

SUPPLEMENTAL INFORMATION

1,2-diacylglycerol choline phosphotransferase catalyzes the final step in the unique *Treponema denticola* phosphatidylcholine biosynthesis pathway

Miguel Ángel Vences-Guzmán¹, M. Paula Goetting-Minesky², Ziqiang Guan³, Santiago Castillo-Ramirez¹, Luz América Córdoba-Castro¹, Isabel M. López-Lara¹, Otto Geiger¹, Christian Sohlenkamp^{1**} and J. Christopher Fenno^{2*}

¹ Centro de Ciencias Genómicas, Universidad Nacional Autónoma de México, Av. Universidad s/n, Apdo. Postal 565-A, Cuernavaca, Morelos, CP62210, Mexico

² Department of Biologic and Materials Sciences, University of Michigan School of Dentistry, Ann Arbor, Michigan 48109, USA

³ Department of Biochemistry, Duke University Medical Center, Durham, North Carolina 27710, U.S.A.

Running title: *CDP-choline pathway for phosphatidylcholine synthesis in bacteria*

* For correspondence. E-mail fenno@umich.edu; Tel. (+1) 734 763 3331; Fax (+1) 734 763 3453.

** For correspondence. E-mail: chsohlen@ccg.unam.mx Tel. 52-777-3291703 Fax. 52-777-3175581

Keywords: choline metabolism, *Treponema*, phosphatidylcholine, phospholipid metabolism
horizontal gene transfer

TABLE S1. Strains and plasmids used in this study

Organism	Strain	Relevant features	Source/Reference
<i>T. denticola</i>	35405	parent strain	ATCC (Chan <i>et al.</i> , 1993)
<i>T. denticola</i>	LBE3	35405 Δ <i>licCA</i>	(Kent <i>et al.</i> , 2004)
<i>T. denticola</i>	CF819	35405 Δ TDE0021	this study
<i>E. coli</i>	DH5 α	cloning strain	(Hanahan, 1983)
<i>E. coli</i>	JM109	cloning strain	(Yanisch-Perron <i>et al.</i> , 1985)
<i>E. coli</i>	JM110	cloning strain	(Yanisch-Perron <i>et al.</i> , 1985)
<i>E. coli</i>	BL21(DE3)/pLysS	expression strain	(Studier, 1991)
<i>S. cerevisiae</i>	HJ091	α <i>ura3-52 his3-1 leu 2-3, 112 trp1-289 cpt1::LEU2 ept1</i>	(McMaster <i>et al.</i> , 1996, Hjelmstad <i>et al.</i> , 1994)

Plasmid	Host	Relevant genes	Source/Reference
pCF737	<i>E. coli</i>	TDE0020-21-22	this study
pCF789	<i>E. coli</i>	TDE0020- <i>aphA2</i> - TDE0022	this study
pET17b- TDE0021	<i>E. coli</i>	TDE0021	this study
pCDF- Duet1- LicCA	<i>E. coli</i>	<i>T. denticola licCA</i>	this study
pYEp352	<i>S. cerevisiae</i>	empty vector	(Hill <i>et al.</i> , 1986)
pRH150	<i>S. cerevisiae</i>	CPT1 in pYEp352	(Hjelmstad & Bell, 1990) (Rodríguez-
pSP-Gm2	<i>S. cerevisiae</i>	empty vector	Limas <i>et al.</i> , 2011)
pSP-Gm2- Trepo	<i>S. cerevisiae</i>	TDE0021 in pSP-Gm2	this study

Table S1: Strains and plasmids used in this study.

Figure S1: TLC of the aqueous phase from the lipid extraction from the yeast complementation experiment (labeling with [¹⁴C]choline).

Figure S2: TLCs of the organic and aqueous phases from the lipid extraction from the yeast complementation experiment (labeling with [¹⁴C]ethanolamine).

Figure S3: SDS-PAGE of extracts from *E. coli* BL21(DE3).pLysS expressing TDE0021 and/or LicCA.

Figure S4: Positive ion collision-induced dissociation mass spectra of ion m/z 706.5 detected in the lipid extract from *E. coli* BL21(DE3).pLysS expressing LicCA and TDE0021.

Figure S5: pH dependency of CDP-choline phosphotransferase activity of Cpt (TDE0021) from *Treponema denticola*

Figure S1. TLC separation of the aqueous phases from the lipid extraction from the yeast complementation experiment (labeling with [¹⁴C]choline). *S.cerevisiae* HJ091 cells harboring pRH150 (yeast CPT1, lane 1), pSP-Gm2-Trepo (TDE0021, lane 2), empty plasmid pSP-GM2 (lane 3), or empty plasmid pYEp352 (lane 4) were labeled for 30 min with [¹⁴C]-choline. Aqueous phases obtained during lipid extraction were separated by one-dimensional TLC. Identities of [¹⁴C]-choline-containing species are assigned according to the expected R_f values. An unknown compound in the yeast CPT1-complemented strain, possibly glycerophosphocholine, is labeled “?” (Lane 1).

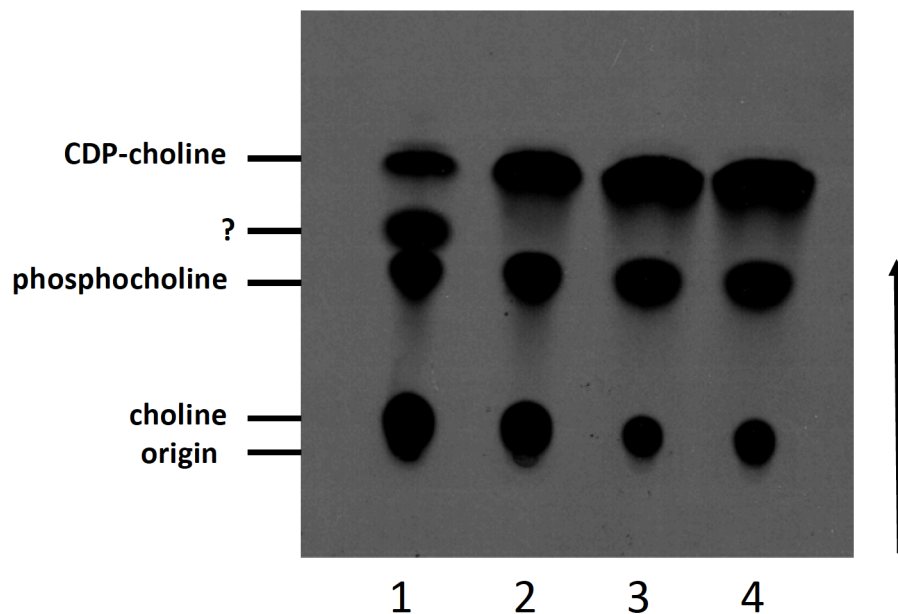


Figure S2. TLCs of the organic and aqueous phases from the yeast complementation experiment (labeling with $[^{14}\text{C}]$ ethanolamine). *S. cerevisiae* HJ091 cells harboring empty plasmid pYEp352 (lane 1), empty plasmid pSP-GM2 (lane 2), pSP-Gm2-Trepo (TDE0021, lane 3), or pYEp352-CPT1 (yeast CPT1, lane 4) were labeled for 30 min with $[^{14}\text{C}]$ -ethanolamine. Lipids were extracted according to Bligh and Dyer and organic (A) and aqueous (B) phases obtained during lipid extraction were separated by one-dimensional TLC. PE- $[^{14}\text{C}]$ phosphatidylethanolamine standard isolated from an *E. coli* strain cultivated in presence of $[^{14}\text{C}]$ acetate.

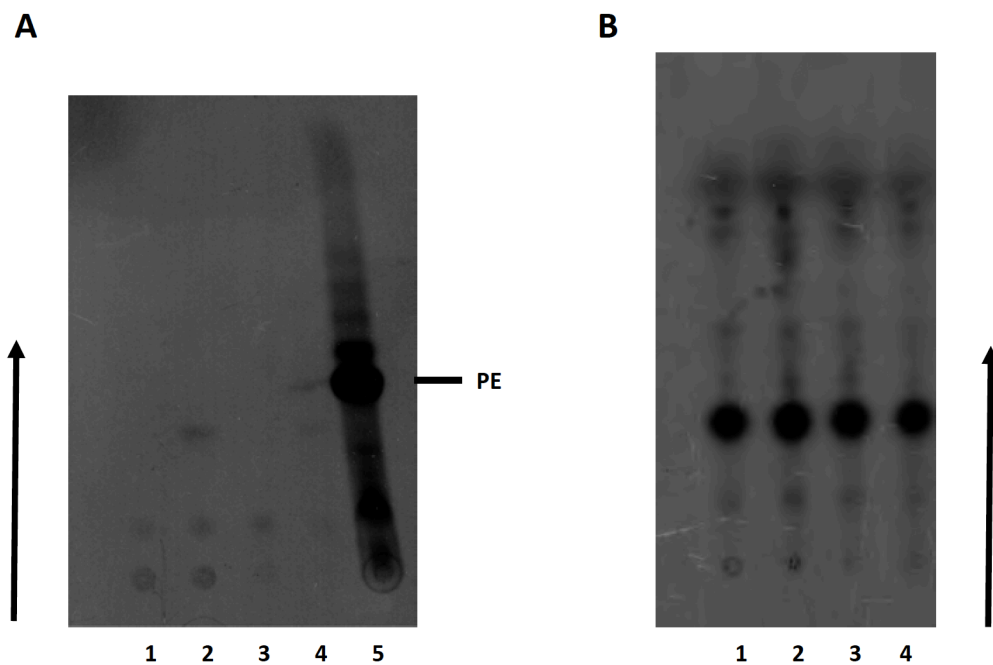


Figure S3. SDS-PAGE of extracts from *E. coli* BL21(DE3).pLysS expressing TDE0021 and/or LicCA. Analysis of cell-free protein extracts and purified membrane fractions from *E. coli* strains. Proteins were separated by 12% SDS-PAGE and stained with Coomassie blue. Cell-free proteins extracts from *E. coli* BL21(DE3).pLysS harboring pCDF-Duet and pET17b (lane1), pCDF-Duet-LicCA and pET17b (lane2), pCDF-Duet and pET17b-TDE0021 (lane3), pCDF-DuetLicCA and pET17b-TDE0021 (lane4). Resuspended membrane fractions from *E. coli* BL21(DE3).pLysS harboring pCDF-Duet and pET17b (lane5), pCDF-Duet-LicCA and pET17b (lane6), pCDF-Duet and pET17b-TDE0021 (lane7), pCDF-DuetLicCA and pET17b-TDE0021 (lane8). 5 micrograms of the membrane fractions were run on the gel (lanes 5 to 8). The molecular weight marker used was from Bio-Rad (#0161-0363).

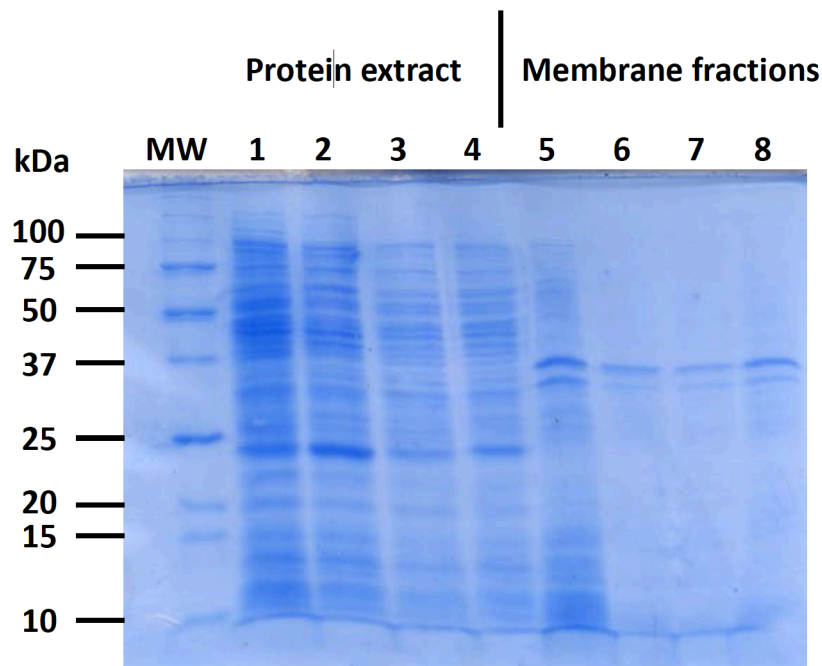


Figure S4. Positive ion collision-induced dissociation mass spectra of ion m/z 706.5 detected in the lipid extract from *E. coli* BL21(DE3).pLysS expressing LicCA and TDE0021. The presence of m/z 184.073 confirms the identity of phosphatidylcholine.

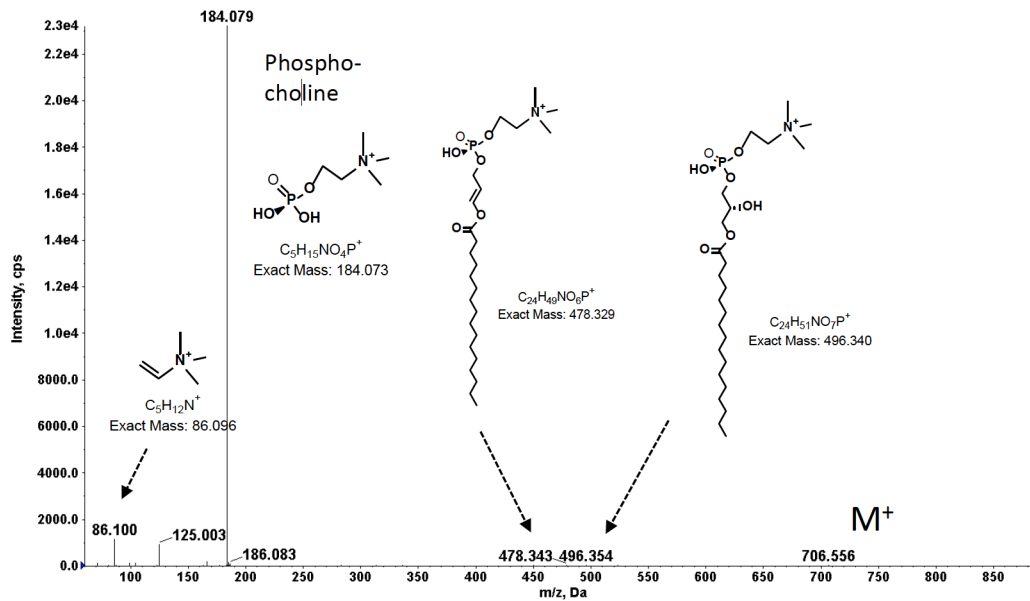


Figure S5. pH dependency of *Treponema denticola* CDP-choline phosphotransferase (TDE0021). TDE0021 activity was assayed at the indicated pH values with 50 mM Bis-Tris/HCl or Tris/HCl. Otherwise, conditions of the standard assay were used. The data points shown are the average of two independent experiments.

