is a potent inhibitor of human T-cell activation: no evidence for B7-H3 and TREML2 interaction. *Eur J Immunol* 2009;**39**:1754–64.
- Nishimura H, Nose M, Hiai H, Minato N, Honjo T. Development of lupus-like autoimmune diseases by disruption of the PD-1 gene encoding an ITIM motif-carrying immunoreceptor. *Immunity* 1999;11:141–51.
- Nishimura H, Okazaki T, Tanaka Y, *et al.* Autoimmune dilated cardiomyopathy in PD-1 receptor-deficient mice. *Science* 2001;291:319–22.
- Wang J, Yoshida T, Nakaki F, Hiai H, Okazaki T, Honjo T. Establishment of NOD-Pdcd1-/- mice as an efficient animal model of type I diabetes. *Proc Natl Acad Sci USA* 2005;**102**:11823–8.
- 40. Hodi FS, O'Day SJ, McDermott DF, *et al.* Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010;**363**:711–23.
- 41. Berger R, Rotem-Yehudar R, Slama G, Landes S, Kneller A, Leiba M, Koren-Michowitz M, Shimoni A, Nagler A. Phase I safety and pharmacokinetic study of CT-011, a humanized antibody interacting with PD-1, in patients with advanced hematologic malignancies. *Clin Cancer Res* 2008;14:3044–51.
- Brahmer JR, Drake CG, Wollner I, *et al.* Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: safety, clinical activity, pharmacodynamics, and immunologic correlates. *J Clin Oncol* 2010;28:3167–75.
- 43. Riley JL. PD-1 signaling in primary T cells. *Immunol Rev* 2009;**229**:114–25.

- 44. Butte MJ, Keir ME, Phamduy TB, Sharpe AH, Freeman GJ. Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity* 2007;**27**:111–22.
- 45. Tsushima F, Yao S, Shin T, Flies A, Flies S, Xu H, Tamada K, Pardoll DM, Chen L. Interaction between B7-H1 and PD-1 determines initiation and reversal of T-cell anergy. *Blood* 2007;**110**:180–5.
- Day CL, Kaufmann DE, Kiepiela P, et al. PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature* 2006;443:350–4.
- Zhang JY, Zhang Z, Wang X, et al. PD-1 up-regulation is correlated with HIV-specific memory CD8+ T-cell exhaustion in typical progressors but not in long-term nonprogressors. Blood 2007;109:4671–8.
- 48. Samimi S, Benoit B, Evans K, Wherry EJ, Showe L, Wysocka M, Rook AH. Increased programmed death-1 expression on CD4+ T cells in cutaneous T-cell lymphoma: implications for immune suppression. *Arch Dermatol* 2010;**146**:1382–8.
- Benson DM Jr, Bakan CE, Mishra A, *et al.* The PD-1/PD-L1 axis modulates the natural killer cell versus multiple myeloma effect: a therapeutic target for CT-011, a novel monoclonal anti-PD-1 antibody. *Blood* 2010;**116**:2286–94.
- 50. Reddy N, Hernandez-Ilizaliturri FJ, Deeb G, Roth M, Vaughn M, Knight J, Wallace P, Czuczman MS. Immunomodulatory drugs stimulate natural killer-cell function, alter cytokine production by dendritic cells, and inhibit angiogenesis enhancing the anti-tumour activity of rituximab in vivo. *Br J Haematol* 2008;140:36–45.
- 51. Wu L, Adams M, Carter T, Chen R, Muller G, Stirling D, Schafer P, Bartlett JB. Lenalidomide enhances natural killer cell and monocyte-mediated antibody-dependent cellular cytotoxicity of rituximab-treated CD20+ tumor cells. *Clin Cancer Res* 2008;**14**:4650–7.
- 52. Wiesmayr S, Webber SA, Macedo C, Popescu I, Smith L, Luce J, Metes D. Decreased NKp46 and NKG2D and elevated PD-1 are associated with altered NK-cell function in pediatric transplant patients with PTLD. *Eur J Immunol* 2011;**42**:541–50.
- Ogasawara K, Yoshinaga SK, Lanier LL. Inducible costimulator costimulates cytotoxic activity and IFNgamma production in activated murine NK cells. J Immunol 2002;169:3676–85.
- Brandt CS, Baratin M, Yi EC, *et al.* The B7 family member B7-H6 is a tumor cell ligand for the activating natural killer cell receptor NKp30 in humans. *J Exp Med* 2009;206:1495–503.
- 55. Joyce MG, Tran P, Zhuravleva MA, Jaw J, Colonna M, Sun PD. Crystal structure of human natural cytotoxicity receptor NKp30 and identification of its ligand binding site. *Proc Natl Acad Sci USA* 2011;**108**:6223–8.

- 56. Li Y, Wang Q, Mariuzza RA. Structure of the human activating natural cytotoxicity receptor NKp30 bound to its tumor cell ligand B7-H6. *J Exp Med* 2011;208: 703–14.
- Wilcox RA. Cancer-associated myeloproliferation: old association, new therapeutic target. *Mayo Clin Proc* 2010;85:656–63.
- Wilcox RA, Ristow K, Habermann TM, et al. The absolute monocyte count is associated with overall survival in patients newly diagnosed with follicular lymphoma. *Leuk Lymphoma* 2011. [Epub ahead of print].
- Wilcox RA, Ristow K, Habermann TM, *et al.* The absolute monocyte and lymphocyte prognostic score predicts survival and identifies high-risk patients in diffuse large-B-cell lymphoma. *Leukemia* 2011;25:1502–9.
- Steidl C, Lee T, Shah SP, *et al.* Tumor-associated macrophages and survival in classic Hodgkin's lymphoma. N Engl J Med 2010;362:875–85.
- Porrata LF, Ristow K, Colgan J, et al. Peripheral blood lymphocyte/monocyte ratio at diagnosis and survival in classical Hodgkin lymphoma. *Haematologica* 2011;97: 262–9.
- Hoechst B, Gamrekelashvili J, Manns MP, Greten TF, Korangy F. Plasticity of human Th17 cells and iTregs is orchestrated by different subsets of myeloid cells. *Blood* 2011;117:6532–41.
- Lin Y, Gustafson MP, Bulur PA, Gastineau DA, Witzig TE, Dietz AB. Immunosuppressive CD14+ HLA-DR(low)/- monocytes in B-cell non-Hodgkin lymphoma. *Blood* 2011;**117**:872–81.
- 64. Wang L, Pino-Lagos K, de Vries VC, Guleria I, Sayegh MH, Noelle RJ. Programmed death 1 ligand signaling regulates the generation of adaptive Foxp3 + CD4 + regulatory T cells. *Proc Nat Acad Sci USA* 2008;**105**:9331–6.
- Dolganiuc A, Paek E, Kodys K, Thomas J, Szabo G. Myeloid dendritic cells of patients with chronic HCV infection induce proliferation of regulatory T lymphocytes. *Gastroenterology* 2008;135:2119–27.
- Kordasti SY, Ingram W, Hayden J, *et al.* CD4 + CD25high Foxp3+ regulatory T cells in myelodysplastic syndrome (MDS). *Blood* 2007;110:847–50.
- 67. Szczepanski MJ, Szajnik M, Czystowska M, Mandapathil M, Strauss L, Welsh A, Foon KA, Whiteside TL, Boyiadzis M. Increased frequency and suppression by regulatory T cells in patients with acute myelogenous leukemia. *Clin Cancer Res* 2009;15:3325–32.
- Relander T, Johnson NA, Farinha P, Connors JM, Sehn LH, Gascoyne RD. Prognostic factors in follicular lymphoma. *J Clin Oncol* 2010;28:2902–13.
- Yang ZZ, Novak AJ, Stenson MJ, Witzig TE, Ansell SM. Intratumoral CD4 + CD25+ regulatory T-cellmediated suppression of infiltrating CD4+ T cells in Bcell non-Hodgkin lymphoma. *Blood* 2006;107:3639–46.
- 70. Han Y, Wu J, Bi L, Xiong S, Gao S, Yin L, Jiang L, Chen C, Yu K, Zhang S. Malignant B cells induce the

conversion of CD4CD25 T cells to regulatory T cells in B-cell non-hodgkin lymphoma. *PLoS One* 2011;6:e28649.

- Zhou Q, Munger ME, Highfill SL, *et al.* Program death- 1 signaling and regulatory T cells collaborate to resist the function of adoptively transferred cytotoxic T lympho- cytes in advanced acute myeloid leukemia. *Blood* 2010;**116**:2484–93.
- Rosenblatt J, Glotzbecker B, Mills H, *et al.* PD-1 blockade by CT-011, anti-PD-1 antibody, enhances ex vivo T-cell responses to autologous dendritic cell/myeloma fusion vaccine. *J Immunother* 2011;34:409–18.
- Azuma T, Yao S, Zhu G, Flies AS, Flies SJ, Chen L. B7-H1 is a ubiquitous antiapoptotic receptor on cancer cells. *Blood* 2008;111:3635–43.
- 74. Pulko V, Harris KJ, Liu X, Gibbons RM, Harrington SM, Krco CJ, Kwon ED, Dong H. B7-h1 expressed by activated CD8 T cells is essential for their survival. J Immunol 2011;187:5606–14.
- Liu H, Tekle C, Chen YW, *et al.* B7-H3 silencing increases paclitaxel sensitivity by abrogating Jak2/Stat3 phosphorylation. *Mol Cancer Ther* 2011;10:960–71.
- 76. Salceda S, Tang T, Kmet M, Munteanu A, Ghosh M, Macina R, Liu W, Pilkington G, Papkoff J. The immunomodulatory protein B7-H4 is overexpressed in breast and ovarian cancers and promotes epithelial cell transformation. *Exp Cell Res* 2005;**306**:128–41.
- Marzec M, Zhang Q, Goradia A, *et al.* Oncogenic kinase NPM/ALK induces through STAT3 expression of immunosuppressive protein CD274 (PD-L1, B7-H1). *Proc Natl Acad Sci USA* 2008;105:20852–7.
- Zhang Q, Wang H, Kantekure K, *et al.* Oncogenic tyrosine kinase NPM-ALK induces expression of the growthpromoting receptor ICOS. *Blood* 2011;118:3062–71.
- 79. Gong AY, Zhou R, Hu G, Li X, Splinter PL, O'Hara SP, LaRusso NF, Soukup GA, Dong H, Chen XM. MicroRNA-513 regulates B7-H1 translation and is involved in IFN-gamma-induced B7-H1 expression in cholangiocytes. *J Immunol* 2009;**182**:1325–33.
- Steidl C, Gascoyne RD. The molecular pathogenesis of primary mediastinal large B-cell lymphoma. *Blood* 2011;118:2659–69.
- Tefferi A, Vainchenker W. Myeloproliferative neoplasms: molecular pathophysiology, essential clinical understanding, and treatment strategies. *J Clin Oncol* 2011;29:573– 82.
- 82. Steidl C, Shah SP, Woolcock BW, *et al.* MHC class II transactivator CIITA is a recurrent gene fusion partner in lymphoid cancers. *Nature* 2011;**471**:377–81.
- Crotty S. Follicular helper CD4 T cells (TFH). Annu Rev Immunol 2011;29:621–63.
- de Leval L, Gisselbrecht C, Gaulard P. Advances in the understanding and management of angioimmunoblastic T-cell lymphoma. *Br J Haematol* 2010;**148**:673–89.
- 85. Piccaluga PP, Agostinelli C, Califano A, *et al.* Gene expression analysis of angioimmunoblastic lymphoma indicates derivation from T follicular helper cells and vas-

cular endothelial growth factor deregulation. *Cancer Res* 2007;**67**:10703–10.

- 86. Dorfman DM, Brown JA, Shahsafaei A, Freeman GJ. Programmed death-1 (PD-1) is a marker of germinal center-associated T cells and angioimmunoblastic T-cell lymphoma. *Am J Surg Pathol* 2006;**30**:802–10.
- Roncador G, Garcia Verdes-Montenegro JF, Tedoldi S, et al. Expression of two markers of germinal center T cells (SAP and PD-1) in angioimmunoblastic T-cell lymphoma. *Haematologica* 2007;92:1059–66.
- Xerri L, Chetaille B, Serriari N, Attias C, Guillaume Y, Arnoulet C, Olive D. Programmed death 1 is a marker of angioimmunoblastic T-cell lymphoma and B-cell small lymphocytic lymphoma/chronic lymphocytic leukemia. *Hum Pathol* 2008;**39**:1050–8.
- Yu H, Shahsafaei A, Dorfman DM. Germinal-center T-helper-cell markers PD-1 and CXCL13 are both expressed by neoplastic cells in angioimmunoblastic T-cell lymphoma. *Am J Clin Pathol* 2009;131:33–41.
- Baseggio L, Traverse-Glehen A, Berger F, Ffrench M, Jallades L, Morel D, Goedert G, Magaud JP, Salles G, Felman P. CD10 and ICOS expression by multiparametric flow cytometry in angioimmunoblastic T-cell lymphoma. *Mod Pathol* 2011;24:993–1003.
- 91. Huang Y, Moreau A, Dupuis J, et al. Peripheral T-cell lymphomas with a follicular growth pattern are derived from follicular helper T cells (TFH) and may show overlapping features with angioimmunoblastic T-cell lymphomas. Am J Surg Pathol 2009;33:682–90.
- 92. Rodriguez-Justo M, Attygalle AD, Munson P, Roncador G, Marafioti T, Piris MA. Angioimmunoblastic T-cell lymphoma with hyperplastic germinal centres: a neoplasia with origin in the outer zone of the germinal centre? Clinicopathological and immunohistochemical study of 10 cases with follicular T-cell markers. *Mod Pathol* 2009;**22**:753–61.
- 93. Marafioti T, Paterson JC, Ballabio E, et al. The inducible T-cell co-stimulator molecule is expressed on subsets of T cells and is a new marker of lymphomas of T follicular helper cell-derivation. *Haematologica* 2010;95:432–9.
- 94. Choi YS, Kageyama R, Eto D, Escobar TC, Johnston RJ, Monticelli L, Lao C, Crotty S. ICOS receptor instructs T follicular helper cell versus effector cell differentiation via induction of the transcriptional repressor Bcl6. *Immunity* 2011;34:932–46.
- 95. Grimbacher B, Hutloff A, Schlesier M, et al. Homozygous loss of ICOS is associated with adult-onset common variable immunodeficiency. *Nat Immunol* 2003;4:261–8.
- Salzer U, Maul-Pavicic A, Cunningham-Rundles C, et al. ICOS deficiency in patients with common variable immunodeficiency. *Clin Immunol* 2004;113:234–40.
- Vinuesa CG, Cook MC, Angelucci C, *et al.* A RINGtype ubiquitin ligase family member required to repress follicular helper T cells and autoimmunity. *Nature* 2005;435:452–8.

- Good-Jacobson KL, Szumilas CG, Chen L, Sharpe AH, Tomayko MM, Shlomchik MJ. PD-1 regulates germinal center B cell survival and the formation and affinity of long-lived plasma cells. *Nat Immunol* 2010;11:535–42.
- Rodriguez Pinilla SM, Roncador G, Rodriguez-Peralto JL, et al. Primary cutaneous CD4+ small/medium-sized pleomorphic T-cell lymphoma expresses follicular T-cell markers. Am J Surg Pathol 2009;33:81–90.
- 100. Cetinozman F, Jansen PM, Willemze R. Expression of programmed death-1 in primary cutaneous CD4-Positive small/medium-sized pleomorphic T-cell lymphoma, cutaneous pseudo-T-cell lymphoma, and other types of cutaneous T-cell lymphoma. *Am J Surg Pathol* 2012;**36**:109– 16.
- 101. Churchill HR, Roncador G, Warnke RA, Natkunam Y. Programmed death 1 expression in variant immunoarchitectural patterns of nodular lymphocyte predominant Hodgkin lymphoma: comparison with CD57 and lymphomas in the differential diagnosis. *Hum Pathol* 2010;**41**:1726–34.
- 102. Nam-Cha SH, Roncador G, Sanchez-Verde L, Montes-Moreno S, Acevedo A, Dominguez-Franjo P, Piris MA. PD-1, a follicular T-cell marker useful for recognizing nodular lymphocyte-predominant Hodgkin lymphoma. *Am J Surg Pathol* 2008;**32**:1252–7.
- 103. Ame-Thomas P, Le PriolJ, Yssel H, et al. Characterization of intratumoral follicular helper T cells in follicular lymphoma: role in the survival of malignant B cells. *Leukemia* 2011. [Epub ahead of print].
- 104. Hilchey SP, Rosenberg AF, Hyrien O, *et al.* Follicular lymphoma tumor-infiltrating T-helper (T(H)) cells have the same polyfunctional potential as normal nodal T(H) cells despite skewed differentiation. *Blood* 2011;**118**:3591–602.
- 105. Wahlin BE, Aggarwal M, Montes-Moreno S, Gonzalez LF, Roncador G, Sanchez-Verde L, Christensson B, Sander B, Kimby E. A unifying microenvironment model in follicular lymphoma: outcome is predicted by programmed death-1 – positive, regulatory, cytotoxic, and helper T cells and macrophages. *Clin Cancer Res* 2010;**16**:637–50.
- 106. Yao S, Wang S, Zhu Y, et al. PD-1 on dendritic cells impedes innate immunity against bacterial infection. Blood 2009;113:5811–8.
- 107. Zhu G, Augustine MM, Azuma T, *et al.* B7-H4-deficient mice display augmented neutrophil-mediated innate immunity. *Blood* 2009;**113**:1759–67.
- 108. Starkebaum G. Chronic neutropenia associated with autoimmune disease. *Semin Hematol* 2002;**39**:121–7.
- 109. Coakley G, Iqbal M, Brooks D, Panayi GS, Lanchbury JS. CD8+, CD57+ T cells from healthy elderly subjects suppress neutrophil development *in vitro*: implications for the neutropenia of Felty's and large granular lymphocyte syndromes. *Arthritis Rheum* 2000;**43**:834–43.

- 110. Liu JH, Wei S, Lamy T, Epling-Burnette PK, Starkebaum G, Djeu JY, Loughran TP. Chronic neutropenia mediated by fas ligand. *Blood* 2000;95:3219–22.
- 111. Song H, Park G, Kim YS, et al. B7-H4 reverse signaling induces the apoptosis of EBV-transformed B cells through Fas ligand up-regulation. Cancer Lett 2008;266:227–37.
- 112. Karadimitris A, Li K, Notaro R, *et al.* Association of clonal T-cell large granular lymphocyte disease and paroxysmal nocturnal haemoglobinuria (PNH): further evidence for a pathogenetic link between T cells, aplastic anaemia and PNH. *Br J Haematol* 2001;**115**: 1010–4.
- 113. Fozza C, Contini S, Galleu A, Simula MP, Virdis P, Bonfigli S, Longinotti M. Patients with myelodysplastic syndromes display several T-cell expansions, which are mostly polyclonal in the CD4(+) subset and oligoclonal in the CD8(+) subset. *Exp Hematol* 2009;**37**:947–55.
- 114. Zheng Z, Qianqiao Z, Qi H, Feng X, Chunkang C, Xiao L. In vitro deprivation of CD8(+)CD57(+)T cells promotes the malignant growth of bone marrow colony cells in patients with lower-risk myelodysplastic syndrome. *Exp Hematol* 2010;**38**:677–84.
- 115. Behl D, Porrata LF, Markovic SN, *et al.* Absolute lymphocyte count recovery after induction chemotherapy predicts superior survival in acute myelogenous leukemia. *Leukemia* 2006;**20**:29–34.
- 116. Behl D, Ristow K, Markovic SN, *et al.* Absolute lymphocyte count predicts therapeutic efficacy of rituximab therapy in follicular lymphomas. *Br J Haematol* 2007;**137**:409–15.
- 117. Porrata LF, Inwards DJ, Ansell SM, Micallef IN, Johnston PB, Gastineau DA, Litzow MR, Winters JL, Markovic SN. Early lymphocyte recovery predicts superior survival after autologous stem cell transplantation in non-Hodgkin lymphoma: a prospective study. *Biol Blood Marrow Transplant* 2008;14:807–16.
- 118. Porrata LF, Litzow MR, Tefferi A, Letendre L, Kumar S, Geyer SM, Markovic SN. Early lymphocyte recovery is a predictive factor for prolonged survival after autologous hematopoietic stem cell transplantation for acute myelogenous leukemia. *Leukemia* 2002;16:1311–8.
- 119. Porrata LF, Ristow K, Habermann TM, Witzig TE, Inwards DJ, Markovic SN. Absolute lymphocyte count at the time of first relapse predicts survival in patients with diffuse large B-cell lymphoma. *Am J Hematol* 2009;84:93–7.
- 120. Porrata LF, Ristow K, Witzig TE, Tuinistra N, Habermann TM, Inwards DJ, Ansell SM, Micallef IN, Johnston PB, Markovic SN. Absolute lymphocyte count predicts therapeutic efficacy and survival at the time of radioimmunotherapy in patients with relapsed follicular lymphomas. *Leukemia* 2007;21:2554–6.

- 121. Porrata LF, Rsitow K, Inwards DJ, et al. Lymphopenia assessed during routine follow-up after immunochemotherapy (R-CHOP) is a risk factor for predicting relapse in patients with diffuse large B-cell lymphoma. *Leukemia* 2010;**24**:1343–9.
- 122. Porrata LF, Inwards DJ, Ansell SM, Micallef IN, Johnston PB, Hogan WJ, Markovic SN. New-onset lymphopenia assessed during routine follow-up is a risk factor for relapse postautologous peripheral blood hematopoietic stem cell transplantation in patients with diffuse large B-cell lymphoma. *Biol Blood Marrow Transplant* 2010;16:376–83.
- 123. Wilcox RA, Frigola Baro X, Porrata LF, Kwon ED, Maurer MJ, Micallef IN, Witzig TE, Ansell SM. Association of serum B7-H1 level and lymphopenia in diffuse large B-cell non-Hodgkin lymphoma. ASCO Meeting Abstracts. 2010: 8080.
- 124. Frigola X, Inman BA, Lohse CM, Krco CJ, Cheville JC, Thompson RH, Leibovich B, Blute ML, Dong H, Kwon ED. Identification of a soluble form of B7-H1 that retains immunosuppressive activity and is associated with aggressive renal cell carcinoma. *Clin Cancer Res* 2011;17:1915–23.
- 125. Zhang L, Gajewski TF, Kline J. PD-1/PD-L1 interactions inhibit antitumor immune responses in a murine acute myeloid leukemia model. *Blood* 2009;**114**:1545–52.
- 126. Saudemont A, Quesnel B. In a model of tumor dormancy, long-term persistent leukemic cells have increased B7-H1 and B7.1 expression and resist CTL-mediated lysis. *Blood* 2004;**104**:2124–33.
- 127. Norde WJ, Maas F, Hobo W, *et al.* PD-1/PD-L1 interactions contribute to functional T-cell impairment in patients who relapse with cancer after allogeneic stem cell transplantation. *Cancer Res* 2011;**71**:5111–22.
- 128. Koestner W, Hapke M, Herbst J, et al. PD-L1 blockade effectively restores strong graft-versus-leukemia effects without graft-versus-host disease after delayed adoptive transfer of T-cell receptor gene-engineered allogeneic CD8 + T cells. Blood 2011;117:1030–41.
- 129. Flies DB, Wang S, Xu H, Chen L. Cutting edge: a monoclonal antibody specific for the programmed death-1 homolog prevents graft-versus-host disease in mouse models. J Immunol 2011;187:1537–41.
- 130. Blazar BR, Carreno BM, Panoskaltsis-Mortari A, Carter L, Iwai Y, Yagita H, Nishimura H, Taylor PA. Blockade of programmed death-1 engagement accelerates graft-versus-host disease lethality by an IFN-gamma-dependent mechanism. *J Immunol* 2003;**171**:1272–7.
- 131. Fanoni D, Tavecchio S, Recalcati S, Balice Y, Venegoni L, Fiorani R, Crosti C, Berti E. New monoclonal antibodies against B-cell antigens: possible new strategies for diagnosis of primary cutaneous B-cell lymphomas. *Immunol Lett* 2011;**134**:157–60.