

Recurrent Deletions and Duplications of Chromosome 2q11.2 and 2q13 are Associated with Variable Outcomes

Kacie N. Riley,^{1,2} Lisa M. Catalano,¹ John A. Bernat,³ Stacie D. Adams,³ Donna M. Martin,^{3,4} Seema R. Lalani,⁵ Ankita Patel,⁵ Rachel D. Burnside,² Jeffrey W. Innis,^{3,4} and M. Katharine Rudd¹*

¹Department of Human Genetics, Emory University School of Medicine, Atlanta, Georgia

²Department of Cytogenetics, Laboratory Corporation of America Holdings, Center for Molecular Biology and Pathology, Research Triangle Park, North Carolina

³Department of Pediatrics and Communicable Diseases, University of Michigan, Ann Arbor, Michigan

⁴Department of Human Genetics, University of Michigan, Ann Arbor, Michigan

⁵Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, Texas

Manuscript Received: 16 November 2014; Manuscript Accepted: 17 July 2015

Copy number variation (CNV) in the long arm of chromosome 2 has been implicated in developmental delay (DD), intellectual disability (ID), autism spectrum disorder (ASD), congenital anomalies, and psychiatric disorders. Here we describe 14 new subjects with recurrent deletions and duplications of chromosome 2q11.2, 2q13, and 2q11.2-2q13. Though diverse phenotypes are associated with these CNVs, some common features have emerged. Subjects with 2q11.2 deletions often exhibit DD, speech delay, and attention deficit hyperactivity disorder (ADHD), whereas those with 2q11.2 duplications have DD, gastroesophageal reflux, and short stature. Congenital heart defects (CHDs), hypotonia, dysmorphic features, and abnormal head size are common in those with 2q13 deletions. In the 2q13 duplication cohort, we report dysmorphic features, DD, and abnormal head size. Two individuals with large duplications spanning 2q11.2-2q13 have dysmorphic features, hypotonia, and DD. This compilation of clinical features associated with 2q CNVs provides information that will be useful for healthcare providers and for families of affected children. However, the reduced penetrance and variable expressivity associated with these recurrent CNVs makes genetic counseling and prediction of outcomes challenging. © 2015 Wiley Periodicals, Inc.

Key words: 2q11.2 deletion; 2q11.2 duplication; 2q13 deletion; 2q13 duplication; recurrent CNV; *FBLN7*; *TMEM87B*

INTRODUCTION

Approximately 15–20% of children referred for chromosomal microarray analysis (CMA) testing have a clinically relevant CNV that in many cases explains their phenotype [Rudd et al., 2009; Cooper et al., 2011; Kaminsky et al., 2011]. However, for almost all pathogenic CNVs there are a range of phenotypic outcomes (variable expressivity)

How to Cite this Article:

Riley KN, Catalano LM, Bernat JA, Adams SD, Martin DM, Lalani SR, Patel A, Burnside RD, Innis JW, Rudd MK. 2015. Recurrent deletions and duplications of chromosome 2q11.2 and 2q13 are associated with variable outcomes.

Am J Med Genet Part A 167A:2664-2673.

and/or unaffected family members who carry the same CNV (incomplete penetrance) [Deak et al., 2011; Moreno-De-Luca et al., 2013; Rosenfeld et al., 2013]. Thus, to capture the spectrum of features associated with a new genomic disorder, it is essential to evaluate multiple individuals with the same CNV. Deletions and duplications mediated by non-allelic homologous recombination (NAHR) are ideal for genotype-phenotype correlations because they have recurrent breakpoints in unrelated individuals [Stankiewicz and Lupski, 2010; Liu et al., 2011; Dittwald et al., 2013].

*Correspondence to:

M. Katharine Rudd, Ph.D., Department of Human Genetics, Emory University School of Medicine, 1518 Clifton Rd 5049 Claudia Nance Rollins, Atlanta, GA 30322.

Email: katie.rudd@emory.edu

Article first published online in Wiley Online Library (wileyonlinelibrary.com): 31 July 2015

DOI 10.1002/ajmg.a.37269

Conflict of interest: None.

Grant sponsor: NIH; Grant number: R01MH092902; Grant sponsor: Emory Genetics Laboratory Director Grant Program; Grant sponsor: Sanofi/Genzyme Corporation Fellowship; Grant sponsor: AHA; Grant number: #13GRNT15810006; Grant sponsor: The University of Michigan; Grant number: NIH R01DC009410.

Large duplications of the long arm of chromosome 2 have been described in children with DD, ID, microcephaly, short stature, and cleft lip and palate [Riegel and Schinzel, 2002; Ounap et al., 2005]. These duplications were originally identified by chromosome banding and span regions as large as 2q11.2-2q21. Six years ago, we reported seven patients with recurrent deletions and duplications of 2q11.2 and 2q13 [Rudd et al., 2009]. Based on CNV inheritance, normal variation in control CNV datasets, and genes within CNVs, we concluded that 2q11.2 CNVs were likely pathogenic whereas 2q13 CNVs had unknown clinical significance. Since then, similar CNVs have been reported in other studies of DD/ID, ASD, CHD, and schizophrenia [Cooper et al., 2011; Soemedi et al., 2012; Yu et al., 2012; Costain et al., 2013; Dittwald et al., 2013; Russell et al., 2014]. Here we present 14 new subjects with genomic imbalances of 2q11.2-2q13 and their phenotypes as compared to published cases, representing the largest study of chromosome 2q11.2-2q13 rearrangements to date.

MATERIALS AND METHODS Human Subjects

This study was approved by the Institutional Review Boards at Emory University, Baylor College of Medicine, and The University of Michigan. Healthcare providers referred families to our research study after testing in diagnostic cytogenetics laboratories. We recruited 14 new subjects: five with deletions of 2q11.2, three with duplications of 2q11.2, three with deletions of 2q13, one with a duplication of 2q13, and two with different duplications of the intervening region between 2q11.2 and 2q13. We compared the clinical features from new subjects and previously reported subjects [Rudd et al., 2009; Yu et al., 2012; Russell et al., 2014]. Subjects range in age from six months to 17 years at the time of assessment and have various ethnic backgrounds. Informed consent was obtained from probands and their family members. Subjects 1, 4, 5, 7-10, 12-14 were consented at Emory University, Subjects 2, 3, and 6 were consented at Baylor College of Medicine, and Subject 11 was consented at The University of Michigan. Once informed consent was provided, medical records and clinical notes were obtained from primary care physicians and relevant specialists.

Chromosomal Microarray Analysis (CMA)

Deletions and duplications were detected by microarray analysis of DNA extracted from peripheral blood. Most subjects were tested with a version of the EmArray oligonucleotide array [Baldwin et al., 2008]. Agilent microarray design identifier (AMADID) numbers correspond to custom array designs (Agilent Technologies, Santa Clara, CA). Subject 10 was tested using the EmArray Cyto6000 version 2 (AMADID 0246141) and Subjects 9, 12, and 13 were tested with the EmArray Cyto60K (AMADID 0246121); both are based on the NCBI36/hg18 genome assembly. Subject 1 was tested using EmArray Cyto (AMADID 0275741), an Oxford Gene Technology (OGT) array based on the GRCh37/hg19 genome assembly. Subjects 4–7 were tested using the Affymetrix CytoScan HD array and Subjects 11 and 14 were tested using the Illumina CytoSNP-850K BeadChip, both based on the GRCh37/hg19 genome assembly. Subjects 2 and 3 were tested using the 180k CMA-HR, based on the NCBI36/hg18 genome assembly. Coordinates from arrays using the NCBI36/hg18 build were converted to GRCh37/hg19 using the LiftOver tool (http://genome.ucsc.edu) [Kent et al., 2002].

DNA digestion, labeling, purification, hybridization, array scanning, and analysis were performed following manufacturers' instructions. To determine the inheritance of CNVs, fluorescence in situ hybridization (FISH) analysis was performed on peripheral blood samples from parents using standard cytogenetic procedures with probes corresponding to the proband's CNV.

RESULTS

2q11.2 Deletions

Speech delay, ADHD, and dysmorphic features are common in children with deletions of chromosome 2q11.2 (Fig. 1; Table I). Subject 1's father carries the 2q11.2 deletion, and describes "trouble in school" and two nephews with ADHD. Subject 2's deletion occurred de novo and Subject 3's deletion was maternally inherited. Subjects 4 and 5 are brothers with the same 2q11.2 deletion, but their parents have not been tested. The 2q11.2 deletion is not present in databases of normal CNV [Itsara et al., 2009; Shaikh et al., 2009; Macdonald et al., 2014].

2q11.2 Duplications

DD, dysmorphic features, gastroesophageal reflux, and short stature are present in individuals with 2q11.2 duplications (Table II). The three children who were assessed exhibited frontal bossing with other minor facial features. Subject 6 has begun growth hormone therapy. Her mother does not carry the duplication; her father had learning difficulties and short stature, but was not available for testing. Subject 7 inherited her duplication from her unaffected mother, and her half-brother (Subject 8) carries the same 2q11.2 duplication. Both children have short stature, feeding difficulties, and gastroesophageal reflux with vomiting. One duplication of 2q11.2 was reported in an individual from the 1000 Genomes project [Consortium et al., 2010]; however, 2q11.2 duplications are not found in other databases of normal CNV [Itsara et al., 2009; Shaikh et al., 2009; Macdonald et al., 2014].

2q13 Deletions

Eleven subjects have 1.62–1.71-Mb deletions of chromosome 2q13 flanked by the same cluster of segmental duplications. Patient 2's atypical 1.35-Mb deletion partially overlaps the common 2q13 deletion region, but is shifted distally (Fig. 2) [Russell et al., 2014]. Both regions are distal to the smaller 2q13 CNV that includes the *NPHP1* gene [Konrad et al., 1996]. Congenital heart defects (7/12), dysmorphic features (10/12), hypotonia (7/12), and abnormal head size (9/12) are common in those with 2q13 deletions (Table III). Cognitive impairment or ASD was reported in 4 of the 7 subjects who were evaluated; however, several subjects in our study were less than two years of age at the time of assessment so their abilities could not be determined. Five



FIG. 1. Features of subjects with 2q CNVs. Subjects 6-8 have duplications of 2q11.2. A: Subject 6 at 10 years. B: Subject 7 at 21 months. C-D: Subject 8 at 21 months and three years, respectively. E: Subject 1 (2q11.2 deletion) at four years. F: Subject 12 (2q13 duplication) at eight years.

out of eight males with 2q13 deletions in our combined studies exhibited hypogonadism or hypospadias [Yu et al., 2012; Russell et al., 2014]. One 1.71-Mb deletion of 2q13 has been described in the CHOP study of normal CNV [Shaikh et al., 2009], and a

similar 2q13 deletion was described in one individual from the Cooper et al. control group [Cooper et al., 2011]. The atypical distal deletion of 2q13 has not been described in databases of normal CNV [Russell et al., 2014].

			1				
	Patient 2	Subject 1	Subject 2	Subject 3	Subject 4	Subject 5	Total
Deference				This study	(ranny 1) Thio otudu	(rannig I) This studu	TUCAI
Reference	[2009]	This study	mis study	This study	This study	This study	
Age at assessment	16 yrs	4 yrs	4 yrs, 10 mos	7 yrs, 1 mos	17 yrs	15 yrs	
Gender	М	М	М	F	М	М	
Ethnicity	_	Caucasian	_	Caucasian	African American	African American	
Size of CNV	1.25 Mb	1.25 Mb	913 kb	913 kb	1.61 Mb	1.60 Mb	
Coordinates	96,766,561-	96,766,565-	96,739,309-	96,739,309-	96,097,383-	96,097,383-	
	98,013,866	98,013,954	97,652,262	97,652,262	97,703,370	97,693,317	
Inheritance	Unknown	Paternal	De Novo	Maternal	Unknown, familial	Unknown, familial	
DD/ID	_	+	+	_	+	+	4/6
ASD	_	+	-	-	+	+	3/6
Speech delay	-	+	+	+	-	+	4/6
ADHD	+	+	+	+	-	-	4/6
Hypotonia	-	+	+	-	-	+	3/6
Dysmorphic	+	Midface hypoplasia,	Malformed ears,	Malformed ears	-	-	4/6
features		coarse features, hupertelorism	midface hypoplasia				
		depressed nasal bridge					
Skeletal	Scoliosis	-	Pectus	_	Mild scoliosis	Kyphoscoliosis,	4/6
			excavatum			kyphosis, lordosis, microcephaly	
Other clinical	Café-au-lait	Recurrent ear	Inversion of the	Inversion of the	Encephalopathy,	Encephalopathy,	
findings	spots, aortic coarctation	infections, hip joint hypermobility	foot, recurrent ear infections	foot, uncontrolled eating	sleep problems, mood disorder, aggression, mild	asthma, ichthyosis, chiari malformation, mood disorder,	
					motor delays	aggiession	

TABLE I. Clinical Features of Subjects with Deletions of Chromosome 2011.2

+, feature present; --, feature absent; DD, developmental delay; ID, intellectual disability; ASD, autism spectrum disorder; ADHD, attention deficit hyperactivity disorder. Genomic coordinates for chromosome 2 are based on the GRCh37/hg19 build of the human genome.

2q13 Duplications

Four subjects have a duplication of 2q13 that is reciprocal to the common 2q13 deletion. Dysmorphic features, DD, and microcephaly are common findings in this group (Table IV). Two of our subjects with 2q13 duplications are members of the same family (Fig. 3). Patient 4 [Rudd et al., 2009] has a nephew (Subject 12) with DD, learning disabilities, and dysmorphic features. Subject 12's mother, who also carries the duplication, reported difficulty with math, but strengths in expressive language and reading. She also described some difficulties with fine motor skills and visual integration. Duplications of this region of chromosome 2q13 are not reported in normal CNV databases [Itsara et al., 2009; Shaikh et al., 2009; Macdonald et al., 2014].

2q11.2–2q13 Duplications

Two subjects have large duplications between the common CNV regions in chromosome 2q11.2 and 2q13. Subject 13 has a 9.22-Mb duplication of 2q11.2–2q13 (hg19, chr2:102,327,289–111,548,995) and subject 14 has a 3.33-Mb duplication of 2q12.2–2q13 (hg19, chr2:107,132,930–110,465,307) (Fig. 2). The distal breakpoint of both duplications lies in a cluster of segmental duplications in

2q13, but the proximal breakpoints are different. Subject 14's duplication is flanked by paralogous segmental duplications in 2q12.2 and 2q13, whereas Subject 13's proximal 2q11.2 breakpoint is not bounded by segmental duplications. Developmental delay, hypotonia, and various dysmorphic features were present in Subject 13 and 14. As for the parents of our subjects who also carry these duplications, Subject 13's mother reported fainting spells and chest pain with suspected mitral valve prolapse. Subject 14's father is reportedly healthy, though he did not finish high school for unspecified reasons.

DISCUSSION

In this study, we describe 14 new subjects with pathogenic CNVs involving chromosome 2q11.2, 2q13, and 2q11.2–2q13 and compare their features to others in the literature. Evaluating multiple individuals with the same CNV can shed light on the range of phenotypes associated with a deletion or duplication.

Chromosome 2q13 deletions are the most common CNV in our study, and between our subjects and others, a consistent phenotype is beginning to emerge. Dysmorphic features, cranial abnormalities, DD, and CHD are the most common features. In all of the

			Subject 7	Subject 8	
	Patient 1	Subject 6	(Family 2)	(Family 2)	Total
Reference	Rudd et al. [2009]	This study	This study	This study	
Age at assessment	16 mos	10 yrs	13 months	10 years	
Gender	М	F	F	М	
Ethnicity	Indian	Hispanic	Caucasian	Caucasian	
Size of CNV	1.47 Mb	1.38 Mb	1.49 Mb	Not determined	
Coordinates	96,545,351-	96,734,646-	96,732,520-	FISH confirmed	
Inhoritanco		Joknown	Maternal	Matarnal	
		UIKIIOWII	Materia	NA	2/2
Hupotopia	+ +	т _		NA	2/3
Dusmorphic	+ +	Prominent foreboad	Prominent forebead	NA	2/3
features	Ŧ	flattened nasal bridge, protuberant ears	triangular face, epicanthal folds, short philtrum,	NA	275
-			high arched palate		
Short stature	+	+		+	4/4
Skeletal	Macrocephaly, dolicocephaly, frontal bossing	Microcephaly (2nd% tile), scoliosis, frontal bossing	Frontal bossing, mild leg asymmetry	NA	3/3
Gastroesophageal reflux	+	+	+	+	4/4
Other clinical findings	Café-au-lait spots	FTT, low weight (<5th% tile), joint pain, hypermobility, speech delau	Feeding difficulties, reflux with vomiting, FTT, low weight (5th% tile), PF0	Feeding difficulties, reflux with vomiting, eosinophilic esophagitis, ADHD	
Other cytogenetic findings	inv(Y) (p11.2q11.2)				
FTT, failure to thrive; NA, not	assessed; PFO, patent foramen	ovale.			

TABLE II. Clinical Features of Subjects with Duplications of Chromosome 2q11.2

Genomic coordinates for chromosome 2 are based on the GRCh37/hg19 build of the human genome.

families where parents were tested, the 2q13 deletion was inherited. However, little or no phenotypic information was available from the parents who carried the deletion. The 2q13 deletion is enriched in other cohorts of individuals with DD/ID [Bisgaard et al., 2007; Cooper et al., 2011], ASD [Szatmari et al., 2007], and CHD [Soemedi et al., 2012] as compared to controls, consistent with a pathogenic CNV. Recent knockdown experiments in zebrafish point to two genes involved in the 2q13 deletion phenotype. Abnormal head size and dysmorphic features may be due to heterozygous loss of *FBLN7* since depletion of this gene leads to craniofacial defects in fish. In addition, knockdown of *FBLN7* or *TMEM87B* individually and synergistically recapitulated heart abnormalities in fish [Russell et al., 2014].

Dysmorphic features and DD, but not CHDs, are present in those with the reciprocal duplication of 2q13. These data suggest that haploinsufficiency, but not triplosensitivity, of 2q13 cause CHDs. Within Family 3, there is variable expressivity of the 2q13 duplication in three individuals with DD, learning disabilities, and/ or dysmorphic features. Duplications of 2q13 have been reported in other children with DD, congenital anomalies, dysmorphic features, and/or ASD [Cooper et al., 2011]. Two subjects with 2q13 duplications have also been described in a schizophrenia cohort [Costain et al., 2013]. All three of the subjects in our studies inherited their duplications, as did three from Cooper et al., and others have unknown inheritance [Cooper et al., 2011; Yu et al., 2012; Costain et al., 2013]. In addition, 2q13 duplications are not reported in normal CNV databases [Itsara et al., 2009; Shaikh et al., 2009; Macdonald et al., 2014]. This, along with the segregation of the duplication in Family 3, suggests that 2q13 duplications are pathogenic with variable expressivity.

Deletions and duplications of 2q11.2 are also considered pathogenic. The 2q11.2 deletion is significantly enriched in children with developmental delay as compared to controls [Coe et al., 2014]. In our cohort, the deletion was inherited from a mildly affected parent (Subject 1), de novo (Subject 2), or present in two similarly affected siblings (Family 1), consistent with pathogenicity. The 2q11.2 duplication was found to be de novo (Patient 1) or present in siblings with similar clinical presentations (Family 2). The short stature observed in all four children with 2q11.2 duplications is striking and prompted other genetic testing. Subject 7 was



FIG. 2. CNVs in chromosome 2q11.1–2q13. Minimum CNV sizes are represented by red (deletion) or blue (duplication) bars and segmental duplications are shown below. The recurrent *NPHP1* deletion/duplication region is shown in green. RefSeq genes (blue) are indicated at the bottom. This figure was made with the PDF/PS tool on the UCSC genome browser (GRCh37/hg19) [Kent et al., 2002].

referred for Russell-Silver testing and Patient 1 was referred for *FGFR3* sequencing; both tests were negative.

Dittwald et al. recently described eight patients with deletions of the 2q12.2–2q13 region [Dittwald et al., 2013]. These deletions are 502 kb–1.91 Mb, lie between paralogous segmental duplications, and overlap the larger duplications in Subjects 13 and 14. Like Subjects 13 and 14, several of the patients with deletions exhibited DD and dysmorphic features. Our Subject 14 and Dittwald et al.'s patient 1 have temporal bone narrowing and Subject 13 has dolichocephaly and frontal bossing. Since all three of these CNVs overlap, it is possible that they include a gene involved in cranial development. However, none of the genes within the smallest region of overlap have been implicated in craniofacial development. Some of the deletions reported by Dittwald et al. are present in databases of normal CNV, but the larger duplications in Subjects 13 and 14 are not [Itsara et al., 2009; Shaikh et al., 2009; Macdonald et al., 2014].

Like other recurrent CNVs [Deak et al., 2011; Moreno-De-Luca et al., 2013; Rosenfeld et al., 2013], deletions and duplications of 2q11.2 and 2q13 exhibit incomplete penetrance and variable expressivity. It is possible that parents who carry the same CNV as their affected child have milder clinical presentations that were not recognized during childhood. On the other hand, affected children may carry additional "second hit" CNVs that exacerbate penetrance of the phenotype [Girirajan et al., 2012]. Patient 5 has a 2q13 deletion and an unbalanced translocation between chromo-

	Patient 5	Patient 6	Patient 7	Subject 9	Subject 10	Subject 11	Patient 1	Patient 2	Case 1	Case 2	Case 3	Case 4	Total
Reference	Rudd et al. [2009]	Rudd et al. [2009]	Rudd et al. [2009]	This study	This study	This study	Russell et al. [2014]	Russell et al. [2014]	Yu et al. [2012]	Yu et al. [2012]	Yu et al. [2012]	Yu et al. [2012]	
Age at	4 yrs	2 yrs	d. <1 yr	<1 yr	5 yrs	6 mos	2 mos	3 yrs	2 yrs	6 yrs	6 yrs	2 yrs	
Gender	Σ	Σ	Σ	Ŀ	Σ	Σ	(a. o mos) F	Σ	Ŀ	Σ	L	Σ	
Ethnicity	Caucasian	Hispanic	Caucasian/	Caucasian	Caucasian	Caucasian	African-	African-	- 1	- I	- 1	1	
			Hispanic				American	American					
Size of CNV	1.62 Mb	1.62 Mb	1.62 Mb	1.62 Mb	1.62 Mb	1.70 Mb	1.62 Mb	1.35 Mb	1.71 Mb	1.71 Mb	1.71 Mb	1.71 Mb	
Coordinates	111,442,131- 112 DEE 220	111,442,131- 112 DEE 230	111,442,131- 112 DEE 230	111,442,176- 112 065 241	111,442,131- 112065 770	111,675,789-	111,442,131- 112,055 730	112,592,088- 112,027,615	111,392,198- 112102 EQA	111,392,198- 112,102 E04	111,392,198- 112,102,504	111,392,198– 112,102,E04	
Inheritance	Unknown	Paternal	llnknown	Paternal	Paternal	De Novo	Maternal	llnknown	Maternal	Uhknown	Maternal	Paternal	
DD/ID	NA		NA	NA	+	NA	NA	+		1	+	+	4/7
ASD	NA	I	NA	NA	NA	NA	NA	NA	I	+	· +	NA	2/4
Speech delau	I	+	NA	NA	+	NA	NA	+	I	• 1	• +	I	3/8
Hypotonia	I	I	+	I	+	+	+	I	I	+	+	+	7/12
Dysmorphic	+	+	+	+	+	I	+	+	+	+	I	+	10/12
sampai			010										0
CHU	IAPVK, VSU, PDA.	I	VSU, PFU, 2/6 murmur	AVSU, HLV, ASD. aortic	Ventricular hupertrophu.	I	HLV, ASU, PDA, DORV.	IUF, VSU, PA. RSAA.	USV	1	I	I	<i>د/</i> 12
	heterotavii			coarctation	slight murmur		PA 2/6 murmur	2/6 miltmilt					
	aortic			hypoplastic aortic									
	coarctation			arch, 2/6									
				murmur									
Skeletal	Ι	Ι	Microcephaly	Microcephaly	Microcephaly	Macrocephaly	Microcephaly	Microcephaly [Mild],	Macrocephaly	I	Macrocephaly	Macrocephaly,	9/12
			[<1st% tile]	[2nd% tile]			(Slight)	absent right 5th metatarsals/				scoliosis	
Other clinical		GERD.	Esophageal atresia.	GERD. short	Duspha <i>g</i> ia, recurrent	Hupospadias.	Cleft lip/palate.	Hupospadias.	Anterior anus	Hupovonadism.	Lack of	Tall stature.	
findings		hupopiamented	hunoronadism.	stature	bronchitis.	seizures.	stiff Joints	bronchomalacia	displacement.	tall stature.	coordination	hupogonadism.	
0		spots	stiff joints,	[<5th% tile]	7th nerve palsy	sleep apnea			hip displacement	obesity, seizures, panhupopituitarism		molar tooth malformation	
Other			der[1] t[1;9]						arr[hg19]	- 5	arr[hg19]		
cytogenetic			[q43;p21.3].						2p22[32,480,016-		22q11.21		
findings			arr[hg18]						33,189,373]×3		[17,270,271-		
)			1q43q44								19,891,514) × 3		
			[237,788,353]										
			×1,9p24.3p21.3										
			[194,193–										
			22,076857) × 3										
CHD, congenital	I heart defect, TAI	PVR, total anomalc	bus pulmonary venou:	s return; VSD, ventric	ular septal defect; PD/	A, patent ductus ar	teriosis; AVSD, atric	oventricular septal defe	ct; HLV, hypoplastic	left ventricle; ASD, at	rial septal defect; l	DORV, double outle	et right
ventricle; PA, p Genomic coordi	ulmonary atresion inates for chrom	a; IUF, tetralogy (osome 2 are bas	of fallot; KSAA, right-s ied on the GRCh37/h	ided aortic arch; שב 19 build of the hun	.KU, gastroesophagea nan genome.	l reflux disorder.							

TABLE III. Clinical Features of Subjects With Deletions of Chromosome 2q13

2670

Reference	Patient 3 Rudd et al. [2009]	Patient 4 (Family 3) Rudd et al. [2009]	Subject 12 (Family 3) This study	Case 5 Yu et al. [2012]	Total
Age at assessment	3 yrs	13 yrs	8 yrs	6 yrs	
Gender	M	M	M	M	
Ethnicity	Caucasian	Caucasian	Caucasian	-	
Size of CNV	1.62 Mb	1.62 Mb	1.62 Mb	1.71 Mb	
Coordinates	111,442,131-	111,442,131-	111,442,131-	111,392,198-	
	113,065,779	113,065,779	113,065,779	113,102,594	
Inheritance	Paternal	Paternal	Maternal	Unknown	
DD/ID	+	+	+	+	4/4
ASD	_	+	-	+	2/4
Hypotonia	+	-	NA	+	2/3
Dysmorphic features	+	+	+	-	3/4
Skeletal	Microcephaly (Mild)	Microcephaly (Mild)	NA	Plagiocephaly	3/3

TABLE IV. Clinical Features of Subjects With Duplications of Chromosome 2q13

Genomic coordinates for chromosome 2 are based on the GRCh37/hg19 build of the human genome.

somes 1 and 9 [Rudd et al., 2009]. Two patients with 2q13 deletions reported in Yu et al. also carry additional CNVs. Case 1 has a 709-kb duplication of chromosome 2p22 and Case 3 has a 2.62-Mb duplication of chromosome 22q11.21, which overlaps with the 22q11.2 Duplication syndrome critical region [Yu et al., 2012]. It is possible that these additional CNVs contribute to or modify the phenotypes of those with 2q13 deletions. Since none of the CNVs in our study are common in databases of normal variation, it is



FIG. 3. 2q13 duplication family. Variable phenotypes associated with the 2q13 duplication in Family 3. Those tested for the duplication are indicated by 2q13 + or 2q13—for positive and negative results, respectively. Subject 12's brothers and aunt were not tested (NT). DD = developmental delay, LD = learning disability, DF = dysmorphic features.

unlikely that they are benign variants without phenotypic consequences.

Though we have expanded the phenotypes associated with these genomic rearrangements of 2q11.2 and 2q13, without a recognizable set of specific features, clinical diagnosis of microdeletions and microduplications of the long arm of chromosome 2 is not possible. However, this "genotype-first" approach is fast becoming the new standard for diagnosis of children with DD/ID, ASD, and congenital anomalies [Watson et al., 2014]. Though these CNVs exhibit variable expressivity, recognizing the spectrum of associated features and conditions is helpful for parents and healthcare providers. Detecting clinically relevant CNVs early is important to monitor conditions, anticipate future difficulties, and potentially intervene at a young age.

ACKNOWLEDGMENTS

The authors thank the participating families. We also thank Karlene Coleman for help consenting families and Cheryl Strauss for editorial assistance. Kate Garber provided helpful comments for the manuscript. This study was supported by a grant from the NIH (R01MH092902 to M.K.R.) and the Emory Genetics Laboratory Director Grant Program. J.A.B. is supported by a Sanofi/Genzyme Corporation fellowship. J.W.I. is supported by AHA grant #13GRNT15810006 and the Morton S. and Henrietta K. Sellner Professorship in Human Genetics. D.M.M. is supported by NIH R01DC009410 and The University of Michigan Donita B. Sullivan MD Research Professorship Funds. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

REFERENCES

Baldwin EL, Lee JY, Blake DM, Bunke BP, Alexander CR, Kogan AL, Ledbetter DH, Martin CL. 2008. Enhanced detection of clinically rele-

vant genomic imbalances using a targeted plus whole genome oligonucleotide microarray. Genet Med 10:415–429.

- Bisgaard AM, Kirchhoff M, Nielsen JE, Brandt C, Hove H, Jepsen B, Jensen T, Ullmann R, Skovby F. 2007. Transmitted cytogenetic abnormalities in patients with mental retardation: Pathogenic or normal variants? Eur J Med Genet 50:243–255.
- Coe BP, Witherspoon K, Rosenfeld JA, van Bon BW, Vulto-van Silfhout, Friend P, Baker KL, Buono C, Vissers S, Schuurs-Hoeijmakers LE, Hoischen JH, Pfundt A, Krumm R, Carvill N, Li GL, Amaral D, Brown D, Lockhart N, Scheffer PJ, Alberti IE, Shaw A, Pettinato M, Tervo R, de Leeuw R, Reijnders N, Torchia MR, Peeters BS, O'Roak H, Fichera BJ, Hehir-Kwa M, Shendure JY, Mefford J, Haan HC, Gecz E, de Vries J, Romano BB, Eichler C, Eichler EE. 2014. Refining analyses of copy number variation identifies specific genes associated with developmental delay. Nat Genet 46:1063–1071.
- Consortium Genome Project, Abecasis GR, Altshuler D, Auton A, Brooks LD, Durbin RM, Gibbs RA, Hurles ME, McVean GA. 2010. A map of human genome variation from population-scale sequencing. Nature 467:1061–1073.
- Cooper GM, Coe BP, Girirajan S, Rosenfeld JA, Vu TH, Baker C, Williams C, Stalker H, Hamid R, Hannig V, Abdel-Hamid H, Bader P, McCracken E, Niyazov D, Leppig K, Thiese H, Hummel M, Alexander N, Gorski J, Kussmann J, Shashi V, Johnson K, Rehder C, Ballif BC, Shaffer LG, Eichler EE. 2011. A copy number variation morbidity map of developmental delay. Nat Genet 43:838–846.
- Costain G, Lionel AC, Merico D, Forsythe P, Russell K, Lowther C, Yuen T, Husted J, Stavropoulos DJ, Speevak M, Chow EW, Marshall CR, Scherer SW, Bassett AS. 2013. Pathogenic rare copy number variants in community-based schizophrenia suggest a potential role for clinical microarrays. Hum Mol Genet 22:4485–4501.
- Deak KL, Horn SR, Rehder CW. 2011. The evolving picture of microdeletion/microduplication syndromes in the age of microarray analysis: Variable expressivity and genomic complexity. Clin Lab Med 31:543–564, viii.
- Dittwald P, Gambin T, Szafranski P, Li J, Amato S, Divon MY, Rodríguez Rojas LX, Elton LE, Scott DA, Schaaf CP, Torres-Martinez W, Stevens AK, Rosenfeld JA, Agadi S, Francis D, Kang SH, Breman A, Lalani SR, Bacino CA, Bi W, Milosavljevic A, Beaudet AL, Patel A, Shaw CA, Lupski JR, Gambin A, Cheung SW, Stankiewicz P. 2013. NAHR-mediated copynumber variants in a clinical population: Mechanistic insights into both genomic disorders and Mendelizing traits. Gen Res 23:1395–1409.
- Girirajan S, Rosenfeld JA, Coe BP, Parikh S, Friedman N, Goldstein A, Filipink RA, McConnell JS, Angle B, Meschino WS, Nezarati MM, Asamoah A, Jackson KE, Gowans GC, Martin JA, Carmany EP, Stockton DW, Schnur RE, Penney LS, Martin DM, Raskin S, Leppig K, Thiese H, Smith R, Aberg E, Niyazov DM, Escobar LF, El-Khechen D, Johnson KD, Lebel RR, Siefkas K, Ball S, Shur N, McGuire M, Brasington CK, Spence JE, Martin LS, Clericuzio C, Ballif BC, Shaffer LG, Eichler EE. 2012. Phenotypic heterogeneity of genomic disorders and rare copy-number variants. N Engl J Med 367:1321–1331.
- Itsara A, Cooper GM, Baker C, Girirajan S, Li J, Absher D, Krauss RM, Myers RM, Ridker PM, Chasman DI, Mefford H, Ying P, Nickerson DA, Eichler EE. 2009. Population analysis of large copy number variants and hotspots of human genetic disease. Am J Hum Genet 84:148–161.
- Kaminsky EB, Kaul V, Paschall J, Church DM, Bunke B, Kunig D, Moreno-De-Luca D, Moreno-De-Luca A, Mulle JG, Warren ST, Richard G, Compton JG, Fuller AE, Gliem TJ, Huang S, Collinson MN, Beal SJ, Ackley T, Pickering DL, Golden DM, Aston E, Whitby H, Shetty S, Rossi MR, Rudd MK, South ST, Brothman AR, Sanger WG, Iyer RK, Crolla JA, Thorland EC, Aradhya S, Ledbetter DH, Martin CL. 2011. An evidencebased approach to establish the functional and clinical significance of copy number variants in intellectual and developmental disabilities. Genet Med 13:777–784.

- Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM, Haussler D. 2002. The human genome browser at UCSC. Gen Res 12:996–1006.
- Konrad M, Saunier S, Heidet L, Silbermann F, Benessy F, Calado J, Le Paslier D, Broyer M, Gubler MC, Antignac C. 1996. Large homozygous deletions of the 2q13 region are a major cause of juvenile nephronoph-thisis. Hum Mol Genet 5:367–371.
- Liu P, Lacaria M, Zhang F, Withers M, Hastings PJ, Lupski JR. 2011. Frequency of nonallelic homologous recombination is correlated with length of homology: evidence that ectopic synapsis precedes ectopic crossing-over. Am J Hum Genet 89:580–588.
- Macdonald JR, Ziman R, Yuen RK, Feuk L, Scherer SW. 2014. The Database of Genomic Variants: a curated collection of structural variation in the human genome. Nucleic Acids Res 42:D986–D992.
- Moreno-De-Luca D, Sanders SJ, Willsey AJ, Mulle JG, Lowe JK, Geschwind DH, State MW, Martin CL, Ledbetter DH. 2013. Using large clinical data sets to infer pathogenicity for rare copy number variants in autism cohorts. Mol Psychiatry 18:1090–1095.
- Ounap K, Ilus T, Laidre P, Uibo O, Tammur P, Bartsch O. 2005. A new case of 2q duplication supports either a locus for orofacial clefting between markers D2S1897 and D2S2023 or a locus for cleft palate only on chromosome 2q13–q21. Am J Med Genet A 137A:323–327.
- Riegel M, Schinzel A. 2002. Duplication of (2)(q11.1–q13.2) in a boy with mental retardation and cleft lip and palate: another clefting gene locus on proximal 2q? Am J Med Genet 111:76–80.
- Rosenfeld JA, Coe BP, Eichler EE, Cuckle H, Shaffer LG. 2013. Estimates of penetrance for recurrent pathogenic copy-number variations. Genet Med 15:478–481.
- Rudd MK, Keene J, Bunke B, Kaminsky EB, Adam MP, Mulle JG, Ledbetter DH, Martin CL. 2009. Segmental duplications mediate novel, clinically relevant chromosome rearrangements. Hum Mol Genet 18:2957–2962.
- Russell MW, Raeker MO, Geisler SB, Thomas PE, Simmons TA, Bernat JA, Thorsson T, Innis JW. 2014. Functional analysis of candidate genes in 2q13 deletion syndrome implicates FBLN7 and TMEM87B deficiency in congenital heart defects and FBLN7 in craniofacial malformations. Hum Mol Genet 23:4272–4284.
- Shaikh TH, Gai X, Perin JC, Glessner JT, Xie H, Murphy K, O'Hara R, Casalunovo T, Conlin LK, D'Arcy M, Frackelton EC, Geiger EA, Haldeman-Englert C, Imielinski M, Kim CE, Medne L, Annaiah K, Bradfield JP, Dabaghyan E, Eckert A, Onyiah CC, Ostapenko S, Otieno FG, Santa E, Shaner JL, Skraban R, Smith RM, Elia J, Goldmuntz E, Spinner NB, Zackai EH, Chiavacci RM, Grundmeier R, Rappaport EF, Grant SF, White PS, Hakonarson H. 2009. High-resolution mapping and analysis of copy number variations in the human genome: a data resource for clinical and research applications. Genome Res 19:1682–1690.
- Soemedi R, Wilson IJ, Bentham J, Darlay R, Topf A, Zelenika D, Cosgrove C, Setchfield K, Thornborough C, Granados-Riveron J, Blue GM, Breckpot J, Hellens S, Zwolinkski S, Glen E, Mamasoula C, Rahman TJ, Hall D, Rauch A, Devriendt K, Gewillig M, J OS, Winlaw DS, Bu'Lock F, Brook JD, Bhattacharya S, Lathrop M, Santibanez-Koref M, Cordell HJ, Goodship JA, Keavney BD. 2012. Contribution of global rare copynumber variants to the risk of sporadic congenital heart disease. Am J Hum Genet 91:489–501.
- Stankiewicz P, Lupski JR. 2010. Structural variation in the human genome and its role in disease. Annu Rev Med 61:437–455.
- Szatmari P, Paterson AD, Zwaigenbaum L, Roberts W, Brian J, Liu XQ, Vincent JB, Skaug JL, Thompson AP, Senman L, Feuk L, Qian C, Bryson SE, Jones MB, Marshall CR, Scherer SW, Vieland VJ, Bartlett C, Mangin LV, Goedken R, Segre A, Pericak-Vance MA, Cuccaro ML, Gilbert JR, Wright HH, Abramson RK, Betancur C, Bourgeron T, Gillberg C, Leboyer M, Buxbaum JD, Davis KL, Hollander E, Silverman JM,

Hallmayer J, Lotspeich L, Sutcliffe JS, Haines JL, Folstein SE, Piven J, Wassink TH, Sheffield V, Geschwind DH, Bucan M, Brown WT, Cantor RM, Constantino JN, Gilliam TC, Herbert M, Lajonchere C, Ledbetter DH, Lese-Martin C, Miller J, Nelson S, Samango-Sprouse CA, Spence S, State M, Tanzi RE, Coon H, Dawson G, Devlin B, Estes A, Flodman P, Klei L, McMahon WM, Minshew N, Munson J, Korvatska E, Rodier PM, Schellenberg GD, Smith M, Spence MA, Stodgell C, Tepper PG, Wijsman EM, Yu CE, Roge B, Mantoulan C, Wittemeyer K, Poustka A, Felder B, Klauck SM, Schuster C, Poustka F, Bolte S, Feineis-Matthews S, Herbrecht E, Schmotzer G, Tsiantis J, Papanikolaou K, Maestrini E, Bacchelli E, Blasi F, Carone S, Toma C, Van Engeland H, de Jonge M, Kemner C, Koop F, Langemeijer M, Hijmans C, Staal WG, Baird G, Bolton PF, Rutter ML, Weisblatt E, Green J, Aldred C, Wilkinson JA, Pickles A, Le Couteur A, Berney T, McConachie H, Bailey AJ, Francis K, Honeyman G, Hutchinson A, Parr JR, Wallace S, Monaco AP, Barnby G, Kobayashi K, Lamb JA, Sousa I, Sykes N, Cook EH, Guter SJ, Leventhal BL, Salt J, Lord C, Corsello C, Hus V, Weeks DE, Volkmar F, Tauber M, Fombonne E, Shih A, Meyer KJ. 2007. Mapping autism risk loci using genetic linkage and chromosomal rearrangements. Nat Genet 39:319–328.

- Watson CT, Marques-Bonet T, Sharp AJ, Mefford HC. 2014. The genetics of microdeletion and microduplication syndromes: an update. Annu Rev Genomics Hum Genet 15:215–244.
- Yu HE, Hawash K, Picker J, Stoler J, Urion D, Wu BL, Shen Y. 2012. A recurrent 1.71 Mb genomic imbalance at 2q13 increases the risk of developmental delay and dysmorphism. Clin Genet 81:257–264.