

EFFECT OF STEREOCHEMISTRY ON ESTER HYDROLYSIS BY
CHOLINESTERASES: IMPLICATIONS FOR RADIOTRACER
DESIGN

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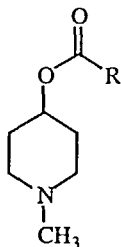
There is currently great interest in developing radiotracers to estimate cortical acetylcholinesterase (AChE) and butyrylcholinesterase (BuChE) enzymatic activity in Alzheimer's disease patients using *in vivo* imaging. As part of an ongoing effort to define structure-activity relationships (SAR) for cholinesterase substrates, we have recently published *in vitro* and *in vivo* kinetic results for a series of 1-[¹¹C]methyl-4-piperidinyl esters (1). These data exhibited the expected preference of AChE for short-chain esters (acetate and propionate) and the specificity of longer-chain esters (butyrate and pentanoate) for BuChE. We have also shown that this general pattern of selectivity extends to the 1-methyl-3-pyrrolidinyl esters, **1a,b** (2). As these latter compounds are chiral, it was of interest to extend the SAR to include the effects of stereochemistry on enzyme selectivity and cleavage rates.

Irie *et al.* previously reported that the AChE-mediated cleavage rates of both the acetate and propionate esters of (R)-1-methyl-3-piperidinol were much more rapid than those of the corresponding (S)-enantiomers (3). Beckett *et al.* have reported a similar relationship for the stereoisomers of α - and β -methylcholine esters (4,5). In our initial study, racemic mixtures of **1a** and **1b** exhibited *in vitro* AChE-mediated cleavage rates similar to the corresponding 4-piperidinyl esters, but the individual enantiomers were not tested (2).

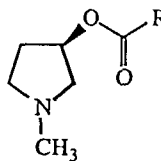
To more clearly define the structure-activity relationships for AChE and BuChE with respect to stereochemistry α to the ester functionality, we have prepared enantiomerically pure (R)-1-methyl-3-pyrrolidinyl propionate, **1a** and butyrate, **1b**, using the methods previously described for 1-methyl-4-piperidinyl propionate, PMP (6). Relative rates of substrate cleavage by purified enzymes (AChE or BuChE) were determined using a simple *in vitro* spectrophotometric assay (1). Esters **1a** and **1b** were also labeled with carbon-11, using methods similar to those reported for [¹¹C]PMP (6), and the *in vivo* biodistribution for each in CD-1 mice was determined.

Based on the work of Beckett *et al.* (4,5), it would be expected that a chiral center β to the ester functionality would also influence enzyme-mediated cleavage rates. To address this question we have also prepared

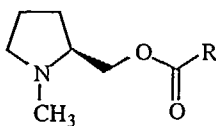
chiral esters of (S)-1-methyl-2-pyrrolidinemethanol, **2a-c**, and (\pm)-1-methyl-2-piperidinemethanol, **3a-c**. *In vitro* and *in vivo* investigations similar to those described above are currently underway.



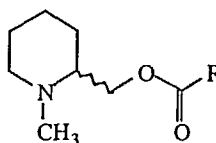
AMP, MP4A R = CH₃
 PMP R = CH₂CH₃
 nBMP R = CH₂CH₂CH₃



1a R = CH₂CH₃
 1b R = CH₂CH₂CH₃



2a R = CH₃
 2b R = CH₂CH₃
 2c R = CH₂CH₂CH₃



3a R = CH₃
 3b R = CH₂CH₃
 3c R = CH₂CH₂CH₃

The preliminary *in vitro* spectrophotometric data for these investigations is given in Table 1. (R)-1-methyl-3-pyrrolidinyl propionate, (R)**1a**, is cleaved by AChE nearly three times faster than both racemic (\pm)**1a** and PMP (Table 1), implying that the (S)-enantiomer of **1a** may act as a weak AChE inhibitor. A much less dramatic stereochemical preference was observed for the BuChE-mediated cleavage of **1a**. This is also consistent with the *in vivo* distribution in mouse brain shown in Table 2. The retention fraction for (R)**1a** was higher than that for both PMP and (\pm)**1a** in all brain areas except striatum. Similarly, the retention fractions for (R)**1b** were higher than those of nBMP in all brain areas.

The racemic primary ester **3b** is also cleaved by AChE more rapidly than is PMP. This preference for the primary ester likely arises from the shorter amine-to-carbonyl distance in **3b**, which better approximates that of acetylcholine. In contrast, (S)-1-methyl-2-pyrrolidinemethyl acetate, **2a**, is cleaved by AChE at a rate only 1.5 times faster than PMP and about one-third the rate of 1-methyl-4-piperidinyl acetate (AMP, MP4A). These results indicate that AChE has a preference for the (R)-configuration at both the α - and β -positions whereas BuChE has little or no stereochemical preference.

Table 1. Relative *in vitro* ChE-mediated cleavage rates. Rates are determined as a change in absorbance (420 nm) of m-nitrophenol per minute per unit of enzyme using the indicated substrates and purified AChE or BuChE. Data are expressed relative to PMP=100 for AChE and nBMP=100 for BuChE.

| Substrate | AChE (rate relative to PMP) | BuChE (rate relative to nBMP) |
|-----------|--------------------------------|----------------------------------|
| AMP,MP4A | 100±0.5 | 33±1 |
| PMP | 20±0.3 | 115±2 |
| (±)1a | 17.2±0.3 | 76±13 |
| (R)1a | 52±1 | 115±8 |
| (S)2a | 32±1.5 | n.d. |
| (±)3b | 81±1.5 | n.d. |
| nBMP | 0.00 | 100±3 |
| (±)1b | 0.00 | 144±22 |

Table 2. Retention fractions calculated from *in vivo* regional distribution in mouse brain at 30 min. post-injection relative to initial uptake at 1 min. post-injection. Values for PMP and (R)1a are an average of 3 experiments (n = 12 total animals per time point), nBMP and 1b are an average of 2 experiments (n = 8) and (±)1a values are from a single study (n = 4).

| Substrate | striatum | cortex | cerebellum | hippocampus | thalamus |
|-----------|-----------|-----------|------------|-------------|-----------|
| PMP | 0.71±0.05 | 0.41±0.03 | 0.26±0.01 | 0.41±0.03 | 0.36±0.10 |
| (±)1a | 0.54±0.07 | 0.40±0.05 | 0.31±0.05 | 0.41±0.05 | 0.39±0.03 |
| (R)1a | 0.55±0.10 | 0.51±0.09 | 0.38±0.08 | 0.53±0.08 | 0.44±0.09 |
| nBMP | 0.11±0.03 | 0.08±0.02 | 0.10±0.02 | 0.10±0.02 | 0.12±0.04 |
| (R)1b | 0.20±0.04 | 0.15±0.02 | 0.18±0.03 | 0.18±0.03 | 0.23±0.03 |

References

- 1) Snyder SE et al. *J Cereb Blood Flow Metab* 21:132-143 (2001).
- 2) Bryan TA et al. *J Labelled Compds Radiopharm* 42:S207-S209 (1999).
- 3) Irie T et al. *J Labeled Compds Radiopharm* 37:214-216 (1995).
- 4) Beckett AH et al. *J Pharm Pharmacol* 15:362-371 (1963).
- 5) Beckett AH et al. *Biochem Pharmacol* 17:1601-1607 (1968).
- 6) Snyder SE et al. *Nucl Med Biol* 25:751-754 (1998).

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