X-rays and virtual taphonomy resolve the first Cissus (Vitaceae) macrofossils from Africa as early-diverging members of the genus

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PREMISE OF THE STUDY: Fossilized seeds similar to Cissus (Vitaceae) have been recognized from the Miocene of Kenya, though some were previously assigned to the Menispermaceae. We undertook a comparative survey of extant African Cissus seeds to identify the fossils and consider their implications for the evolution and biogeography of Cissus and for African early Miocene paleoenvironments.

METHODS: Micro-computed tomography (μCT) and synchrotron-based X-ray tomographic microscopy (SRXTM) were used to study seed morphology and anatomy. Virtual taphonomy, using SRXTM data sets, produced digital fossils to elucidate seed taphonomy. Phylogenetic relationships within Cissus were reconstructed using existing and newly produced DNA sequences for African species. Paleobiology and paleoecology were inferred from African nearest living relatives.

KEY RESULTS: The fossils were assigned to four new Cissus species, related to four modern clades. The fossil plants were interpreted as climbers inhabiting a mosaic of riverine woodland and forest to more open habitats. Virtual taphonomy explained how complex mineral infill processes concealed key seed features, causing the previous taxonomic misidentification. Newly sampled African species, with seeds most similar to the fossils, belong to four clades within core Cissus, two of which are early diverging.

CONCLUSIONS: Virtual taphonomy, combined with X-ray imaging, has enabled recognition of the first fossil Cissus and Vitaceae from Africa. Early-divergent members of the core Cissus clade were present in Africa by at least the early Miocene, with an African origin suggested for the Cissus sciaphila clade. The fossils provide supporting evidence for mosaic paleoenvironments inhabited by early Miocene hominoids.

KEY WORDS: Cissus; Hiwegi Formation; liana; Menispermicarpum; microCT; Miocene; paleoecology; seeds; SRXTM; virtual taphonomy
2009), including *Ekembo* (previously *Proconsul*, see McNulty et al., 2015), which mark the transition between Paleogene arboreal primates, thought to inhabit tropical forests (Andrews, 1992; Janis, 1993), and Neogene bipedal hominids, often associated with open savanna grassland (Robinson, 1963; Reed, 1997; Pickford, 2002). Study of the Hiwegi Formation flora is essential to understand the paleoenvironments in which these transitional hominoids evolved.

The fruit and seed flora was partly described by Chesters (1957) from surface-picked collections, but these lacked a sedimentological and stratigraphic context. These issues of specimen provenance prompted in situ excavations at the new site of R117 (Collinson et al., 2009), where over 360 fruits and seeds were collected, including several specimens tentatively identified as cf. *Cissus* sp. 1 nov. (Vitaceae). During that study, three other morphotypes with similarity to seeds of extant *Cissus* L. species were recognized among the collections originally studied by Chesters (1957, 1958). If these four fossil records of *Cissus* can be verified, they would constitute the first records of the Vitaceae in the flora, the earliest reported record of Vitaceae from the African continent and could provide evidence for arid- or rainforest-adapted taxa in the African Miocene vegetation (De Santo et al., 1987; Verdcourt, 1993; Lombardi, 2000; Manchester et al., 2012a).

Recent molecular phylogenetic analyses of the genus *Cissus* (Liu et al., 2013; Rodrigues et al., 2014) showed that modern species could be assigned to several distinct clades. All African species fell within the core *Cissus* clade, but within that were distributed in more than seven distinct subclades, two of which also included Asian species (Liu et al., 2013). However, many African species were missing from these phylogenies, including those with external seed morphology most similar to the putative *Cissus* fossils. If the fossils are *Cissus*, molecular study of these neglected modern species will be essential to place them in their phylogenetic context.

This paper therefore aims to (1) confirm or refute the identification of Hiwegi Formation fossils to *Cissus* and, if confirmed, identify the clades to which the fossil species are likely related by comparing external and internal seed morphology of the fossils to extant African species; (2) place the nearest living relatives of the fossils into the existing phylogenetic framework; and (3) evaluate the fossils’ paleoenvironmental and biogeographic significance.

**MATERIALS AND METHODS**

**Specimens studied**—The fossil fruits and seeds from the Hiwegi Formation are composed of carbonate minerals, which derive from highly alkaline ash erupted from the nepheline-carbonatite Kisingiri volcano during the Miocene (Bestland et al., 1995; Harris and Van Couvering, 1995), and which replaced biological structures during fossilization (Collinson et al., 2009). The specimens were collected from the Hiwegi Formation (for wider stratigraphic context, see Drake et al., 1988; Collinson et al., 2009) by surface-picking and in situ excavation and are stored in collections at the Natural History Museum, London (NHMUK, specimen numbers prefixed V) and the National Museums of Kenya, Nairobi (KNM). Three fossil seed morphotypes in NHMUK, which Chesters (1957, 1958) had placed in the Menispermaceae (due to their bisymmetry, horseshoe-shaped curvature, and sculptured margins), were noted by M. E. Collinson (personal observations) to have strikingly similar exterior seed coat morphology to modern African *Cissus* species, as illustrated in African floras (e.g., Dewit and Willems, 1960; Descoings, 1967, 1972; Verdcourt, 1993). Collinson et al. (2009) also listed several specimens from the R117 site assigned to ‘cf. *Cissus* sp. 1 nov.’ (Vitaceae) based on similarity to modern *Cissus* seeds. All these fossils were re-examined during the current study.

Fifteen modern species of African *Cissus* and three species of African *Cyphostemma* (Planch.) Alston, which have comparable seed ornamentation to the putative *Cissus* fossils, were sampled from loose fruits on herbarium sheets in the Royal Botanic Gardens, Kew Herbarium (K) to study seed anatomy. The most visibly mature and undamaged specimens were selected. Additionally, fruits of *Cissus* *dasantha* were obtained from the herbarium at the Botanic Garden Meise, Belgium (BR). Herbarium sheet information for the species sampled is provided in Appendix S1 (see Supplemental Data with the online version of this article).

**Macrophotography and VP-SEM**—Photographs of the specimens described by Chesters (1957, 1958) were provided by the NHMUK Photographic Unit. The smaller specimens, assigned to ‘cf. *Cissus* sp. 1 nov.’ by Collinson et al. (2009), were examined uncoated under a Leo 1455 vapor pressure scanning electron microscope (VP-SEM) at the Imaging and Analysis Centre, NHMUK. Specimens were placed loose onto a sheet of black paper in a small tray, moved into appropriate orientation and turned over using a fine (size 00000) artist’s brush. A small amount of Blu-Tack (Bostik, Paris, France) was used, when unavoidable, to orient specimens for apical and basal views. Images were obtained using the back scatter detector (BSD), a chamber pressure of 14–15 Pa, current of 20 kV, spot size 500, and working distance 38–39 mm. Images were adjusted uniformly for contrast and brightness using Adobe (San Jose, California, USA) Photoshop CS2 or CS6.

**Synchrotron-based X-ray tomographic microscopy (SRXTM)**—The traditional method of boiling and scrubbing modern fruits to study their seeds is problematic. Depending on tissue toughness, seed features may not be revealed in a repeatable or comparable manner across different species or genera. Cutting or histological sectioning to study internal anatomy also has limitations: it is destructive, may introduce artifacts (tears, gaps), and multiple planes of section through the same specimen cannot be acquired. X-ray imaging solves these problems because multiple planes of section through a single specimen can be easily and nondestructively obtained (e.g., Smith et al., 2009). Modern *Cissus* and *Cyphostemma* fruits were scanned using SRXTM, as this technique provides the necessary quality of resolution to enable distinction of cellular details in the fruit wall and seed coat layers for systematic study and for virtual taphonomy (Smith et al., 2009; Collinson et al., 2013). The SRXTM was performed on the TOMCAT beamline at the Swiss Light Source, Paul Scherrer Institut, Villigen, Switzerland (Stampanoni et al., 2006). Specimens were mounted onto brass pin stubs using polyvinyl acetate glue and were scanned during one session of beamtime in July 2014. X-rays transmitted by the specimens were converted into visible light by a 300 μm-thick Ce-doped LAG scintillator screen. A microscope objective of 1.25x or 2x (depending on fruit size) magnified the projection data, which were then digitized by a high-resolution scientific CMOS camera (PCO.edge; PCO GmbH, Kelheim, Germany), giving a resultant voxel size of 3 to 5 μm. The energy was set at 17.5 keV, and the exposure time per projection was 50 ms. For each scan, 1501 projections (2560 × 2160 pixels with PCO.edge camera) were acquired over 180°. Reconstruction algorithms were then used to combine the projections...
and obtain a three-dimensional volume, reconstruction was performed on a dedicated Linux PC cluster using a highly optimized routine based on the Fourier transform method and a gridding procedure (Marone et al., 2010; Marone and Stampanoni, 2012). Multiple stacked scans were used if the specimens did not completely fit within the field of view. Three-dimensional data sets were visualized, and images and videos were captured, in the program Avizo 8.1 (FEI Visualization Science Group, Bordeaux, France). Images were adjusted uniformly for contrast and brightness using Adobe Photoshop CS2 or CS6. Videos of digital SRXTM tomograms in transverse section (DTS) through fruits of each of these modern species are available from the Dryad Digital Repository (http://dx.doi.org/10.5061/dryad.g9r36).

**Micro-computed tomography (μCT)**—Externally visible ventral in- folds are a characteristic feature of Cissus seeds (Chen and Manchester, 2011), but are not evident in the fossils (Fig. 1). Information on internal structure of the fossils (including holotypes) is required to test whether these characteristic ventral in folds are (1) genuinely absent, which would exclude affinity with Vitaceae; (2) present but externally obscured by a seed coat layer, which would indicate affinity with Cyphostemma (Chen and Manchester, 2011); or (3) obscured as a consequence of taphonomic processes, such as mineral infilling during fossilization, which would support identification to Cissus. For holotypes and rare fossils, this information must be obtained nondestructively. Therefore, three fossil specimens (V33753, V68501, V68506) from collections studied by Chesters (1957, 1958) and stored in NHMUK and two specimens (R117.1981.314, R117.1981.476) from the R117 site, identified by Collinson et al. (2009) as ‘cf. Cissus sp. 1. nov.’ and housed in KNM were scanned by μCT using a Nikon Metrology HMX ST 225 at the Imaging and Analysis Centre, NHMUK. Specimens were stabilized by inserting them into blocks of OASIS Floral Foam (Smithers-Oasis Company, Kent, Ohio, USA) within a plastic tube. Specimens were wrapped for protection in cling film: a thin film of PVC (polyvinyl chloride) or LDPE (low density polyethylene). A voltage of 200 kV was used with a current of 180 μA, a tungsten reflection target, a 0.5 or 0.25 mm copper filter and an exposure time of 708 ms, resulting in a voxel size of 12 μm. Four modern Cissus fruits scanned by SRXTM (one C. dinklagei, one C. populnea, two C. integrifolia) were also scanned using μCT. This duplicative scanning aimed to ensure that μCT scans of the fossils could be interpreted in the context of directly comparable scans of modern seeds (a comparison of imaging methods is provided in Appendix S2 with the online Supplemental Data). A voltage of 125 kV was used with a current of 200 μA, a molybdenum reflection target, no filter and an exposure time of 708 ms, resulting in a voxel size of 8 to 15 μm. μCT data sets were reconstructed using CT Pro (Nikon Metrology, Tring, UK) and were visualized in Avizo 8.1. Images and videos were obtained as for SRXTM data sets. Videos are available from the Dryad Digital Repository (http://dx.doi.org/10.5061/dryad.g9r36).

**Virtual taphonomy**—The technique of virtual taphonomy, developed by Smith et al. (2009), solves the problem of potential variability in tissue removal with traditional boiling or scrubbing methods in seed preparation. Virtual taphonomy uses X-ray data sets to digitally remove specific tissue or cell layers from modern fruits or seeds, thereby creating digital fossils, the surfaces of which can be directly compared with real fossils to determine which layers are preserved. Digital seed infills can also be produced using this technique, mimicking the mineral infill of fruits and seeds that can occur during fossilization (Smith et al., 2009; Collinson et al., 2013). For modern *Cissus populnea* Guill. & M.Brandt, a digital infill of the space inside the inner seed coat (the endotesta) was produced (a virtual fossil), to mimic mineral infill during fossilization. The ventral infolds were then digitally infilled to mimic processes that might have led to mineral obscuring the ventral infolds during fossilization.

**Molecular phylogeny**—Phylogenetic relationships within *Cissus* and the placement of species assigned to this genus within Vitaceae were assessed using available sequence data from the plastid genome (trnL intron, trnL-F spacer, atpB-rbcL spacer, trnC-petN spacer and rps16 intron), as well as newly produced sequences of trnL-F and rps16 for eight African Cissus species with seeds most similar to the putative *Cissus* fossils, which were not represented in previous molecular phylogenetic studies (see Appendix 1). Sequence data obtained from public repositories comprise 91 *Cissus* species and 92 species from other genera of Vitaceae (see Appendix 2).

Total genomic DNA was extracted using a standard CTAB-based protocol (Doyle and Doyle, 1987) and purified using a combined cesium chloride/ethidium bromide gradient and dialysis procedure. The trnL intron/trnL-F spacer and the rps16 intron were amplified using the primers designed by Taberlet et al. (1991) and Shaw et al. (2005), respectively. Further details regarding the polymerase chain reactions, amplification procedures, PCR product purifications, and cycle sequencing reactions are provided in online Appendix S3.

Matrices (including sequences obtained from public repositories and those produced for the current study) were aligned using MUSCLE (Edgar, 2004) in the program Geneious; alignments are available from the TreeBASE depository (https://treebase.org; study ID 18491). A phylogenetic analysis was performed on a combined matrix using the maximum likelihood criterion as implemented in the program RAxML v8.1.24 (Stamatakis, 2014) using the rapid bootstrap algorithm with 1000 replicates and a search for the best-scoring tree. Divergence time estimates were obtained using the Bayesian inference approach implemented in the package BEAST v.1.8.2 (Drummond and Rambaut, 2007). All analyses were run on the Cipres Science Gateway portal (www.phylo.org). Further details of the phylogenetic analyses are provided in Appendix S3.

Calibration was performed using three fossils. The first is the oldest known fossil securely identified to the Vitaceae family (*Indovitis*) from the latest Cretaceous/earliest Paleogene Deccan traps of India (ca. 66 Ma, based on radiometric dating and biostratigraphy; Manchester et al., 2013) and was used as calibration on the crown node of subfamily Vitioideae (calibration A), comprising all genera of Vitaceae except the genus *Leea*, which is assigned to subfamily Lleoideae. A log-normal distribution was used, which allows the age to vary (given the uncertainty in fossil age estimation and given that a fossil’s age is considered a minimum age for a given group), with an offset value of 65 and a standard deviation of 1.0. The second calibration point comes from fossil seeds assigned to *Ampe- locissus parvisepima* Chen & Manchester from the late Paleocene of North America at the Beicegel Creek locality of the Sentinel Butte Formation, Fort Union Group, North Dakota (Chen and Manchester, 2007), considered by Zetter et al. (2011) to be late Paleocene (61.7–56.8 Ma) in age based on molluscan and mammalian (Kihm and Hartman, 1991; Hartman and Kihm, 1995) biostratigraphy and pollen zonation (Nichols and Ott, 1978). It was assigned to the stem node of the clade comprising genera *Ampe locissus* Planch., *Nothocissus* (Miq.) Latiff, *Parthenocissus* Planch., *Pterisanthes* Blume, *Vitis*...
TABLE 1. Seed morphotypes of selected single-seeded, modern African Cissus species based on internal and external morphology obtained from synchrotron X-ray data sets and descriptions and illustrations in floras (Dewit and Willems, 1960; Descoings, 1972; Verdcourt, 1993). H = height, W = width.

<table>
<thead>
<tr>
<th>Seed morphotype</th>
<th>Species</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 (Figs. 6D, 6E, 7A)</td>
<td>C. integrifolia, C. populnea</td>
<td>Two-layered, thick fruit wall, with a denser outer layer and more porous inner layer</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seeds laterally flattened (height/width [H/W] ratio of 1.5–1.8 in median digital transverse section [DTS]), elongate in the dorsiventral dimension</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Obvious break in seed coat near the chalaza</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Outer endotesta consists of thin layer with different X-ray attenuation (possibly high mineral content)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Dense and thin seed coat</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seed surface with two marginal ridges, one on each lateral face, a ridged and faceted marginal area, long ridges radiating from ventral margin across lateral faces</td>
</tr>
<tr>
<td>2 (Figs. 6B, 7B)</td>
<td>C. barbeyana, C. dasyantha</td>
<td>Thin to moderately thick fruit wall</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seeds slightly laterally flattened (H/W ratio of 1.3–1.5 in median DTS), elongate in the dorsiventral dimension</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Indistinct break in seed coat near the chalaza</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Endotesta has uniform X-ray attenuation</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Dense and moderately thick seed coat</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seed surface with two marginal ridges; a ridged and faceted marginal area, where ridges sometimes fuse to form a reticulum; short ridges radiating across part of the lateral faces</td>
</tr>
<tr>
<td>3 (Figs. 6H, 7C)</td>
<td>C. lebrunii, C. sciaphila, C. tiliifolia</td>
<td>Thin fruit wall</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seeds not, or slightly, laterally flattened (H/W ratio of 1.0–1.6 in median DTS), short in the dorsiventral dimension</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Indistinct break in seed coat near the chalaza</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Endotesta has uniform X-ray attenuation</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Thin seed coat of variable texture</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seed surface covered in ridges, forming a complete to incomplete reticulum across the lateral faces</td>
</tr>
<tr>
<td>4 (Figs. 6M, 7D)</td>
<td>C. petiolata</td>
<td>Thick fruit wall</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seeds laterally flattened (H/W ratio of 1.5–1.8 in median DTS), short in the dorsiventral dimension</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Thickened chalaza with no break in seed coat</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Endotesta has uniform X-ray attenuation</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Dense and thick seed coat</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Seed surface smooth</td>
</tr>
</tbody>
</table>

L., and Yua C.L.Li, following Nie et al. (2012) and Liu et al. (2016) (calibration B). As for the previous calibration, a log-normal distribution was used, with an offset value of 55.8 and a standard deviation of 1.0. The third calibration point is the oldest fossil unequivocally assigned to genus Cissus, from the Belén flora (North Coastal Peru) of the Oligocene, with a maximum age of 30–28.5 Ma based on diatom biostratigraphy (Manchester et al., 2012a). This fossil has features shared with species from morphotype 1 and the Cissus integrifolia clade (see below); it was therefore assigned to the stem node of this group (calibration C) with a log-normal distribution with an offset value of 27.5 and a standard deviation of 1.0.

Extant Cissus seed morphological information—In addition to the 16 SRXTM data sets of African Cissus seeds most similar to the Hiwegi Formation fossils, published seed illustrations and descriptions were sourced, where available, for all African Cissus species included in the molecular phylogeny (Table 1; online Appendix S4). Together, these data were used to determine the modern species with seeds most similar to those of the fossils and to place these nearest living relatives into the existing phylogenetic framework.

Ecological and biogeographic information—Data on the habit, habitat, and biogeographic distributions of extant Cissus species were gathered for extant species in clades containing similar seed morphotypes to the fossils (Table 2). Georeferenced occurrence data from illustrated herbarium sheets were obtained from the Global Biodiversity Information Facility (GBIF, 2013), and floras and other herbarium sheets were also used. Herbarium sheet sources are given in online Appendix S5.

FIGURE 1 External morphology of fossilized Cissus seeds from the Hiwegi Formation, Rusinga Island, Kenya. (A) Lateral, (B) apical, (C) basal, (D) ventral views of the Cissus crenulata (Chesters) comb. nov. holotype (V33753). (E) Ventral, (F) lateral, (G) apical, (H) basal views of the Cissus andrewsii sp. nov. holotype. (I) Lateral and (J) basal views of a paratype of C. andrewsii (V68500), and (K) lateral and (L) apical views of another paratype (V68502), demonstrating intraspecific variation in seed size, shape, ornamentation and basal/apical width. (M) Lateral, (N) basal and (O) apical views of the Cissus rusingensis sp. nov. holotype (R117.1981.314). (P, S, V) Lateral, (Q, T, W) basal and (R, U, X) apical views of paratypes of C. rusingensis (R117.1981.476, R117.1981.604, R117.1981.605 respectively), illustrating intraspecific variation in seed shape (subrounded to pyriform); the number of muri and enclosed lumina, the extent of the perichalazal rib and the length of the basal projection. (Y) Lateral, (Z) apical, (AA) basal, (BB) ventral views of the Cissus psilata sp. nov. holotype (V68506). Scale bar = 10 mm, in 1 mm increments.
TABLE 2. Distribution, habit, and habitat data of modern Cissus species.

<table>
<thead>
<tr>
<th>Species</th>
<th>Distribution</th>
<th>Habit</th>
<th>Habitat</th>
<th>Sources</th>
</tr>
</thead>
<tbody>
<tr>
<td>C. adnata Roxb.</td>
<td>Australasia; E, SE, and S Asia</td>
<td>Scrambling or climbing shrub, or woody liana, up to 10 m</td>
<td>Primary lowland monsoon forests, riparian forest, disturbed semiopen scrub and deciduous forest, shrubland and thickets</td>
<td>1–4</td>
</tr>
<tr>
<td>C. albiporcula Masinde &amp; L.E.Newton</td>
<td>E Africa</td>
<td>Climber</td>
<td>Bushland in rocky areas</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. aphyllantha Gilg</td>
<td>E Africa</td>
<td>Shrub, scrambler or woody climber, 1–4 m tall</td>
<td>Acacia scrub or desert thornbush, scrubby woodland, rocky outcrops</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. aralioides (Welw. ex Baker) Planch.</td>
<td>C, E and W Africa</td>
<td>Vigorous, succulent liana or herbaceous climber, a strong, lofty climber, up to 25 m</td>
<td>Coastal and riverine evergreen forest, rainforest, coastal bushland, Acacia bushland, grassland, thickets</td>
<td>3, 5–9</td>
</tr>
<tr>
<td>C. barboyana De Wild. &amp; T.Durand</td>
<td>C and W Africa</td>
<td>Herbaceous to woody, low, small liana</td>
<td>Rainforest, forest clearings, forest fragments and clusters</td>
<td>3, 6, 7, 9</td>
</tr>
<tr>
<td>C. cachiformis Gilg</td>
<td>E and S Africa</td>
<td>Succulent climber or scrambler, 1.2–4.5 m long</td>
<td>Woodland, mixed bushland, usually in stony places, rock domes</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. dasyantha Gilg &amp; M.Brandt</td>
<td>W Africa</td>
<td>Liana, up to 6 m</td>
<td>Occasionally flooded forest, gallery forests</td>
<td>3, 7</td>
</tr>
<tr>
<td>C. faucicola Wild &amp; R.B.Drumm.</td>
<td>E Africa</td>
<td>Herbaceous climber, several meters long</td>
<td>Evergreen rainforest, especially edges by waterfalls and in grassy clearings</td>
<td>5, 8</td>
</tr>
<tr>
<td>C. flornbunda (Baker) Planch.</td>
<td>E Africa</td>
<td>Thin, woody liana or climber, several meters long</td>
<td>Rainforest, dense moist tropical forest, disturbed forest or forest edge</td>
<td>4, 10</td>
</tr>
<tr>
<td>C. integroida (Baker) Planch.</td>
<td>C and E Africa</td>
<td>Herbaceous vigorous climber to 5 m, reaching tops of trees or thicket-forming</td>
<td>Evergreen riverine forest, woodland, bushland and savanna grassland, often on rocky scarps</td>
<td>3, 5, 7, 8, 11</td>
</tr>
<tr>
<td>C. lebrunii Dewit</td>
<td>C Africa</td>
<td>Herbaceous climber</td>
<td>Rainforest</td>
<td>3, 7</td>
</tr>
<tr>
<td>C. oliver (Engl.) Gilg ex Engl.</td>
<td>C and E Africa</td>
<td>Herbaceous to woody climbing shrub, up to 6 m long</td>
<td>Riverine and gallery forest, marshy areas, papyrus swamps, wet grassland with scattered trees</td>
<td>3, 5, 7</td>
</tr>
<tr>
<td>C. oorephiila Gilg &amp; M.Brandt</td>
<td>C and W Africa</td>
<td>Large herbaceous liana</td>
<td>Gallery and riverine forest, forest edges, swamp areas</td>
<td>3, 4, 6, 9</td>
</tr>
<tr>
<td>C. petiolaris Hook.f.</td>
<td>C, E and W Africa</td>
<td>Large, somewhat succulent liana, vigorous climber or scrambler, to at least 10 m</td>
<td>Riverine forest, occasionally flooded forest edges, thickets, Acacia mixed bushland, rocky ground with scattered trees and shrubs, woody/shrub savanna</td>
<td>3, 5–9, 11</td>
</tr>
<tr>
<td>C. phyomatocarpa Masinde &amp; L.E.Newton</td>
<td>E Africa</td>
<td>Climber</td>
<td>Thickets on forest edges and coastal bushland</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. polita Desc.</td>
<td>E Africa</td>
<td>Herbaceous liana, 1.5–3 m</td>
<td>Calcareous hills and plateaux, dry, deciduous seasonal forest, savanna grassland with dry forest</td>
<td>4, 10</td>
</tr>
<tr>
<td>C. polyantha Gilg &amp; M.Brandt</td>
<td>C, E and W Africa</td>
<td>Herbaceous to woody climber/liana, up to 15 m</td>
<td>Riverine and gallery rainforest, rocky hollows in grassland and thickets</td>
<td>3, 5–7, 9</td>
</tr>
<tr>
<td>C. populnea Guill. &amp; Perr.</td>
<td>C, E and W Africa</td>
<td>Bushy liana, up to 4.5 m</td>
<td>Wooded savanna, rocky outcrops and scree, bushland, lowland forest edges</td>
<td>3, 5–7, 9</td>
</tr>
<tr>
<td>C. pseudoguineensis Verdc.</td>
<td>E Africa</td>
<td>Spreading herb, at least 60 cm long</td>
<td>Woodland, low shrubs on sand, swampy places</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. quadrangularis L.</td>
<td>Arabia; C, E, N, S, and W Africa; SE and S Asia</td>
<td>Succulent bushy liana 1–15 m long, or succulent climbing shrub</td>
<td>Xerophitic thickets, thorny savanna, Acacia woodland, grassland, riverine thicket, coastal forest edges</td>
<td>3, 5, 8–10</td>
</tr>
<tr>
<td>C. quamet Dewit</td>
<td>C and E Africa</td>
<td>Erect herb or herbaceous climber, 0.6–1 m tall</td>
<td>Riverine vegetation, Brachystegia woodland</td>
<td>3, 5, 7, 8</td>
</tr>
<tr>
<td>C. rhodothica (Baker) Desc.</td>
<td>E Africa</td>
<td>Scrambling and climbing strong liana or erect shrub</td>
<td>Rocky outcrops, deciduous seasonal forest, wooded savanna</td>
<td>10</td>
</tr>
<tr>
<td>C. rondoensis Verdc.</td>
<td>E Africa</td>
<td>Herbaceous to semiwoody climber to around 4 m</td>
<td>Moist, (semi) evergreen forest, dense forest thicket</td>
<td>3–5</td>
</tr>
<tr>
<td>C. rostrata (Miq.) Korth. ex Planch.</td>
<td>SE Asia</td>
<td>Climber/liana, up to 10–15 m</td>
<td>Fringe and understorey forest, forest river banks, peat swamp/ marshy forest</td>
<td>3, 4, 12</td>
</tr>
<tr>
<td>C. rotundifolia Vahl</td>
<td>Arabia; C, E and S Africa</td>
<td>Succulent, herbaceous to woody, vigorous climber/liana, to 5 m</td>
<td>Dry woodland and bush, thorny savanna, Acacia scrub, bushland, thickets, dry forest and forest edges particularly on rocky outcrops</td>
<td>3, 5, 7, 8</td>
</tr>
<tr>
<td>C. sagittifera Desc.</td>
<td>E Africa</td>
<td>Creeping or climbing, thin liana</td>
<td>Woodland edges, limestone cliffs, quartzite outcrops</td>
<td>3, 4, 10</td>
</tr>
<tr>
<td>C. scaphila Gilg</td>
<td>E Africa</td>
<td>Woody climber/liana, 3–12 m long or shrubbery</td>
<td>Lowland riverine forest fringes, woodland slopes above river valleys</td>
<td>3, 5, 8</td>
</tr>
<tr>
<td>C. smithiana (Baker) Planch.</td>
<td>C and W Africa</td>
<td>Large liana/climber</td>
<td>Rainforest, forest galleries and edges</td>
<td>3, 7, 9</td>
</tr>
<tr>
<td>C. sylvicola Masinde &amp; L.E.Newton</td>
<td>E Africa</td>
<td>Herbaceous, somewhat succulent and fl leshy liana, to 12 m</td>
<td>Evergreen forest, also forest on rocky hills, coralline limestone and thicket</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. tiliifolia Planch.</td>
<td>C and E Africa</td>
<td>Herbaceous to woody climber, up to 10 m</td>
<td>Forest and thickets in swampy areas, particularly near lake shores, swampy grassland</td>
<td>3, 5</td>
</tr>
<tr>
<td>C. welwitschii (Baker) Planch.</td>
<td>C and E Africa</td>
<td>Vigorous, woody climber, 2–9 m long, or shrubbery</td>
<td>Semievergreen bushland, riverine fringes, thickets and termite mounds in Brachystegia woodland, rocky outcrops</td>
<td>3, 5, 8</td>
</tr>
</tbody>
</table>

Sources: (1) Lu (1993); (2) Chen et al. (2007); (3) GBIF (2013); (4) herbarium sheet data (online Appendix S5); (5) Verdcourt (1993); (6) Keay (1958); (7) Dewit and Willems (1960); (8) Wild and Drummond (1966); (9) Descoings (1972); (10) Descoings (1967); (11) Beentje (1994); (12) Yeo et al. (2012).
SYSTEMATICS

Definitions of lateral, ventral, dorsal, apical, and basal views and seed height, width, and dorsiventral dimension are given in online Appendix S6.

Family—Vitaceae Juss. 1789.

Genus—Cissus L. 1753.

Species—Cissus crenulata (Chesters) Adams, Collinson, S.Y.Smith & Bamford comb. nov.

Basionym—Menispermicarpum crenulatum Chesters (1957, pl. 19, figs. 19, 20).

Emended diagnosis—Seed bilaterally symmetrical, 19 mm in dorsoventral dimension, laterally flattened, suboval to D-shaped in lateral view, narrowly elliptical and 7 mm wide in apical and basal views, and elliptical in ventral view. Center of lateral face crossed by four pronounced, long ridges radiating from adjacent to the ventral infolds; longest ridge almost as long as seed; curved ridge, 2–3 mm from dorsal and basal margins of lateral faces, delineates faceted marginal area; prominent median ridge (rib perichalaza) extends from beneath ventral infolds, around base, over dorsal margin, and almost full length of apical margin as far as ventral infolds. Upper portion of ventral surface concave, forming acute angle with long axis of seed; pair of very deep, narrow ventral infolds present.

Holotype—Seed: V33753 (Fig. 1A–D; μCT DTS video available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36).

Excluded specimens—Seeds: KNMP-RU7787 (8 specimens with field number 60’52, formerly P. B. 8, designated as paratypes of Menispermicarpum crenulatum by Chesters, 1957).

Type locality—Rusinga Island, Lake Victoria, Kenya.

Geological horizon and age—Hiweger Formation, early Miocene.

Repository—Natural History Museum, London, UK.

Description—The seed is bilaterally symmetrical around a prominent median perichalazal rib (Fig. 1B, C), laterally flattened with a height/width ratio of 1.7 (Fig. 1A, B), suboval to D-shaped in lateral view (Fig. 1A), 19 mm in the dorsiventral dimension and 11 mm in height, narrowly elliptical and 7 mm wide in apical (Fig. 1B) and basal (Fig. 1C) views, and elliptical in ventral view (Fig. 1D). The seed apex is rounded, with no indication of an apical notch or chalazal grooves (Fig. 1A–C), and the seed narrows to the ventral margin (Fig. 1A–C). The seed ornamentation is most clearly visible on one lateral face (Fig. 1A). A curved ridge, 2–3 mm from the basal and dorsal margins of the lateral faces, delineates a faceted outer margin with facets spaced at 2.5 to 4.5 mm (Fig. 1A). There are short ridges (ca. 2 mm long), roughly perpendicular to the long curved ridge, within the outer margin, which define the facets (Fig. 1A). There are at least four ridges crossing the center of the lateral face, with one long (10 mm) ridge perpendicular to the ventral surface, and three curved ridges, which radiate away from the ventral surface toward the basal margin, butting the curved ridge (Fig. 1A). The reverse lateral face is partly obscured by mineral encrustation in the holotype (the only specimen), making the ornamentation less clear. Nevertheless, at least three clearly distinguishable ridges cross the center of the face radiating from the ventral surface, with one longer and more pronounced than the others. In lateral view, the upper portion of the ventral surface is nearly straight for three-quarters of its length, but the basalmost part is indented, forming an angle of ca. 65° with the long axis of the seed (Fig. 1A). The seed narrows into a rounded point on the ventral surface, possibly equivalent to the beak in typical Vitaceae (see fig. 1 of Chen and Manchester, 2011). The lateral flattening, near-straight ventral surface and suboval, or near elliptical, outline give the seed a very different shape from typical Vitaceae genera (e.g., Chen and Manchester, 2011). Externally the ventral infolds are only tentatively identifiable from a pink mineral infill from the apical and ventral views (Fig. 1B, D). However, in μCT digital transverse section (DTS), a pair of very deep (4.5 mm) and narrow (0.5 mm) ventral infolds are clearly delineated by a very thin gap (black in Fig. 2C) between the inferred outer surface of the endotesta and the mineral infill of the infolds (outlined in solid yellow in Fig. 2D).

Comments—Seeds of modern Cissus integrifolia (Baker) Planch. are very similar to the holotype of C. crenulata, being narrow, laterally flattened and suboval in lateral view with a line of bisymmetry passing through median longitudinal rib perichalaza and having a similar seed coat ornamentation. However, the greater number of ridges across the lateral faces and different orientation of the ventral surface relative to the long axis of the seed in Cissus integrifolia support the recognition of a separate species. Chesters (1957) listed P. B. 8 as a paratype of Menispermicarpum crenulatum. Currently, the number KNMP-RU7787 (P. B. 8) includes eight specimens, which have here been transferred to Cissus andrewsii sp. nov. (see below). Therefore, Cissus crenulata is represented only by a single specimen.

Species—Cissus andrewsii Adams, Collinson, S.Y.Smith, & Bamford sp. nov.

Etymology—The species epithet andrewsii is named in honor of Dr. Peter Andrews, in recognition of his extensive work on the Kenyan Miocene and the invaluable support he provided, which enabled one of us (Collinson) to undertake fieldwork on Rusinga and Mfangano Islands in 1980–1981.

Diagnosis—Seeds bilaterally symmetrical, 16–19 mm in dorsiventral dimension, laterally flattened, suboval in lateral view, narrowly elliptical and 5–8 mm wide in apical and basal views, elliptical in ventral view. Centers of lateral faces ornamented by 4–5 short ridges in radial pattern; curved ridge 1–2.5 mm from margins of lateral faces extends around most of seed and delineates faceted to reticulate marginal area; prominent median ridge (rib perichalaza) extends from base of ventral surface, around base, over dorsal surface, and almost full length of apical surface. Upper portion of ventral surface forms acute angle with long axis of the seed. Pair of deep, wide ventral infolds present.

Holotype hic designatus—Seed: V68501 (Fig. 1E–H; μCT DTS video available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36).
Type locality—Rusinga Island, Lake Victoria, Kenya.

Paratypes—Seeds: V68500 (Fig. 1 I, J); V68502 (Fig. 1 K, L); KNMP-RU7787 (8 specimens with field number 60×52, formerly P. B. 8, previously listed as paratypes of Menispermicarpum crenulatum by Chesters, 1957; re-examined for this study by Bamford).

Geological horizon and age—Hiwegi Formation, early Miocene.

Repository—Natural History Museum, London, UK (holotype and paratype specimens with prefix V); National Museums of Kenya, Nairobi, Kenya (other paratypes).

Description—Seeds are bismetrametrically arranged around a prominent median pericalaza rib (Fig. 1 H, J) and vary from 16 to 19 mm in dorsiventral dimension, 11 to 15 mm in height and 5 to 8 mm in width (Fig. 1 E–L). Seeds are laterally flattened with height/width ratios of 1.9–2.4 and are narrow in apical and basal views (Fig. 1 G, H, J, L). The seed apex is rounded, with no indication of an apical notch or chalazal grooves (Fig. 1 E–G), and narrows to the ventral margin (Fig. 1 F, G). The ventral surface is slightly rounded, not straight (Fig. 1 F, K). The lateral surfaces have a curved ridge, delimiting a sculptured outer margin, which extends from approximately half way along the apical margin, around the dorsal margin, and fully along the basal margin (Fig. 1 F). The sculptured marginal band is faceted, facets spaced at 2 to 3.5 mm, and ridged with ridges sometimes fusing to form a reticulum (Fig. 1 F, I, K). The central areas of the lateral faces are ornamented by four to five short (2–3 mm) ridges in a radial arrangement (Fig. 1 F). Where undamaged, the upper portion of the ventral surface forms an angle of 52° to 55° with the long axis of the seed (Fig. 1 F, K). The ventral infolds are not visible on the outside of the seed (Fig. 1 E, G, L), but by using μCT, deep (2.5 mm), broad (0.6 mm) ventral infolds can be identified in V68501 (Fig. 2 F) by differences in X-ray attenuation (gray level) due to variation in mineral density and mineral texture in the infold infills. A very clear gap (black in Fig. 2 E) demarcates the inferred original position of the endotesta outer surface in the areas away from the infolds, where endotesta would have been originally thicker based on observations in modern seeds. Some additional outer mineral (possibly representing exotesta or fruit wall remnants) is also present in the holotype (Fig. 2 E).

Comments—In her unpublished thesis, Chesters (1958) assigned the specimen shown in Fig. 1 E–H (V68501) to Menispermicarpum crenulatum, here revised to Cissus crenulata, but there are clear differences between C. crenulata and V68501, supporting assignment of this specimen, and other similar specimens, to a new species. Although there is now only a single specimen of Cissus crenulata (making it impossible to assess intraspecific variation), there are 11 specimens of C. andrewsii, all with consistent morphology distinct from that of the single specimen of C. creulata. In Cissus andrewsii the central portion of the lateral faces is crossed by short ridges in a radial pattern, unlike C. crenulata, and the lateral faces have a greater number of short transverse ridges in the outer margins, some of which fuse to form a reticulum. In addition, the curved ridge, delineating the sculptured margin from the central flat area, extends farther down the ventral surface and the ventral margin is curved not straight. Cissus andrewsii is therefore more similar to seeds of modern Cissus dasyantha Gilg & M.Brandt than C. integrifolia, but is sufficiently different from these extant species to warrant assignment to a new species, and differs in several ways from the fossil C. crenulata.

Species—Cissus rusingensis Adams, Collinson, S.Y.Smith, & Bamford sp. nov.

Synonymy—’cf. Cissus sp. 1 nov.’ in Collinson et al. (2009).

Etymology—The epithet rusingensis refers to the type locality on Rusinga Island from which the specimens were collected during in situ excavations.

Diagnosis—Seeds bilaterally symmetrical, 7–8 mm in dorsiventral dimension, slightly laterally flattened, subrounded to pyriform in lateral view, broadly elliptical in apical, basal, and ventral views. Lateral faces ornamented by ridges and reticula with 7–11 wide muri radiating to margins and enclosing lumina in central area; prominent median ridge (rib perichalet) extends from ventral margin, around base, over dorsal surface, and almost full length of the apex. Upper portion of ventral surface forms acute angle with long axis of the seed. Pair of deep, very wide ventral infolds present.

Holotype hic designatus—Seed: R117.1981.314 (Fig. 1 M–O; μCT DTS video available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36).

Type locality—R117 site (see Collinson et al., 2009), Rusinga Island, Lake Victoria, Kenya.

Paratypes—Seeds: KNMP-RU9647 (field number R117.1981.422); R117.1981.476 (Fig. 1 P–R; μCT DTS video available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36); R117.1981.604 (Fig. 1 S–U); R117.1981.605 (Fig. 1 V–X).

Geological horizon and age—Grit Member, Hiwegi Formation, early Miocene.

FIGURE 2 Ventral infolds revealed in a seed of modern African Cissus populnea Guill. & Perr. (A, B) by synchrotron-based X-ray tomographic microscopy (SRXTM) and in fossil seeds from the Hiwegi Formation, Rusinga Island, Kenya (C–J), assigned to Cissus herein, by micro-computed tomography (μCT). (A, B) SRXTM digital transverse section (DTS) through modern C. populnea, infill of the endotesta in yellow in (B) highlighting the position of the ventral infolds. (C, D) μCT DTS through the holotype (V33753) of Cissus crenulata comb. nov., with (D) showing inferred position of the ventral infolds (solid yellow lines) and the margins of the endotesta (dotted yellow lines). (E, F) μCT DTS through the holotype (V68501) of Cissus andrewsii sp. nov., with (F) showing infill of the inferred endotesta in yellow, highlighting two parallel, broad ventral infolds. (G, H) μCT DTS through the holotype (R117.1981.314) of Cissus rusingensis sp. nov., with (H) showing infill of inferred embryo cavity within endotesta in yellow, highlighting two parallel, very broad ventral infolds. (I, J) μCT DTS through holotype (V68506) of Cissus psilata sp. nov., with (J) showing inferred position of pair of very short, narrow parallel ventral infolds (solid yellow lines). All μCT sections obtained from near ventral part of seeds, where ventral infolds were most likely to be evident, if concealed externally, based on their position in modern Cissus seeds. Scale bars = 1 mm.

Description—Seeds are bisymmetrical around a prominent median perichalazal rib (Fig. 1N, O, Q, R, T, U, W, X), and they vary from 7 to 8 mm in dorsiventral dimension, 5 to 6 mm in height, and 3 to 4 mm in width (Fig. 1M–X). The seeds are laterally flattened with a height/width ratio varying between 1.5 and 1.8 and are broadly elliptical in apical and basal views (Fig. 1N, O, Q, R, T, U, W, X). The seeds have a rounded apex (Fig. 1M, P, S, V) but are variable in their lateral shape (Fig. 1M, P). The perichalazal rib extends around less of the specimen in R117.1981.476 (Fig. 1P) and R117.1981.604 (Fig. 1S), resulting in a reduced lateral width higher up the seed, a longer ventral projection and a more pyriform shape. By contrast, the holotype (Fig. 1M) and R117.1981.605 (Fig. 1V) are both subrounded with only small ventral projections. The lateral surfaces have a reticulate ornamentation with one or two centrally positioned lumina (Fig. 1M, P, S, V) and 7 to 11 muri or ridges ranging in width from 0.2 to 0.6 mm (e.g., Fig. 1M). Some ridges radiate to the edges of the specimen and join up with the strong perichalazal rib producing marginal lumina (Fig. 1O, X). Others terminate before reaching the margin without forming a reticulum (bottom right in Fig. 1S; top left in Fig. 1V), resulting in a radiating pattern of marginal ridges and unenclosed marginal depressions. The upper portion of the ventral surface forms an angle of 35° to 55° with the long axis of the seed (Fig. 1M, P, S, V). The ventral infolds, although not externally visible (Fig. 1N, Q, T, W), are readily identifiable in the holotype by differences in contrast and mineral density in μCT scans (Fig. 2G). A distinct pale area (highlighted yellow in Fig. 2H) marks the position of the embryo cavity, while the deep (1.5 mm) and very broad (0.75 mm) ventral infolds are infilled with mineral with lower X-ray attenuation and hence darker gray color (Fig. 2H).

Comments—Cissus rusingensis is distinctive in lacking a distinct curved ridge delineating a marginal region and in having muri in the central area of the lateral faces forming a reticulum. This species is very similar to seeds of modern Cissus lebrunii Dewit, but C. rusingensis has fewer muri and at most two (rather than three) centrally positioned lumina.

Species—Cissus psilata Adams, Collinson, S.Y.Smith, & Bamford sp. nov.

Etymology—The epithet psilata derives from the ancient Greek adjective psilos, meaning bare or smooth, and refers to the lack of seed coat ornamentation on the smooth surface of the lateral faces.

Diagnosis—Seed bilaterally symmetrical, 7.5 mm in dorsiventral dimension, subrounded in lateral view, broadly elliptical in apical, basal, and ventral views. Seed smooth, lacks external ornamentation (psilate), except for prominent median ridge (rib perichalaza) that extends all around dorsal surface, over apex, and almost full length of ventral surface. Base of ventral surface concave, forming acute angle with long axis of seed. Pair of very short, narrow ventral infolds present.

Holotype hic designatus—Seed: V68506 (Fig. 1Y-BB; μCT DTS video available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36).

Type locality—Rusinga Island, Lake Victoria, Kenya.

Geological horizon and age—Hiwegi Formation, early Miocene.

Repository—Natural History Museum, London, UK.

Description—Seed inferred to be originally bilaterally symmetrical around a prominent median perichalazal rib, although the single specimen is slightly deformed (Fig. 1Z, AA), probably due to abnormal development in life or distortion during fossilization. The seed is subrounded in lateral view (Fig. 1Y), 7.5 mm in dorsiventral dimension and 6 mm in height, and broadly elliptical in apical and basal (Fig. 1Z, AA) views, 4.5 mm in width. The seed is not laterally flattened and has an inflated morphology with a height/width ratio of 1.3. The perichalazal rib is ca. 0.3–0.45 mm thick around the dorsal and basal margins (Fig. 1Y, Z) but thicker at the seed base (ca. 0.8 mm), forming a strong point (Fig. 1Y, AA), equivalent to the beak in typical Vitaceae (see fig. 1 of Chen and Manchester, 2011). The lateral surfaces are smooth (Fig. 1Y). The upper portion of the ventral surface forms an angle of 45° with the long axis of the seed (Fig. 1Y). The ventral infolds are not visible on the outside of the seed (Fig. 1Z, BB). μCT scans of the holotype (Fig. 2I) show very little internal information compared to the other fossils, except for two very short (less than 0.5 mm) grooves at the base of the ventral surface (Fig. 2I), which correspond to the position of the ventral infolds characteristic of Cissus.

Comments—in her unpublished thesis, Chesters (1958) suggested that V68506 could be assigned to the genus Menispermicarpum as a new, but never published, species. However, the fossil morphology differs from endocarps of Menispermaceae. It is almost identical to seeds of modern Cissus petiolaris Hook.f., differing in the greater extent of the rib perichalaza on the dorsal surface. In modern Cissus petiolaris seeds, the ventral infolds are very shallow and short which, in combination with mineralization effects, can explain the very limited evidence for this diagnostic feature in V68506 (Fig. 2I).

RESULTS

Identification of fossils to genus Cissus—The lack of cellular and tissue detail in the fossils (Fig. 2C–I), and the complexities of mineralization during fossilization, make it difficult to judge if the external morphology of a fossil represents the external morphology of a living equivalent. The SRXTM videos of modern Cissus seeds (available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36) show that the inner and outer surfaces of the endotesta are parallel to one another, and hence, a mineral infill of the endotesta will have a very similar external morphology to a mineral replacement of the endotesta itself. Therefore, it is justifiable to compare the external surface of the Hiwei Formation fossils with that of modern seeds for purposes of identification. These comparisons show that several modern African Cissus species have seeds with almost identical shape and ornamentation to the Hiwei Formation fossil seeds (see Phylogenetic context of seed morphotypes). However, the ventral infolds that characterize modern Cissus seeds are not visible on the fossils. The taphonomy of the fossils may explain the absence of these key features.

A novel approach in virtual taphonomy was used to produce a digital infill of the endotesta of a modern Cissus seed (Fig. 3A). This virtual fossil showed all the key characteristics of Cissus seeds (i.e., a long, linear chalaza, the perichalaza; a thickened ridge of seed coat along the perichalaza, the perichalazal rib; deep, narrow, and linear
Phylogenetic context of seed morphotypes—Seeds of four distinct Cissus species are now recognized from the early Miocene Hiwegi Formation. Given that claims of Vitaceae pollen from the Oligocene of Cameroon remain unconfirmed (Salard-Cheboldaeff, 1978, 1981; Muller, 1981), these seeds represent the first confirmed fossil record of Vitaceae and of the genus Cissus in Africa. The four fossil species represent distinct morphotypes, which are also found in seeds of extant species (Table 1), as revealed by SRXTM imaging of modern fruits and by a literature survey of published seed descriptions. The phylogenetic position of these extant species provides a phylogenetic context for the early Miocene fossils.

Morphotype 1—Cissus crenulata (Fig. 1A–E) has laterally flattened, suboval seeds with a faceted marginal area, several long radiating lateral ridges and two marginal ridges. The extant species Cissus integrifolia (Figs. 6D, 7A) and C. populnea (Figs. 2A, 6E) share this distinctive morphotype (Table 1). These two species are recovered in our phylogenetic analyses with strong support (100% BS) as a distinct clade, the Cissus integrifolia clade, which is one of the two earliest-diverging clades in core Cissus (Fig. 5). It is, therefore, likely that Cissus crenulata was a member of the early-diverging Cissus integrifolia clade.

Morphotype 2—Cissus andrewsi has seeds that are laterally flattened and have a ridged and faceted marginal area where ridges sometimes fuse to form a reticulum, two prominent marginal ridges, and short radiating lateral ridges. The extant species Cissus barbeyana (Fig. 6B) and C. dasyantha (Fig. 7B and fig. 13J of Dewit and Willems, 1960) are most similar to C. andrewsi, in that they are slightly laterally flattened (height/width ratio of 1.3–1.5) with a similarly ridged and faceted marginal reticulum, two marginal ridges and short radiating lateral ridges (Table 1). Although the sampled specimen of Cissus dasyantha failed to amplify material for phylogenetic analysis, C. barbeyana was found in the basal clade of core Cissus, the Cissus barbeyana clade, with C. floribunda and C. sagittifera.
Seeds of *Cissus floribunda* (Fig. 6A) and *C. sagittifera* (fig. 14 of Descoings, 1967) also possess the morphotype 2 characters, (except that *C. sagittifera* lacks the ridged and faceted marginal area), suggesting that morphotype 2 is characteristic of the basal *Cissus barbeyana* clade and that *C. andrewsii* can be placed in this clade. Future work determining if *Cissus dasyantha* is in the *C. barbeyana* clade would test this hypothesis.

Morphotype 3—*Cissus rusingensis* has slightly laterally flattened seeds and ridges forming a complete to incomplete reticulum across the lateral faces. Extant species sharing these features (Table 1) include: *Cissus lebrunii* (fig. 13K of Dewit and Willems, 1960), *C. oreophila* (pl. 39, fig. 11 of Descoings, 1972), *C. sciaphila* (Fig. 6H), *C. smithiana* (Fig. 6G), and *C. tilifolia* (Fig. 7C and Verdcourt, 1993). *Cissus sciaphila* and *C. tilifolia* are found together in the *C. sciaphila* clade (Fig. 5). *Cissus aralioides* (Fig. 6L), *C. rotundifolia*, and *C. sylvicola* have morphotype 4 seeds (online Appendix S4). *Cissus oliveri*, with coarsely

Morphotype 4—Seeds of *Cissus psilata* are subrounded with smooth lateral faces almost identical to seeds of extant *C. petiolata* (Table 1; Figs. 6M, 7D). *Cissus petiolata* is found in a clade with seven solely African species (Fig. 5): *C. aralioides*, *C. oliveri*, *C. polita*, *C. rondoenensis*, *C. rotundifolia*, *C. sylvicola*, and *C. welwitschii*. Of these, *Cissus aralioides* (Fig. 6L), *C. rotundifolia*, *C. sylvicola*, and *C. welwitschii* have morphotype 4 seeds (online Appendix S4). *Cissus oliveri*, with coarsely
First fossil seeds of Cissus (Vitaceae) from Africa

[Diagram showing a phylogenetic tree or classification system related to Cissus species, with annotations and labels indicating different morphological types and key for scale and other seeds studied and unlike fossils.]
pitted seeds and very strong radial and lateral ribs (Verdcourt, 1993), and *C. rondoensis*, with seeds bearing two to three, faint transverse ridges (Verdcourt, 1993), lack the smooth seed coat typical of morphotype 4. These two species occupy contrasting habitats to the rest of the clade (Table 2), so contrasting seed morphology may reflect different environmental pressures.

Extant species in the *Cissus pseudoguerkeana-C. albipor cata* clade, for which seed descriptions were available, also share seed morphotype 4: *C. faucicola* and *C. quadrangularis* have smooth seeds (online Appendix S4), and *C. cactiformis* also has the smooth seed coat (Verdcourt, 1993). Furthermore, the majority of species in the *Cissus elongata-C. subtetragona* clade (Fig. 5) share morphotype 4 characters (online Appendix S4). This evidence suggests that morphotype 4 occurs throughout the broader *Cissus petiolata-C. albipor cata* clade (Fig. 5) and that *C. psilata* belongs in this clade.

Further morphological and molecular analyses, with expanded geographic and taxonomic sampling (including those species that failed to amplify for this study), are needed to fully evaluate the systematic significance of these seed morphotypes. This study focused on modern African species to place the African Miocene fossils in context. On the basis of extant species that have morphologically comparable seeds and their phylogenetic position, species from four clades of *Cissus* (or from the ends of their stem lineages) were present during the early Miocene in East Africa. These species were related to the two early-diverging clades of core *Cissus* (the *Cissus integrifolia* and *C. barbeyana* clades) and two later-diverging clades with mainly African species today, the *C. sciaphila* clade and the *C. petiolata-C. albipor cata* clade (Fig. 6).

**Fossil plant biology and ecology** — Habit and habitat data (Table 2) for nearest living relatives of the fossil *Cissus* (Fig. 6) can be used to consider the likely paleobiology and paleoecology of the fossil plants. In the *Cissus integrifolia* clade, the plants are lianas or herba ceous climbers in wide-ranging habitats, from evergreen forest and woodland to bushland and savanna grassland. Members of the *Cissus barbeyana* clade are herbaceous to woody lianas and predominantly occupy rainforest, gallery and riverine forest fringes. Species in the *Cissus sciaphila* clade are also herbaceous to woody climbers in rainforests or are scrambling shrubs or woody climbers in drier deciduous forest and woodland. The extant species of the *Cissus aralioides-C. albipor cata* clade are herbaceous to woody lianas or climbing shrubs, a number of which are succulent (*C. aralioides, C. cactiformis, C. petiolata, C. quadrangularis, C. rotundifolia, C. sylvicol a*). They occur in riverine forest and wooded savanna to rocky outcrops, xerophilic thickets in Acacia mixed bushland and grassland (Table 2). The two members of the *Cissus petiolata* subclade with different seeds (see previous section) also differ in their habitat preferences, being most often found in moist rainforests.

These modern ecologies suggest that a diversity of climbers (either herbaceous or woody or both) were present in the early Miocene on Rusinga Island. These climbers may have occupied gallery or riverine forest (*Cissus andrewsii* related to the *C. barbeyana* clade) and evergreen forest, through woodland to savanna (*C. rusingensis* related to the *C. sciaphila* clade and *C. crenulata* related to the *C. integrifolia* clade). The plant producing the *Cissus psilata* seeds may have been somewhat succulent and hence able to occupy arid habitats as well as riverine forest or savanna, based on the habitats of a number of related extant species in the *C. aralioides-C. albipor cata* clade.

**DISCUSSION**

**Homology of the ventral surface in Vitaceae seeds** — The current convention for descriptive terminology of Vitaceae seeds (Chen and Manchester, 2011) and fossil *Cissus* seeds (Manchester et al., 2012b) has been followed here (online Appendix S6) to allow for ease of comparison with their work. However, an alternative interpretation is possible, particularly for the strongly flattened seeds (e.g., Figs. 1A–L, 2A–D, 3, 6A–F), whereby the ventral surface incorporates both ventral and apical (sensu online Appendix S6) and the ventral grooves are short occupying less than half of the dimension of that surface. This alternative seed orientation is followed in all the floras to which we refer in this paper (i.e., the modern seed illustrations in Fig. 6 are all rotated 90° clockwise). These two alternative homologies would best be investigated by a developmental study.

**Cissus origins, phylogeny, and Miocene diversity in Africa** — Relationships among genera in the Vitaceae are mostly comparable between our study and the most recent previous studies of *Cissus* (Liu et al., 2013, 2016; Rodrigues et al., 2014), with limited support for the backbone of the trees (but see Wen et al., 2013 and Zhang et al., 2015). *Cissus* species are found in three distinct clades in all analyses, identified by Rodrigues et al. (2014) as the *Cissus striata* clade (clade III of Liu et al., 2013), the *C. trianae* clade (clade V of Liu et al., 2013) and the core *Cissus* clade, which contains most of the species. As in Liu et al. (2013), the earliest-diverging clades comprise mainly African species, although this is more evident in our study in which the first branches are exclusively African species. This topology provides support for an African origin for the genus, as Liu et al. (2013) also concluded.

The new African fossils are entirely consistent with an African origin for *Cissus*. The notable similarities in seed morphology between extant *Cissus* species and Hiwegi Formation fossils across multiple seed morphotypes suggest the presence of four clades or members of their stem lineages, indicating diversity of *Cissus*, by the early Miocene in Africa. Based on the divergence times (online Appendix S9), it appears that all four clades originated much earlier than the ca. 18 Ma age of the Miocene fossils: 41.5 Ma (HPD 31.5–51.0 Ma) for the split of the *Cissus barbeyana* clade from the rest of core *Cissus*; 36.0 Ma (HPD 27.6–44.7 Ma) for the divergence of the *C. integrifolia* clade; 24.6 Ma (HPD 18.1–32.1 Ma) for the divergence of the *C. sciaphila* clade, and 22.8 Ma (HPD ca. 16.6–30.0 Ma) for the divergence of the *C. aralioides-C. albipor cata* clade. The time-calibrated phylogeny (online Appendix S9) therefore suggests that several ghost lineages of *Cissus* await discovery in the fossil record. The new African Miocene seeds, and those from the Oligocene of Peru (see **Comparison with other fossil Cissus** below), provide fossil evidence consistent with the suggested phylogenetic history of *Cissus*.

**Comparison with other fossil Cissus** — The fossil record of the grape family (Vitaceae) extends back to the latest Cretaceous or earliest Paleogene in central India (Manchester et al., 2013) and to the Paleogene in North America and Europe (e.g., Manchester, 1994; Fairon-Demaret and Smith, 2002; Chen and Manchester, 2007; Collinson et al., 2012). However, for *Cissus*, the oldest, and only other, currently recognized examples are from the late early Oligocene Belén flora of northern Peru (Manchester et al., 2012b). This flora contains two *Cissus* species: *Cissus willardii* Berry and *Cissus lombardii* Manchester, Chen, & Lott. *Cissus willardii* is small
and globose with smooth lateral faces and large ventral infolds (fig. 2A–Q of Manchester et al., 2012b), unlike any of the fossils described above from Rusinga Island.

Conversely, *Cissus lombardii* shares some characteristics with African Miocene *C. crenulata* and modern *C. integrifolia* (seed morphotype 1), being bilaterally symmetrical, laterally flattened, elliptical in lateral view with a pronounced median perichalazal rib and a faintly faceted marginal area defined by a marginal ridge on each lateral face (fig. 3A–N of Manchester et al., 2012b). Thus, *Cissus lombardii* might be related to the *C. integrifolia* clade, extending its fossil record to the early Oligocene (30–28.5 Ma). However, *Cissus lombardii* is smaller in all dimensions than *C. crenulata* and *C. integrifolia* and lacks the distinct ornamenting ridges that cross the lateral faces in seeds of these species. The specimens of *Cissus lombardii* from the Belén flora are internal casts, which could result in a more subdued surface ornamentation than if the fossils were seeds themselves. However, as has been shown in extant *Cissus* seeds in this study using SRXTM (SRXTM videos of modern *Cissus* seeds are available from the Dryad Digital Repository, http://dx.doi.org/10.5061/dryad.g9r36), the inner surface of the endotesta closely parallels the outer surface, which would result in a similar pattern of ornamentation whether a fossil is an internal cast or a replacement of the endotesta itself. There is no indication of even faint ridges across the lateral faces in *Cissus lombardii* (fig. 3A and B of Manchester et al., 2012b), suggesting that the ornamentation of the original seed was significantly different from morphotype 1. The portion of the ventral surface containing the ventral infolds is “more or less planar (not markedly concave)” in *Cissus lombardii* (Manchester et al., 2012b, p. 936), rather than weakly to strongly concave as in *C. integrifolia* and *C. crenulata*. Therefore, *Cissus lombardii* is clearly distinct from the African Miocene species.

**Biogeographic implications**—The position of previously unsampled modern African species near the base of the phylogeny (Fig. 5) reinforces the African origin for the core *Cissus* clade, suggested by Liu et al. (2013). The new African Miocene fossil *Cissus crenulata* has seed morphotype 1 as do both extant species of the *C. integrifolia* clade, confirming the presence of early-divergent members of the core *Cissus* clade in Africa by at least the Miocene.

The two modern species outside Africa in the *Cissus sciaphila* clade (*C. rostrata* and *C. adnata*) are advanced within the clade. Six of the seven modern African species in this clade are characterized by seeds of morphotype 3, and the seventh includes specimens with this
seed morphotype. The new African Miocene fossil *Cissus rusingensis* also has seeds of morphotype 3. These data suggest that this clade originated in Africa. Dispersal during or after the Neogene to Australasia resulted in the modern pantropical intercontinental disjunct distribution. The calibrated phylogeny presented here (online Appendix S9) suggests that the divergence of Australasian species occurred near the end of the Pliocene at 2.7 Ma (HPD 0.8–5.5 Ma) differing from, although within the error of, the late Miocene estimate of 7.8 Ma (HPD 3.0–15.1 Ma) made by Liu et al. (2013).

Liu et al. (2013) argued that transoceanic long-distance dispersal, rather than terrestrial mammalian dispersal, was the most likely explanation for pantropical intercontinental disjunctions in *Cissus* because *Cissus* fruits are fleshy and, although in some instances dispersed by mammals, are predominantly bird-dispersed, enabling long-distance transport by bird migration. Multiple large islands across the Indian Ocean may have facilitated an out-of-Africa migration by acting as migratory “stepping stones”, as invoked for dispersal of other vitaceous genera (e.g., *Cayratia*; Lu et al., 2013).

The distributions of modern species in all four clades containing nearest living relatives to the fossils extend across Africa from East to West (Table 2). The fossil seeds suggest that the clades containing these living relatives may have had their origins in East Africa with subsequent spread to the rest of the continent. However, additional African fossil records of *Cissus* are needed to document dispersal patterns.

**Paleoenvironmental implications**—Previous paleoenvironmental reconstructions from the Hiweti Formation have inferred a habitat mosaic inhabited by early hominoids, such as *Ekembo* (see McNulty et al., 2015). Evidence for mosaic habitats comes from gastropod (Verdcourt, 1963; Pickford, 1995) and mammal faunas (e.g., Andrews and Van Couvering, 1975), paleosols (Retallack et al., 1995) and paleobotany (Collinson et al., 2009; Maxbauer et al., 2013; Michel et al., 2014). Collinson et al. (2009) concluded that the overall paleoenvironmental signal, considering the evidence from plants, mammals, gastropods and paleosols, was one of “mixed habitats

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**FIGURE 7** Digital transverse sections, produced by synchrotron-based X-ray tomographic microscopy, through fruits of modern African *Cissus*, using representative specimens to illustrate typical features of each seed morphotype listed in Table 1. (A) Morphotype 1, *Cissus integrifolia* Guill. & Perr. (B) Morphotype 2, *Cissus dasyantha* Gilg & M.Brandt. (C) Morphotype 3, *Cissus tiliifolia* Planch. (D) Morphotype 4, *Cissus petiolata* Hook.f. Transverse sections were obtained from near ventral part of fruits to best show features of ventral infolds and characteristics of seed coat layers. Scale bars = 1 mm.
dominated by woodlands, with waterside environments and small patches of forest big enough to support forest faunas” (p. 161). This conclusion is very similar to that derived from the possible fossil *Cissus* paleoecologies based on nearest living relatives (see *Fossil plant paleobiology and paleoecology*).

The vegetation in which *Cissus andrewsii*, *C. crenulata*, and *C. psilata* lived cannot be inferred from associated fossils, as the specimens were surface-picked. However, the context of the *Cissus rusingensis* fossils is well understood as they derive from in situ excavations of plant litter assemblages at the R117 site (Collinson et al., 2009). These litter assemblages were interpreted to have accumulated under a continuous canopy in deciduous, broad-leaved woodland bordering a river, based on the fossil fruits and seeds and their taphonomy (Collinson et al., 2009). *Cissus rusingensis* is therefore known to have inhabited a closed riverine woodland, consistent with interpretations made for this species from inferred near living relatives.

Recent studies have revealed temporal paleoenvironmental changes through the Hiwegi Formation (Michel et al., 2013, 2014; Garrett et al., 2015) and have suggested that interpretations of mosaic paleoenvironments may be based on time-averaged faunal and floral assemblages that confine separate, more homogeneous habitats. These studies suggest that more open, drier woodland habitats low in the Hiwegi Formation (e.g., Grit Member) gave way to dense, closed canopy forest further up (e.g., Fossil Bed and Kibanga Members), with early hominoid fossils recovered from both paleoenvironments (Garrett et al., 2015). Since the stratigraphic context of *Cissus andrewsii*, *C. crenulata*, and *C. psilata* are unknown and they are not associated with *C. rusingensis*, the four new species of *Cissus* described may or may not have existed contemporaneously. Despite this uncertainty, it is known from in situ excavations in the Fruit and Nut Bed (Collinson et al., 2009) and stratigraphically associated leaf assemblages (Maxbauer et al., 2013) that a riverine mosaic habitat of woodland and forest existed during the deposition of the Grit Member of the lower Hiwegi Formation. The paleoecology of the new fossil species of *Cissus*, inferred from living relatives and supported (for *Cissus rusingensis*) by associated fossils, provides new evidence for mosaic landscapes on Rusinga Island during the early Miocene, ranging from gallery or riverine forest to woodland, bushland, and savanna.

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**LITERATURE CITED**


APPENDIX 1
Voucher information and GenBank accession numbers of Cissus species, for which new sequences were produced for the present phylogenetic analysis.

**Taxon:** tmlF; rps16; trnC-petN; atpB-rbcL; voucher information; Kew DNA Bank accession number.

*Cissus barbeyana* De Wild. & T.Durand; KX131178; - ; Lisowski, S. 16406 (K); 31957. *Cissus bosseri* Desc.; KX131174; - ; KX131172; Phillipson, P.B. & Rabeshianaka, S. 3140 (K); 31965. *Cissus petiolaris* Hook.f.; KX131175; - ; Luke, P.A. & WRQ 9365 (K); 31952. *Cissus polyantha* Gilg & M.Brandt; KX131176; - ; Deighton, F.C. 5208 (K); 31955. *Cissus populnea* Guill. & Perr.; KX131179; - ; Daramola, B.O. 221 (K); 31968. *Cissus rondoensis* Verdc.; KX131170; - ; Bigdood, S., Abdallah, R. & Vollesen, K. 1553 (K); 31961. *Cissus smithiana* (Baker) Planch.; KX131177; - ; Louis, J. 559 (K); 31959. *Cissus tillifolia* Planch.; KX131173; - ; KX131171; Eilu, G. 240 (K); 31964.

APPENDIX 2
Species included in the phylogenetic analysis of family Vitaceae for which sequences were obtained from GenBank, with a particular focus on genus *Cissus*.

**Taxon:** tmlF; rps16; trnC-petN; atpB-rbcL.

*Cissus adnata* Roxb.; JX476585; JX476547; JX476673; JX476429. *Cissus albiporata* Masinde & L.E. Newton; JF437304; JX476648; JF437201; JX476640. *Cissus amazonica* Lindl.; JX476659; JX476649; JX476674; JX476431. *Cissus anamica* Lombardi & B.; JX476502; JX476650; JX476675; JX476432. *Cissus annamica* Gagnep.; - ; JX476676; - ; *Cissus antarctica* Vent.; JX476696; JX476651; JX476677; JX476433. *Cissus appendiculata* Lombardi; JX476513; - ; *Cissus aphyllantha* Gilg; JX476682; JX476553; JX476658; JX476645. *Cissus araguainensis* Lombardi; JX314315; - ; *Cissus aralioides* (Welw. ex Baker) Planch.; JF437305; - ; JF437202; - ; *Cissus assamica* (M.A. Lawson) Craib; JF437307; JX476559; JF437204; JX476441. *Cissus auricoma* Desc.; JX476686; JX476566; JX476682; JX476644. *Cissus baihensis* Lombardi; JX476562; JX476684; JX476644. *Cissus blandichiana* Planch.; JX314317; - ; *Cissus castelliformis* Gilg; JX476688; JX476563; JX476685; JX476645. *Cissus campestris* (Baker) Planch.; JX314318; - ; *Cissus cardiophylla* Standley; EF179089; - ; *Cissus cornifolia* (Baker) Planch.; JF437308; JX476657; JF437205; JX476440; *Cissus decidua* Lombardi; JX314319; - ; *Cissus descoingsii* Lombardi; JX314320; - ; *Cissus diffusa* (Miq.) Amshoff; JX476871; JX476565; JX476689; JX476451. *Cissus diffusifolia* (Baker) Planch.; JX476872; JX476570; - ; JX476452. *Cissus discolor* Blume; JF437309; - ; JF437206. *Cissus duarteana* Cambess.; JX314321; - ; *Cissus eloengata* Roxb.; - ; JX476573; JX476691; JX476555. *Cissus eversa* Rich; HMS58942; HMS58502; JX476693; HMS58526. *Cissus fauciaca* Wild & R.B.Drumm.; JX476784; JX476764; JX476458. *Cissus floribunda* (Baker) Planch.; JX476785; JX476577; JX476695; JX476459. *Cissus gongylodes* (Burch. ex Baker) Planch.; JX476877; JX476779; JX476697; JX476647. *Cissus granulosa* Ruiz & Pav.; JX476680; JX476562; JX476703; JX476644. *Cissus hastata* Miq.; AB235012; - ; JX476701; JX476465. *Cissus hypoglaucia* Duranq.; JX476881; JX476583; JX476702; JX476466. *Cissus incisa* (Nutt.) Des Moul. Ex S.Watson; HMS58949; HMS58804; JX475521. *Cissus integrifolia* (Baker) Planch.; JX476882; JX476584; JX476703; JX476467. *Cissus javana* DC.; JX476685; JX476689; JX476468. *Cissus janea* Desc.; JX476884; JX476685; JX476705; JX476469. *Cissus leucophaeus* (Scott-Elliot) Suess.; JX476885; JX476587; JX476706; JX476470. *Cissus madecassica* Desc.; JX476886. *Cissus macrocarpa* Vahl.; JX476888; JX476590; JX476709; JX476471. *Cissus microdonta* Vahl.; JX476889; JX476651; JX476710; JX476474. *Cissus neei* Croat; JX313424; - ; *Cissus nodosa* Blume; HMS58945; JX476592; JX476711; JX476475. *Cissus obliqua* Ruiz & Pav.; JX476890; JX476693; JX476712; JX476467. *Cissus oblonga* (Benth.) Planch.; EF179083; - ; *Cissus oliveri* Gilg. ex Eng. & Gilg; JX476892; JX476595; JX476714; JX476478. *Cissus paraensis* Lombardi; JX314327; - ; *Cissus paulinifolia* Velloz.JX313426; - ; *Cissus penninervis* (F.Muell.) Planch.; AF300309; - ; *Cissus pentaedra* Jackes; EF179084; - ; *Cissus phymatocarpa* Masinde & L.E. Newton; JF437311; JX476659; JF437209; JX476647. *Cissus piletosa* Desc.; JX476893; JX476576; JX476715; JX476480. *Cissus polita* Desc.; JX476894; JX476598; JX476716; JX476481.
delavayana Planch.; HM232253; ; ; . *Ampelopsis rubifolia* (Wall.) Planch.; JF437293; JX476546; JF437186; JX476428. *Cayratia acris* F. Muell.; EI197070; ; ; . *Cayratia clematidea* (F.Muell.) Domini; EF197072; ; . *Cayratia cordifolia* C.Y. Wu ex C.L. Li; HM585535; HM585794; JX476668; HM585818. *Cayratia debilis* (Baker)Suess.; JF437296; ; . *Cayratia euryema* B.L.Burtt; EF197073; ; ; . *Cayratia gracilis* (Guill. & Perr.)Suess.; JF437297; ; ; . *Cayratia imerimensis* (Baker) Desc.; HM585936; HM585796; JX476659; HM585520. *Cayratia japonica* (Thunb.) Gagnep.; HM585927; ; HM585521. *Cayratia maritima* Jackes; EF197074; ; ; . *Cayratia mollissima* Gagnep.; HM585938; HM585798; JX476671; HM585522. *Cayratia pedata* Gagnep.; AB235005; ; ; . *Cayratia saponaria* (Seem. Ex Benth.) Domini; EF197075; ; ; . *Cayratia trifolia* (L.) Domini; HM585940; JX476672; HM585524. *Cayratia tridentata* (Baker) Desc.; HM585941; ; .

*Clanoticissus angustissima* (F.Muell.) Planch.; EF197091; ; .
