

Themed Section: Opioids: New Pathways to Functional Selectivity

# INTERNATIONAL UNION OF BASIC AND CLINICAL PHARMACOLOGY REVIEW

# Challenges for opioid receptor nomenclature: IUPHAR Review 9

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Recent developments in the study of the structure and function of opioid receptors raise significant challenges for the definition of individual receptor types and the development of a nomenclature that precisely describes isoforms that may subserve different functions *in vivo*. Presentations at the 2013 meeting of the International Narcotics Research Conference in Cairns, Australia, considered some of the new discoveries that are now unravelling the complexities of opioid receptor signalling. Variable processing of opioid receptor messenger RNAs may lead to the presence of several isoforms of the  $\mu$  receptor. Each opioid receptor type can function either as a monomer or as part of a homo- or heterodimer or higher multimer. Additionally, recent evidence points to the existence of agonist bias in the signal transduction pathways activated through  $\mu$  receptors, and to the presence of regulatory allosteric sites on the receptors. This brief review summarizes the recent discoveries that raise challenges for receptor definition and the characterization of signal transduction pathways activated by specific receptor forms.

# **LINKED ARTICLES**

This article is part of a themed section on Opioids: New Pathways to Functional Selectivity. To view the other articles in this section visit http://dx.doi.org/10.1111/bph.2015.172.issue-2

## **Abbreviations**

DOP,  $\delta$  opioid receptor; INRC, International Narcotics Research Conference; IUPHAR, International Union of Basic and Clinical Pharmacology; KOP,  $\kappa$  opioid receptor; MOP,  $\mu$  opioid receptor; N/OFQ, nociceptin/orphanin FQ; NC-IUPHAR, International Union of Basic and Clinical Pharmacology Receptor Nomenclature Committee; NOP, N/OFQ peptide receptor; OP, opioid receptor family; ORNS, Opioid Receptor Nomenclature Subcommittee

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This article, written by members of the International Union of Basic and Clinical Pharmacology Committee on Receptor Nomenclature and Drug Classification (NC-IUPHAR) subcommittee for the opioid receptors, confirms the existing nomenclature for these receptors and reviews our current understanding of their structure, pharmacology and functions and their likely physiological roles in health and disease. More information on this receptor family can be found in the Concise Guide to Pharmacology (http://onlinelibrary.wiley.com/ doi/10.1111/bph.12445/abstract) and for each member of the family in the corresponding database. http://www .guidetopharmacology.org/ GRAC/FamilyDisplayForward? familyId=50&familyType=GPCR



# Links to online information in IUPHAR-DB and the BPS Concise Guide to PHARMACOLOGY

Targets	Ligands
5-Hydroxytryptamine receptors (serotonin receptors)	α-neoendorphin
μ receptor	β-endorphin
$\delta$ receptor	dynorphin A
κ receptor	dynorphin B
NOP receptor	enkephalins
Chemokine receptors	N/OFQ
GPCR	naloxone
orexin-A	
orexin-B	
OT	
LIMK1	
LIMK2	
ROCK1	
ROCK2	
TLR4	

This table lists chemical names, words and phrases that are hyperlinked to relevant entries in http://www.guidetopharmacology.org, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Pawson *et al.*, 2014; PMID: 24234439) and the Concise Guide to PHARMACOLOGY 2013/14 (http://onlinelibrary.wiley.com/doi/10.1111/bph.2013.170.issue-8/issuetoc, Alexander *et al.*, 2013).

Almost two decades ago, the International Union of Basic and Clinical Pharmacology (IUPHAR) established a nomenclature committee to standardize the definitions and characterize the properties of receptors activated by neurotransmitters, hormones, cytokines and many drugs – this committee is known by the acronym NC-IUPHAR. In turn, NC-IUPHAR established subcommittees to make recommendations on specific receptors and to develop a database defining the receptor systems and drug targets coded by the human genome with references to the most appropriate experimental models and the best selective radioligands, agonists and antagonists. This database can be found at http://www.iuphar-db.org; it is a mine of useful information for almost all receptors and ion channels.

The well-established Greek symbol terminology,  $\mu$ ,  $\delta$  and  $\kappa$ , for the first three types of opioid receptors to be identified was proposed by Bill Martin, Hans Kosterlitz and their co-workers (Gilbert and Martin, 1976; Martin et al., 1976; Lord et al., 1977) in the mid-1970s. In 1996, an alternative terminology was proposed by an NC-IUPHAR subcommittee (Dhawan et al., 1996), but this terminology was not accepted by the field and is no longer used. A reconstituted Opioid Receptor Nomenclature Subcommittee (ORNS) proposed a return to the original nomenclature for opioid receptors, and added additional recommendations relating to the opioid receptor family (OP) and the receptor that is selectively activated by the endogenous ligand, nociceptin/orphanin FQ (abbreviated here as N/OFQ). The recommended revised terminology and abbreviations were accepted by NC-IUPHAR, and can be found on the NC-IUPHAR website: http://

www.guidetopharmacology.org/GRAC/FamilyIntroduction Forward?familyId=50. The recommended nomenclature is briefly summarized in Table 1.

The close structural homologies between the three classic types of opioid receptors,  $\mu$ ,  $\delta$  and  $\kappa$ , and the more recently discovered receptor for N/OFQ have been confirmed by the recent reports of the crystal structures of each of these receptors when complexed with antagonists (Granier et al., 2012; Manglik et al., 2012; Thompson et al., 2012; Wu et al., 2012). They are clearly members of one family of proteins, with differences between the receptor types arising by gene duplication events during evolution. It is thus appropriate to group these receptors as a single receptor family. NC-IUPHAR policy is to name receptors after their endogenous ligands, and to require that the abbreviation selected to represent a receptor family is two letters when there would be potential for confusion with other receptors if a single letter were to be used. Given the existence of receptors for oxytocin and orexins, the family name selected by NC-IUPHAR for opioid receptors is OP (i.e. opioid peptide receptors). The Greek symbol terminology for the three receptors of the OP that were first discovered,  $\mu$ ,  $\delta$  and  $\kappa$ , is retained, so these become the  $\mu$ receptor, the  $\delta$  receptor and the  $\kappa$  receptor (or  $\mu OP$  receptor, δOP receptor and κOP receptor). Because it is sometimes inconvenient or impractical to use the Greek symbols, alternative abbreviations recognized by NC-IUPHAR are MOP, DOP and KOP. By analogy, the fourth member of the family becomes the NOP receptor (for nociceptin opioid peptide receptor). Note that in the NC-IUPHAR system the letter R for receptor is never used as part of the receptor name as this



 Table 1

 NC-IUPHAR-approved nomenclature for opioid peptide receptors

Current NC-IUPHAR- approved nomenclature <sup>a</sup>	Other (non-approved) nomenclature	Presumed endogenous ligand(s)
μ, mu or MOP	MOR, OP <sub>3</sub>	β-Endorphin (not selective <sup>c</sup> )
		Enkephalins (not selective <sup>c</sup> )
		Endomorphin-1 <sup>b</sup>
		Endomorphin-2 <sup>b</sup>
$\delta$ , delta or DOP	DOR, OP <sub>1</sub>	Enkephalins (not selective <sup>c</sup> )
		β-Endorphin (not selective <sup>c</sup> )
κ, kappa or KOP	KOR, OP <sub>2</sub>	Dynorphin A
		Dynorphin B
		α-Neoendorphin
NOP	ORL1, OP₄	Nociceptin/orphanin FQ
		(N/OFQ)

Receptors activated by opiate drugs respond physiologically to endogenous opioid peptides; they are therefore opioid peptide receptors, the receptor family being designated by the two-letter abbreviation OP.

adds no information to the terminology; the context usually makes clear that the terminology refers to a receptor. The widely used abbreviations MOR, DOR and KOR are therefore inconsistent with the NC-IUPHAR standards for receptor nomenclature; the ORNS recommends that these abbreviations should not be used to describe opioid receptor types. A summary of the recommended nomenclature and abbreviations for opioid receptor types is presented in Table 1.

Some investigators have questioned whether the NOP receptor should be classified as a member of the OP family, perhaps influenced by Hans Kosterlitz's dictum, frequently repeated by him at International Narcotics Research Conference (INRC) meetings during the 1970s and 1980s, that if a receptor-mediated action is not antagonized by naloxone, then the action should not be called an opioid receptormediated effect. This insistence on a rigid procedural definition of an 'opioid' was valuable at the time. For example, it became apparent that the actions of drugs at the sigma receptor, originally identified by Bill Martin as an opioid receptor (Gilbert and Martin, 1976; Martin et al., 1976), were not antagonized by naloxone (in contrast to Martin's original claim) and should not be called opioid. Subsequent studies have established that the sigma receptor exists, but as a protein that is very different in structure and function from the  $\mu$ ,  $\delta$  and  $\kappa$  receptors (Seth *et al.*, 1998). Furthermore, many of the ligands that activate this receptor have very different structures from the endogenous ligands for the opioid receptors (Hayashi and Su, 2005). Kosterlitz's dictum need not be applied to the entire OP receptor family. The NOP receptor, unlike sigma receptors, is very similar in structure and in

most functions to the other OP receptors. Actions of N/OFQ through the NOP are not antagonized by naloxone, but the amino acid sequence of N/OFQ indicates that this peptide is closely related structurally to the endogenous opioid peptides, probably derived during evolution by gene duplications among the opioid peptide gene family in much the same way as the various OP receptor forms diverged by gene duplication during evolution (Nothacker et al., 1996). As noted earlier, the NOP receptor crystal structure closely resembles the crystal structures of the  $\mu$ ,  $\delta$  and  $\kappa$  receptors and is closer to these than to other GPCRs. Thus, in contrast to sigma receptors, NOP receptors display primary, secondary and tertiary structural similarity to other members of the OP receptor family, and are activated by an endogenous ligand that has a primary structure that is closely related to that of the endogenous ligands for the  $\mu$ ,  $\delta$  and  $\kappa$  receptors. Additionally, the NOP receptor employs a repertoire of signal transduction pathways that is very similar to the set of pathways activated by the three classic opioid receptors. These structural and functional considerations trump the absence of sensitivity to naloxone antagonism and clearly necessitate the assignment of the NOP receptors to the OP receptor family. The NOP receptor should be considered a subcategory of the OP receptor family with atypical low affinity for the classic opioid peptides (the enkephalins, β-endorphin and dynorphin) and insensitivity to antagonism by naloxone.

Several issues that have implications for opioid receptor classification and nomenclature were discussed during the 2013 INRC meeting. An area with potential significance for OP receptor classification is the growing evidence that the

<sup>&</sup>lt;sup>a</sup>The well-established Greek terminology for opioid receptor types using the descriptors  $\mu$ ,  $\delta$  and  $\kappa$  is recommended, but where Greek symbols are not permitted or impractical, the use of mu, delta or kappa, or MOP, DOP or KOP is permissible.

 $<sup>^{</sup>b}$ No mechanism for the endogenous synthesis of endormorphins has been identified; their status as *endogenous* ligands for the  $\mu$  receptor is tentative.

c'Not selective' indicates that these ligands are not strongly selective for the specific receptor types indicated; they may have sufficient affinity and efficacy at other opioid receptors to exert pharmacological effects through the non-preferred site. For example, the enkephalins are listed as non-selective ligands for both  $\mu$  and  $\delta$  receptors. However, all ligands in this table have very low affinity and efficacy at non-opioid GPCRs.

signal transduction pathways that are activated by agonists acting at the same receptor type are not always identical. Evidence that individual agonist ligands may preferentially direct the functional response elicited by their common receptor to different transduction pathways was the subject of a plenary lecture by Arthur Christopoulos on biased agonism at GPCRs. The main emphasis of Christopoulos's talk was on other GPCRs, not specifically on opioid receptors, but other speakers addressed biased agonism at OP receptors. Eamonn Kelly from Bristol showed unambiguously that certain agonists at MOP receptors bias the response towards either G-protein- or  $\beta$ -arrestin-mediated transduction pathways. The signalling pathway repertoire that can be activated by these transducers is also expanding. For example, Wendy Walwyn presented evidence that  $\delta$  and NOP receptors can activate cofilin, an actin-modulating protein, via β-arrestin, Rho-kinase (ROCK) and Lim domain kinase (LIMK). Until recently, it had been assumed that any ligand that could activate a receptor would induce essentially the same cellular response, with the major differences in response relating to the relative efficacies of different agonists. Now that biased agonism at OP receptors is an established fact, apparent differences in the responses induced by agonists that act at the same receptor type do not require the postulation of separate receptor subtypes for each agonist; the same receptor may be differentially biased by each agonist to mediate different transduction pathways.

Functional studies of OP receptors in the 1980 and 1990s suggested the existence of subtypes of the major OP receptor forms; specifically, differences in the relative potencies of selected agonists at  $\delta$  receptors and their differential sensitivities to certain antagonists led to claims of the existence of subtypes of  $\delta$  receptor (see review by Zaki et al., 1996). At  $\mu$ receptors, the actions of some agonists are reported to be more readily antagonized by the irreversible antagonist, naloxonazine, than others (Pasternak and Wood, 1986; Paul et al., 1989). These observations led to the proposal that there are subclasses of  $\delta$  and  $\mu$  receptor, named  $\delta_1$ ,  $\delta_2$ ,  $\mu_1$  and  $\mu_2$ , but no evidence for the existence of more than one gene for the δ or μ receptors exists despite careful homology searches of the genome. Knockout of the  $\delta$  receptor gene is reported to abolish the activity of ligands preferentially acting at both  $\delta_1$ and  $\delta_2$  sites (Filliol *et al.*, 2000). There are also proposals for the existence of subtypes of  $\kappa$  receptor, based on relative agonist potencies for selected actions that appear to be mediated by  $\kappa$  receptors (Rothman et al., 1989). However, a triple knockout of  $\mu$ ,  $\delta$  and  $\kappa$  receptors completely abolishes binding and function of all opioid ligands (Clarke et al., 2002; Martin et al., 2003), indicating that these ligands require at least one of the three receptor members of the OP receptor family for activity.

It is possible that some or all of the data leading to the proposal that there are subtypes of  $\mu$ ,  $\delta$  and  $\kappa$  receptors might be explained by biased agonism. Agonist potency ratios are now only interpretable if the experimental system from which the data are obtained is fully defined, including not only the receptor type mediating the actions, but also the cell type(s), the experimental conditions and the signal transduction systems mediating the measured effects. Examining differences in the relative potencies of a series of agonists in different cell or tissue preparations or *in vivo* was historically

an important approach to the identification of heterogeneity of many receptor types. It was this type of evidence that was used in part to support the proposed  $\delta$  receptor subtypes (Zaki *et al.*, 1996). However, because of the possibility of biased agonism, differential agonist potency or efficacy can no longer be regarded as strong enough evidence to postulate the existence of non-identical receptors as the mediators of these actions, although antagonist dissociation constants continue to provide more robust evidence of receptor heterogeneity. These conclusions have significance for the receptor databases. Agonist potency ratios are of value in the context of highly defined experimental systems but must be interpreted with caution. The possible existence of opioid receptor subtypes should be re-examined in the light of recent studies demonstrating biased agonism at opioid receptors.

Apparent receptor heterogeneity might also be induced by interactions of receptors with interacting proteins or modulating ligands. Cvejic and Devi (1997) and her colleagues reported that OP receptors could form homodimers, and in 1999 they showed the formation of functional heterodimers with ligand-binding properties that differed from those displayed by either of the individual receptor types (Jordan and Devi, 1999). In the case of the  $\delta$ - $\kappa$  receptor heterodimer, the ligand-binding properties were found to match the properties of the putative  $\kappa_2$  subtype (Zukin et al., 1988). Subsequently, many other groups have confirmed the existence of opioid receptor dimers and higher order forms (oligomers), and the observation has been extended to many other GPCRs (Milligan, 2009). Indeed, several non-opioid GPCRs including chemokine and serotonin receptors have now been reported to form functional heterodimers with opioid receptors (Hebert, 2008; Rozenfeld and Devi, 2011; Cussac et al., 2012). But to date it has not been unambiguously demonstrated that the reported  $\mu$ ,  $\delta$  and  $\kappa$  receptor heterogeneity can be accounted for by receptor heterodimerization. Towards this end, reagents that allow detection and evaluation of the endogenous OP heterodimers are being generated and these have begun to show promising results;  $\delta$ - $\mu$  heterodimer-selective antibodies have been useful in revealing morphine-induced up-regulation of this heterodimer in the brain and in demonstrating heterodimerdirected signal trafficking (Rozenfeld and Devi, 2007; Gupta et al., 2010). Ligands selectively targeting the heterodimer have helped demonstrate allosteric modulation of ligand binding and signalling by heterodimerization (Gomes et al., 2011; 2013) as well as the exploration of the pharmacological properties of heterodimers in vivo (Daniels et al., 2005; Milan-Lobo et al., 2013). Finally, cell-permeable peptides that selectively disrupt the  $\delta$ - $\mu$  heterodimer have helped address the contribution of this heterodimer to opioid pharmacology (He et al., 2011). Reagents such as these will be valuable in addressing the extent to which receptor heterogeneity could be attributed to opioid receptors heterodimers in biological systems.

Agonist actions at many GPCRs are additionally subject to either positive or negative regulation by ligands acting through allosteric regulatory sites on the GPCR. The existence of allosteric modulators of the  $\mu$  receptor (Burford *et al.*, 2013) was discussed at the meeting by Andrew Alt and John Traynor. Positive and negative allosteric compounds binding to a GPCR change receptor conformation to either enhance



or inhibit orthosteric agonist binding and receptor activation; positive allosteric modulators may also show agonist effects, such compounds are allosteric agonists. A key finding is that the allosteric modulator-occupied receptor can have differential affinity for some, but not all orthosteric ligands, resulting in probe dependence; moreover, the allosteric modulator may induce, or change the direction of, signalling bias. Thus, differential sensitivity of the activation of receptors by diverse agonists to allosteric regulation offers another potential explanation for the apparent differences in the actions of different agonists at the same receptor. These observations point to the need for additional research describing more completely allosteric modulatory sites on each OP receptor. Because there are changes in relative agonist affinity and/or efficacy, and perhaps induction of a signalling bias, the allosteric modulator-bound receptor may be considered a novel entity. On the other hand, allosteric modulators only subtly alter receptor conformation and so an OP receptor bound to an allosteric modulator remains an OP receptor, based on the structural and functional arguments discussed earlier. Nonetheless, the fact that allosteric modulators can differentially change the ability of agonists to bind to and activate the receptor and may have agonist actions on their own presents new challenges for OP receptor nomenclature, and in particular for opioid ligand nomenclature.

The role of alternative transcription from a single gene as a potential basis for OP receptor subtypes is also controversial. Gavril Pasternak and others have shown that there is more than one transcription start site on the μ receptor gene and multiple alternative mRNA splicing pathways, resulting in multiple transcripts coding for proteins with different primary structures (Abbadie et al., 2000; Koch et al., 2001; Kvam et al., 2004). It is suggested that these variant receptor forms can account for the apparent functional heterogeneity of  $\mu$  receptors (Pasternak and Pan, 2013). There are no reports of multiple start sites and alternative transcripts for the  $\delta$  and  $\kappa$  receptor genes. Thus, the alternative transcript hypothesis is only plausible as a potential explanation for the apparent heterogeneity of  $\mu$  receptors but, with few exceptions (e.g. Liu et al., 2011), there remains much uncertainty about the levels of expression of the variant mRNA forms for this receptor, their stability in the cell and the properties of any proteins expressed from these mRNA variants. The presence of functional receptor proteins derived from variant transcripts (arising from different transcription start sites or alternative mRNA splicing) from a single receptor gene requires further study. If confirmed then a consistent nomenclature differentiating the variant forms of a single receptor will need to be developed.

Sequence variations within opioid receptor genes might cause the expressed receptors to display properties that distinguish these receptors from those of the most frequently expressed receptor form. There are numerous single nucleotide polymorphisms in human opioid receptor genes, but most are rare and none are known to alter the conformations of the expressed receptor (Mague and Blendy, 2010). Only one polymorphism in the coding region of human opioid receptor genes is known to occur with relatively high frequency (rs 1799971; varying from 40 to 50% in some Asian populations to 5% or less in African Americans; Gelernter et al., 1999), and its known functional consequences are

limited. A change of adenosine to guanosine in position 118 (A118G) of the coding region (exon 1) of the human MOP gene results in the expression of a receptor with aspartic acid (Asp) in position 40 instead of asparagine (Asn); this change removes a potential glycosylation site. A transgenic mouse line in which guanosine is replaced by adenosine in the equivalent position of the mouse  $\mu$  receptor gene (A112G, expressing N38D) resulted in expression of receptors with similar ligand-binding properties but reduced levels of expression of the receptor mRNA and reduced receptor protein levels relative to the wild-type receptor (Mague et al., 2009). This is consistent with other reports that downstream signalling is impaired in the variant form relative to the wild-type human μ receptor (Mague and Blendy, 2010; Oertel et al., 2012). The mechanism for the reduced level of expression may relate to increases in methylation of the 118G  $\mu$ receptor gene. Oertel et al. (2012) report that the degree of gene methylation at positions +117 and adjacent downstream methylation sites was higher in heroin-using subjects expressing the 118G variant than in 118A expressing subjects. In 118G subjects, chronic heroin use was not associated with elevated levels of  $\mu$  receptor expression, but in 118A subjects chronic heroin use induced an up-regulation of μ receptor expression in the thalamus and a cortical region. Increased receptor methylation in the 118G receptor gene may impede receptor up-regulation in response to drug exposure, suggesting an epigenetic regulation of the level of expression of  $\mu$  receptors (Oertel et al., 2012). This work requires confirmation in a larger set of subjects. To date, however, there are no reports indicating that functional opioid receptors with altered ligand binding or signal transduction properties are produced as a result of polymorphisms in opioid receptor genes.

Some opioid drugs have been reported to bind to nonopioid receptor proteins such as filamin A, which interacts directly with  $\mu$  receptors (Wang et al., 2008), or to the Toll-like receptor-4 (TLR-4) (Hutchinson et al., 2011) that does not interact directly with opioid receptors. Naturally, the structural requirements for interaction of opioids with these proteins are very different from their binding to classical opioid receptors. Nonetheless, the interactions might be important, for example, direct actions of opioids on the TLR-4 complex have been proposed to activate microglia to mediate many of the adverse effects of morphine (Hutchinson et al., 2011). However, this potential mechanism has been ruled out in other studies of morphine-induced microglial activation (Ferrini et al., 2013; Fukagawa et al., 2013), and the affinity of interaction of opioids with TLR-4 mechanisms is at least several orders of magnitude weaker than their interaction with MOP (Wang et al., 2012), questioning their pharmacological relevance. Thus, the functional relevance of binding of some opioid drugs to proteins other than the opioid receptors is not clearly established. As these interactions do not involve direct binding to opioid receptors, it is not feasible to define them within the framework of an opioid receptor nomenclature, but investigators need to be aware that ligands for opioid receptors, like many other receptor ligands, can interact with other proteins with possible functional consequences, whether with very high affinity as for filamin A (Wang et al., 2008) or with low affinity as for TLR-4 (Wang et al., 2012).



# Concluding comments

There has been a growing consensus on the use of recommended opioid receptor nomenclature shown in Table 1 since the publication of the most recent NC-IUPHAR recommendation. If and until the accepted nomenclature for opioid receptors is revised to encompass the proposed variant forms of  $\mu$ ,  $\delta$  and  $\kappa$  receptors based on more stringent criteria that take into account the additional variables in receptor properties outlined earlier, we suggest that the simple classification in Table 1 be used by all authors.

Beyond  $\mu$ ,  $\delta$ ,  $\kappa$  and NOP receptors, a description of opioid receptor subtypes such as  $\mu_1$  or  $\mu_2$  is not recommended unless they are described as putative. We do not consider that the evidence for opioid receptor subtypes that has been offered to date provides unambiguous evidence of independent functional receptors that are not variant forms of the major opioid receptors. Until a new nomenclature for opioid receptor variants encompassing possible alternative transcription start sites or splice sites, receptor homo- and heteromultimers, positive and negative allosteric regulation, and biased agonism is established and accepted by the research community, any such proposed variants should be fully described. Evidence of activation of signalling pathway(s) not previously associated with opioid receptors, the identification of novel allosteric regulatory site(s) and the establishment of novel polymeric structures should not be considered sufficient justification for modification of the current nomenclature.

# **Conflict of interest**

The authors all assert that they have no financial conflicts of interest.

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