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The Cytokine Network in Women with an Asymptomatic Short Cervix and the Risk of Preterm Delivery

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Abstract

Problem: To characterize the amniotic fluid (AF) inflammatory-related protein (IRP) network in patients with a sonographic short cervix (SCx) and to determine its relation to early preterm delivery (ePTD).

Method of Study: A retrospective cohort study included women with a SCx ≤ 25 mm) (n=223) who had amniocentesis and were classified according to gestational age (GA) at diagnosis and at delivery (ePTD < 32 weeks of gestation).

Results: 1) In women with a SCx $< 22 \frac{1}{7}$ weeks, the concentration of most IRPs increased as the cervix shortens; those with ePTD had a higher rate of increase of MIP-1a, MCP-1 and IL-6 concentrations than those delivering later; 2) concentration of most IRPs, and correlation between several IRP pairs, was higher in the ePTD group vs. those delivering later.

Conclusions: Women with a SCx at 16 to $22 \frac{1}{7}$ weeks have a unique AF cytokine network correlating with cervical length at diagnosis and GA at delivery. This network may assist in predicting ePTD.

Keywords: amniocentesis, cervical insufficiency, cytokine, macrophage inflammatory protein (MIP), monocyte chemoattractant protein-1 (MCP), network analysis, preterm birth

Running Head: AF Cytokine Network in Women with Short Cervix

Introduction

A sonographic short cervix is a strong predictor of spontaneous preterm delivery (1-15). The earlier the diagnosis of a short cervix, the more likely the patient will deliver preterm (16-33). Compelling evidence supports the view that this condition is syndromic in nature and has multiple etiologies (34, 35, 13, 36, 37), such as a decline in progesterone action (38-48), prior

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cervical surgery (49-57), intra-amniotic inflammation and/or infection (58-63), and genuine cervical insufficiency (35, 64). Patients with a mid-trimester sonographic short cervix can be offered treatment with vaginal progesterone (65-73) and, for those with a history of prior preterm birth, a cervical cerclage (74-80) may be placed.

A subset of patients with a mid-trimester sonographic short cervix [cervical length (CL) \leq 25 mm] may have microbial invasion of the amniotic fluid (MIAC) with or without inflammation (62, 81, 82). A sterile intra-amniotic inflammation appears to be more common than MIAC (10% versus 2.2%, respectively) and is associated with an increased risk of spontaneous preterm delivery <34 weeks of gestation (82). The presence of intra-amniotic inflammation (both microbial and sterile) is associated with adverse pregnancy outcomes (83-89, 35, 90, 91) in patients presenting with preterm labor and intact membranes (92-108), preterm prelabor rupture of the membranes (PPROM) (109-114), a sonographic short cervix (115, 60, 62, 82), and cervical insufficiency (58, 59, 116, 64, 117). Thus, we and others have explored the behavior of cytokines and other inflammatory-related markers in amniotic (100, 118, 119, 106, 120-123, 107, 82, 108, 124-128) or vaginal (129-140) fluid.

Disease states are generally caused by the interaction of a group of correlated molecules (or network) and *not* by the effect of a single or individual molecule (141-147). For example, in the context of inflammation, one cytokine can induce the expression of others, which can modulate the immune response and lead to the development of feedback regulatory networks (148). Thus, in order to understand the pathogenesis of inflammatory-related diseases, cytokines should ideally be studied as groups of interacting molecules/proteins (149-151). Indeed, recent evidence suggests that biomarkers derived from network analysis are able to achieve better diagnostic performance than those derived using a single-molecule approach (152-155, 61, 156, 157, 148, 158).

Women with microbial-associated or sterile intra-amniotic inflammation had a more coordinated cytokine network than those without intra-amniotic inflammation (127). By using a network approach, we were able to demonstrate that microbial and sterile intra-amniotic inflammation differ in their cytokine networks (127). Therefore, the objective of this study was to examine the amniotic fluid inflammatory-related protein network in asymptomatic patients with a mid-

trimester sonographic short cervix according to gestational age at the time of diagnosis and delivery.

Materials and methods

Study population

A retrospective cohort study was conducted to include 223 patients with an asymptomatic sonographic short cervix. Amniotic fluid samples of participants were selected from the clinical database and Bank of Biological Materials of Wayne State University, the Detroit Medical Center, and the Perinatology Research Branch of the *Eunice Kennedy Shriver* National Institute of Child Health and Human Development (NICHD) if the patients met the following criteria: 1) singleton gestation; 2) asymptomatic sonographic cervical length ≤ 25 mm; 3) transabdominal amniocentesis performed for molecular microbiological studies; and 4) available pregnancy outcomes. Exclusion criteria were: 1) rupture of membranes or preterm labor symptoms before amniotic fluid collection; 2) chromosomal or structural fetal anomaly; or 3) placenta previa. A subset of patients in the current study was included in a prior study (82), which provides description of the molecular microbiologic findings of the amniotic cavity and amniotic fluid IL-6 concentrations. All patients provided written informed consent, and the use of biological specimens as well as clinical and ultrasound data for research purposes was approved by the Institutional Review Boards of Wayne State University and NICHD.

Clinical definitions, methods of sonographic assessment of the cervix, sample collection, and the detection of microorganisms as well as amniotic fluid IL-6 concentrations were previously reported (82). Briefly, spontaneous preterm labor was defined by the presence of regular uterine contractions occurring at least twice every 10 minutes, associated with cervical changes before 37 completed weeks of gestation. Early spontaneous preterm delivery was defined as spontaneous preterm delivery before 32 completed weeks of gestation. Patients classified as having spontaneous preterm delivery included those with spontaneous preterm labor, preterm prelabor rupture of the membranes (PPROM), and those whose labor was induced or augmented due to clinical chorioamnionitis (159). Patients who were lost to follow-up or who had unavailable delivery data were censored.

Intra-amniotic inflammation was characterized by an amniotic fluid IL-6 concentration ≥ 2.6 ng/mL (100, 118, 106, 120, 122, 123, 107, 82, 108, 124, 114). Microbial invasion of the amniotic cavity was defined according to the results of amniotic fluid culture and broad-range polymerase chain reaction (PCR) coupled with electrospray ionization mass spectrometry (PCR/ESI-MS) analysis (105, 113). Acute histologic chorioamnionitis was diagnosed based on the presence of inflammatory cells in the chorionic plate and/or chorioamniotic membranes, and acute funisitis was diagnosed by the presence of neutrophils in the wall of the umbilical vessels and/or Wharton's jelly, using previously described criteria (86, 160-169).

Multiplex determination of inflammatory-related proteins

Amniotic fluid concentrations of 33 inflammatory-related proteins were determined using a multiplex bead array assay developed by the investigators as previously described in detail (see Supplementary Methods) (127).

Statistical analysis

The goal of the statistical analysis was to a) assess the relation between the risk of preterm delivery and gestational age at amniocentesis, b) assess the relation between inflammatory-related protein concentration and cervical length, c) evaluate differences in inflammatory-related protein concentration between groups (delivery ≤ 32 or >32 weeks of gestation) and d) evaluate differences in correlation of pairs of proteins between groups and build correlation networks. To accomplish these goals, women were divided into three groups based on the gestational age at amniocentesis with the two cut-off points representing tertiles of the distribution: 16 5/7 to 22 1/7, 22 2/7 to 26 1/7, and 26 2/7 to 31 5/7 weeks of gestation. In addition to creating equal-sized subgroups of patients, this sub-classification of samples is based on i) previous observations that the populations of patients diagnosed with short cervix are different depending whether or not the diagnosis is made earlier as opposed to later in gestation (81) and also on ii) the observation that pro-inflammatory protein concentrations may vary with gestational age at amniocentesis (127).

The association between the interval of gestational age at amniocentesis and risk of preterm delivery was tested using a chi-squared test for trend in proportions. To test for differential

analyte concentration between groups within each gestational age interval, a linear model was fit to the log (base 2) of analyte concentration as a function of the group indicator (patient delivered ≤ 32 or > 32 weeks of gestation) while adjusting for gestational age at amniocentesis. Similarly, to test the association between analyte concentration and cervical length, a linear model was fit to log (base 2) of analyte concentration as a function of cervical length and the group indicator (patient delivered ≤ 32 or > 32 weeks of gestation) while adjusting for the gestational age at amniocentesis. An interaction term was allowed between the group indicator and cervical length to accommodate possible differences in the rate of change of analyte concentration with cervical length between groups.

Significance test p values for the group (delivery ≤ 32 or > 32 weeks of gestation) coefficient as well as for the cervical length coefficient were adjusted with the Benjamini-Hochberg method over all 33 analytes to compute q-values. Significance of differences in concentration was determined based on a q-value < 0.1 and a fold change > 1.5 . For changes with cervical length, the minimum fold change cut-off of 1.5 was defined over 1cm change in cervical length.

The difference in concentration correlation of each possible pair of analytes (e.g., IL-1 α and IL-6, IL-1 α and IL-33, etc.) between groups was assessed as follows. A linear model was fit to the log-transformed data of each protein as a function of gestational age using samples of each group separately in each gestational-age interval. The residuals (actual value - fitted value) were then used to compute the Pearson correlation for each pair of analytes (called partial correlations). To test for differences in partial correlations between groups within each interval, the partial correlations were first converted into an intermediary statistic using Fisher's z transformation. Under the null hypothesis (partial correlations are equal between groups), the standardized differences in z values among groups were assumed to follow a standard normal distribution. Significant differences in partial correlations were inferred based on $p < 0.01$ and the magnitude of correlation differences was at least 0.2.

A network of differential correlation between groups of women was constructed for each gestational-age interval by linking/connecting the proteins with significant differences in partial correlations (also referred to as perturbed correlation). For each node (protein) in the network,

we calculated the degree (number of perturbed correlation of a given protein), and the average absolute difference in correlations as previously described (127).

To identify confounding variables that could impact the analyses described above, we have used Chi-square tests to determine the association of covariates (administration of steroids, antibiotics, and progesterone before amniocentesis) and preterm delivery within each interval. Covariates with significant association with the outcome were adjusted for in the differential concentration and differential correlation analyses.

Results

Characteristics of the study population

Two hundred twenty-three patients who had a sonographic short cervix (≤ 25 mm) were included in this study. Demographic and clinical characteristics of the study population are displayed in Table 1. The median gestational age at amniocentesis was 24 $\frac{3}{7}$ weeks. Forty-four percent (99/223) of patients with a short cervix delivered at term. Most patients did not have intra-amniotic inflammation [75.3% (168/223)]. Among patients with intra-amniotic inflammation, the frequency of sterile intra-amniotic inflammation was higher than microbial-associated intra-amniotic inflammation [10.3% (23/223) and 2.2% (5/223), respectively] (Table 1). Patients with a sonographic short cervix were classified according to gestational age at delivery, and 28.7% (64/223) delivered ≤ 32 weeks of gestation while the remaining 71.3% (159/223) delivered > 32 weeks of gestation.

The samples included in this study were collected before the publication of randomized trials reporting that vaginal progesterone reduces the rate of preterm delivery and neonatal morbidity; therefore, none of our patients received vaginal progesterone. Eight percent (18/221) of patients underwent cerclage. There were no significant differences in the frequencies with which 17-alpha-hydroxyprogesterone caproate, antibiotic, and antenatal corticosteroids were administered within 7 days before amniocentesis between women with a short cervix who delivered ≤ 32 weeks of gestation and those who delivered > 32 weeks of gestation (Table 1).

The prevalence of intra-amniotic inflammation (defined as amniotic fluid IL-6 concentration ≥ 2.6 ng/mL) including both microbial-associated and sterile intra-amniotic inflammation, was

significantly higher in patients with a sonographic short cervix who delivered ≤ 32 weeks of gestation than in those who delivered >32 weeks of gestation ($p < 0.05$ for all; Table 1). Additionally, the median amniotic fluid IL-6 concentrations (ng/mL) and the frequency of acute inflammatory lesions of the placenta were significantly higher and more frequent in patients with a short cervix who delivered ≤ 32 weeks of gestation than in those who delivered >32 weeks of gestation [IL-6: median (IQR); 1.11 (0.31-4.15) vs. 0.59 (0.28-0.99); $p = 0.002$; acute inflammatory lesion of the placenta: 73.3% (44/60) vs. 33.3% (48/144); $p < 0.001$; Table 1].

The risk of preterm delivery according to the presence of a sonographic short cervix and gestational age at diagnosis

Patients with a short cervix who delivered ≤ 32 weeks of gestation had a significantly lower gestational age at amniocentesis/ at diagnosis with a short cervix and they had a shorter sonographic cervical length than those who delivered >32 weeks of gestation ($p < 0.001$ for all; Table 1). When patients were stratified according to the tertiles of gestational age at amniocentesis, the rate of delivery at ≤ 32 was 50% in those with asymptomatic short cervix identified at 16 5/7 - 22 1/7, 21% in those identified at 22 2/7 - 26 1/7, and 15% in those at 26 2/7-31 5/7 weeks of gestation (Chi-squared test for trend $p < 0.0001$).

Potential effects of confounding variables

We examined whether the administration of steroids, antibiotics, and progesterone before amniocentesis could have had a confounding effect on the differential concentration analysis of proteins. The association of these covariates with the “group indicator” (whether a patient delivered ≤ 32 or >32 weeks of gestation) in each gestational age interval at amniocentesis was assessed and was found to be significant ($p = 0.014$) only for steroids in the gestational age window between 26 2/7 and 31 5/7 weeks interval of gestational age at amniocentesis. Therefore, we adjusted for exposure to steroids in the third interval of gestational age at amniocentesis and found little effect on the differential concentration and correlation analysis results, as described below.

The inflammatory protein concentrations and sonographic cervical length

We have tested the association between inflammatory-related protein concentrations and cervical length within the group of patients in which the diagnosis with short cervix/ gestational age at amniocentesis was made early in pregnancy (16 5/7 - 22 1/7 weeks) while controlling for the gestational age at amniocentesis. This set of patients include an equal number of deliveries at ≤ 32 and > 32 weeks of gestation. The concentration of all inflammatory-related proteins were inversely related to cervical length (i.e. the shorter the cervix, the higher the concentration the proteins) ($q < 0.1$ for all). Only IP-10 and IL-18 did not reach the required minimum of 1.5 fold change in abundance per cm change in cervical length to be considered statistically significant. Fold changes for significant proteins ranged from 1.5 to 8.5 fold per cm (IL-33 and IL-8, respectively) (see Figure 1 and Table 2). For three proteins (MIP-1 α , MCP-1, IL-6) the rate of change in abundance with cervical length was higher in patients who delivered at ≤ 32 weeks compared to those who delivered at > 32 weeks of gestation (Figure 2).

Differences in amniotic fluid inflammatory-related protein concentrations between patients with a sonographic short cervix who delivered before and after 32 weeks of gestation

The amniotic fluid concentrations of 33 inflammatory-related proteins were compared between patients with a sonographic short cervix who delivered ≤ 32 and > 32 weeks of gestation. Since the amniotic fluid concentration of these analytes has a non-linear relationship with gestational age and their discriminatory power may vary as a function of duration of pregnancy, we assessed differential concentration within three separate intervals and adjusting for gestational age at amniocentesis.

Amniotic fluid concentration of inflammatory proteins in patients with a short cervix diagnosed between 16 5/7 and 22 1/7 weeks of gestation

All inflammatory proteins but five (IL-18, IL-33, IP-10, MIG, and MIP-3 α) had a significantly higher mean concentrations in patients who delivered ≤ 32 weeks of gestation than in those who delivered > 32 weeks of gestation (fold change > 1.5 ; q -value < 0.1). The highest changes of amniotic fluid protein concentration were observed for IL-8, MIP-1 β , IL-6, and IL-10 (5.6, 5.1, 4.5, and 4.4-fold, respectively) (Table 3). Of the 28 inflammatory-related proteins whose concentrations were significantly different between groups in this interval, 18 would remain

significant when the analysis was restricted to patients without intra-amniotic inflammation (see highlighted proteins in Table 3).

Amniotic fluid concentration of inflammatory proteins in patients with a short cervix diagnosed between 22 2/7 and 26 1/7 weeks of gestation

In this gestational age window, the differences in amniotic fluid concentrations of inflammatory-related proteins between groups was of a lesser magnitude than those in the first gestational age interval (16 5/7 to 22 1/7 weeks); for nine analytes, the mean amniotic fluid concentration was significantly higher in patients who delivered ≤ 32 weeks of gestation compared to those who delivered >32 weeks of gestation (fold change >1.5 ; q-value <0.1). The three analytes with the highest concentration were IL-8, IL-6, and MIP-1 β (5.4, 4.2, and 2.6-fold, respectively, Table 3).

Amniotic fluid concentration of inflammatory proteins in patients with a short cervix diagnosed between 26 2/7 and 31 5/7 weeks of gestation

The difference in amniotic fluid concentration of inflammatory-related proteins between groups was lesser than that in the other two gestational age groups. The only protein with a significant difference in median concentration between patients who delivered ≤ 32 weeks of gestation compared to those who delivered >32 weeks of gestation was IL-8 (fold change of 2.3 and nominal p value <0.05 , Table 3). However, this significance was lost when adjusting for the administration of steroids, which was a confounder variable in the analysis of patients in this interval of gestational age (as described above).

Amniotic fluid cytokine network analysis according to gestational age at diagnosis of a short cervix

Since the concentration of pro-inflammatory proteins changes with gestational age at amniocentesis, (see Figure 3, top panel for IL-10 and MIP-1 β) differences in correlation patterns of pairs of proteins between groups were assessed within three intervals of gestational age, after removing the linear trend with gestational age from log (base 2) protein concentration (Figure 3, bottom panel). The example presented in Figure 3 shows that in the interval 16 5/7 and 22 1/7

weeks, the partial correlation between MIP-1 β and IL-10 is 0.65 in patients who delivered at ≤ 32 weeks and only 0.05 in patients who delivered >32 weeks (difference in correlation coefficient of $0.65-0.05=0.60$, $p=0.0055$, see Table S1).

Patients with a sonographic short cervix diagnosed between 16 5/7 and 22 1/7 weeks of gestation who delivered ≤ 32 weeks of gestation had a higher concentration of inflammatory-related proteins than those who delivered >32 weeks of gestation (22 perturbed proteins). IL-8, MIP-1 β , and MIP-1 α had the highest number of perturbed correlations (degrees of 6, 5, and 5, respectively), while IL-8, MIP-1 β , and IL-6 showed the largest magnitude of change between groups (Figure 4A).

Patients with a sonographic short cervix diagnosed between 22 2/7 and 26 1/7 weeks of gestation who delivered ≤ 32 weeks of gestation had higher correlation among 16 amniotic fluid inflammatory-related proteins than those who delivered >32 weeks of gestation (35 perturbed correlations). RANTES, IL-8, IL-6, MCP-1, and MIP-1 β had the highest number of perturbed correlations (degree of 11, 8, 8, 8, and 6, respectively), whereas IL-6 and IL-8 demonstrated the highest magnitude of change in concentration (Figure 4B).

Among patients with a sonographic short cervix diagnosed between 26 2/7 and 31 5/7 weeks of gestation who delivered ≤ 32 weeks of gestation had the less degree of perturbation in correlation and expression in amniotic fluid cytokines compared to those who delivered >32 weeks of gestation (only 5 perturbed correlations).

Discussion

Principal findings of the study: 1) The earlier the short cervix was diagnosed, the higher the rate of preterm delivery ≤ 32 weeks of gestation, reaching 50% in the 16 5/7 to 22 1/7 weeks

interval; 2) In patients with short cervix diagnosed at 16 5/7 to 22 1/7 weeks, the concentration of most inflammatory-related proteins increases as the cervix shortens in both delivery groups, yet the rate of increase of MIP-1 α , MCP-1 and IL-6 is higher in women who delivered at ≤ 32 weeks compared to those who delivered at >32 weeks of gestation 3) The concentration of most inflammatory-related proteins studied herein was higher in patients who delivered at ≤ 32 weeks compared to those who delivered at >32 weeks of gestation, yet the magnitude of the difference decreases with increasing gestational age at amniocentesis and diagnosis of a short sonographic cervix. 4) The concentration correlation of several pairs of inflammatory-related proteins was overall higher in patients who delivered at ≤ 32 weeks compared to those who delivered at >32 weeks of gestation.

Inflammatory-related protein network connectivity of patients who had preterm delivery ≤ 32 weeks of gestation and had been diagnosed with a short cervix between 16 and 22 weeks

Patients with a sonographic short cervix and a subsequent early preterm delivery had more coordinated amniotic fluid cytokine concentrations than those who had late preterm delivery. IL-8 and MIP-1 β are the key mediators derived from both differential correlation and concentration analysis. Interestingly, MIP-1 α is the top-ranked protein obtained from the differential correlation approach despite its small magnitude of change in concentration (fold change=1.56; $p=0.00016$).

The 2 top-ranked proteins, IL-8 and MIP-1 β , obtained from the analysis of both correlation network and differential concentration have been implicated in the pathogenesis of intra-amniotic inflammation/infection in preterm labor (170-172), preterm prelabor rupture of the membranes (172), and clinical chorioamnionitis (173). Both IL-8 (neutrophil attractant/activating peptide-1) and MIP-1 β (CCL4) are chemotactic cytokines that activate neutrophils and other human granulocytes in response to inflammation or infection (174, 175). These chemokines are mainly produced by macrophages, mononuclear cells, and lymphocytes (176, 175). Previous studies demonstrated that amniotic fluid concentrations of IL-8 and MIP-1 β increase in patients with preterm labor who had intra-amniotic inflammation/infection and clinical chorioamnionitis at term with proven intra-amniotic infection, and they can predict the likelihood of spontaneous

preterm delivery (177, 170, 171, 178, 172, 179-183). In addition, both chemokines are significantly higher in patients with a mid-trimester short cervix who delivered <34 weeks than in those who delivered ≥ 34 weeks of gestation (155, 148), and their concentrations correlated with cervical length (61).

In contrast, MIP-1 α (or CCL-3) is the top-ranked protein identified from correlation analysis *but not* differential concentration analysis. Interestingly, this cytokine has been implicated in the pathogenesis of intra-amniotic inflammation/infection in patients with preterm delivery and clinical chorioamnionitis (183). MIP-1 α is produced by macrophages (184) and activates human granulocytes in response to inflammation and infection (185-193). These observations suggest that proteins ranked by network correlation analysis are of value and have biological plausibility in the pathogenesis of intra-amniotic inflammation/infection in patients with a sonographic short cervix, despite having a small magnitude of change from a simple comparison of mean concentrations.

Inflammatory-related protein network connectivity between patients who delivered ≤ 32 and >32 weeks of gestation with a short cervix diagnosed between 22 2/7 and 26 1/7 weeks of gestation

The top-ranked proteins obtained from differential concentration analysis (IL-8, IL-6, and MIP-1 β) are similar to those derived from correlation analysis. Moreover, IL-8 and MIP-1 β are also the top-ranked proteins in the network of perturbed inflammatory-related protein concentrations between 16 5/7 and 22 1/7 weeks of gestation. RANTES and MCP-1 are the two additional proteins that are key mediators during this interval derived from correlation analysis. MCP-1 is significantly lower in patients who delivered ≤ 32 weeks than in those who delivered >32 weeks when a sonographic short cervix was diagnosed between 22 2/7 and 26 1/7 weeks of gestation (fold change=1.97, $p=0.022$). The amniotic fluid concentrations of RANTES were not significantly different ($p=0.156$). Interestingly, RANTES had no perturbed correlations of inflammatory-related protein concentration in women with a short cervix diagnosed between 16 5/7 and 22 1/7 weeks, but it had the highest number of perturbed correlations in women with a

short cervix diagnosed between 22.2 and 26.1 weeks of gestation (n=11), suggesting that gestational age is a factor that may modulate the nature of the cytokine response.

It is well established that amniotic fluid IL-8, IL-6, and MIP-1 β are involved in the pathogenesis of intra-amniotic inflammation/infection in preterm delivery (194, 170, 195, 171, 196, 178, 172, 101, 181, 182, 197, 108), preterm prelabor rupture of the membranes (179, 180, 111, 198-201), clinical chorioamnionitis (183), and cervical insufficiency (202-204). In patients with a mid-trimester short cervix, the concentrations of these cytokines/chemokines in the amniotic cavity are correlated with sonographic cervical length (61) and gestational age at delivery (148). In addition, all three cytokines are included in the amniotic fluid inflammatory score model, which can predict gestational age at delivery (148).

In the current study, the information obtained from network analysis is of value, since there is evidence that suggests that amniotic fluid MCP-1 (also known as CCL-2) and RANTES play a role in intra-amniotic inflammation/infection in patients with preterm labor (132, 205, 155, 206), preterm prelabor rupture of the membranes (132), a sonographic short cervix (155, 61), spontaneous labor at term (207, 208), and histological chorioamnionitis (205).

Keeler et al. reported that patients with a mid-trimester sonographic short cervix demonstrated that an elevation of amniotic fluid MCP-1 concentration (>1500 pg/mL) had 60% sensitivity, 100% specificity, and a positive predictive value for delivery <34 weeks of gestation (155). Moreover, MCP-1 was the most predictive cytokine of spontaneous preterm delivery in patients with a mid-trimester short cervix among the other 25 cytokines (155). MCP-1 is capable of recruiting macrophages and other leukocytes into sites of inflammation (209-212). Similarly, RANTES is a pro-inflammatory chemokine secreted from T cells, which recruit monocytes, lymphocytes, basophils, and eosinophils in the host response to inflammation/infection (213-217).

Collectively, patients with a sonographic short cervix who subsequently delivered early had a more coordinated cytokine network than those who had a late preterm delivery. The ranked proteins derived from correlation analysis are informative, have known biological properties relevant to parturition, and can be potentially useful in the future for the development of a

biomarker pipeline to identify patients with a sonographic short cervix who subsequently deliver early.

Inflammatory-related protein network connectivity between patients who delivered ≤ 32 and >32 weeks of gestation with a short cervix diagnosed between 26 2/7 and 31 5/7 weeks of gestation

In this interval, the number of perturbed correlations is small ($n=5$), and none of the inflammatory-related protein concentrations differed between patients who delivered early or late preterm. However, we found a connection between the antimicrobial peptide lactoferrin and T cell-associated cytokines IL-16 and IL-13. Elevated IL-16 and lactoferrin amniotic fluid concentrations in women with preterm labor and intra-amniotic infection/inflammation were previously reported by our group and others (218-220).

Gestational age at the diagnosis of a sonographic short cervix and the behavior of the cytokine network

It is known that the earlier the diagnosis of a short cervix is made, the greater the degree of intra-amniotic inflammation and the earlier the delivery (17-21, 23-27, 30, 61, 62). In addition, Moroz et al, reported that in women with a sonographic short cervix (≤ 25 mm) diagnosed before 25 weeks of gestation, there was a significant association between cervical shortening and maternal systemic inflammation (determined by C-reactive protein) ($r^2 = 0.34$, $p=0.014$). However, this association was not observed in patients without a short cervix or a short cervix diagnosed after 25 weeks of gestation (221). We present here for the first time that the earlier the occurrence of a short cervix, the more orchestrated is the inflammatory response associated with it in women who subsequently deliver prior to 32 weeks of gestation. These findings suggest that the risk of early spontaneous preterm delivery in asymptomatic women with a sonographic short cervical length changes as a function of gestational age at diagnosis and is associated with the magnitude and character of the intra-amniotic inflammatory processes characterized by network analysis.

The importance of study of protein networks in the “great obstetrical syndromes”

The improved understanding of the immune response and its soluble mediators (cytokines) coupled with the application of molecular biology has led to substantial gains in the description of the behavior of multiple inflammatory-related proteins in health and disease. The initial emphasis in the study of inflammatory molecules was on the individual changes in the concentration and expression of these molecules. This is understandable as discoveries of virtually every cytokine occur one at a time. Now, decades later, a more comprehensive and detailed map of the protein inflammatory network is available, as well as an improved understanding of the nature of the protein-protein interaction and biological function of these molecules. A major advance in the understanding of the biology of the inflammatory response is that cytokines are organized in complex and redundant networks and the realization that global analysis is required to improve insights into the biology and that this approach is superior to a simple catalogue in the changes of concentration of individual cytokines. Indeed, the cellular response during inflammation represents the interaction between the input derived from several cytokines that activate different receptors on the cell surface, leading to generation of several intracellular processes. In addition, each cytokine can attach to different receptors on the cell surface, and this has implications on the type of cellular response, cellular activation, number and profile of cytokine receptors expressed on the cell membrane (222-224). Thus, in order to understand the effect of different cytokines an integrative model of cytokine activity is needed. There are several reports documenting that information derived from correlation network analysis can: 1) improve classification of disease (155, 61, 148); 2) chart the cytokine interaction in terms of their effect on specific inflammatory cell types (B and T cells, macrophages) in single and multiple cell interactions (223, 224); 3) identify new interfaces between signaling molecules in uni- or multi-scale models that incorporates several cell populations (225, 226); 4) be the basis of hypothesis generating studies (227); and 5) identify potential therapeutic targets (228, 229). This has been the case with breast cancer (157) and chronic fatigue syndrome (230). The description of the cytokine network presented herein is novel and may assist in identifying the key cytokines involved in the amniotic fluid inflammatory response at different gestational age windows which in turn may have diagnostic, prognostic, and therapeutic implications.

Strengths and limitations of the study

A major strength of the study is that it is the first one to study the inflammatory protein network in asymptomatic women at risk for preterm birth because of a sonographic short cervix. Second, the study focuses on amniotic fluid, the biological fluid in which major changes in cytokine changes are observed in the context of a short cervix. Third, the characterization of the protein inflammatory network was not restricted to cytokines, but also included antimicrobial proteins, such as lactoferrin, which have been implicated in premature labor. Fourth, the results of the cytokine network were unknown at the time of patient management and, therefore, could not have biased the clinicians and investigators. As with any other studies, replication of the findings is desirable.

Conclusions

We have characterized for the first time the amniotic fluid pro-inflammatory protein network in women with asymptomatic short cervix who are at risk for early preterm birth (<32 weeks of gestation). Importantly, the shorter the cervical length, the greater are the perturbations in the amniotic fluid inflammatory network, and the higher the risk of early preterm delivery. Characterization of the amniotic fluid inflammatory network has implications for the taxonomy of disease for patients with a short cervix and identification of those at risk for early premature birth.

Figure Legends

Figure 1: Association between pro-inflammatory protein concentration and cervical length for patients diagnosed with a short cervix at 16 5/7 - 22 1/7 weeks gestational age. The figure shows the concentration (\log_2 , thereof) as a function of cervical length for 4 of the 33 proteins that increase with cervical shortening (see Table 2) overall both women who delivered at ≤ 32 weeks of gestation (red) and those >32 weeks of gestation (black).

Figure 2: Three proteins that increase in abundance with cervical shortening at a higher rate in women who delivered at ≤ 32 weeks of gestation (red) compared to those who delivered at >32 weeks of gestation (black). As in Figure 1 only patients diagnosed with a short cervix at 16 5/7 -

22 1/7 weeks gestational age (GA) are included in this analysis. Solid lines represent the best linear fit of the \log_2 protein concentration as a function of GA at amniocentesis.

Figure 3: Differential correlation analyses. The figure shows \log_2 concentration (pg/mL) of IL-10 (upper left panel) and MIP1 β (upper right panel) as a function of gestational age at amniocentesis in patients who had a sonographic short cervix and who delivered ≤ 32 weeks of gestation (red) and those >32 weeks (blue). Dashed lines denote the two tertiles of the distribution of gestational age at amniocentesis splitting patients into 3 intervals, and the scatterplot of the \log_2 concentration of IL-10 and MIP1 β in the first interval is shown (bottom left panel). A linear model was fit to the \log_2 concentration of each analyte as a function of gestational age in each group within each gestational-age interval, and residuals were used to compute partial correlations between analytes (bottom right panel). The partial correlation between the two analytes was significantly increased in patients who delivered ≤ 32 weeks of gestation than those >32 weeks.

Figure 4: Network of perturbed cytokine concentration correlations between patients who had a sonographic short cervix and who delivered ≤ 32 weeks of gestation and those who delivered >32 weeks in the first (left panel, A) and second (right panel, B) gestational-age intervals. Each node (sphere) represents one of the 33 analytes, with a link (line) between two nodes representing a significantly perturbed correlation. The node color represents the direction of concentration change between delivery ≤ 32 compared to delivery >32 weeks (red=increased, blue=decreased, and white=no change), while the color intensity is proportional to the magnitude of concentration change. The color of links gives the direction of correlation change (red=increased correlation and blue=decreased correlation), while the color intensity is proportional to the magnitude of correlation change. The numbers inside/outside the dotted black circle represent the node degree/average absolute difference in correlations.

References

1. Andersen HF, Nugent CE, Wanty SD, Hayashi RH. Prediction of risk for preterm delivery by ultrasonographic measurement of cervical length. *American journal of obstetrics and gynecology*. 1990;163(3):859-67.
2. Iams JD, Goldenberg RL, Meis PJ, Mercer BM, Moawad A, Das A, et al. The length of the cervix and the risk of spontaneous premature delivery. National Institute of Child Health and Human Development Maternal Fetal Medicine Unit Network. *The New England journal of medicine*. 1996;334(9):567-72.
3. Goldenberg RL, Iams JD, Mercer BM, Meis PJ, Moawad AH, Copper RL, et al. The preterm prediction study: the value of new vs standard risk factors in predicting early and all spontaneous preterm births. NICHD MFMU Network. *American journal of public health*. 1998;88(2):233-8.
4. Heath VC, Southall TR, Souka AP, Elisseou A, Nicolaides KH. Cervical length at 23 weeks of gestation: prediction of spontaneous preterm delivery. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 1998;12(5):312-7
5. Berghella V, Daly SF, Tolosa JE, DiVito MM, Chalmers R, Garg N, et al. Prediction of preterm delivery with transvaginal ultrasonography of the cervix in patients with high-risk pregnancies: does cerclage prevent prematurity? *American journal of obstetrics and gynecology*. 1999;181(4):809-15.
6. Watson WJ, Stevens D, Welter S, Day D. Observations on the sonographic measurement of cervical length and the risk of premature birth. *The Journal of maternal-fetal medicine*. 1999;8(1):17-9.
7. Cook CM, Ellwood DA. The cervix as a predictor of preterm delivery in 'at-risk' women. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2000;15(2):109-13.
8. Hassan SS, Romero R, Berry SM, Dang K, Blackwell SC, Treadwell MC, et al. Patients with an ultrasonographic cervical length \leq 15 mm have nearly a 50% risk of early spontaneous preterm delivery. *American journal of obstetrics and gynecology*. 2000;182(6):1458-67.
9. To MS, Skentou C, Liao AW, Cacho A, Nicolaides KH. Cervical length and funneling at 23 weeks of gestation in the prediction of spontaneous early preterm delivery. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2001;18(3):200-3.
10. Owen J, Yost N, Berghella V, MacPherson C, Swain M, Dildy GA, 3rd, et al. Can shortened midtrimester cervical length predict very early spontaneous preterm birth? *American journal of obstetrics and gynecology*. 2004;191(1):298-303.

11. Tekesin I, Eberhart LH, Schaefer V, Wallwiener D, Schmidt S. Evaluation and validation of a new risk score (CLEOPATRA score) to predict the probability of premature delivery for patients with threatened preterm labor. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2005;26(7):699-706.
12. DeFranco EA, Lewis DF, Odibo AO. Improving the screening accuracy for preterm labor: is the combination of fetal fibronectin and cervical length in symptomatic patients a useful predictor of preterm birth? A systematic review. *American journal of obstetrics and gynecology*. 2013;208(3):233 e1-6.
13. Romero R, Yeo L, Miranda J, Hassan SS, Conde-Agudelo A, Chaiworapongsa T. A blueprint for the prevention of preterm birth: vaginal progesterone in women with a short cervix. *Journal of perinatal medicine*. 2013;41(1):27-44.
14. Boots AB, Sanchez-Ramos L, Bowers DM, Kaunitz AM, Zamora J, Schlattmann P. The short-term prediction of preterm birth: a systematic review and diagnostic metaanalysis. *American journal of obstetrics and gynecology*. 2014;210(1):54 e1- e10.
15. McIntosh J, Feltovich H, Berghella V, Manuck T. The role of routine cervical length screening in selected high- and low-risk women for preterm birth prevention. *American journal of obstetrics and gynecology*. 2016;215(3):B2-7.
16. Kushnir O, Vigil DA, Izquierdo L, Schiff M, Curet LB. Vaginal ultrasonographic assessment of cervical length changes during normal pregnancy. *American journal of obstetrics and gynecology*. 1990;162(4):991-3.
17. Okitsu O, Mimura T, Nakayama T, Aono T. Early prediction of preterm delivery by transvaginal ultrasonography. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 1992;2(6):402-9.
18. Iams JD, Paraskos J, Landon MB, Teteris JN, Johnson FF. Cervical sonography in preterm labor. *Obstetrics and gynecology*. 1994;84(1):40-6.
19. Tongsong T, Kamprapanth P, Srisomboon J, Wanapirak C, Piyamongkol W, Sirichotiyakul S. Single transvaginal sonographic measurement of cervical length early in the third trimester as a predictor of preterm delivery. *Obstetrics and gynecology*. 1995;86(2):184-7.
20. Hasegawa I, Tanaka K, Takahashi K, Tanaka T, Aoki K, Torii Y, et al. Transvaginal ultrasonographic cervical assessment for the prediction of preterm delivery. *The Journal of maternal-fetal medicine*. 1996;5(6):305-9.

21. Rozenberg P, Goffinet F, Malagrida L, Giudicelli Y, Perdu M, Houssin I, et al. Evaluating the risk of preterm delivery: a comparison of fetal fibronectin and transvaginal ultrasonographic measurement of cervical length. *American journal of obstetrics and gynecology*. 1997;176(1 Pt 1):196-9.
22. Tongsong T, Kamprapanth P, Pitaksakorn J. Cervical length in normal pregnancy as measured by transvaginal sonography. *International journal of gynaecology and obstetrics: the official organ of the International Federation of Gynaecology and Obstetrics*. 1997;58(3):313-5.
23. Guzman ER, Mellon C, Vintzileos AM, Ananth CV, Walters C, Gipson K. Longitudinal assessment of endocervical canal length between 15 and 24 weeks' gestation in women at risk for pregnancy loss or preterm birth. *Obstetrics and gynecology*. 1998;92(1):31-7.
24. Taipale P, Hiilesmaa V. Sonographic measurement of uterine cervix at 18-22 weeks' gestation and the risk of preterm delivery. *Obstetrics and gynecology*. 1998;92(6):902-7.
25. Andrews WW, Copper R, Hauth JC, Goldenberg RL, Neely C, Dubard M. Second-trimester cervical ultrasound: associations with increased risk for recurrent early spontaneous delivery. *Obstetrics and gynecology*. 2000;95(2):222-6.
26. Hibbard JU, Tart M, Moawad AH. Cervical length at 16-22 weeks' gestation and risk for preterm delivery. *Obstetrics and gynecology*. 2000;96(6):972-8.
27. Owen J, Yost N, Berghella V, Thom E, Swain M, Dildy GA, 3rd, et al. Mid-trimester endovaginal sonography in women at high risk for spontaneous preterm birth. *JAMA : the journal of the American Medical Association*. 2001;286(11):1340-8.
28. Durnwald CP, Walker H, Lundy JC, Iams JD. Rates of recurrent preterm birth by obstetrical history and cervical length. *American journal of obstetrics and gynecology*. 2005;193(3 Pt 2):1170-4.
29. Gomez R, Romero R, Medina L, Nien JK, Chaiworapongsa T, Carstens M, et al. Cervicovaginal fibronectin improves the prediction of preterm delivery based on sonographic cervical length in patients with preterm uterine contractions and intact membranes. *American journal of obstetrics and gynecology*. 2005;192(2):350-9.
30. Matijevic R, Grgic O, Vasilj O. Is sonographic assessment of cervical length better than digital examination in screening for preterm delivery in a low-risk population? *Acta obstetrica et gynecologica Scandinavica*. 2006;85(11):1342-7.
31. Zhou M, Cool D, Grunwald W, Khamis H, McKenna D. Abstract No.516: Clinical findings in amniotic fluid of women with asymptomatic short cervix in the midtrimester. *American journal of obstetrics and gynecology*. 2013;208(1):S222.

32. Raiche E, Ouellet A, Berthiaume M, Rousseau E, Pasquier JC. Short and inflamed cervix predicts spontaneous preterm birth (COLIBRI study). *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2014;27(10):1015-9.
33. Melamed N, Pittini A, Hirsch L, Yogev Y, Korzeniewski SJ, Romero R, et al. Do serial measurements of cervical length improve the prediction of preterm birth in asymptomatic women with twin gestations? *American journal of obstetrics and gynecology.* 2016;215(5):616.e1-.e14.
34. Romero R, Gomez R, Mazor M, Ghezzi F, Yoon BH. The preterm labor syndrome. In: Elder MG, Romero R, Lamont RF, editors. *Preterm labor.* New York: Churchill Livingstone; 1997. p. 29-49.
35. Romero R, Espinoza J, Kusanovic JP, Gotsch F, Hassan S, Erez O, et al. The preterm parturition syndrome. *BJOG : an international journal of obstetrics and gynaecology.* 2006;113 Suppl 3:17-42.
36. Romero R, Dey SK, Fisher SJ. Preterm labor: one syndrome, many causes. *Science (New York, NY).* 2014;345(6198):760-5.
37. Lockwood CJ. Risk factors for preterm birth and new approaches to its early diagnosis. *Journal of perinatal medicine.* 2015;43(5):499-501.
38. Hegele-Hartung C, Chwalisz K, Beier HM, Elger W. Ripening of the uterine cervix of the guinea-pig after treatment with the progesterone antagonist onapristone (ZK 98.299): an electron microscopic study. *Hum Reprod.* 1989;4(4):369-77.
39. Wolf JP, Sinosich M, Anderson TL, Ulmann A, Baulieu EE, Hodgen GD. Progesterone antagonist (RU 486) for cervical dilation, labor induction, and delivery in monkeys: effectiveness in combination with oxytocin. *American journal of obstetrics and gynecology.* 1989;160(1):45-7.
40. Norman J. Antiprogesteroes. *British journal of hospital medicine.* 1991;45(6):372-5.
41. Chwalisz K. The use of progesterone antagonists for cervical ripening and as an adjunct to labour and delivery. *Hum Reprod.* 1994;9 Suppl 1:131-61.
42. Elliott CL, Brennand JE, Calder AA. The effects of mifepristone on cervical ripening and labor induction in primigravidae. *Obstetrics and gynecology.* 1998;92(5):804-9.
43. Stenlund PM, Ekman G, Aedo AR, Bygdeman M. Induction of labor with mifepristone--a randomized, double-blind study versus placebo. *Acta obstetrica et gynecologica Scandinavica.* 1999;78(9):793-8.
44. Word RA, Li XH, Hnat M, Carrick K. Dynamics of cervical remodeling during pregnancy and parturition: mechanisms and current concepts. *Seminars in reproductive medicine.* 2007;25(1):69-79.

45. Timmons B, Akins M, Mahendroo M. Cervical remodeling during pregnancy and parturition. *Trends in endocrinology and metabolism: TEM*. 2010;21(6):353-61.
46. Mahendroo M. Cervical remodeling in term and preterm birth: insights from an animal model. *Reproduction (Cambridge, England)*. 2012;143(4):429-38.
47. Ahn KH, Bae NY, Hong SC, Lee JS, Lee EH, Jee HJ, et al. The safety of progestogen in the prevention of preterm birth: meta-analysis of neonatal mortality. *Journal of perinatal medicine*. 2016.
48. Areia AL, Vale-Pereira S, Vaz-Ambrosio A, Alves V, Rodrigues-Santos P, Rosa MS, et al. Does progesterone administration in preterm labor influence Treg cells? *Journal of perinatal medicine*. 2016;44(6):605-11.
49. Moinian M, Andersch B. Does cervix conization increase the risk of complications in subsequent pregnancies? *Acta obstetrica et gynecologica Scandinavica*. 1982;61(2):101-3.
50. Blomfield PI, Buxton J, Dunn J, Luesley DM. Pregnancy outcome after large loop excision of the cervical transformation zone. *American journal of obstetrics and gynecology*. 1993;169(3):620-5.
51. Kristensen J, Langhoff-Roos J, Wittrup M, Bock JE. Cervical conization and preterm delivery/low birth weight. A systematic review of the literature. *Acta obstetrica et gynecologica Scandinavica*. 1993;72(8):640-4.
52. Raio L, Ghezzi F, Di Naro E, Gomez R, Luscher KP. Duration of pregnancy after carbon dioxide laser conization of the cervix: influence of cone height. *Obstetrics and gynecology*. 1997;90(6):978-82.
53. Berghella V, Pereira L, Gariepy A, Simonazzi G. Prior cone biopsy: prediction of preterm birth by cervical ultrasound. *American journal of obstetrics and gynecology*. 2004;191(4):1393-7.
54. Bruinsma FJ, Quinn MA. The risk of preterm birth following treatment for precancerous changes in the cervix: a systematic review and meta-analysis. *BJOG : an international journal of obstetrics and gynaecology*. 2011;118(9):1031-41.
55. Orzechowski K, Nicholas S, Berghella V. Abstract No. 504: Does cervical conization increase the risk of a sonographic short cervix in the second trimester of pregnancy? *American journal of obstetrics and gynecology*. 2013;208(1):S217.
56. Miller ES, Grobman WA. The association between cervical excisional procedures, midtrimester cervical length, and preterm birth. *American journal of obstetrics and gynecology*. 2014;211(3):242 e1-4.
57. Miller ES, Sakowicz A, Grobman WA. The association between cervical dysplasia, a short cervix, and preterm birth. *American journal of obstetrics and gynecology*. 2015;213(4):543 e1-4.

58. Romero R, Gonzalez R, Sepulveda W, Brandt F, Ramirez M, Sorokin Y, et al. Infection and labor. VIII. Microbial invasion of the amniotic cavity in patients with suspected cervical incompetence: prevalence and clinical significance. *American journal of obstetrics and gynecology*. 1992;167(4 Pt 1):1086-91.
59. Mays JK, Figueroa R, Shah J, Khakoo H, Kaminsky S, Tejani N. Amniocentesis for selection before rescue cerclage. *Obstetrics and gynecology*. 2000;95(5):652-5.
60. Hassan S, Romero R, Hendler I, Gomez R, Khalek N, Espinoza J, et al. A sonographic short cervix as the only clinical manifestation of intra-amniotic infection. *Journal of perinatal medicine*. 2006;34(1):13-9.
61. Kiefer DG, Keeler SM, Rust OA, Wayock CP, Vintzileos AM, Hanna N. Is midtrimester short cervix a sign of intraamniotic inflammation? *American journal of obstetrics and gynecology*. 2009;200(4):374 e1-5.
62. Vaisbuch E, Hassan SS, Mazaki-Tovi S, Nhan-Chang CL, Kusanovic JP, Chaiworapongsa T, et al. Patients with an asymptomatic short cervix (≤ 15 mm) have a high rate of subclinical intraamniotic inflammation: implications for patient counseling. *American journal of obstetrics and gynecology*. 2010;202(5):433 e1-8.
63. Choi J, Park JW, Kim BJ, Choi YJ, Hwang JH, Lee SM. Funisitis is more common in cervical insufficiency than in preterm labor and preterm premature rupture of membranes. *Journal of perinatal medicine*. 2016;44(5):523-9.
64. Lee SE, Romero R, Park CW, Jun JK, Yoon BH. The frequency and significance of intraamniotic inflammation in patients with cervical insufficiency. *American journal of obstetrics and gynecology*. 2008;198(6):633 e1-8.
65. DeFranco EA, O'Brien JM, Adair CD, Lewis DF, Hall DR, Fusey S, et al. Vaginal progesterone is associated with a decrease in risk for early preterm birth and improved neonatal outcome in women with a short cervix: a secondary analysis from a randomized, double-blind, placebo-controlled trial. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2007;30(5):697-705.
66. Fonseca EB, Celik E, Parra M, Singh M, Nicolaides KH. Progesterone and the risk of preterm birth among women with a short cervix. *The New England journal of medicine*. 2007;357(5):462-9.
67. O'Brien JM, DeFranco EA, Adair CD, Lewis DF, Hall DR, How H, et al. Effect of progesterone on cervical shortening in women at risk for preterm birth: secondary analysis from a multinational,

randomized, double-blind, placebo-controlled trial. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2009;34(6):653-9.

68. Hassan SS, Romero R, Vidyadhari D, Fusey S, Baxter JK, Khandelwal M, et al. Vaginal progesterone reduces the rate of preterm birth in women with a sonographic short cervix: a multicenter, randomized, double-blind, placebo-controlled trial. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2011;38(1):18-31.

69. Romero R, Nicolaides K, Conde-Agudelo A, Tabor A, O'Brien JM, Cetingoz E, et al. Vaginal progesterone in women with an asymptomatic sonographic short cervix in the midtrimester decreases preterm delivery and neonatal morbidity: a systematic review and metaanalysis of individual patient data. *American journal of obstetrics and gynecology*. 2012;206(2):124 e1-19.

70. Conde-Agudelo A, Romero R, Nicolaides K, Chaiworapongsa T, O'Brien JM, Cetingoz E, et al. Vaginal progesterone vs. cervical cerclage for the prevention of preterm birth in women with a sonographic short cervix, previous preterm birth, and singleton gestation: a systematic review and indirect comparison metaanalysis. *American journal of obstetrics and gynecology*. 2013;208(1):42 e1-e18.

71. Romero R, Yeo L, Chaemsaitong P, Chaiworapongsa T, Hassan SS. Progesterone to prevent spontaneous preterm birth. *Seminars in fetal & neonatal medicine*. 2014;19(1):15-26.

72. Conde-Agudelo A, Romero R. Vaginal Progesterone to Prevent Preterm Birth in Pregnant Women with a Sonographic Short Cervix: Clinical and Public Health Implications. *American journal of obstetrics and gynecology*. 2015.

73. Suhag A, Saccone G, Berghella V. Vaginal progesterone for maintenance tocolysis: a systematic review and metaanalysis of randomized trials. *American journal of obstetrics and gynecology*. 2015;213(4):479-87.

74. Romero R, Espinoza J, Erez O, Hassan S. The role of cervical cerclage in obstetric practice: can the patient who could benefit from this procedure be identified? *American journal of obstetrics and gynecology*. 2006;194(1):1-9.

75. Keeler SM, Kiefer D, Rochon M, Quinones JN, Novetsky AP, Rust O. A randomized trial of cerclage vs. 17 alpha-hydroxyprogesterone caproate for treatment of short cervix. *Journal of perinatal medicine*. 2009;37(5):473-9.

76. Owen J, Hankins G, Iams JD, Berghella V, Sheffield JS, Perez-Delboy A, et al. Multicenter randomized trial of cerclage for preterm birth prevention in high-risk women with shortened midtrimester cervical length. *American journal of obstetrics and gynecology*. 2009;201(4):375 e1-8.
77. Berghella V, Mackeen AD. Cervical length screening with ultrasound-indicated cerclage compared with history-indicated cerclage for prevention of preterm birth: a meta-analysis. *Obstetrics and gynecology*. 2011;118(1):148-55.
78. Berghella V, Rafael TJ, Szychowski JM, Rust OA, Owen J. Cerclage for short cervix on ultrasonography in women with singleton gestations and previous preterm birth: a meta-analysis. *Obstetrics and gynecology*. 2011;117(3):663-71.
79. Childress KS, Flick A, Dickert E, Gavard J, Bolanos R, Gross G. Abstract No 173: A comparison of cervical cerclage and vaginal pessaries in the prevention of spontaneous preterm birth in women with short cervix. *American journal of obstetrics and gynecology*. 2015;212(1):S101.
80. Kiefer DG, Peltier MR, Keeler SM, Rust O, Ananth CV, Vintzileos AM, et al. Efficacy of Midtrimester Short Cervix Interventions is Conditional on Intra-Amniotic Inflammation. *American journal of obstetrics and gynecology*. 2015.
81. Vaisbuch E, Romero R, Erez O, Kusanovic JP, Mazaki-Tovi S, Gotsch F, et al. Clinical significance of early (< 20 weeks) vs. late (20-24 weeks) detection of sonographic short cervix in asymptomatic women in the mid-trimester. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*. 2010;36(4):471-81.
82. Romero R, Miranda J, Chaiworapongsa T, Chaemsaihong P, Gotsch F, Dong Z, et al. Sterile intra-amniotic inflammation in asymptomatic patients with a sonographic short cervix: prevalence and clinical significance. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2014:1-17.
83. Romero R, Kadar N, Hobbins JC, Duff GW. Infection and labor: the detection of endotoxin in amniotic fluid. *American journal of obstetrics and gynecology*. 1987;157(4 Pt 1):815-9.
84. Romero R, Roslansky P, Oyarzun E, Wan M, Emamian M, Novitsky TJ, et al. Labor and infection. II. Bacterial endotoxin in amniotic fluid and its relationship to the onset of preterm labor. *American journal of obstetrics and gynecology*. 1988;158(5):1044-9.

85. Gravett MG, Witkin SS, Haluska GJ, Edwards JL, Cook MJ, Novy MJ. An experimental model for intraamniotic infection and preterm labor in rhesus monkeys. *American journal of obstetrics and gynecology*. 1994;171(6):1660-7.
86. Yoon BH, Romero R, Kim CJ, Jun JK, Gomez R, Choi JH, et al. Amniotic fluid interleukin-6: a sensitive test for antenatal diagnosis of acute inflammatory lesions of preterm placenta and prediction of perinatal morbidity. *American journal of obstetrics and gynecology*. 1995;172(3):960-70.
87. Hitti J, Tarczy-Hornoch P, Murphy J, Hillier SL, Aura J, Eschenbach DA. Amniotic fluid infection, cytokines, and adverse outcome among infants at 34 weeks' gestation or less. *Obstetrics and gynecology*. 2001;98(6):1080-8.
88. Romero R, Gomez R, Chaiworapongsa T, Conoscenti G, Kim JC, Kim YM. The role of infection in preterm labour and delivery. *Paediatric and perinatal epidemiology*. 2001;15 Suppl 2:41-56.
89. Romero R, Erez O, Espinoza J. Intrauterine infection, preterm labor, and cytokines. *Journal of the Society for Gynecologic Investigation*. 2005;12(7):463-5.
90. Romero R, Espinoza J, Goncalves LF, Kusanovic JP, Friel L, Hassan S. The role of inflammation and infection in preterm birth. *Seminars in reproductive medicine*. 2007;25(1):21-39.
91. Romero R, Gotsch F, Pineles B, Kusanovic JP. Inflammation in pregnancy: its roles in reproductive physiology, obstetrical complications, and fetal injury. *Nutrition reviews*. 2007;65(12 Pt 2):S194-202.
92. Romero R, Mazor M, Wu YK, Sirtori M, Oyarzun E, Mitchell MD, et al. Infection in the pathogenesis of preterm labor. *Seminars in perinatology*. 1988;12(4):262-79.
93. Romero R, Sirtori M, Oyarzun E, Avila C, Mazor M, Callahan R, et al. Infection and labor. V. Prevalence, microbiology, and clinical significance of intraamniotic infection in women with preterm labor and intact membranes. *American journal of obstetrics and gynecology*. 1989;161(3):817-24.
94. Gibbs RS, Romero R, Hillier SL, Eschenbach DA, Sweet RL. A review of premature birth and subclinical infection. *American journal of obstetrics and gynecology*. 1992;166(5):1515-28.
95. Watts DH, Krohn MA, Hillier SL, Eschenbach DA. The association of occult amniotic fluid infection with gestational age and neonatal outcome among women in preterm labor. *Obstetrics and gynecology*. 1992;79(3):351-7.
96. Romero R, Yoon BH, Kenney JS, Gomez R, Allison AC, Sehgal PB. Amniotic fluid interleukin-6 determinations are of diagnostic and prognostic value in preterm labor. *American journal of reproductive immunology*. 1993;30(2-3):167-83.

97. Romero R, Galasso M, Gomez R, Ramirez M, Sorokin Y, Behnke E, et al. A comparative study of the value of amniotic fluid interleukin-6, white blood cell count and gram stain in the diagnosis of microbial invasion of the amniotic cavity in patients with spontaneous labor at term. Annual Meeting of the Society of Perinatal Obstetricians; Las Vegas, NV.1994. p. A250.
98. Greci LS, Gilson GJ, Nevils B, Izquierdo LA, Qualls CR, Curet LB. Is amniotic fluid analysis the key to preterm labor? A model using interleukin-6 for predicting rapid delivery. American journal of obstetrics and gynecology. 1998;179(1):172-8.
99. Maymon E, Romero R, Chaiworapongsa T, Berman S, Conoscenti G, Gomez R, et al. Amniotic fluid matrix metalloproteinase-8 in preterm labor with intact membranes. American journal of obstetrics and gynecology. 2001;185(5):1149-55.
100. Yoon BH, Romero R, Moon JB, Shim SS, Kim M, Kim G, et al. Clinical significance of intra-amniotic inflammation in patients with preterm labor and intact membranes. American journal of obstetrics and gynecology. 2001;185(5):1130-6.
101. Jacobsson B, Mattsby-Baltzer I, Andersch B, Bokstrom H, Holst RM, Wennerholm UB, et al. Microbial invasion and cytokine response in amniotic fluid in a Swedish population of women in preterm labor. Acta obstetrica et gynecologica Scandinavica. 2003;82(2):120-8.
102. Yoon BH, Romero R, Lim JH, Shim SS, Hong JS, Shim JY, et al. The clinical significance of detecting *Ureaplasma urealyticum* by the polymerase chain reaction in the amniotic fluid of patients with preterm labor. American journal of obstetrics and gynecology. 2003;189(4):919-24.
103. Friel LA, Romero R, Edwin S, Nien JK, Gomez R, Chaiworapongsa T, et al. The calcium binding protein, S100B, is increased in the amniotic fluid of women with intra-amniotic infection/inflammation and preterm labor with intact or ruptured membranes. Journal of perinatal medicine. 2007;35(5):385-93.
104. Lee SE, Romero R, Jung H, Park CW, Park JS, Yoon BH. The intensity of the fetal inflammatory response in intraamniotic inflammation with and without microbial invasion of the amniotic cavity. American journal of obstetrics and gynecology. 2007;197(3):294 e1-6.
105. DiGiulio DB, Romero R, Amogan HP, Kusanovic JP, Bik EM, Gotsch F, et al. Microbial prevalence, diversity and abundance in amniotic fluid during preterm labor: a molecular and culture-based investigation. PloS one. 2008;3(8):e3056.
106. Romero R, Chaiworapongsa T, Alpay Savasan Z, Xu Y, Hussein Y, Dong Z, et al. Damage-associated molecular patterns (DAMPs) in preterm labor with intact membranes and preterm PROM: a

study of the alarmin HMGB1. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2011;24(12):1444-55.

107. Romero R, Miranda J, Chaiworapongsa T, Chaemsaihong P, Gotsch F, Dong Z, et al. A novel molecular microbiologic technique for the rapid diagnosis of microbial invasion of the amniotic cavity and intra-amniotic infection in preterm labor with intact membranes. *Am J Reprod Immunol.* 2014;71(4):330-58.

108. Romero R, Miranda J, Chaiworapongsa T, Korzeniewski SJ, Chaemsaihong P, Gotsch F, et al. Prevalence and clinical significance of sterile intra-amniotic inflammation in patients with preterm labor and intact membranes. *Am J Reprod Immunol.* 2014;72(5):458-74.

109. Romero R, Quintero R, Oyarzun E, Wu YK, Sabo V, Mazor M, et al. Intraamniotic infection and the onset of labor in preterm premature rupture of the membranes. *American journal of obstetrics and gynecology.* 1988;159(3):661-6.

110. Gomez R, Romero R, Edwin SS, David C. Pathogenesis of preterm labor and preterm premature rupture of membranes associated with intraamniotic infection. *Infectious disease clinics of North America.* 1997;11(1):135-76.

111. Jacobsson B, Mattsby-Baltzer I, Andersch B, Bokstrom H, Holst RM, Nikolaitchouk N, et al. Microbial invasion and cytokine response in amniotic fluid in a Swedish population of women with preterm prelabor rupture of membranes. *Acta obstetrica et gynecologica Scandinavica.* 2003;82(5):423-31.

112. Shim SS, Romero R, Hong JS, Park CW, Jun JK, Kim BI, et al. Clinical significance of intra-amniotic inflammation in patients with preterm premature rupture of membranes. *American journal of obstetrics and gynecology.* 2004;191(4):1339-45.

113. DiGiulio DB, Romero R, Kusanovic JP, Gomez R, Kim CJ, Seok KS, et al. Prevalence and diversity of microbes in the amniotic fluid, the fetal inflammatory response, and pregnancy outcome in women with preterm pre-labor rupture of membranes. *Am J Reprod Immunol.* 2010;64(1):38-57.

114. Romero R, Miranda J, Chaemsaihong P, Chaiworapongsa T, Kusanovic JP, Dong Z, et al. Sterile and microbial-associated intra-amniotic inflammation in preterm prelabor rupture of membranes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2015;28(12):1394-409.

115. Gomez R, Romero R, Nien JK, Chaiworapongsa T, Medina L, Kim YM, et al. A short cervix in women with preterm labor and intact membranes: a risk factor for microbial invasion of the amniotic cavity. *American journal of obstetrics and gynecology*. 2005;192(3):678-89.
116. Bujold E, Morency AM, Rallu F, Ferland S, Tetu A, Duperron L, et al. Bacteriology of amniotic fluid in women with suspected cervical insufficiency. *Journal of obstetrics and gynaecology Canada : JOGC = Journal d'obstetrique et gynecologie du Canada : JOGC*. 2008;30(10):882-7.
117. Oh KJ, Lee SE, Jung H, Kim G, Romero R, Yoon BH. Detection of ureaplasmas by the polymerase chain reaction in the amniotic fluid of patients with cervical insufficiency. *Journal of perinatal medicine*. 2010;38(3):261-8.
118. Kim KW, Romero R, Park HS, Park CW, Shim SS, Jun JK, et al. A rapid matrix metalloproteinase-8 bedside test for the detection of intraamniotic inflammation in women with preterm premature rupture of membranes. *American journal of obstetrics and gynecology*. 2007;197(3):292 e1-5.
119. Kiefer D, Peltier M, Keeler S, Rust O, Ananth C, Hanna N, et al. Does intra-amniotic inflammation influence pregnancy outcome after cerclage or progesterone (17OHP-C) therapy for midtrimester short cervix? *American journal of obstetrics and gynecology*. 2009;201(6 Supplement):S61.
120. Gervasi MT, Romero R, Bracalente G, Erez O, Dong Z, Hassan SS, et al. Midtrimester amniotic fluid concentrations of interleukin-6 and interferon-gamma-inducible protein-10: evidence for heterogeneity of intra-amniotic inflammation and associations with spontaneous early (<32 weeks) and late (>32 weeks) preterm delivery. *Journal of perinatal medicine*. 2012;40(4):329-43.
121. Lee SY, Park KH, Jeong EH, Oh KJ, Ryu A, Kim A. Intra-amniotic infection/inflammation as a risk factor for subsequent ruptured membranes after clinically indicated amniocentesis in preterm labor. *Journal of Korean medical science*. 2013;28(8):1226-32.
122. Combs CA, Gravett M, Garite TJ, Hickok DE, Lapidus J, Porreco R, et al. Amniotic fluid infection, inflammation, and colonization in preterm labor with intact membranes. *American journal of obstetrics and gynecology*. 2014;210(2):125 e1- e15.
123. Romero R, Kadar N, Miranda J, Korzeniewski SJ, Schwartz AG, Chaemsaitong P, et al. The diagnostic performance of the Mass Restricted (MR) score in the identification of microbial invasion of the amniotic cavity or intra-amniotic inflammation is not superior to amniotic fluid interleukin-6. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2014;27(8):757-69.

124. Chaemsaitong P, Romero R, Korzeniewski SJ, Dong Z, Yeo L, Hassan SS, et al. A point of care test for the determination of amniotic fluid interleukin-6 and the chemokine CXCL-10/IP-10. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2015;28(13):1510-9.
125. Chaemsaitong P, Romero R, Korzeniewski SJ, Martinez-Varea A, Dong Z, Yoon BH, et al. A point of care test for interleukin-6 in amniotic fluid in preterm prelabor rupture of membranes: a step toward the early treatment of acute intra-amniotic inflammation/infection. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2015:1-8.
126. Chaemsaitong P, Romero R, Korzeniewski SJ, Martinez-Varea A, Dong Z, Yoon BH, et al. A rapid interleukin-6 bedside test for the identification of intra-amniotic inflammation in preterm labor with intact membranes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2015:1-11.
127. Romero R, Grivel JC, Tarca AL, Chaemsaitong P, Xu Z, Fitzgerald W, et al. Evidence of perturbations of the cytokine network in preterm labor. *American journal of obstetrics and gynecology.* 2015.
128. Kunze M, Klar M, Morfeld CA, Thorns B, Schild RL, Markfeld-Erol F, et al. Cytokines in noninvasively obtained amniotic fluid as predictors of fetal inflammatory response syndrome. *American journal of obstetrics and gynecology.* 2016;215(1):96.e1-8.
129. Rizzo G, Capponi A, Vlachopoulou A, Angelini E, Grassi C, Romanini C. Interleukin-6 concentrations in cervical secretions in the prediction of intrauterine infection in preterm premature rupture of the membranes. *Gynecologic and obstetric investigation.* 1998;46(2):91-5.
130. Jun JK, Yoon BH, Romero R, Kim M, Moon JB, Ki SH, et al. Interleukin 6 determinations in cervical fluid have diagnostic and prognostic value in preterm premature rupture of membranes. *American journal of obstetrics and gynecology.* 2000;183(4):868-73.
131. Jacobsson B, Holst RM, Mattsby-Baltzer I, Nikolaitchouk N, Wennerholm UB, Hagberg H. Interleukin-18 in cervical mucus and amniotic fluid: relationship to microbial invasion of the amniotic

fluid, intra-amniotic inflammation and preterm delivery. *BJOG : an international journal of obstetrics and gynaecology*. 2003;110(6):598-603.

132. Jacobsson B, Holst RM, Wennerholm UB, Andersson B, Lilja H, Hagberg H. Monocyte chemotactic protein-1 in cervical and amniotic fluid: relationship to microbial invasion of the amniotic cavity, intra-amniotic inflammation, and preterm delivery. *American journal of obstetrics and gynecology*. 2003;189(4):1161-7.

133. Holst RM, Mattsby-Baltzer I, Wennerholm UB, Hagberg H, Jacobsson B. Interleukin-6 and interleukin-8 in cervical fluid in a population of Swedish women in preterm labor: relationship to microbial invasion of the amniotic fluid, intra-amniotic inflammation, and preterm delivery. *Acta obstetrica et gynecologica Scandinavica*. 2005;84(6):551-7.

134. Jacobsson B, Mattsby-Baltzer I, Hagberg H. Interleukin-6 and interleukin-8 in cervical and amniotic fluid: relationship to microbial invasion of the chorioamniotic membranes. *BJOG : an international journal of obstetrics and gynaecology*. 2005;112(6):719-24.

135. Holst RM, Laurini R, Jacobsson B, Samuelsson E, Savman K, Doverhag C, et al. Expression of cytokines and chemokines in cervical and amniotic fluid: relationship to histological chorioamnionitis. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2007;20(12):885-93.

136. Chandiramani M, Seed PT, Orsi NM, Ekbote UV, Bennett PR, Shennan AH, et al. Limited relationship between cervico-vaginal fluid cytokine profiles and cervical shortening in women at high risk of spontaneous preterm birth. *PloS one*. 2012;7(12):e52412.

137. Cobo T, Jacobsson B, Kacerovsky M, Hougaard DM, Skogstrand K, Gratacos E, et al. Systemic and local inflammatory response in women with preterm prelabor rupture of membranes. *PloS one*. 2014;9(1):e85277.

138. Combs CA, Garite TJ, Lapidus JA, Lapointe JP, Gravett M, Rael J, et al. Detection of microbial invasion of the amniotic cavity by analysis of cervicovaginal proteins in women with preterm labor and intact membranes. *American journal of obstetrics and gynecology*. 2015;212(4):482 e1- e12.

139. Hadzi-Lega M, Markova AD, Stefanovic M, Tanturovski M. Correlation of cervical length, fetal fibronectin, pHIGFBP-1, and cytokines in spontaneous preterm birth up to 14 days from sampling. *Journal of perinatal medicine*. 2015;43(5):545-51.

140. Kacerovsky M, Musilova I, Jacobsson B, Drahosova M, Hornychova H, Janku P, et al. Cervical fluid IL-6 and IL-8 levels in pregnancies complicated by preterm prelabor rupture of membranes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2015;28(2):134-40.
141. Oltvai ZN, Barabasi AL. Systems biology. Life's complexity pyramid. *Science (New York, NY).* 2002;298(5594):763-4.
142. Barabasi AL, Oltvai ZN. Network biology: understanding the cell's functional organization. *Nature reviews Genetics.* 2004;5(2):101-13.
143. Klemm K, Bornholdt S. Topology of biological networks and reliability of information processing. *Proceedings of the National Academy of Sciences of the United States of America.* 2005;102(51):18414-9.
144. Dong J, Horvath S. Understanding network concepts in modules. *BMC systems biology.* 2007;1:24.
145. Schadt EE. Molecular networks as sensors and drivers of common human diseases. *Nature.* 2009;461(7261):218-23.
146. Liu R, Wang X, Aihara K, Chen L. Early diagnosis of complex diseases by molecular biomarkers, network biomarkers, and dynamical network biomarkers. *Medicinal research reviews.* 2014;34(3):455-78.
147. Menche J, Sharma A, Kitsak M, Ghiassian SD, Vidal M, Loscalzo J, et al. Disease networks. Uncovering disease-disease relationships through the incomplete interactome. *Science (New York, NY).* 2015;347(6224):1257601.
148. Kiefer DG, Keeler SM, Rust O, Chow SS, Craig ME, Peltier MR, et al. Amniotic fluid inflammatory score is associated with pregnancy outcome in patients with mid trimester short cervix. *American journal of obstetrics and gynecology.* 2012;206(1):68 e1-6.
149. Kitano H. Systems biology: a brief overview. *Science (New York, NY).* 2002;295(5560):1662-4.
150. Chen L, Wang R, Zhang X. Biomolecular networks: Methods and application in systems biology. Hoboken: New Jersey: John Wiley and Sons; 2009. 391 p.
151. Liu ZP, Wang Y, Zhang XS, Chen L. Network-based analysis of complex diseases. *IET systems biology.* 2012;6(1):22-33.

152. Bowen JM, Chamley L, Keelan JA, Mitchell MD. Cytokines of the placenta and extra-placental membranes: roles and regulation during human pregnancy and parturition. *Placenta*. 2002;23(4):257-73.
153. Segal E, Friedman N, Kaminski N, Regev A, Koller D. From signatures to models: understanding cancer using microarrays. *Nature genetics*. 2005;37 Suppl:S38-45.
154. Orsi NM. Cytokine networks in the establishment and maintenance of pregnancy. *Human fertility (Cambridge, England)*. 2008;11(4):222-30.
155. Keeler SM, Kiefer DG, Rust OA, Vintzileos A, Atlas RO, Bornstein E, et al. Comprehensive amniotic fluid cytokine profile evaluation in women with a short cervix: which cytokine(s) correlates best with outcome? *American journal of obstetrics and gynecology*. 2009;201(3):276 e1-6.
156. Slavov N, Dawson KA. Correlation signature of the macroscopic states of the gene regulatory network in cancer. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;106(11):4079-84.
157. Taylor IW, Linding R, Warde-Farley D, Liu Y, Pesquita C, Faria D, et al. Dynamic modularity in protein interaction networks predicts breast cancer outcome. *Nature biotechnology*. 2009;27(2):199-204.
158. Pinheiro MB, Martins-Filho OA, Mota AP, Alpoim PN, Godoi LC, Silveira AC, et al. Severe preeclampsia goes along with a cytokine network disturbance towards a systemic inflammatory state. *Cytokine*. 2013;62(1):165-73.
159. Gibbs RS, Blanco JD, St Clair PJ, Castaneda YS. Quantitative bacteriology of amniotic fluid from women with clinical intraamniotic infection at term. *The Journal of infectious diseases*. 1982;145(1):1-8.
160. Yoon BH, Romero R, Park JS, Kim M, Oh SY, Kim CJ, et al. The relationship among inflammatory lesions of the umbilical cord (funisitis), umbilical cord plasma interleukin 6 concentration, amniotic fluid infection, and neonatal sepsis. *American journal of obstetrics and gynecology*. 2000;183(5):1124-9.
161. Park JS, Romero R, Yoon BH, Moon JB, Oh SY, Han SY, et al. The relationship between amniotic fluid matrix metalloproteinase-8 and funisitis. *American journal of obstetrics and gynecology*. 2001;185(5):1156-61.
162. Pacora P, Chaiworapongsa T, Maymon E, Kim YM, Gomez R, Yoon BH, et al. Funisitis and chorionic vasculitis: the histological counterpart of the fetal inflammatory response syndrome. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2002;11(1):18-25.

163. Yoon BH, Romero R, Shim JY, Shim SS, Kim CJ, Jun JK. C-reactive protein in umbilical cord blood: a simple and widely available clinical method to assess the risk of amniotic fluid infection and funisitis. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2003;14(2):85-90.
164. Redline RW, Heller D, Keating S, Kingdom J. Placental diagnostic criteria and clinical correlation--a workshop report. *Placenta.* 2005;26 Suppl A:S114-7.
165. Lee SE, Romero R, Kim CJ, Shim SS, Yoon BH. Funisitis in term pregnancy is associated with microbial invasion of the amniotic cavity and intra-amniotic inflammation. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet.* 2006;19(11):693-7.
166. Redline RW. Inflammatory responses in the placenta and umbilical cord. *Seminars in fetal & neonatal medicine.* 2006;11(5):296-301.
167. Park CW, Lee SM, Park JS, Jun JK, Romero R, Yoon BH. The antenatal identification of funisitis with a rapid MMP-8 bedside test. *Journal of perinatal medicine.* 2008;36(6):497-502.
168. Kim CJ, Romero R, Chaemsaitong P, Chaiyasit N, Yoon BH, Kim YM. Acute chorioamnionitis and funisitis: definition, pathologic features, and clinical significance. *American journal of obstetrics and gynecology.* 2015;213(4 Suppl):S29-52.
169. Redline RW. Classification of placental lesions. *American journal of obstetrics and gynecology.* 2015;213(4 Suppl):S21-8.
170. Romero R, Ceska M, Avila C, Mazor M, Behnke E, Lindley I. Neutrophil attractant/activating peptide-1/interleukin-8 in term and preterm parturition. *American journal of obstetrics and gynecology.* 1991;165(4 Pt 1):813-20.
171. Cherouny PH, Pankuch GA, Romero R, Botti JJ, Kuhn DC, Demers LM, et al. Neutrophil attractant/activating peptide-1/interleukin-8: association with histologic chorioamnionitis, preterm delivery, and bioactive amniotic fluid leukoattractants. *American journal of obstetrics and gynecology.* 1993;169(5):1299-303.
172. Ghezzi F, Gomez R, Romero R, Yoon BH, Edwin SS, David C, et al. Elevated interleukin-8 concentrations in amniotic fluid of mothers whose neonates subsequently develop bronchopulmonary dysplasia. *European journal of obstetrics, gynecology, and reproductive biology.* 1998;78(1):5-10.

173. Romero R, Chaemsaithong P, Korzeniewski SJ, Tarca AL, Bhatti G, Xu Z, et al. Clinical chorioamnionitis at term II: the intra-amniotic inflammatory response. *Journal of perinatal medicine*. 2016;44(1):5-22.
174. Zeilhofer HU, Schorr W. Role of interleukin-8 in neutrophil signaling. *Current opinion in hematology*. 2000;7(3):178-82.
175. Maurer M, von Stebut E. Macrophage inflammatory protein-1. *The international journal of biochemistry & cell biology*. 2004;36(10):1882-6.
176. Mukaida N. Interleukin-8: an expanding universe beyond neutrophil chemotaxis and activation. *International journal of hematology*. 2000;72(4):391-8.
177. Casey ML, Cox SM, Word RA, MacDonald PC. Cytokines and infection-induced preterm labour. *Reproduction, fertility, and development*. 1990;2(5):499-509.
178. Arntzen KJ, Kjollesdal AM, Halgunset J, Vatten L, Austgulen R. TNF, IL-1, IL-6, IL-8 and soluble TNF receptors in relation to chorioamnionitis and premature labor. *Journal of perinatal medicine*. 1998;26(1):17-26.
179. Hsu CD, Meaddough E, Aversa K, Copel JA. The role of amniotic fluid L-selectin, GRO-alpha, and interleukin-8 in the pathogenesis of intraamniotic infection. *American journal of obstetrics and gynecology*. 1998;178(3):428-32.
180. Hsu CD, Meaddough E, Aversa K, Hong SF, Lu LC, Jones DC, et al. Elevated amniotic fluid levels of leukemia inhibitory factor, interleukin 6, and interleukin 8 in intra-amniotic infection. *American journal of obstetrics and gynecology*. 1998;179(5):1267-70.
181. Figueroa R, Garry D, Elimian A, Patel K, Sehgal PB, Tejani N. Evaluation of amniotic fluid cytokines in preterm labor and intact membranes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2005;18(4):241-7.
182. Witt A, Berger A, Gruber CJ, Petricevic L, Apfalter P, Husslein P. IL-8 concentrations in maternal serum, amniotic fluid and cord blood in relation to different pathogens within the amniotic cavity. *Journal of perinatal medicine*. 2005;33(1):22-6.
183. Romero R, Chaemsaithong P, Korzeniewski SJ, Tarca AL, Bhatti G, Xu Z, et al. Clinical chorioamnionitis at term II: the intra-amniotic inflammatory response. *Journal of perinatal medicine*. 2015.

184. Sherry B, Tekamp-Olson P, Gallegos C, Bauer D, Davatelis G, Wolpe SD, et al. Resolution of the two components of macrophage inflammatory protein 1, and cloning and characterization of one of those components, macrophage inflammatory protein 1 beta. *The Journal of experimental medicine*. 1988;168(6):2251-9.
185. Wolpe SD, Davatelis G, Sherry B, Beutler B, Hesse DG, Nguyen HT, et al. Macrophages secrete a novel heparin-binding protein with inflammatory and neutrophil chemokinetic properties. *The Journal of experimental medicine*. 1988;167(2):570-81.
186. Saukkonen K, Sande S, Cioffe C, Wolpe S, Sherry B, Cerami A, et al. The role of cytokines in the generation of inflammation and tissue damage in experimental gram-positive meningitis. *The Journal of experimental medicine*. 1990;171(2):439-48.
187. Alam R, Forsythe PA, Stafford S, Lett-Brown MA, Grant JA. Macrophage inflammatory protein-1 alpha activates basophils and mast cells. *The Journal of experimental medicine*. 1992;176(3):781-6.
188. Christman JW, Blackwell TR, Cowan HB, Shepherd VL, Rinaldo JE. Endotoxin induces the expression of macrophage inflammatory protein 1 alpha mRNA by rat alveolar and bone marrow-derived macrophages. *American journal of respiratory cell and molecular biology*. 1992;7(4):455-61.
189. Rot A, Krieger M, Brunner T, Bischoff SC, Schall TJ, Dahinden CA. RANTES and macrophage inflammatory protein 1 alpha induce the migration and activation of normal human eosinophil granulocytes. *The Journal of experimental medicine*. 1992;176(6):1489-95.
190. Schall TJ, Bacon K, Camp RD, Kaspari JW, Goeddel DV. Human macrophage inflammatory protein alpha (MIP-1 alpha) and MIP-1 beta chemokines attract distinct populations of lymphocytes. *The Journal of experimental medicine*. 1993;177(6):1821-6.
191. Wang JM, Sherry B, Fivash MJ, Kelvin DJ, Oppenheim JJ. Human recombinant macrophage inflammatory protein-1 alpha and -beta and monocyte chemoattractant and activating factor utilize common and unique receptors on human monocytes. *J Immunol*. 1993;150(7):3022-9.
192. Dudley DJ, Hunter C, Mitchell MD, Varner MW. Elevations of amniotic fluid macrophage inflammatory protein-1 alpha concentrations in women during term and preterm labor. *Obstetrics and gynecology*. 1996;87(1):94-8.
193. Ren M, Guo Q, Guo L, Lenz M, Qian F, Koenen RR, et al. Polymerization of MIP-1 chemokine (CCL3 and CCL4) and clearance of MIP-1 by insulin-degrading enzyme. *The EMBO journal*. 2010;29(23):3952-66.

194. Romero R, Avila C, Santhanam U, Sehgal PB. Amniotic fluid interleukin 6 in preterm labor. Association with infection. *The Journal of clinical investigation*. 1990;85(5):1392-400.
195. Romero R, Sepulveda W, Kenney JS, Archer LE, Allison AC, Sehgal PB. Interleukin 6 determination in the detection of microbial invasion of the amniotic cavity. *Ciba Foundation symposium*. 1992;167:205-20; discussion 20-3.
196. Romero R, Yoon BH, Kenney JS, Gomez R, Allison AC, Sehgal PB. Amniotic fluid interleukin-6 determinations are of diagnostic and prognostic value in preterm labor. *Am J Reprod Immunol*. 1993;30(2-3):167-83.
197. Marconi C, de Andrade Ramos BR, Peracoli JC, Donders GG, da Silva MG. Amniotic fluid interleukin-1 beta and interleukin-6, but not interleukin-8 correlate with microbial invasion of the amniotic cavity in preterm labor. *Am J Reprod Immunol*. 2011;65(6):549-56.
198. Kacerovsky M, Drahosova M, Hornychova H, Pliskova L, Bolehovska R, Forstl M, et al. Value of amniotic fluid interleukin-8 for the prediction of histological chorioamnionitis in preterm premature rupture of membranes. *Neuro endocrinology letters*. 2009;30(6):733-8.
199. Cobo T, Kacerovsky M, Holst RM, Hougaard DM, Skogstrand K, Wennerholm UB, et al. Intra-amniotic inflammation predicts microbial invasion of the amniotic cavity but not spontaneous preterm delivery in preterm prelabor membrane rupture. *Acta obstetrica et gynecologica Scandinavica*. 2012;91(8):930-5.
200. Kacerovsky M, Musilova I, Khatibi A, Skogstrand K, Hougaard DM, Tambor V, et al. Intraamniotic inflammatory response to bacteria: analysis of multiple amniotic fluid proteins in women with preterm prelabor rupture of membranes. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2012;25(10):2014-9.
201. Kacerovsky M, Musilova I, Andrys C, Hornychova H, Pliskova L, Kostal M, et al. Prelabor rupture of membranes between 34 and 37 weeks: the intraamniotic inflammatory response and neonatal outcomes. *American journal of obstetrics and gynecology*. 2014;210(4):325 e1- e10.
202. Endres LK, Wang EY. Interleukin-6 and tumor necrosis factor alpha as predictors of success after emergent cerclage. *American journal of perinatology*. 2004;21(8):477-81.
203. Lee KY, Jun HA, Kim HB, Kang SW. Interleukin-6, but not relaxin, predicts outcome of rescue cerclage in women with cervical incompetence. *American journal of obstetrics and gynecology*. 2004;191(3):784-9.

204. Jung EY, Park KH, Lee SY, Ryu A, Oh KJ. Non-invasive prediction of intra-amniotic infection and/or inflammation in patients with cervical insufficiency or an asymptomatic short cervix (≤ 15 mm). *Archives of gynecology and obstetrics*. 2015;292(3):579-87.
205. Esplin MS, Romero R, Chaiworapongsa T, Kim YM, Edwin S, Gomez R, et al. Monocyte chemotactic protein-1 is increased in the amniotic fluid of women who deliver preterm in the presence or absence of intra-amniotic infection. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2005;17(6):365-73.
206. Polettini J, Cobo T, Kacerovsky M, Vinturache AE, Laudanski P, Peelen MJ, et al. Biomarkers of spontaneous preterm birth: a systematic review of studies using multiplex analysis. *Journal of perinatal medicine*. 2016.
207. Esplin MS, Romero R, Chaiworapongsa T, Kim YM, Edwin S, Gomez R, et al. Amniotic fluid levels of immunoreactive monocyte chemotactic protein-1 increase during term parturition. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2003;14(1):51-6.
208. Mittal P, Romero R, Tarca AL, Gonzalez J, Draghici S, Xu Y, et al. Characterization of the myometrial transcriptome and biological pathways of spontaneous human labor at term. *Journal of perinatal medicine*. 2010;38(6):617-43.
209. Proost P, Wuyts A, Van Damme J. Human monocyte chemotactic proteins-2 and -3: structural and functional comparison with MCP-1. *Journal of leukocyte biology*. 1996;59(1):67-74.
210. Gu L, Rutledge B, Fiorillo J, Ernst C, Grewal I, Flavell R, et al. In vivo properties of monocyte chemoattractant protein-1. *Journal of leukocyte biology*. 1997;62(5):577-80.
211. Wuyts A, Van Osselaer N, Haelens A, Samson I, Herdewijn P, Ben-Baruch A, et al. Characterization of synthetic human granulocyte chemotactic protein 2: usage of chemokine receptors CXCR1 and CXCR2 and in vivo inflammatory properties. *Biochemistry*. 1997;36(9):2716-23.
212. Muller WA. New mechanisms and pathways for monocyte recruitment. *The Journal of experimental medicine*. 2001;194(9):F47-51.
213. Schall TJ, Bacon K, Toy KJ, Goeddel DV. Selective attraction of monocytes and T lymphocytes of the memory phenotype by cytokine RANTES. *Nature*. 1990;347(6294):669-71.

214. Alam R, Stafford S, Forsythe P, Harrison R, Faubion D, Lett-Brown MA, et al. RANTES is a chemotactic and activating factor for human eosinophils. *J Immunol*. 1993;150(8 Pt 1):3442-8.
215. Adams DH, Lloyd AR. Chemokines: leucocyte recruitment and activation cytokines. *Lancet*. 1997;349(9050):490-5.
216. Campbell EM, Proudfoot AE, Yoshimura T, Allet B, Wells TN, White AM, et al. Recombinant guinea pig and human RANTES activate macrophages but not eosinophils in the guinea pig. *J Immunol*. 1997;159(3):1482-9.
217. Appay V, Rowland-Jones SL. RANTES: a versatile and controversial chemokine. *Trends in immunology*. 2001;22(2):83-7.
218. Athayde N, Romero R, Maymon E, Gomez R, Pacora P, Yoon BH, et al. Interleukin 16 in pregnancy, parturition, rupture of fetal membranes, and microbial invasion of the amniotic cavity. *American journal of obstetrics and gynecology*. 2000;182(1 Pt 1):135-41.
219. Pacora P, Maymon E, Gervasi MT, Gomez R, Edwin SS, Yoon BH, et al. Lactoferrin in intrauterine infection, human parturition, and rupture of fetal membranes. *American journal of obstetrics and gynecology*. 2000;183(4):904-10.
220. Hsu TY, Lin H, Lan KC, Ou CY, Tsai CC, Cheng BH, et al. High interleukin-16 concentrations in the early second trimester amniotic fluid: an independent predictive marker for preterm birth. *The journal of maternal-fetal & neonatal medicine : the official journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstet*. 2013;26(3):285-9.
221. Moroz LA, Simhan HN. Rate of sonographic cervical shortening and biologic pathways of spontaneous preterm birth. *American journal of obstetrics and gynecology*. 2014;210(6):555 e1-5.
222. Moreau JL, Chastagner P, Tanaka T, Miyasaka M, Kondo M, Sugamura K, et al. Control of the IL-2 responsiveness of B lymphocytes by IL-2 and IL-4. *J Immunol*. 1995;155(7):3401-8.
223. Morel BF, Burke MA, Kalagnanam J, McCarthy SA, Tweardy DJ, Morel PA. Making sense of the combined effect of interleukin-2 and interleukin-4 on lymphocytes using a mathematical model. *Bulletin of mathematical biology*. 1996;58(3):569-94.
224. Burke MA, Morel BF, Oriss TB, Bray J, McCarthy SA, Morel PA. Modeling the proliferative response of T cells to IL-2 and IL-4. *Cellular immunology*. 1997;178(1):42-52.

225. Palsson S, Hickling TP, Bradshaw-Pierce EL, Zager M, Jooss K, O'Brien PJ, et al. The development of a fully-integrated immune response model (FIRM) simulator of the immune response through integration of multiple subset models. *BMC systems biology*. 2013;7:95.
226. Gong C, Linderman JJ, Kirschner D. Harnessing the heterogeneity of T cell differentiation fate to fine-tune generation of effector and memory T cells. *Frontiers in immunology*. 2014;5:57.
227. Kwok CK, Ng PY. Network analysis approach for biology. *Cellular and molecular life sciences : CMLS*. 2007;64(14):1739-51.
228. Araujo RP, Liotta LA, Petricoin EF. Proteins, drug targets and the mechanisms they control: the simple truth about complex networks. *Nature reviews Drug discovery*. 2007;6(11):871-80.
229. Hopkins AL. Network pharmacology: the next paradigm in drug discovery. *Nature chemical biology*. 2008;4(11):682-90.
230. Broderick G, Fuite J, Kreitz A, Vernon SD, Klimas N, Fletcher MA. A formal analysis of cytokine networks in chronic fatigue syndrome. *Brain, behavior, and immunity*. 2010;24(7):1209-17.

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Table 1 Clinical and demographic characteristics of the study population

Variable	Overall cohort (N = 223)	Gestational age at delivery		p-value*
		≤32 weeks of gestation (N = 64)	>32 weeks of gestation (N = 159)	
Maternal age (years)	23.5 (20-28)	25 (21-32.8)	23 (20-27)	0.03
Pre-pregnancy body mass index (kg/m ²)**	25.8 (21.5-32.6)	29.1 (23.0-35.4)	24.7 (21.1-31.0)	0.004
Race, % (n)				0.64
African American	91.5% (204)	93.7% (60)	90.6% (144)	
Hispanic	3.1% (7)	3.1% (2)	3.1% (5)	
Caucasian	2.7% (6)	1.6% (1)	3.1% (5)	
Asian	0.9% (2)	1.6% (1)	0.6% (1)	
Other	1.8% (4)	0	2.5% (4)	
Gestational age at amniocentesis (weeks)	24.4 (21.1-27.4)	21.7 (19.9-24.5)	25.4 (22.9-28.4)	<0.001
Sonographic cervical length at diagnosis (mm)	13 (7-18)	7 (2.3-14)	14 (10-19)	<0.001
Diagnosis				
No intra-amniotic inflammation, % (n)	75.3% (168)	59.4% (38)	81.8% (130)	<0.001
Microbial invasion of the amniotic cavity, % (n)	12.1% (27)	9.4% (6)	13.2% (21)	0.43
Sterile intra-amniotic inflammation, % (n)	10.3% (23)	25.0% (16)	4.4% (7)	<0.001
Microbial-associated intra-amniotic inflammation, % (n)	2.2% (5)	6.3% (4)	0.6% (1)	0.01
Intra-amniotic inflammation (ELISA IL-6 ≥2.6 ng/mL), % (n)	12.6% (28)	31.3% (20)	5.0% (8)	<0.001
Treatment				
Cerclage, % (n/N)	8.1% (18/221)	9.4% (6/64)	7.6% (12/157)	0.67
Progesterone supplementation before amniocentesis, % (n/N)	1.4% (3/214)	0%	2.0% (3/152)	0.27
Administration of antenatal corticosteroids within 7 days before amniocentesis, % (n)	11.7% (26)	12.5% (8)	11.3% (18)	0.80
Antibiotics before amniocentesis, % (n)	5.8% (13)	6.3% (4)	5.7% (9)	0.87
Amniotic fluid				
Amniotic fluid IL-6 concentration	0.65 (0.29-1.28)	1.11 (0.31-4.15)	0.59 (0.28-0.99)	0.002

(ng/mL)				
Amniotic fluid glucose concentration (mg/dL) ^{***}	30 (25-35) (N=221)	31 (24.3-36) (N=64)	30 (25-35) (N=157)	0.52
Amniotic fluid white blood cell count (cells/mm ³) ^{****}	2 (0-7.5) (N=218)	2 (0-10) (N=63)	1 (0-6) (N=155)	0.06
Placenta				
Acute inflammatory lesion of placenta, % (n/N)	45.1% (92/204)	73.3% (44/60)	33.3% (48/144)	<0.001
Acute histologic chorioamnionitis, % (n/N)	44.1% (90/204)	73.3% (44/60)	31.9% (46/144)	<0.001
Acute funisitis, % (n/N)	32.4% (66/204)	51.7% (31/60)	24.3% (35/144)	<0.001
Delivery				
Amniocentesis to delivery interval (days)	68 (36-95)	23 (10-35)	85 (64-105)	<0.001
Gestational age at delivery (weeks)	35.7 (30.6-38.7)	26.6 (22.1-29.5)	38.0 (35.1-39.1)	<0.001
Delivery ≥37weeks of gestation, % (n)	44.4% (99)	0%	62.3% (99)	<0.001

Data are given as median (interquartile range) and percent (n)

*Comparison between patients with a diagnosis of short cervix who delivered before and after 32 weeks of gestation

** missing data 18 cases, *** missing data 2 cases, **** missing data 5 cases

ELISA, enzyme-linked immunosorbent assay; IL, Interleukin

Acute inflammatory lesion of placenta is defined as acute histologic chorioamnionitis and/or acute funisitis

Table 2: Changes in protein concentration with cervical shortening

Analyte	Overall change with CXL			Interaction between group and CXL		
	Fold change ¹	p	q	Fold change ²	p	q
MCP-1	-2.3	0.000	0.000	-2.4	0.003	0.098
MIP-1a	-2.7	0.000	0.000	-2.7	0.006	0.098
IL-6	-5.0	0.000	0.000	-4.6	0.009	0.098
MIP-1b	-3.7	0.000	0.000	-3.5	0.014	0.115
MIP-3a	-5.4	0.000	0.000	-2.7	0.135	0.735
HMGB-1	-5.0	0.000	0.000	2.0	0.177	0.735
I-TAC/CXCL11	-4.5	0.000	0.000	2.4	0.190	0.735
Gro-a/CXCL1	-2.5	0.000	0.000	-1.5	0.226	0.735
Calgranulin C	-2.1	0.041	0.043	-2.5	0.241	0.735
IL-8	-8.5	0.000	0.000	-4.0	0.241	0.735
IL-1a	-2.7	0.001	0.001	-2.0	0.273	0.735
IL-15	-4.3	0.000	0.000	1.9	0.292	0.735
Eotaxin	-4.2	0.000	0.000	1.7	0.299	0.735
IL-12	-4.4	0.000	0.000	1.9	0.330	0.735
RANTES	-2.3	0.004	0.005	1.8	0.334	0.735
IL-1b	-4.9	0.000	0.000	-1.6	0.392	0.808
IL-18	-1.4	0.004	0.005	1.2	0.444	0.862
IL-16	-2.5	0.000	0.000	1.2	0.535	0.897
IL-10	-4.0	0.001	0.002	-1.7	0.558	0.897
GM-CSF	-3.0	0.001	0.001	1.5	0.571	0.897
IFN-g	-3.7	0.000	0.000	1.3	0.658	0.971
IP-10	-1.4	0.042	0.043	1.1	0.742	0.971
M-CSF	-3.0	0.000	0.000	1.2	0.749	0.971
IL-2	-3.9	0.000	0.000	1.1	0.809	0.971
IL-33	-1.5	0.013	0.014	1.1	0.819	0.971
IL-7	-6.2	0.000	0.000	1.1	0.846	0.971

IL-4	-2.2	0.000	0.000	1.1	0.860	0.971
IL-13	-6.3	0.000	0.000	-1.1	0.920	0.971
TGF-b	-7.8	0.000	0.000	-1.1	0.940	0.971
TNF-a	-4.0	0.000	0.000	1.0	0.942	0.971
Calgranulin A	-4.8	0.000	0.000	1.1	0.949	0.971
Lactoferrin	-2.4	0.000	0.000	1.0	0.971	0.971

CXL= Cervix length;

¹Refers to the change in average protein concentration per cm increase in cervical length.

Minus sign denotes decrease in concentration with increasing cervical length (or increase with cervical shortening). Both groups of patients (delivery ≤ 32 and > 32 weeks) were included in this analysis.

²Refers to the ratio of change in average protein concentration per cm of cervical length in patients with delivery at ≤ 32 weeks vs those who delivered at > 32 weeks. Minus sign denotes that the decrease in protein concentration with cervical length (or increase in concentration with cervical shortening) is larger in patients who delivered at ≤ 32 weeks compared to those who delivered at > 32 weeks.

Table 3: Differential protein concentration analysis for different intervals of gestational age at amniocentesis

Gene	16.7-22.1 weeks			22.1-26.1 weeks			26.1-31.7 weeks		
	FC	p	q	FC	p	q	FC	p	q
Calgranulin A	<u>3.8</u>	0.0006	0.002	1.4	0.0382	0.097	-1.2	0.4503	0.803
Calgranulin C	3.3	0.0023	0.004	1.3	0.4633	0.493	1.4	0.4294	0.803
Eotaxin	<u>2.4</u>	0.0021	0.004	1.3	0.0937	0.155	1.1	0.5669	0.813
GM-CSF	<u>2.5</u>	0.0023	0.004	1.3	0.3937	0.433	2.2	0.0686	0.479
Gro-a/CXCL1	<u>2.2</u>	0.0003	0.001	1.6	0.0058	0.030	1.2	0.3670	0.803
HMGB-1	<u>3.0</u>	0.0008	0.002	1.8	0.0064	0.030	1.1	0.7594	0.878
I-TAC/CXCL-11	2.4	0.0062	0.009	1.4	0.0484	0.114	1.2	0.4836	0.803
IFN-g	<u>2.4</u>	0.0050	0.008	1.3	0.0942	0.155	1.1	0.7037	0.878
IL-10	<u>4.4</u>	0.0001	0.000	1.4	0.0650	0.134	1.1	0.6757	0.878
IL-12	<u>2.9</u>	0.0005	0.001	1.3	0.1875	0.229	1.2	0.5176	0.803
IL-13	1.9	0.0774	0.088	1.5	0.0884	0.155	1.1	0.7981	0.878
IL-15	1.8	0.0183	0.023	1.1	0.5204	0.537	1.4	0.1785	0.793
IL-16	<u>2.8</u>	0.0000	0.000	1.4	0.0361	0.097	-1.0	0.9394	0.969
IL-18	1.4	0.0231	0.028	1.3	0.0290	0.095	1.3	0.0870	0.479
IL-1a	<u>2.7</u>	0.0006	0.002	1.2	0.5498	0.550	2.1	0.0721	0.479
IL-1b	3.0	0.0034	0.006	1.9	0.0193	0.071	1.2	0.5224	0.803
IL-2	2.0	0.0144	0.020	1.3	0.1661	0.211	1.2	0.3397	0.803
IL-33	1.8	0.3545	0.354	1.8	0.1050	0.156	1.2	0.6707	0.878
IL-4	<u>1.9</u>	0.0033	0.006	1.3	0.0935	0.155	-1.2	0.2510	0.793
IL-6	<u>4.5</u>	0.0002	0.001	4.2	0.0000	0.000	1.0	0.9970	0.997
IL-7	<u>3.0</u>	0.0020	0.004	1.4	0.1122	0.156	1.2	0.4243	0.803
IL-8	<u>5.6</u>	0.0003	0.001	5.4	0.0000	0.000	2.3	0.0331	0.479
IP-10	1.4	0.0881	0.097	1.5	0.1180	0.156	1.2	0.4149	0.803
Lactoferrin	<u>2.8</u>	0.0000	0.000	1.4	0.0318	0.095	1.6	0.3441	0.803
M-CSF	<u>2.8</u>	0.0000	0.000	1.3	0.3386	0.385	1.4	0.2240	0.793
MCP-1	2.4	0.0000	0.000	2.0	0.0027	0.022	1.3	0.2644	0.793

MIG	1.4	0.2105	0.217	1.4	0.2975	0.351	-1.1	0.7902	0.878
MIP-1a	<u>2.9</u>	0.0000	0.000	1.9	0.0075	0.031	1.5	0.0756	0.479
MIP-1b	<u>5.1</u>	0.0000	0.000	2.6	0.0016	0.018	1.4	0.0860	0.479
MIP-3a	1.9	0.1479	0.157	1.9	0.0046	0.030	-1.3	0.2384	0.793
RANTES	1.9	0.0326	0.038	1.4	0.1172	0.156	-1.2	0.5353	0.803
TGF-b	2.5	0.0167	0.022	1.3	0.1060	0.156	-1.0	0.9385	0.969
TNF-a	2.4	0.0109	0.016	1.3	0.0592	0.130	1.1	0.7260	0.878

FC=Fold change. Negative values represent decrease in patients delivered ≤ 32 weeks compared to those delivered > 32 weeks. p=nominal p-values; q=False Discovery Rate adjusted p-values; Underlined fold changes are for proteins for which the differential abundance remains significant if only the patients with no intra-amniotic inflammation are used in the analysis.

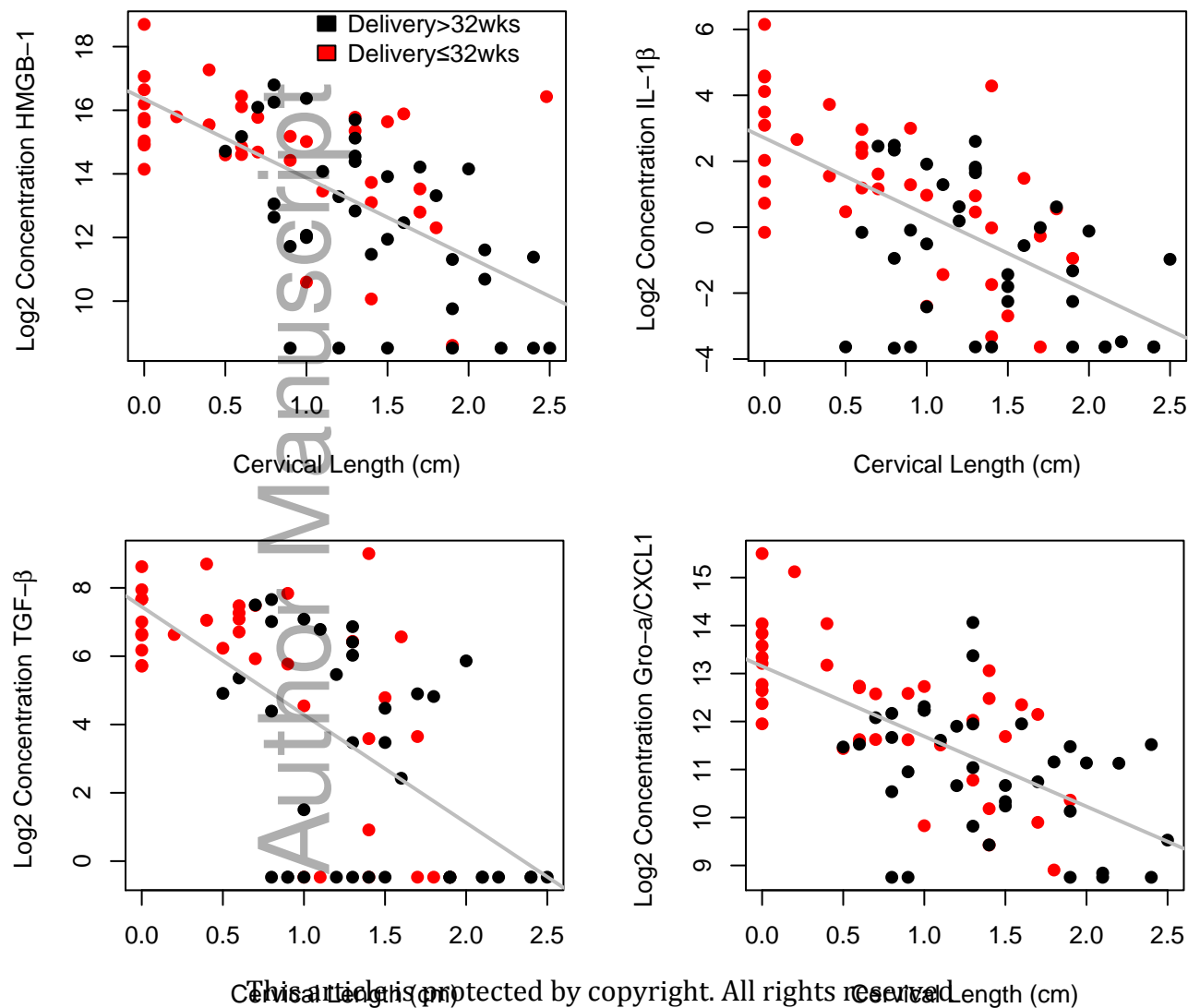
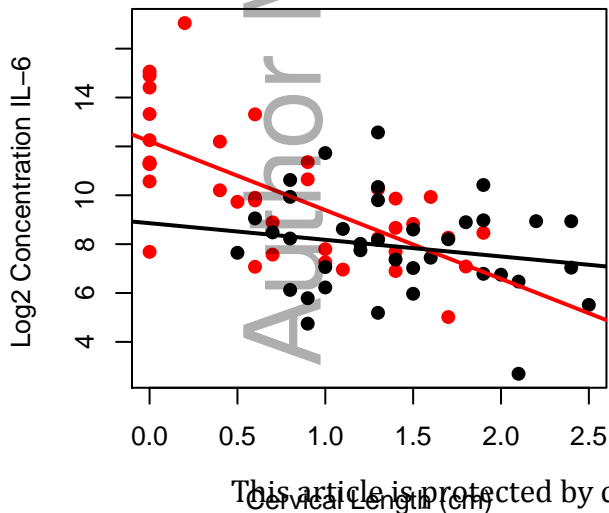
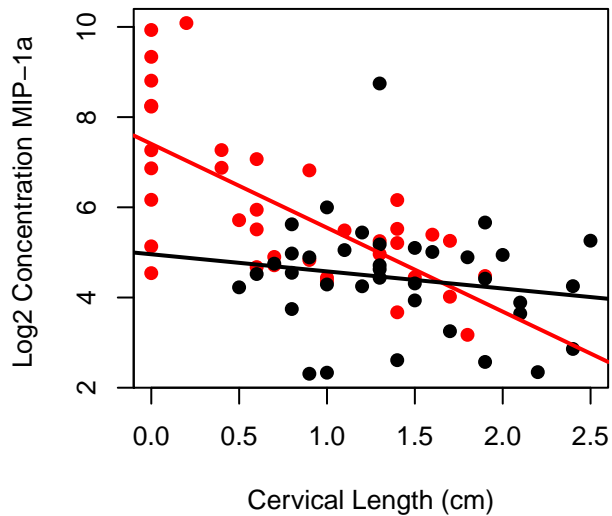
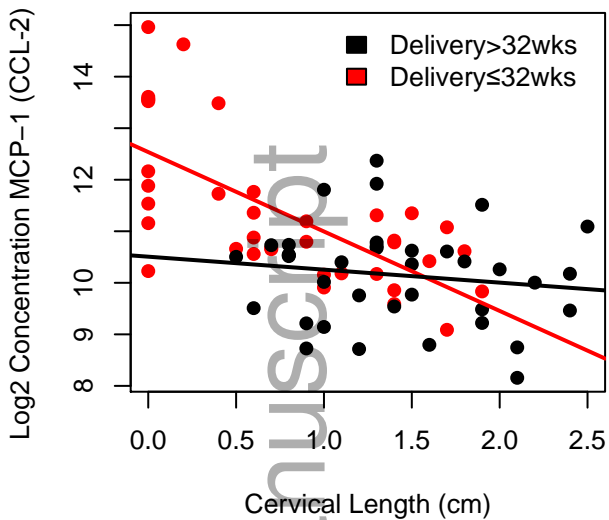


Figure 1



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Figure 2

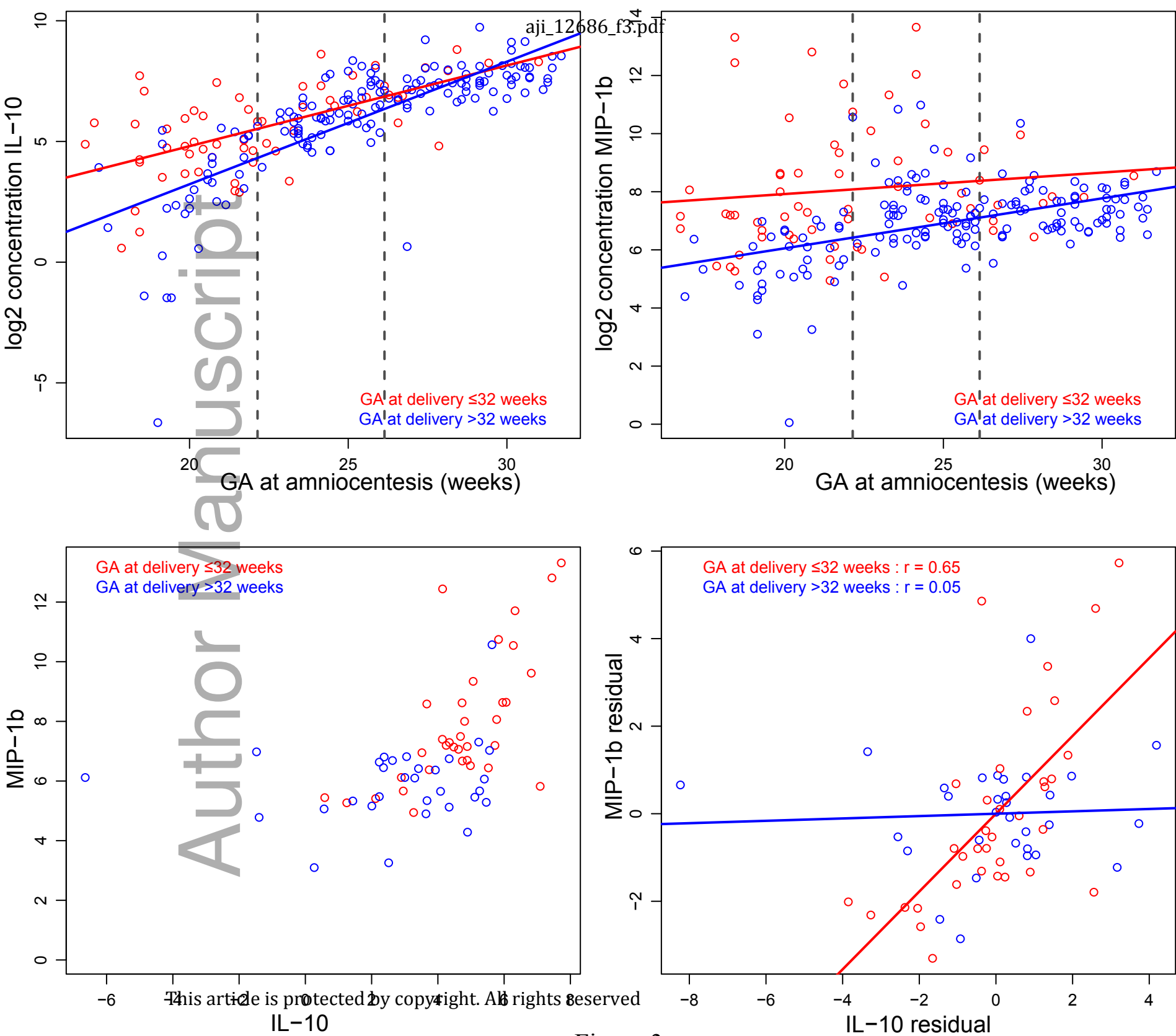
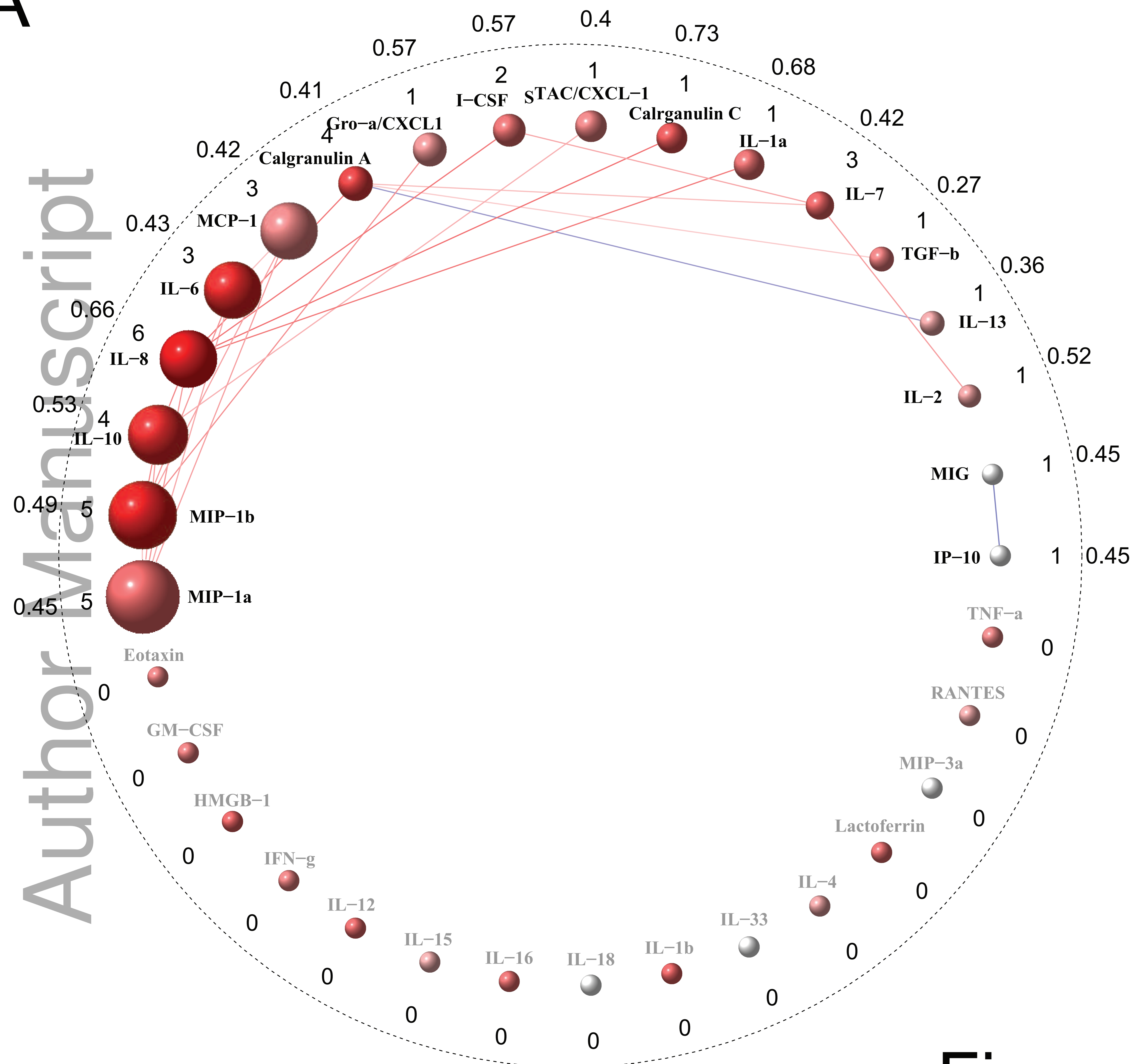


Figure 3

A



B

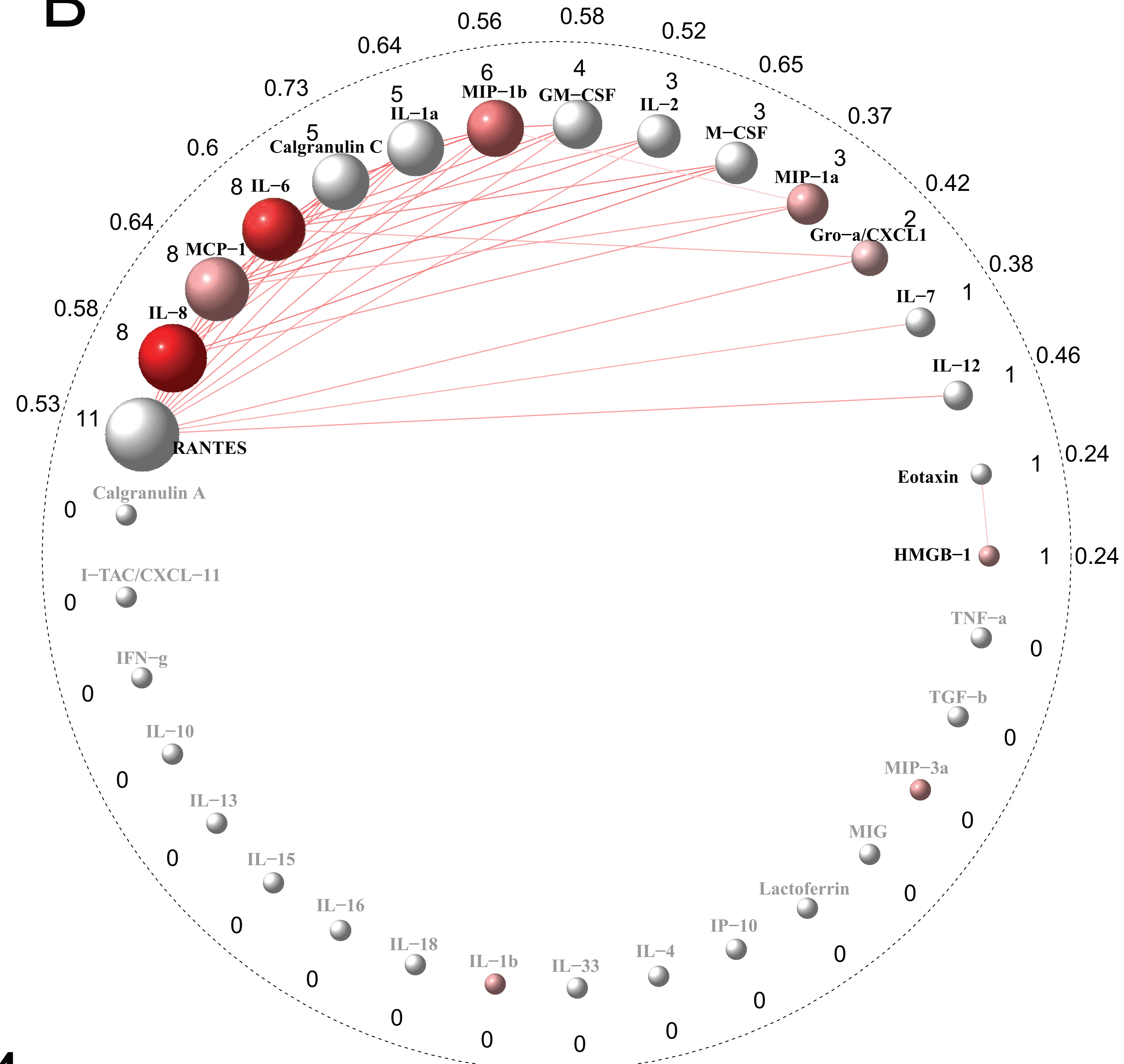


Figure 4