

Supplemental Figure S1

The USCS Genome Browser alignment of the *D. melaogaster* *Ugt36Bc* WRE region with several other *Drosophila* species. Several species with particularly poor conservation have been omitted. The TCF binding sites identified by DNA footprinting in *D. melanogaster* (Figure 4A,B) are over-scored with the AGAWAW sequences shown in bold. DNA sequence conservation breaks down fairly uniformly across the WRE with increasing evolutionary distance. No particularly well-conserved DNA sequences, including the footprinted TCF sites, are obvious in any species.

Supplemental Data

Table S1 Primers for dsRNA, *in situ* probes, real-time PCR, and ChIP analysis

Primers for real-time PCR measurement of gene expression	
<i>naked cuticle</i>	TAAAATTCTCGGCGGCTACAA / CGCACCTGGTGGTACATCAG (#272-273)
<i>CG6234</i>	AGGCGTTGAGGACCCATCTAC / CTGGTTGTCGTCGTGCTCTTC (#157-158)
<i>Ugt36Bc</i>	CCGAATGTGCCTGGAGTAATC / TCCGTGCTTGCCCTTCTC (#161-162)
<i>Tiggrin</i>	AATGGCGGTTATGTGGTGGA / GGAGGCTTTGGTGTAGACAC (#329-330)
<i>Peroxidasin</i>	C TTGCGATG TGGGTGACAA / GTTGCTCCGCTTGGTGTAGG (#327-328)
<i>Ugt58Fa</i>	CGGTGATGCGGACTCTGGT / CTCTTGGGCGAGGAAGTGGT (#331-332)
<i>β-Tubulin56D</i>	AGACCTACTGCATCGACAAAC / GACAAGATGGTTCAGGTCAC (#274-275)
<i>Luciferase</i>	GCCCTTCCGCATAGAACTG / TGATGAAACATTCCAAAACCGTGAT (#384-385)

Primers for real-time PCR measurement of ChIP signals	
<i>Ugt36Bc</i> WRE	CCGAAAGCATGAGTTTGAC / TCGCCCCTTATCTACCCACAAA(#308-309)
Cloned <i>Ugt36Bc</i>	ATTGACCTCGTTGCAGGACAGGATGTGGT /
WRE	ATTGCGGCCGCACATGTTCCAGATGCTGCTCG (#648-649)

Primers used to generate dsRNA

All primers in this section also have the T7 binding sequence
(GCGTTAATACGACTCACTATAGGGAGA) appended to the 5' end.

Ctrl(1) (<i>bla</i>)	CCGTGTCGCCCTTATCCCTT / GACCCACGCTCACCGGCTC (#133-143)
Ctrl(2) (<i>GFP</i>)	AGCTGTTACCGGGTGGT / GGTCACGAACTCCAGCAGG (#804-805)
TCF(1)	GAAGATGACTACGATGATGATAAACTAGGCGGA / AATAGGGTTTCGGGATGTGTTTGCA (Fang et al, 2006)
TCF(2)	TACTGATGAAGTGAAGATTTT / TGCTTCCAGGTTCCCTTGGAT (#743-744)
Arm(1) 3'UTR	CCGAGCTAAGGGTAAGG / GCAAACATTTGTTAATTTTCC (#57-86)
Arm(2) ORF	GGTCACCTGTGCCGCCGGT / AGAGAACAGCGGCGGCGTAT (#184-185)
Axin(1)	ATTCTTGCGCAAGAGTCAGC / CCAGGAGGTCGGATTTCCAG
Axin(2)	GAGACGCCGAAATGCTCTAGAGG / GATCCTGGCGCAGTCCTGACGGC (#404-405)
CtBP ORF	ATGCACAAAGCACCTCCGAAATACACGA / GCACCAGGTCGCATCTGTTTAATTGTGAAT (Fang et al, 2006)
Brinker	TCCAGCCAGCGCTGCAGG / CATCATCATCACCATCACC (#380-381)
Snail	CCGCTCTATACTCGATCCCG / TGGGATGTCATTTGCGACTG (#665-666)
Reptin	AAGGAGGTGGTGACACTGT / AACATGTAGTCATCCTGGTACT (#796-797)

Primers used to generate cRNA *In Situ* probes

<i>Peroxidasin</i>	ATTTAGGTGACACTATAGAAGTGCTGAGCTCACAGAGAACTG / TTAATACGACTCACTATAGGGAGAGTTGCACTTGGTGCAGACGT (#831-832)
<i>Tiggrin</i>	ATTTAGGTGACACTATABAAGTGATGAGGGCTCTGGGCGGGAT / TTAATACGACTCACTATAGGGAGATCCGCTCAAGTCCACGGGTT (#652-653)
<i>Ugt36Bc</i> #1	ATTTAGGTGACACTATAGAAGTGATGACACAAAACCGGAGCAC / TTAATACGACTCACTATAGGGAGATGATTTTGTGATTCAGGAAG (#650-651)
<i>Ugt36Bc</i> #2	ATTTAGGTGACACTATAGAAGTGATGCCCACTTTTGCAGGC / TTAATACGACTCACTATAGGGAGACTACTGCTTCTTACCTTCTTT (#866-867)
