

Hydrogen Sulfide: Redox Metabolism and Signaling

Ruma Banerjee

Abstract

The recognition of hydrogen sulfide (H_2S) as an endogenously produced gas with signaling potential has stimulated research on a multitude of physiological effects mediated in the cardiovascular, immune, gastrointestinal, genitourinary, endocrine, and central nervous systems. The heightened activity in the area of H_2S biology led to convening of the first international conference on H_2S in Shanghai in the summer of 2009 and to two Forum issues published in 2010 by *Antioxidants & Redox Signaling* on the physiological effects of H_2S . Yet, fundamental questions regarding the biogenesis and regulation of H_2S , the bioenergetics of its catabolism, its tissue concentrations, and elucidation of its molecular targets remain. Some of these issues are the subject of the current Forum on H_2S . *Antioxid. Redox Signal.* 15, 339–341.

REACTIONS IN THE SULFUR METABOLIC PATHWAY involving the amino acids cysteine and homocysteine are potential sources of hydrogen sulfide (H_2S) (4, 28). These include the two enzymes in the transsulfuration pathway, cystathionine β -synthase (CBS), and γ -cystathionase (CSE), which successively convert serine and homocysteine to cystathionine and cystathionine, to cysteine. Lax substrate specificity creates the potential for a multitude of alternative reactions to be catalyzed by both enzymes leading to H_2S generation from either cysteine or homocysteine or both (6, 7, 27). A third potential route for H_2S generation is *via* the combined actions of cysteine aminotransferase and mercaptopyruvate sulfurtransferase (26), enzymes involved in cysteine catabolism. However, the product of coupling these two enzymes is persulfide rather than H_2S , which can be liberated in the presence of reductant (15).

An efficient mitochondrial pathway for H_2S oxidation exists (10) and is important for holding steady-state tissue levels of H_2S at very low values. This has led to the hypothesis that H_2S might function as an oxygen sensor since, under conditions of hypoxia, the catabolic removal of H_2S would be impeded, leading to augmented H_2S levels and activation of its signaling responses (23). Similarities between the effects of H_2S and hypoxia and enhancement and abrogation of the hypoxic responses by H_2S precursors and H_2S synthesis inhibitors, respectively, support the proposal that H_2S might be involved in oxygen sensing (23).

The discovery of H_2S as a physiological mediator was made by Abe and Kimura, who first reported its neuromodulatory effects (1) and soon thereafter, its vasorelaxant effect on smooth muscle (11). Today, a plethora of physiological effects are associated with H_2S with some of the major links being to

the cardiovascular and central nervous systems and to inflammation. H_2S modulates a range of cardiovascular effects, including cardioprotection against ischemic and myocardial reperfusion injury and vascular contraction or relaxation (9, 23, 25, 29). The endothelial-derived relaxing factor activity of H_2S in mice lacking the CSE gene is controversial, with two groups reporting conflicting results. Thus, Wang and coworkers reported marked age-dependent hypertension (30), whereas Ishii and coworkers reported a normotensive phenotype in the CSE knockout mice (14). H_2S stimulates opening of K_{ATP} channels, suggesting a mechanism for its vasorelaxant effects in addition to its reported involvement in the Nrf2 and ERK and PI3K/Akt signaling pathways (5, 13, 31). In brain, H_2S has varied effects ranging from enhancing hippocampal long-term potentiation (1), to triggering calcium waves and increasing intracellular calcium levels in astrocytes (22), to increased neuroprotection by stimulating glutathione synthesis (19, 20). A possible mechanism for neuromodulation by H_2S involves induction of cyclic AMP synthesis and activating N-methyl D-aspartate-receptor-mediated excitatory postsynaptic currents (18). Both pro- and anti-inflammatory effects have been ascribed to H_2S and efforts are underway to develop the therapeutic potential of H_2S -releasing drugs to counter inflammatory diseases.

Large discrepancies have been reported in the literature on tissue concentrations of H_2S and H_2S production rates in the presence of exogenous substrate. These stem in part from the technical difficulties associated with handling H_2S , which is redox sensitive, and the considerably larger tissue stores of sulfane sulfur and acid-labile sulfur that can be released if adequate precautions are not taken during sample preparation. Additionally, the complexity in utilization of cysteine

and/or homocysteine for H₂S generation by CBS and CSE leads to a significant underestimation of H₂S production rates and bias against CBS when only cysteine is applied as substrate, which is commonly the case in the field. Finally, the use of compounds such as hydroxylamine and aminooxyacetate that inhibit many pyridoxal phosphate enzymes and are not specific for CBS or CSE produces results that are not readily interpretable. Two articles in this Forum address these quantitative issues surrounding H₂S determination. In one, Levitt and coworkers report that steady-state tissue levels of H₂S are negligibly low with the exception of aorta, which exhibits 20–100 times higher H₂S concentrations (21). In the other article, the author's group uses quantitative Western blot analyses to determine the absolute protein levels of CBS and CSE in liver and kidney and reports the tissue H₂S production capacity in the presence of saturating concentrations of cysteine and homocysteine used alone or in combination as substrates (16). These studies reveal the differential importance of CBS versus CSE in various tissues. From simulations of the liver data adjusted for the difference in protein levels of CBS and CSE, it is estimated that CSE accounts for ~96% of H₂S production at physiologically relevant concentrations of substrate.

Bouillaud and coworkers discuss sulfide bioenergetics in the context of the mitochondrial catabolic pathway and the similarities between cyanide and H₂S inhibition of cytochrome c oxidase (3). They also discuss potential mechanisms for neutralizing sulfide particularly as it pertains to the biology of colonocytes, cells that are routinely exposed to high sulfide concentrations, and to the invertebrates living in sulfide-rich habitats. The first enzyme in the H₂S oxidation pathway, sulfide quinone reductase, has a very high affinity for sulfide, and oxidation activity can be detected at intracellular sulfide concentrations ≥ 10 –20 nM, thus protecting cytochrome c oxidase from inhibition. These data also argue against significant steady-state H₂S levels in tissues under normoxic conditions. However, when oxygen concentrations are limiting, H₂S levels are expected to rise leading to inhibition of cytochrome c oxidase concomitant with enhanced mitochondrial reactive oxygen species generation, which is in fact observed under hypoxic conditions.

Tiranti and coworkers discuss the effects of chronic sulfide exposure as seen in the inherited disorder, ethylmalonic encephalopathy, on degradation of cytochrome c oxidase (8). Ethylmalonic encephalopathy results from mutations in *Eth1*, the gene encoding the second enzyme in the mitochondrial sulfide oxidation pathway, sulfur dioxygenase. Using tissue-specific conditional *Eth1* knockouts, the authors demonstrate that the cytochrome c oxidase deficiency is limited to the tissues targeted for ablation. Hence, H₂S acts locally in tissue with disrupted *Eth1*, leading to heme *a* inhibition and enhanced degradation of cytochrome c oxidase subunits.

López-Garriga and co-workers (24) discuss hemeoproteins such as cytochrome c oxidase, myoglobin, and hemoglobin as targets for H₂S. Inhibition of cytochrome c oxidase is responsible for decreasing metabolic activity and inducing a hibernation-like state (2). The basis for the differential reactivity of H₂S with hemeoproteins that results in formation of stable hexacoordinate low spin Fe^{III}-SH₂ complex versus iron reduction and conversion to an unligated Fe^{II} heme and the influence of the iron oxidation state are discussed.

The remaining two articles focus on the neurophysiological effects of H₂S. One, a comprehensive review by Jin-Song and colleagues (12), covers the current state of our understanding of the neurophysiology and neuropathology of H₂S, discussing the underlying cellular mechanisms and the neuroprotective effects of this gaseous mediator. The original research article by Ichinose and coworkers examines the protective effects of H₂S on a murine model of Parkinson's disease induced by the neurotoxin, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (17). They demonstrate that the neuroprotective effects of H₂S are correlated with induction of antioxidant genes, including glutamate cysteine ligase and heme oxygenase-1.

Acknowledgment

This work was supported in part by a grant from the National Institutes of Health (HL58984 and DK64959).

References

1. Abe K and Kimura H. The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16: 1066–1071, 1996.
2. Blackstone E, Morrison M, and Roth MB. H₂S induces a suspended animation-like state in mice. *Science* 308: 518, 2005.
3. Bouillaud F and Blachier F. Mitochondria and sulfide: a very old story of poisoning, feeding, and signaling? *Antioxid Redox Signal* 15: 379–391, 2011.
4. Braunstein AE and Goryachenkova EV. The beta-replacement-specific pyridoxal-P-dependent lyases. *Adv Enzymol Relat Areas Mol Biol* 56: 1–89, 1984.
5. Calvert JW, Jha S, Gundewar S, Elrod JW, Ramachandran A, Pattillo CB, Kevil CG, and Lefer DJ. Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. *Circ Res* 105: 365–374, 2009.
6. Chen X, Jhee KH, and Kruger WD. Production of the neuromodulator H₂S by cystathionine beta-synthase via the condensation of cysteine and homocysteine. *J Biol Chem* 279: 52082–52086, 2004.
7. Chiku T, Padovani D, Zhu W, Singh S, Vitvitsky V, and Banerjee R. H₂S biogenesis by cystathionine gamma-lyase leads to the novel sulfur metabolites, lanthionine and homolanthionine, and is responsive to the grade of hyperhomocysteinemia. *J Biol Chem* 284: 11601–11612, 2009.
8. Di Meo I, Fagiolari G, Prella A, Viscomi C, Zeviani M, and Tiranti V. Chronic exposure to sulfide causes accelerated degradation of cytochrome c oxidase in ethylmalonic encephalopathy. *Antioxid Redox Signal* 15: 353–362, 2011.
9. Elrod JW, Calvert JW, Morrison J, Doeller JE, Kraus DW, Tao L, Jiao X, Scalia R, Kiss L, Szabo C, Kimura H, Chow CW, and Lefer DJ. Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. *Proc Natl Acad Sci USA* 104: 15560–15565, 2007.
10. Hildebrandt TM and Grieshaber MK. Three enzymatic activities catalyze the oxidation of sulfide to thiosulfate in mammalian and invertebrate mitochondria. *FEBS J* 275: 3352–3361, 2008.
11. Hosoki R, Matsuki N, and Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant

- in synergy with nitric oxide. *Biochem Biophys Res Commun* 237: 527–531, 1997.
12. Hu LF, Lu M, Wong PTH, and Bian JS. Hydrogen sulfide: neurophysiology and neuropathology. *Antioxid Redox Signal* 15: 405–419, 2011.
 13. Hu Y, Chen X, Pan TT, Neo KL, Lee SW, Khin ES, Moore PK, and Bian JS. Cardioprotection induced by hydrogen sulfide preconditioning involves activation of ERK and PI3K/Akt pathways. *Pflugers Arch* 455: 607–616, 2008.
 14. Ishii I, Akahoshi N, Yamada H, Nakano S, Izumi T, and Suematsu M. Cystathionine gamma-Lyase-deficient mice require dietary cysteine to protect against acute lethal myopathy and oxidative injury. *J Biol Chem* 285: 26358–26368, 2010.
 15. Kabil O and Banerjee R. The redox biochemistry of hydrogen sulfide. *J Biol Chem* 285: 21903–21907, 2010.
 16. Kabil O, Vitvitsky V, Xie P, and Banerjee R. The quantitative significance of the transsulfuration enzymes for H₂S production in murine tissues. *Antioxid Redox Signal* 15: 363–372, 2011.
 17. Kida K, Yamada M, Tokuda K, Marutani E, Kakinohana M, Kaneki M, and Ichinose F. Inhaled hydrogen sulfide prevents neurodegeneration and movement disorder in a mouse model of Parkinson's disease. *Antioxid Redox Signal* 15: 343–352, 2011.
 18. Kimura H. Hydrogen sulfide induces cyclic AMP and modulates the NMDA receptor. *Biochem Biophys Res Commun* 267: 129–133, 2000.
 19. Kimura Y, Goto Y, and Kimura H. Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 12: 1–13, 2010.
 20. Kimura Y and Kimura H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 18: 1165–1167, 2004.
 21. Levitt MD, Abdel-Rehim MS, and Furne J. Free and acid-labile hydrogen sulfide concentrations in mouse tissues: anomalously high free hydrogen sulfide in aortic tissue. *Antioxid Redox Signal* 15: 373–378, 2011.
 22. Nagai Y, Tsugane M, Oka J, and Kimura H. Hydrogen sulfide induces calcium waves in astrocytes. *FASEB J* 18: 557–559, 2004.
 23. Olson KR and Whitfield NL. Hydrogen sulfide and oxygen sensing in the cardiovascular system. *Antioxid Redox Signal* 12: 1219–1234, 2010.
 24. Pietri R, Román-Morales E, and López-Garriga J. Hydrogen sulfide and hemoproteins: knowledge and mysteries. *Antioxid Redox Signal* 15: 393–404, 2011.
 25. Predmore BL and Lefer DJ. Development of hydrogen sulfide-based therapeutics for cardiovascular disease. *J Cardiovasc Transl Res* 3: 487–498, 2010.
 26. Shibuya N, Tanaka M, Yoshida M, Ogasawara Y, Togawa T, Ishii K, and Kimura H. 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 11: 703–714, 2009.
 27. Singh S, Padovani D, Leslie RA, Chiku T, and Banerjee R. Relative contributions of cystathionine beta-synthase and gamma-cystathionase to H₂S biogenesis via alternative transsulfuration reactions. *J Biol Chem* 284: 22457–22466, 2009.
 28. Stipanuk MH and Beck PW. Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. *Biochem J* 206: 267–277, 1982.
 29. Xu Z, Prathapasinghe G, Wu N, Hwang SY, Siow YL, and O K. Ischemia-reperfusion reduces cystathionine-beta-synthase-mediated hydrogen sulfide generation in the kidney. *Am J Physiol Renal Physiol* 297: F27–F35, 2009.
 30. Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K, Meng Q, Mustafa AK, Mu W, Zhang S, Snyder SH, and Wang R. H₂S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. *Science* 322: 587–590, 2008.
 31. Zhao W, Zhang J, Lu Y, and Wang R. The vasorelaxant effect of H₂S as a novel endogenous gaseous K(ATP) channel opener. *EMBO J* 20: 6008–6016, 2001.

Address correspondence to:

Prof. Ruma Banerjee
 Department of Biological Chemistry
 University of Michigan Medical School
 1150 W. Medical Center Dr.
 3320B MSRB III
 Ann Arbor, MI 48109-0600

E-mail: rbanerje@umich.edu

Date of first submission to ARS Central, January 25, 2011; date of acceptance, January 29, 2011.

Abbreviations Used

CBS = cystathionine β -synthase
 CSE = cystathionase
 H₂S = hydrogen sulfide

This article has been cited by: