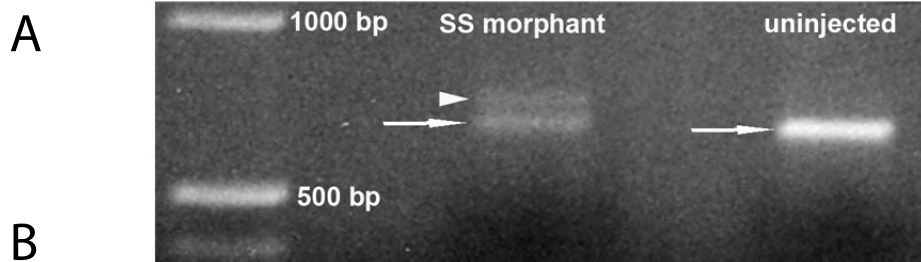


Figure 2.6: *pgrn-a* 5'UTR-targeting morpholinos block *Pgrn-a* translation and gross morphology of *pgrn-a* morphants. (A) Western blot showing *Pgrn-a* and Actin expression in 72hpf uninjected (UI) and morphant (5'UTR MO) embryos. **(B)** Histogram showing quantification of *Pgrn-a* expression in UI controls ($26.2 \pm 2.05\%$) and morphants ($2.7 \pm 0.9\%$); $**p \leq 0.01$. Quantitative data from three biological and technical replicates is normalized to Actin and represented as mean; error bars represent the standard deviation. **(C-H)** Representative whole embryos from control (C-F) and experimental (G-H) groups at 72hpf. 5'UTR MM MO and SS MM MO, embryos injected with 5-nucleotide mismatch control morpholinos; 5'UTR MO, embryos injected with *pgrn-a* 5'UTR-targeted morpholinos; SS MO, embryos injected with *pgrn-a* e3i3 splice site-targeted morpholinos.



B

ATGTTGAGACTGACAGTCTGCCTCGCTGTGGTGACCCTGGTTATTTGCTCGCAGTGCCCCG
ATAATGAAGTCTGTGAAGCAGGCCAGTCTGCTGCCAGGATCCCAGTGGTGGCTTCAGCTG
CTGCCCTTTCCATCATGGAGAGTGTGTGAAGACCATCTGCACTGCTGTCCCGAAGGCATG
TTGTGCAGTGTGAAGGACTTAACATGTACAAACGCAACACATACAGAGCCATTGGCGGACA
GGACACAAGCTAAAAAGCCAGAGTCGCCCCAAAgtgagacattatatcaagcgttaagtgat
agtactcaaagctgatcgtcatgcactaataaaaaaccgtttctcagTCATTCAGAATGAT
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GAGTTCTCCTGTCTGCTGATGTCTACATCATATGGCTGCTGTCCAGTAGCACAGGGCCTTG
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CTGCAGACACTACATGTTGTCAGGCTGAAGATGGCCTATGGGGGTGCTGCCCCATGCCAAA
GGCTGTATGCTGTGATGACAAAATCCATTGCTGCCCAGAGGACACTGTTTGTGACGTCAA
GCCTTGAAATGCATATCCTCAACCAACCAGAGCTGCCCATGTGGGACAAATCCCTGCTCG
CCTTAGGGCTGAATGGGAAGATCACAAACAAAAAAGCCTGAAACTCAACGCACTACA
ACTAGACCTACAGGCACTACAAGCACTAATACAGCTGCCAACCAATGACTACGCTGCCTGCCG
AACACCAAGCGGTGTCTTCAGATGTTCCCTGTAACGACACTGCGGCCTGTGCTGATGGA...

Figure 2.7: *pgrn-a* splice site-targeting morpholinos cause intron retention and knockdown Pgrn-a. (A) PCR confirmation of *pgrn-a* knockdown using morpholino oligonucleotides targeting the e3i3 splice site of *pgrn-a* (SS MO). The PCR product was sequenced and the double band seen in the SS morphant lane represents intron retention. The overall intensity of the Pgrna band is reduced in SS morphant when compared to uninjected control. (B) Sequence of *pgrn-a*. Exon sequence (upper case); intron sequence (lower case); primer sequence (blue); SS MO sequence (green); stop codon (red).

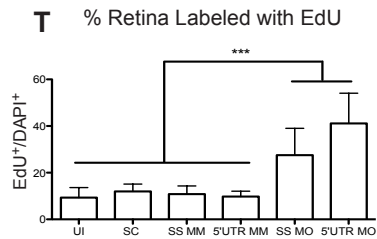
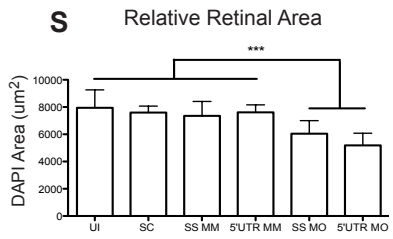
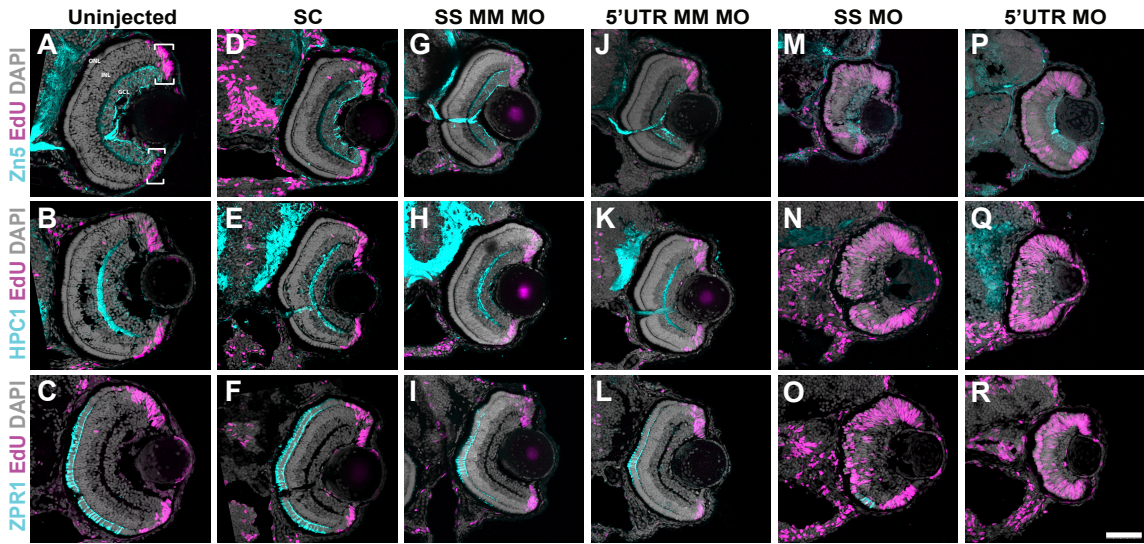


Figure 2.8: Pgrn-a knockdown results in microphthalmia and diminished neuronal differentiation. (A-R) Sections through central retina of uninjected (UI, A-C); standard control morpholino-injected (SC, D-F); SS MM (G-I) and 5'UTR MM (J-L), embryos injected with 5-nucleotide mismatch control morpholinos; SS MO (M-O) and 5'UTR MO (P-R), embryos injected with e3i3 splice site-targeted or 5'UTR-targeted morpholinos, respectively, at 72hpf. Sections are immunolabeled (cyan) with markers of differentiated ganglion cells (Zn5, top row), amacrine cells (HPC1, middle row), and red-green double cone photoreceptor cells (ZPR1, bottom row), EdU (fuscia), and DAPI (gray). **(S)** Histogram showing relative retinal area in UI ($7945.8 \pm 1319.6 \mu\text{m}^2$; $n=11$), SC-injected ($7594.3 \pm 475.2 \mu\text{m}^2$; $n=9$), SS MM MO-injected ($7354 \pm 1062.6 \mu\text{m}^2$; $n=11$), and 5'UTR MM MO-injected ($7610.8 \pm 547.6 \mu\text{m}^2$; $n=10$), SS MO ($6044.7 \pm 956.2 \mu\text{m}^2$; $n=10$), and 5'UTR MO ($5188.7 \pm 891.8 \mu\text{m}^2$; $n=16$) retinas at 72hpf; $***p \leq 0.001$. **(T)** Histogram showing the percent of the retina labeled with EdU in UI ($9.3 \pm 4.3\%$; $n=11$), SC-injected ($12 \pm 3.1\%$; $n=9$), SS MM MO-injected ($10.8 \pm 3.5\%$; $n=11$), 5'UTR MM MO-injected ($9.8 \pm 2.3\%$; $n=10$), SS MO-injected ($27.5 \pm 11.5\%$; $n=10$), and 5'UTR MO-injected ($41.1 \pm 13\%$; $n=16$) retinas at 72hpf; $***p \leq 0.001$. Quantitative data are represented as mean; error bars represent the standard deviation. Outer nuclear layer (ONL), inner nuclear layer (INL), and ganglion cell layer (GCL); ciliary marginal zone (CMZ, brackets). Scale bar equals $50\mu\text{m}$.

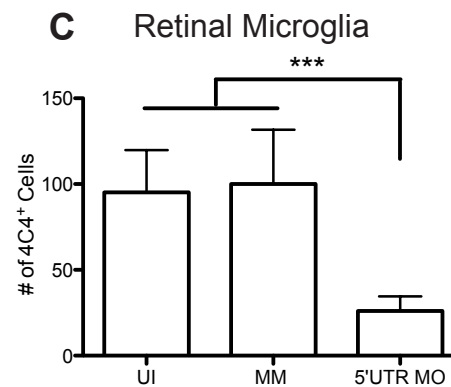
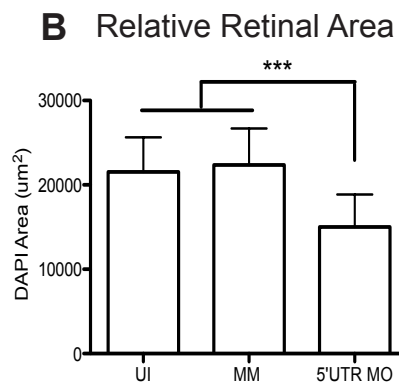
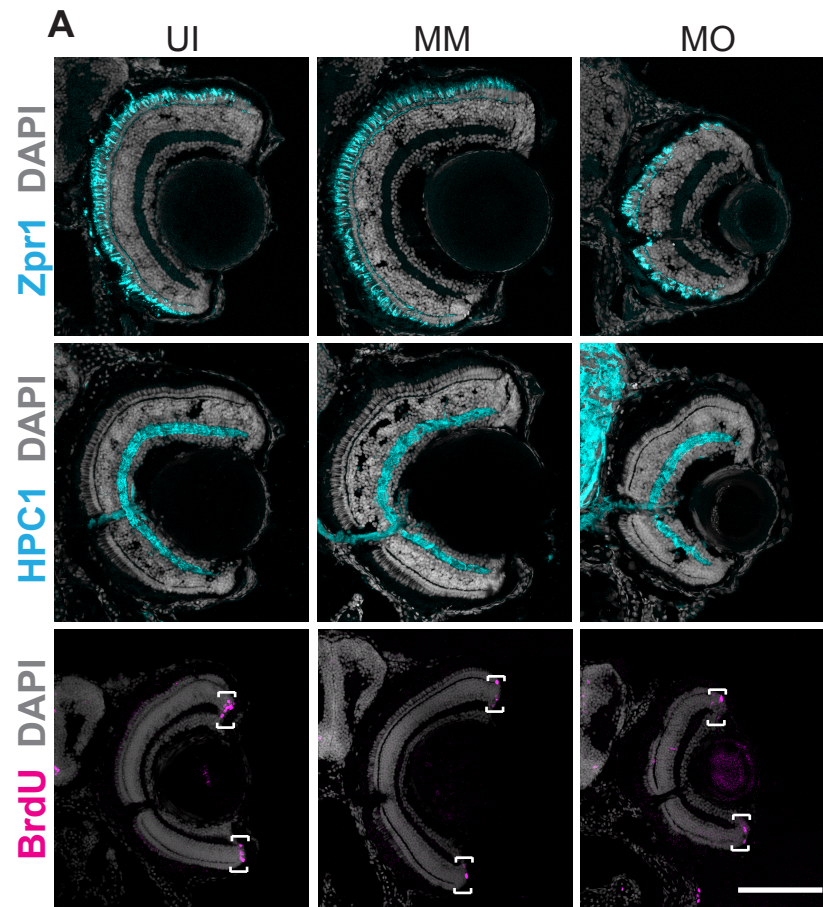


Figure 2.9: Aspects of Pgrn-a knockdown retinal phenotype recover by 8dpf. (A)

Sections through central retina of uninjected (UI, left column), 5'UTR MM MO (middle column), and 5'UTR MO (right column) larvae at 8dpf, immunolabeled (cyan) with markers of red-green cone photoreceptors (ZPR1, top row) and amacrine cells (HCP1, middle row), BrdU (fuscia, bottom row), and DAPI (gray). **(B)** Histogram showing the size of the retina in UI ($21533.35 \pm 4112.8 \mu\text{m}^2$; $n=15$), 5'UTR MM MO ($22360.0 \pm 4324.0 \mu\text{m}^2$; $n=9$), and 5'UTR MO ($15000.4 \pm 3861.2 \mu\text{m}^2$; $n=15$) larvae at 8dpf; $***p<0.001$. **(C)** Histogram showing the number of retinal microglia (4C4⁺ cells) in UI (95.2 ± 24.7 cells; $n=15$), 5'UTR MM MO (100 ± 31.7 cells; $n=9$), and 5'UTR MO (26.1 ± 8.5 cells; $n=15$) larvae at 8dpf; $***p<0.001$. Quantitative data are represented as mean; error bars represent the standard deviation. Outer nuclear layer (ONL), inner nuclear layer (INL), and ganglion cell layer (GCL). Scale bar equals 100 μm .

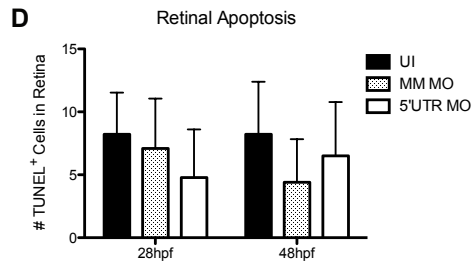
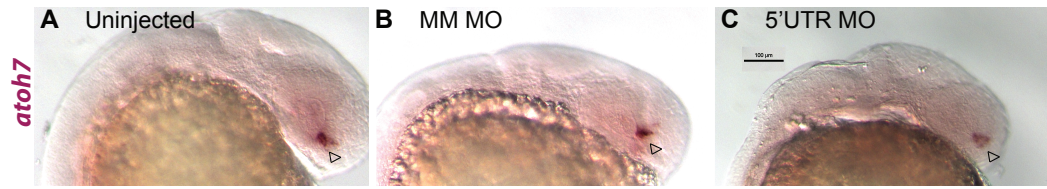


Figure 2.10: No difference in *atoh7* expression and TUNEL-positive cell counts between *pgrn-a* morphant and control embryos. Whole mount *in situ* hybridization showing *atoh7* expression in uninjected **(A)**, 5'UTR MM MO **(B)**, and 5'UTR MO **(C)** embryos at 28hpf. **(D)** Histogram showing the number of TUNEL-positive cells in uninjected, 5'UTR MM MO-injected, and 5'UTR MO-injected retinas at 28 and 48hpf.

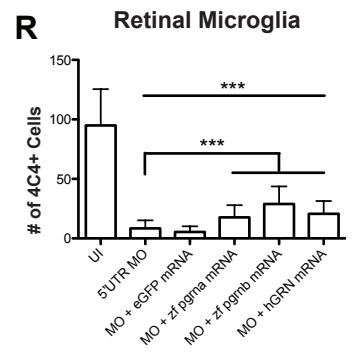
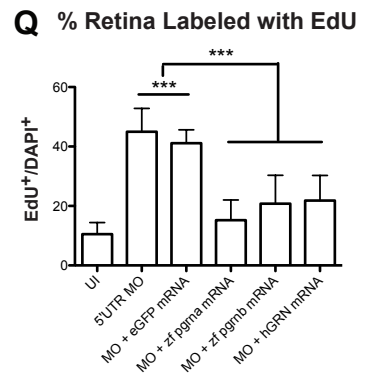
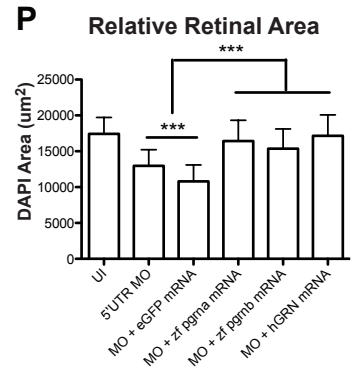
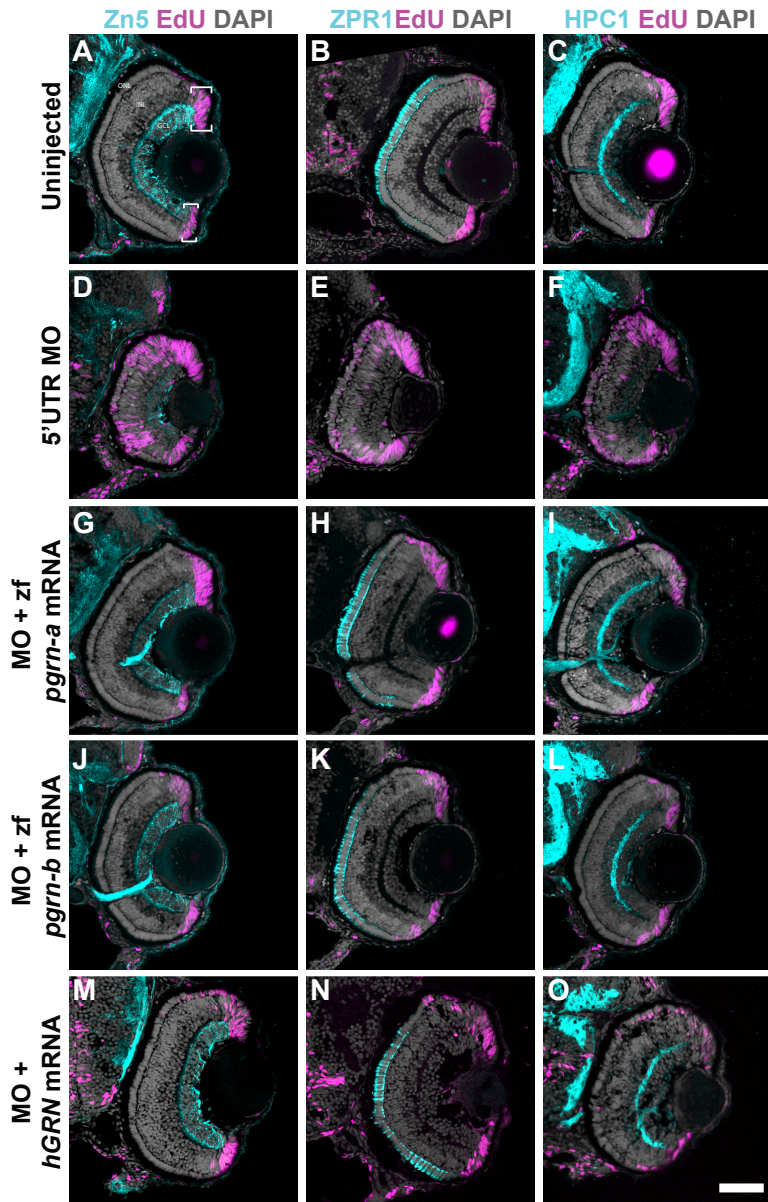


Figure 2.11: Co-injection of 5'UTR MO and zf *pgrn-a*, zf *pgrn-b*, or *hGRN* mRNA

rescues most aspects of knockdown retinal phenotype. (A-O) Cross-sections

through central retina of 72hpf uninjected (UI, A-C), 5'UTR MO-injected (D-F), 5'UTR MO and zf *pgrn-a* mRNA co-injected (G-I), 5'UTR MO and zf *pgrn-b* mRNA co-injected (J-L), and 5'UTR MO and *hGRN* mRNA co-injected (M-O) embryos at 72 hpf. Sections are immunolabeled (cyan) with markers for ganglion cells (Zn-5, left column), red-green double cone photoreceptors (Zpr1, middle column), and amacrine cells (HPC1, right column), EdU (fuscia), and DAPI (gray).

(P) Histogram showing relative retinal area of UI ($17437.4 \pm 2286.1 \mu\text{m}^2$; $n=23$), 5'UTR MO-injected ($12961.2 \pm 2251.8 \mu\text{m}^2$; $n=21$), 5'UTR MO and *eGFP* mRNA co-injected ($10809 \pm 2282.5 \mu\text{m}^2$; $n=8$), 5'UTR MO and zf *pgrn-a* mRNA co-injected ($16435.7 \pm 2882.2 \mu\text{m}^2$; $n=12$), 5'UTR MO and zf *pgrn-b* mRNA co-injected ($15357.8 \pm 2750.2 \mu\text{m}^2$; $n=13$), and 5'UTR MO and *hGRN* mRNA co-injected ($17146.1 \pm 2930.7 \mu\text{m}^2$; $n=12$) embryos at 72hpf; *** $p < 0.001$.

(Q) Histogram showing the percent of the retina labeled with EdU in UI ($10.5 \pm 3.9\%$; $n=7$), 5'UTR MO-injected ($45 \pm 7.9\%$; $n=18$), 5'UTR MO and *eGFP* mRNA co-injected ($41.1 \pm 4.5\%$; $n=6$), 5'UTR MO and zf *pgrn-a* mRNA co-injected ($15.2 \pm 6.8\%$; $n=22$), 5'UTR MO and zf *pgrn-b* mRNA co-injected ($20.8 \pm 9.6\%$; $n=18$), and 5'UTR MO and *hGRN* mRNA co-injected ($21.8 \pm 8.5\%$; $n=12$) embryos at 72hpf; *** $p < 0.001$.

(R) Histogram showing the number of retinal microglia (4C4⁺ cells) at 72hpf in UI (95 ± 30.5 cells; $n=23$), 5'UTR MO-injected (8.4 ± 6.8 cells; $n=21$), 5'UTR MO and *eGFP* mRNA co-injected (5.4 ± 4.8 cells; $n=8$), 5'UTR MO and zf *pgrn-a* mRNA co-injected (17.7 ± 10.2 cells; $n=12$), 5'UTR MO and zf *pgrn-b* mRNA co-injected (28.9 ± 14.7 cells; $n=13$), and 5'UTR MO and *hGRN* mRNA co-injected (20.6 ± 10.8 ; $n=12$) embryos; *** $p < 0.001$. Quantitative data are represented as mean; error bars represent the standard deviation.

Outer nuclear layer (ONL), inner nuclear layer (INL), and ganglion cell layer (GCL); ciliary marginal zone (CMZ, brackets). Scale bar equals 50 μm .

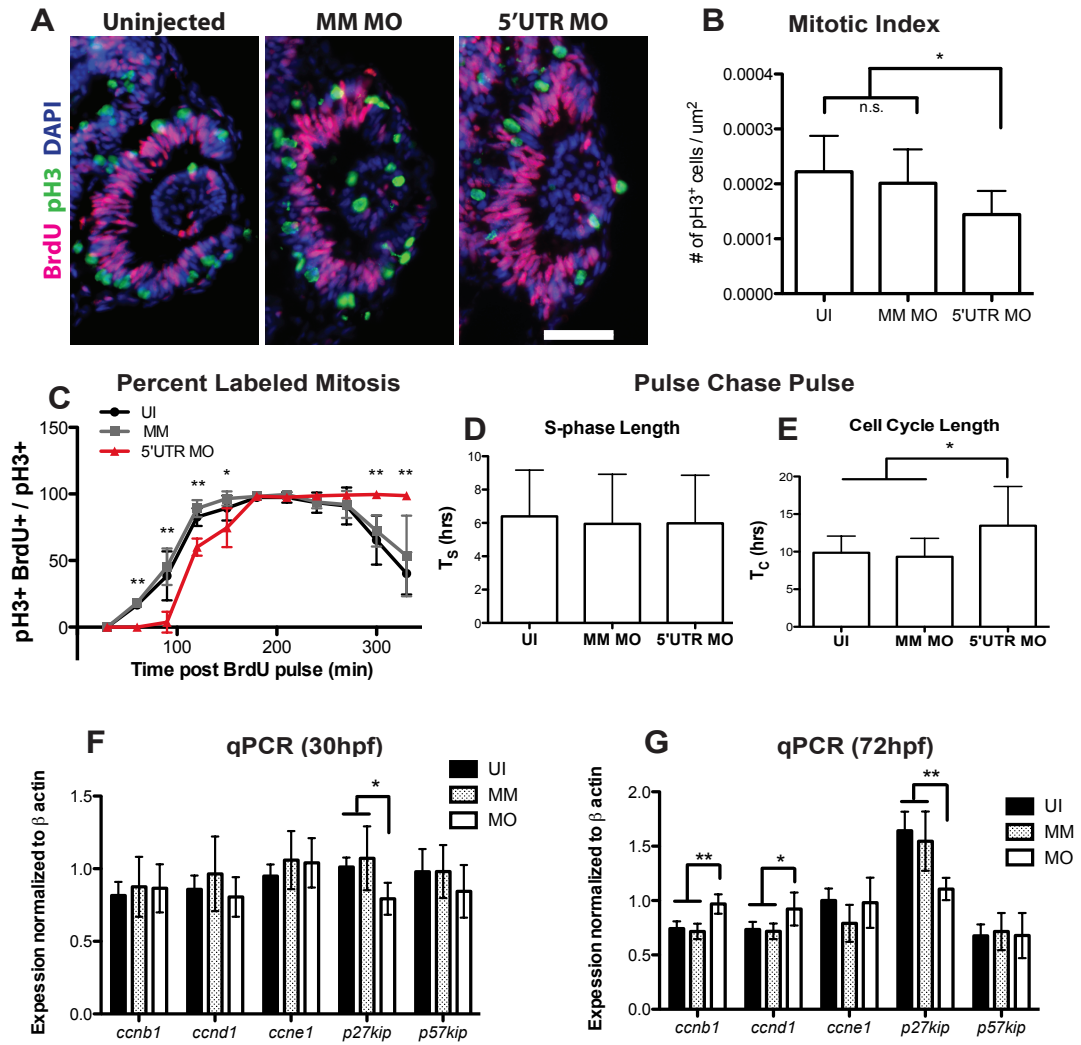


Figure 2.12: Knockdown of *pgrn-a* alters cell cycle kinetics in retinal progenitors.

(A) Representative retinal sections from 28hpf uninjected (left panel), MM MO (center panel), and 5'UTR MO (right panel) embryos stained with antibodies against pH3 (green), BrdU (fuscia), and DAPI (blue) used for quantification of mitotic index. **(B)** Histogram showing the mitotic index at 28hpf. The number of pH3⁺ cells/unit area in UI ($0.0002 \pm 1.973e-005$ cells per μm^2 ; $n=11$), MM MO ($0.0002 \pm 1.947e-005$ cells per μm^2 ; $n=10$), and 5'UTR MO retinas ($0.0001 \pm 1.366e-005$ cells per μm^2 ; $n=10$); $*p \leq 0.01$. **(C)** Graph showing the percent labeled mitosis for UI, MM MO and 5'UTR MO embryos between 28 and 35hpf. **(D)** Histogram showing average S-phase length (T_S) in 26-28hpf UI (6.4 ± 2.8 hrs; $n=16$), MM MO (5.9 ± 3.0 hrs; $n=10$), and 5'UTR MO (6.0 ± 2.9 hrs; $n=7$) embryos. **(E)** Histogram showing average total cell cycle length (T_C) at 26-28hpf in UI (9.9 ± 2.2 hrs; $n=16$), MM MO (9.3 ± 2.5 hrs; $n=10$), and 5'UTR MO (13.5 ± 5.2 hrs; $n=7$); $*p \leq 0.05$. **F-G** Histograms showing relative cyclin B (*ccnb1*), cyclin D (*ccnd1*), cyclin E (*ccne1*), *p27kip* and *p57kip* mRNA expression normalized to beta actin at 30hpf (F) and 72hpf (G); $*p \leq 0.05$ and $**p \leq 0.01$. Quantitative data are represented as mean; error bars represent the standard deviation. Scale bar = 50 μm .

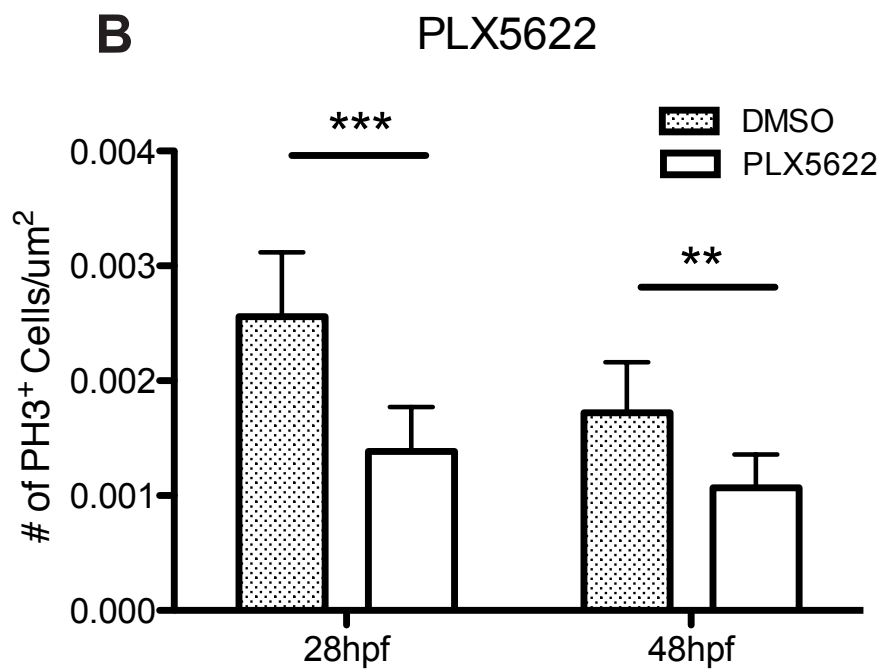
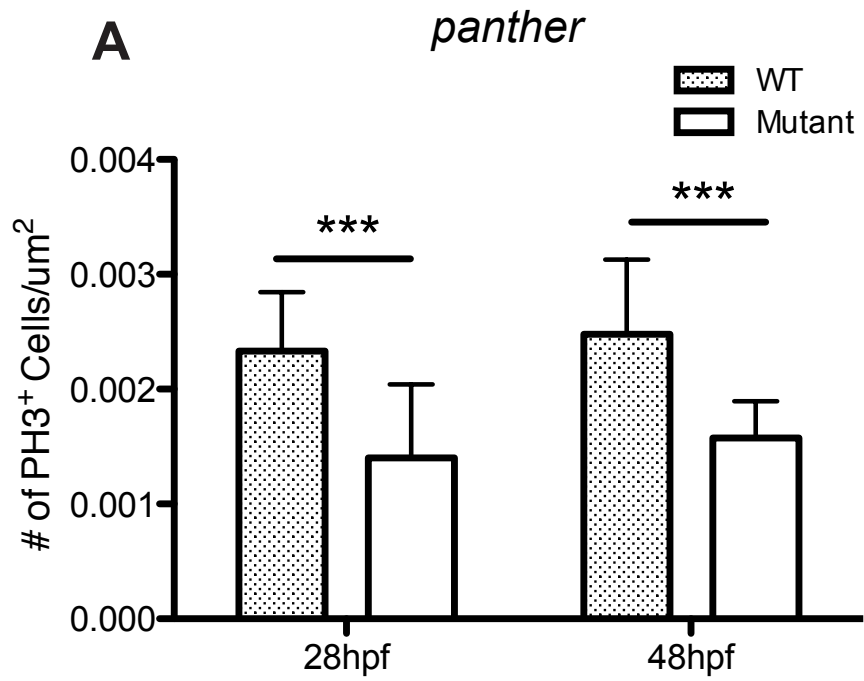


Figure 2.13: Mitotic index in 28 and 48hpf panther mutant and PLX5622-treated embryos. (A) Histogram showing the ratio of pH3⁺ cells in wild type (WT; $.002 \pm .0005$ cells per μm^2 ; $n=14$) and *panther* mutant ($.001 \pm .0006$ cells per μm^2 ; $n=22$) retinas at 28hpf and in WT ($.002 \pm .0006$ cells per μm^2 ; $n=12$) and *panther* mutant ($.002 \pm .0003$ cells per μm^2 ; $n=18$) retinas at 48hpf; *** $p \leq 0.001$. **(B)** Histogram showing the ratio of pH3⁺ cells in DMSO-treated control ($.003 \pm .0006$ cells per μm^2 ; $n=11$) and PLX5622-treated ($.001 \pm .0004$ cells per μm^2 ; $n=12$) retinas at 28hpf, and DMSO-treated control ($.002 \pm .0004$ cells per μm^2 ; $n=10$) and PLX5622-treated ($.001 \pm .0003$ cells per μm^2 ; $n=10$) retinas at 48hpf; *** $p \leq 0.001$. Quantitative data are represented as mean; error bars represent the standard deviation.

Table 2.1: Antibody List

Primary Antibodies	Company	Dilution
Monoclonal anti-Zn5	ZIRC; zfin.org/ZDB-ATB-081002-19	1:200
Monoclonal anti-Zpr1 (anti-Arrestin)	ZIRC; zfin.org/ZDB-ATB-081002-43	1:200
Mouse anti-HPC1 (anti-Syntaxin)	Sigma S0664	1:200
Monoclonal anti-4C4	Umich Hybridoma Core	1:200
Rabbit anti-leucocyte-specific plastin (L-plastin)	Gift from Michael Redd	1:500
Mouse anti-BrdU	BD Biosciences 347580	1:100
Rabbit anti-pH3 (phospho-histone 3 ser10)	Millipore 06-570	1:200
Secondary Antibodies		
Alexa Fluor goat anti-mouse 488 or 555	Invitrogen	1:500
Alexa Fluor goat anti-rabbit 488 or 555	Invitrogen	1:500

Table 2.2: Restriction Enzymes and RNA Polymerases for *pgrn-a*, *atoh7*, and *fms* Riboprobe Synthesis

Probe	Restriction Enzyme	RNA polymerase
<i>pgrn-a</i> sense	Sall	T7
<i>pgrn-a</i> anti-sense	Smal	T3
<i>atoh7</i> anti-sense	EcoRI	T7
<i>fms</i> anti-sense	EcoRI	T7

Table 2.3: Morpholino Oligonucleotide Sequences

Name	Sequence
pgrn-a 5'UTR MO	5'-AGCGGAAAGTAAATGATCAGTCCGT-3'
pgrn-a 5'UTR MM MO	5'-AGCGcAAAcTAtATcATCAcTCCGT-3'
pgrn-a SS MO	5'-TATAATGTCTCACTTTGGGAAGGTC-3'
pgrn-a SS MM MO	5'-TAaAATcTCTgACTTAgcGAAGGTC-3'
fms MO	5'-AAGAGCGCGAAGAACATCTCAGAGC-3'
fms MM MO	5'-AAcAcCGCcAAcAACATCTCAcAGC-3'
p53 MO	5'-GCGCCATTGCTTTGCAAGAATTG-3'
SC MO	5'-CCTCTTACCTCAGTTACAATTTATA-3'

Table 2.4: Primer Sequences for mRNA Rescue Experiments

Primer Name	Sequence
zf_pgrna_FW	ATGTTGAGACTGACAGTCTGCC
zf_pgrna_REV	TTATAGAGTTAGGGCTCGTTTC
zf_pgrnb_FW	ATGGTGCGTGCAGCTTTCATAGC
zf_pgrnb_REV	TTAGAGAGAATTATTCCACCACG

Table 2.5: Primer Sequences for qRT-PCR

Gene	Forward Primer	Reverse Primer
<i>cycB1</i>	AGTTTAGGCTGCTTCAGGAGAC	ATGCACGGTCTGTCACAAAAGC
<i>cycD1</i>	CAAACACGCCCAGACCTTTGTG	GGGTCATTCTGATGACTTGCG
<i>cycE1</i>	GGAAGAGAAAAGCAGACGTGGC	GGTTCCTCGACTTCATCAGGTG
<i>p27kip</i>	AACGGGAATCACGACTGTAGGG	ATGTGGGTGTCCGACTCAATGG
<i>p57kip</i>	CCGGTAGCTCAAGAATCCGAGG	TCGTGGATGTGCCGGCTTGAAG
<i>bactin</i>	GCAATGAGCGTTTCCGTTG	TGTGTTGGCATAACAGGTCC

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