

Role of C-X-C chemokines as regulators of angiogenesis in lung cancer

Robert M. Strieter,* Peter J. Polverini,† Douglas A. Arenberg,* Alfred Walz,§ Ghislain Opdenakker,¶ Jo Van Damme,¶ and Steven L. Kunkel*†

*The University of Michigan Medical School, Departments of Internal Medicine (Division of Pulmonary and Critical Medicine) and †Pathology, Ann Arbor, Michigan; ‡The University of Michigan Dental School, Section of Oral Pathology, Ann Arbor, Michigan; §Theodor Kocher Institut, University of Bern, Bern, Switzerland; ¶Rega Institute, University of Leuven, Leuven, Belgium

Abstract: Lung cancer is the leading cause of malignancy-related mortality in the U.S. and is predicted to increase over the remainder of this decade. Despite attempts to advance early diagnosis and use combination therapies, the clinical response of this cancer yields an overall 5-year survival rate of less than 15%. Clearly, new strategies for therapy are indicated. Although carcinogenesis is complex, tumor growth beyond 1–2 mm³ is dependent on angiogenesis. One of the potential mechanisms that allows for tumorigenesis is dysregulation of the balance of angiogenic and angiostatic factors that favors net neovascularization within the primary tumor. Numerous studies have investigated the role of a variety of molecules in the regulation of angiogenesis. Recently, interleukin-8 (IL-8), a member of the C-X-C chemokine family, has been found to be an angiogenic factor. In contrast, platelet factor 4 (PF4), another C-X-C chemokine, has been shown to have angiostatic properties. It is interesting that the major structural difference between IL-8 and PF4 is the presence of the NH₂-terminal ELR (Glu-Leu-Arg) motif that precedes the first cysteine amino acid residue of IL-8 and is important in ligand/receptor interactions. We hypothesize that angiogenesis associated with tumorigenesis is dependent on members of the C-X-C chemokine family acting as either angiogenic or angiostatic factors. This paradigm predicts that the biological balance in the expression of these C-X-C chemokines dictates whether the neoplasm grows and develops metastatic potential or regresses. In this review we discuss our recent laboratory findings that support this contention and suggest that further elucidation of the biology of C-X-C chemokines in the context of neovascularization of nonsmall cell lung cancer will permit novel targeted therapy aimed specifically at attenuating tumor growth and metastasis. *J. Leukoc. Biol.* 57: 752–762; 1995.

Key Words: tumor genesis • cytokines • neovascularization • chemotaxis

INTRODUCTION

Lung cancer is the leading cause of malignancy-related mortality in the U.S. [1]. Although the incidence of several other malignancies has declined or remained stable, the occurrence of bronchogenic carcinoma has escalated to near-epidemic proportions. Over 150,000 new cases are diagnosed and an equal number of deaths annually are attributable to bronchogenic carcinoma in the U.S.

[1]. Despite attempts to advance early diagnosis and use combination therapies, the clinical response of this tumor yields an overall 5-year survival rate for lung cancer patients of less than 15% [2]. Although lung cancer prevention (i.e., cigarette smoking cessation) should remain a high priority, a projection of lung cancer mortality through the end of the 20th century predicts that even with further reduction in smoking incidence, lung cancer in this decade will increase to a rate of 53.2 deaths per year per 100,000 population [2]. Clearly, new strategies for therapy are necessary.

The process of tumor growth and metastasis is complex and requires the highly orchestrated interactions of transformed neoplastic cells, tissue resident cells (i.e., fibroblasts, macrophages, and endothelial cells), and recruited cells (i.e., platelets, neutrophils, monocytes, and lymphocytes) from the circulation. One of the potential mechanisms that allows for maintenance of tumor growth is dysregulation of the balance of angiogenic and angiostatic factors. This dysregulation allows for the perpetuation of tumor growth and eventual metastasis. Based on a growing body of evidence, it is clear that a multitude of cytokines regulate growth. However, these cytokines may not express this full potential as individual polypeptides but collectively in a "cytokine network." To study all the pathophysiological aspects of cytokine networks during tumor growth and metastasis is a formidable task. Thus, in this review we will focus our studies on the expression and activity of members of the C-X-C chemokine family.

Abbreviations: NSCLC, nonsmall cell lung cancer; PF4, platelet factor 4; IFN, interferon; PBP, platelet basic protein; TP, total protein; SCID, severe combined immunodeficient; CTAP-III, connective tissue-activating protein-III; βTG, β-thromboglobulin; NAP-2, neutrophil-activating protein-2; IL-8, interleukin-8; GRO, growth-related oncogene; MIG, monokine induced by IFN-γ; IP-10, IFN-γ-inducible protein; ENA-78, epithelial neutrophil-activating protein-78; GCP-2, granulocyte chemotactic protein-2; bFGF, basic fibroblast growth factor.

Reprint requests: Robert M. Strieter, Department of Internal Medicine, Division of Pulmonary and Critical Care, Box 0360, University of Michigan Medical Center, 3916 Taubman Drive, Ann Arbor, MI 48109-0360.

Received November 8, 1994; accepted February 8, 1995.

TUMORIGENESIS IS AN EXAMPLE OF EXAGGERATED WOUND REPAIR

For nonsmall cell lung cancer (NSCLC) growth to succeed within the host, a complex interplay must occur between transformed neoplastic cells and nontransformed resident and recruited immune and nonimmune cells (i.e., fibroblasts, endothelial cells, and subpopulations of leukocytes) [4]. Although carcinogenesis or neoplastic transformation is dependent on multiple genetic and epigenetic events [5], the salient feature of all solid tumor growth is the presence of neovascularization [6, 7]. In the absence of local capillary proliferation and delivery of oxygen and nutrients, neoplasms cannot grow beyond 1–2 mm³ [6–8]. Folkman [7] first proposed in 1972 that tumors are angiogenesis dependent, with tumor growth correlating with a concomitant increase in vascular supply. In support of this contention is the finding that tumor cells contiguous with neovascularization have the highest [³H]thymidine-labeling index, whereas, tumor cells further removed from capillaries have the lowest [³H]thymidine-labeling index. Certain tumors have been found to produce factors that are directly angiogenic, whereas others may depend on neovascularization induced by products of resident cells or elicited leukocytes [7, 9–11]. These events are analogous to the formation of granulation tissue during the evolution of wound repair [4].

ANGIOGENESIS IS A PIVOTAL PROCESS OF TUMORIGENESIS

Angiogenesis is one of the most pervasive and essential biological events encountered in the mammalian organism [6, 12–14]. A number of physiological and pathological processes, such as embryonic development, the formation of inflammatory granulation tissue during wound healing, and the growth of malignant solid tumors, are strictly dependent on the recruitment of new capillaries. Normally, physiological angiogenesis occurs infrequently, yet can be rapidly induced in response to a number of diverse physiological stimuli. Among the most extensively studied of these angiogenesis-dependent physiological processes is normal wound repair [15]. An important feature of wound-associated angiogenesis is that it is locally transient and tightly controlled. The rate of capillary endothelial cell turnover is typically measured in months or years [16, 17]. However, when normally quiescent endothelial cells lining venules are stimulated, they will degrade their basement membrane and proximal extracellular matrix, migrate directionally, divide, and organize into new functioning capillaries invested by a new basal lamina all within a matter of days. This dramatic amplification of the microvasculature is nevertheless temporary, for as rapidly as they are formed they virtually disappear with similar swiftness, returning the tissue vasculature to homeostasis. The abrupt termination of angiogenesis that accompanies the resolution of the wound response suggests two possible mechanisms of control, neither of which are mutually exclusive. First, there is probably a marked reduction in angiogenic mediators. Second, a simultaneous increase occurs in the level of angiostatic factors that inhibit new vessel growth [8]. Although angiogenesis under conditions of normal wound repair appears to be under strict control, during

neoplastic transformation neovascularization is exaggerated. It appears that tumors are continually renewing and altering their vascular supply [7]. Interestingly, a normal vascular mass of tissue is approximately 20%, whereas, during tumorigenesis tumor vascular mass may be 50% of the total tumor [7]. These findings are consistent with the observations that angiogenic activity is both a marker of preneoplastic-to-neoplastic transformation as well as an event that perpetuates tumorigenesis. In addition, the magnitude of tumor-derived angiogenesis has been directly correlated with metastasis of melanoma, prostate cancer, breast cancer, and NSCLC [7, 18–23]. Moreover, this would support the notion that tumor-associated angiogenesis is dysregulated in such a manner that a biological imbalance exists that favors either the overexpression of local angiogenic factors or the suppression of endogenous angiostatic factors [7, 18, 24]. Although most investigations studying angiogenesis have focused on the identification and mechanism of action of angiogenic factors, recent evidence suggests that angiostatic factors may play an equally important role in the control of neovascularization [8, 24–32].

ANGIOGENESIS IS REGULATED BY ANGIOSTATIC FACTORS

A role for inhibitors in the control of angiogenesis was first described with the observation that hyaline cartilage was particularly resistant to vascular invasion [24, 25]. These studies reported that a heat labile guanidium chloride extract prepared from cartilage contained an inhibitor of neovascularization. Later an identical extract from rodent neonatal and shark cartilage was able to effectively block neovascularization and growth of tumors in vivo [26, 27]. Similar inhibitors of angiogenesis have been reported for other cell and tissue extracts [26–30] and for a variety of natural and artificial agents including inhibitors of basement membrane biosynthesis [31–34], placental RNase inhibitor [35], lymphotoxin [36], interferons (IFNs) [37], prostaglandin synthetase inhibitors [38], heparin binding fragments of fibronectin [39], protamine [40], angiostatic steroids [41], several antineoplastic and anti-inflammatory agents [42, 43], and platelet factor 4 (PF4) [44]. Many of these compounds have several biological activities. For example, PF4 blocks immunosuppression and inhibits bone resorption. Although most inhibitors can act directly on the endothelial cell to block migration and/or mitogenesis in vitro, their effects in vivo may be considerably more complex, involving additional cells and their products.

AN IMBALANCE IN ANGIOGENIC AND ANGIOSTATIC FACTORS MAY CONTRIBUTE TO THE PATHOLOGY OF CHRONIC DISEASES

Several lines of evidence suggest that a biological imbalance in the production of angiogenic and angiostatic factors contributes to the pathogenesis of several angiogenesis-dependent disorders. For example, in rheumatoid arthritis the unrestrained proliferation of fibroblasts and neovascularization leads to the formation of prolonged and persistent granulation tissue whose degradative enzymes contribute to profound destruction of joint spaces [45–47]. In contrast, monocyte-derived

macrophage from patients with scleroderma fail to stimulate the expected angiogenic activity [48], suggesting that a defect in macrophage responsiveness may contribute to the attenuated neovascularization that is encountered in scleroderma. Psoriasis is a well-known angiogenesis-dependent disorder that is characterized by marked dermal neovascularization. We have recently reported that keratinocytes derived from psoriatic plaques are potentially angiogenic compared with normal keratinocytes. Interestingly, this aberrant phenotype is due, in part, to a combined defect in the overproduction of the angiogenic cytokine, IL-8, and a deficiency in the production of the angiogenesis inhibitor thrombospondin-1 [49-52]. Thus, in diseases such as rheumatoid arthritis, psoriasis, or tumorigenesis, neovascularization appears to be aberrantly up-regulated. Although angiogenic and angiostatic factors may vary among different physiological and pathological settings, the recognition of this dual mechanism for control of angiogenesis is necessary to gain a more thorough understanding of this complex process and its significance in promoting tumor growth.

C-X-C CHEMOKINE FAMILY OF CYTOKINES

The fidelity of inflammation and wound repair is dependent on cellular communication. Although this communication is often accomplished through direct cell-to-cell contact via specific cellular adhesion molecules, cells may signal one another through soluble mediators, such as cytokines. These polypeptide molecules often have pleiotropic effects on a number of biological functions including proliferation, differentiation, recognition, and cellular recruitment. Their actions are mediated through paracrine and autocrine signaling and under certain conditions they can behave as hormones. Recently, a new family of cytokines have been identified that appear to have proinflammatory and reparative activities [53-57]. These cytokines in their monomeric forms are all less than 10 kDa and are characteristically basic heparin binding proteins. This family displays four highly conserved cysteine amino acid residues, with the first two cysteines separated by one nonconserved amino acid residue. In general, these cytokines appear to have specific chemotactic activity for neutrophils. Because of their chemotactic properties and the presence of the C-X-C cysteine motif, these cytokines have been designated the C-X-C chemokine family. Interestingly, these chemokines are all clustered on human chromosome 4 and exhibit between 20% to 50% homology on the amino acid level [53-57]. Over the last decade, 12 different C-X-C chemokines have been identified and include PF4, NH₂-terminal truncated forms of platelet basic protein (PBP; connective tissue-activating protein-III [CTAP-III], β -thromboglobulin [β TG], and neutrophil-activating protein-2 [NAP-2]), interleukin-8 (IL-8), growth-related oncogene (GRO α), GRO β , GRO γ , IFN- γ -inducible protein (IP-10), monokine induced by IFN- γ (MIG), epithelial neutrophil-activating protein-78 (ENA-78), and granulocyte chemotactic protein-2 (GCP-2) [53-60]. The NH₂-terminal truncated forms of PBP are generated when PBP is released from platelet α -granules and undergoes proteolytic cleavage by monocyte-derived proteases. PF4, the first member of the C-X-C chemokine family to be described, was originally identified for its ability to bind to heparin, leading to the inactivation of heparin's anticoagulation function [61]. Both IP-10 and MIG are

IFN-inducible C-X-C chemokines [58, 62]. Although IP-10 appears to be induced by all three interferons (IFN- α , IFN- β , and IFN- γ), MIG is unique in that it appears to be only expressed in the presence of IFN- γ [58]. Although IFN- γ induces the production of IP-10 and MIG, this cytokine has been found to attenuate the expression of both IL-8 and ENA-78 (63; and unpublished observations). These findings would suggest that members of the C-X-C chemokine family demonstrate disparate regulation in the presence of IFN- γ . GRO α , GRO β , and GRO γ and are closely related C-X-C chemokines, with GRO α originally described for its melanoma growth stimulatory activity [64-66]. IL-8, ENA-78, and GCP-2 were all initially identified on the basis of their ability to induce neutrophil activation and chemotaxis [53-60]. IL-8 has been found to be produced by an array of cells including monocytes, alveolar macrophages, neutrophils, keratinocytes, mesangial cells, epithelial cells, hepatocytes, fibroblasts, and endothelial cells [53-57, 67-80]. Interestingly, IL-8 is expressed in neoplasms and produced by a number of transformed neoplastic cells [72, 75, 81-83]. Although numerous investigations have shown both *in vivo* and *in vitro* the importance of IL-8 in acute inflammation, as a chemotactic/activating factor for neutrophils, only recently has it become apparent that this C-X-C chemokine may be important in angiogenesis associated with tumorigenesis.

ROLE OF C-X-C CHEMOKINES IN ANGIOGENESIS

Our laboratory and others have found that IL-8 is a potent angiogenic factor [47, 84, 85]. Recombinant IL-8 mediates both endothelial cell chemotactic and proliferative activity *in vitro* and angiogenic activity *in vivo*. We found that IL-8 induced similar angiogenic activity as basic fibroblast growth factor (bFGF) [47]. Because monocytes/macrophages may represent a major source of angiogenic activity in wounds, other chronic diseases, and solid tumors [13], we extended our studies to determine whether IL-8 was a predominant angiogenic factor liberated by activated human monocytes or by synovial macrophages isolated from rheumatoid arthritis synovial tissues [47]. Conditioned medium from both populations of mononuclear phagocytes induced significant angiogenic activity that was attributable to IL-8 [47]. To further demonstrate that the angiogenic effect was attributable to IL-8, we used an IL-8 antisense oligonucleotide strategy to inhibit the production of IL-8 at the pretranslational level [47]. The conditioned medium from monocytes treated in the presence of IL-8 antisense inhibited endothelial chemotactic activity by 84% compared with the IL-8 sense oligonucleotide-treated monocytes. Similar results were found in the *in vivo* corneal micropocket model of angiogenesis. These findings indicated that IL-8, at concentrations of approximately 1 nM, functioned as a mediator of angiogenesis. This amount of IL-8 compares with amounts reported for the induction of corneal angiogenic activity by tumor necrosis factor- α , aFGF, bFGF, angiotropin, angiotropin, and endothelial cell growth factor [47].

Another member of the C-X-C chemokine family, PF4, has been shown to have angiostatic properties in nanomolar to micromolar concentrations [44] and to attenuate the growth of tumors *in vivo* [86]. Interestingly, these studies had initially demonstrated that the angiostatic ac-

tivity of PF4 was due to its heparin binding domain (within the COOH-terminal of the molecule) [44, 86]. However, recent studies have now shown that a PF4 mutant that lacks the heparin binding domain, and functional heparin binding is equipotent *in vivo* to native PF4 for the attenuation of tumor growth [87]. The above findings would suggest that members of the C-X-C chemokine family function as either angiogenic or angiostatic factors in regulating neovascularization, and the biological balance in the magnitude of expression of these angiogenic and angiostatic C-X-C chemokines within a tumor could dictate overall tumor-derived angiogenic activity. Although it remains unclear whether the COOH-terminal of these chemokines dictates their biological role in angiogenesis, the differences in C-X-C chemokine function could also be explained by other structural domains. Recently, both Hébert et al. [88] and Clark-Lewis et al. [89] have demonstrated a salient amino acid sequence in the primary structure of the C-X-C chemokine family that appears, in part, to account for the ability of these chemokines to function in neutrophil chemotaxis and activation. They demonstrated that the three amino acid residues that immediately preceded the first cysteine amino acid are critically important in the binding and activation of neutrophils. These amino acids are Glu-Leu-Arg, the ELR motif, which is absent in certain members of the C-X-C chemokine family (PF4, IP-10, and MIG) that display markedly reduced potency in mediating neutrophil chemotaxis. Interestingly, when the ELR motif was introduced into PF4, this chemokine gained 1000-fold potency in mediating neutrophil chemotaxis [89]. Thus, these structural differences, in part, may explain the disparity of angiogenic activity of the C-X-C chemokine family and support the hypothesis that a biological imbalance in the expression of angiogenic and angiostatic C-X-C chemokines may lead to the perpetuation of neovascularization during tumor growth and metastasis.

ROLE OF IL-8 AS AN ANGIOGENIC FACTOR IN NSCLC

Previous studies have demonstrated that human tumors and neoplastic cell lines may directly elaborate IL-8 [72, 75, 81–83, 90–92]. Because angiogenesis is critical to tumorigenesis and metastasis, we extended our initial discovery of IL-8 as an angiogenic factor to assess whether this C-X-C chemokine was present in NSCLC and whether it contributed to overall tumor-derived angiogenic activity [93]. We found significantly elevated IL-8 levels in natural human NSCLC, by using a specific ELISA, that were four-fold greater than normal lung tissue [93]. Results from IL-8 immunohistochemistry of tumor sections confirmed a heterogeneous pattern of tumor cell production of IL-8 from both adenocarcinomas and squamous cell carcinomas of the lung [93]. The heterogeneity of the tumor cell expression was of interest, especially because we had previously shown that only 36% to 38% of adenocarcinoma cells (A549 cell line) expressed IL-8 [75]. The findings of heterogeneous expression of IL-8 by tumor cells suggests that specific subclones of neoplastic cells may exist and function as the primary cellular source of tumor-derived IL-8. Our results also revealed that nontransformed stromal cells within the host desmoplastic response to the tumor were also serving as significant cellular sources for IL-8, especially in response to the squamous cell carcinoma

[93]. Importantly, these specific findings may reflect the different clinical behavior of squamous cell and adenocarcinomas. The more aggressive course of adenocarcinomas could be related to their capacity to generate a sufficient angiogenic signal [IL-8], independent from the surrounding host-responding immune and nonimmune cells. The observation of a minimal inflammatory cell infiltrate in the tumor specimens, despite the presence of IL-8, was unexpected and supported the contention that IL-8 may have other biological functions in the context of NSCLC.

Because it was apparent that NSCLC was associated with significantly elevated levels of IL-8, we next determined whether IL-8 contributed to overall tumor-derived angiogenic activity. By using neutralizing antibodies to IL-8, we found that IL-8 accounted for 42% to 80% of the angiogenic activity for each of the tumor specimens, as determined by bioassays of angiogenesis [93]. The above data suggested that a significant portion of tumor-derived angiogenic activity was mediated directly by IL-8. Although IL-8-dependent angiogenic activity represented a significant proportion of overall NSCLC-derived angiogenesis, we wanted to compare the relative contribution of IL-8 to other known angiogenic factors in NSCLC. Neutralizing antibodies to IL-8 resulted in a significant reduction of endothelial cell chemotactic activity in response to NSCLC tissue, with a decline to 75%, 39%, and 61% of the standard bioactivity, respectively, for adenocarcinoma, squamous cell carcinoma, and A549 (adenocarcinoma) samples. In contrast, anti-bFGF antibodies had no significant effect on the endothelial cell chemotaxis in response to samples of A549 cells/tissue or squamous cell carcinoma tissue; however, neutralizing anti-bFGF antibodies reduced the endothelial cell chemotactic activity from adenocarcinoma tissue by 35% of the standard bioactivity. Interestingly, the neutralization of transforming growth factor- α had no significant effect on the chemotaxis in response to adenocarcinoma or to the A549 cell/tissue; however, these antibodies resulted in a significant reduction (45%) in the endothelial cell chemotactic response to squamous cell carcinoma tissue. Although bFGF and transforming growth factor- α have been previously described as potential angiogenic factors involved in tumor angiogenesis [4, 7, 92, 94, 95], these studies were the first to demonstrate that a primary angiogenic signal for NSCLC neovascularization was directly mediated by tumor-associated IL-8.

ROLE OF C-X-C CHEMOKINES IN MEDIATING ANGIOGENESIS IN THE CONTEXT OF TUMORIGENESIS: OUR HYPOTHETICAL MODEL

The experiments designed in our laboratories have focused on the role of angiogenic and angiostatic C-X-C chemokines and whether a biological imbalance in their expression favors tumor-derived angiogenic activity. We hypothesize that angiogenesis associated with NSCLC tumor growth is dependent on members of the C-X-C chemokine family acting as either angiogenic or angiostatic factors (Fig. 1). This paradigm predicts that a shift in the balance of expression of these C-X-C chemokines dictates whether the neoplasm grows and develops metastatic potential or regresses. The net angiogenic activity during the progression of tumorigenesis is mediated by the biological imbalance that favors the expression of

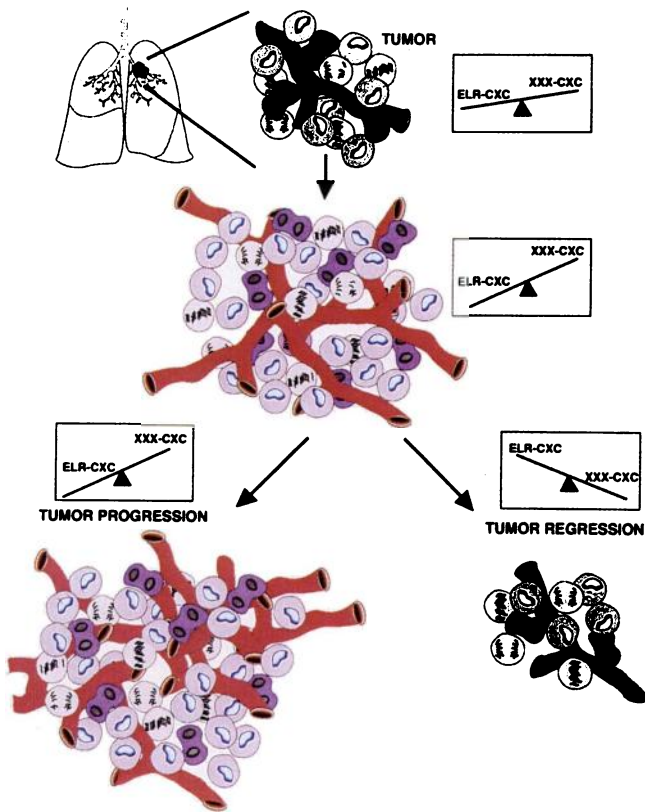


Fig. 1. The role of C-X-C chemokines in mediating angiogenesis in the context of tumor genesis: our hypothetical model. ELR, chemokines containing the ELR motif; XXX, chemokines that lack the ELR motif.

angiogenic C-X-C chemokines (ELR-C-X-C) compared with the angiostatic C-X-C chemokines (XXX-C-X-C). However, the magnitude of expression and the biological imbalance will favor the production of angiogenic C-X-C chemokines (ELR-C-X-C). This effect results in tumor growth, invasion, and metastasis beyond the confines of its primary site of origin. In contrast, the generation of IP-10 and MIG will have a negative effect on tumorigenesis through a reduction in tumor-derived angiogenic activity. These C-X-C chemokines are primarily induced by IFN- γ , which is known to be attenuated by IL-10. The production of IL-10 by both neoplastic cells and surrounding nontransformed immune and nonimmune cells will have a significant, yet indirect, impact on the generation of angiostatic (XXX-C-X-C) chemokines (IP-10 and MIG). Under these circumstances, IL-10 expression in the context of tumorigenesis can be viewed as an indirect promoter of angiogenesis via the down-regulation of IFN- γ . The experimental model of human NSCLC/SCID mouse chimera has provided the opportunity to test whether either attenuation of angiogenic (ELR-C-X-C) chemokines or accentuation of angiostatic (XXX-C-X-C) chemokines promotes tumor regression.

PRELIMINARY STUDIES

The major aims of our experiments were to study the role of C-X-C chemokines as they relate mechanistically to the pathophysiological consequences of tumorigenesis. The

following preliminary data demonstrates that specific members of the C-X-C chemokine family of chemotactic cytokines are either angiogenic or angiostatic mediators of neovascularization. In addition, these data support the notion that these C-X-C chemokines are likely candidates to target novel future specific therapies.

C-X-C CHEMOKINES (ELR MOTIF) INDUCE AND IP-10 AND PF4 (NON-ELR MOTIF) ATTENUATE ANGIOGENESIS

We postulated that the members of the C-X-C chemokine family may exert disparate effects in mediating angiogenesis for primarily four reasons. First, members of the C-X-C chemokine family that display binding and activation of neutrophils share the same ELR motif homology that immediately precedes the first cysteine amino acid residue, whereas, PF4, IP-10, and MIG lack this motif [88, 89]. Second, IL-8 (contains ELR motif) is angiogenic and PF4 (lacks the ELR motif) is angiostatic. Third, the IFNs (IFN- α , IFN- β , and IFN- γ) are all known inhibitors of wound repair, especially angiogenesis and reepithelialization [37, 92, 95-110], however, these cytokines up-regulate IP-10 and MIG from a number of cells, including keratinocytes, fibroblasts, endothelial cells, and mononuclear phagocytes [58, 62]. We hypothesize that IFNs may be exerting their negative influence in wound repair by up-regulating the production of IP-10 and MIG, which may act in an autocrine and paracrine manner to suppress angiogenesis. Finally, we and others have found that IFN- α , IFN- β , and IFN- γ are potent inhibitors of both monocyte-derived IL-8 and ENA-78 (63; and unpublished observations), supporting the notion that IFN- α , IFN- β , and IFN- γ may shift the biological balance of C-X-C chemokines toward a preponderance of angiostatic C-X-C chemokines. To test this postulate, we performed endothelial cell chemotaxis in the presence or absence of C-X-C chemokines that contain the ELR motif (IL-8, ENA-78, GCP-2, GRO α , GRO β , GRO γ , PBP, CTAP-III, and NAP-2) and C-X-C chemokines lacking the ELR motif (IP-10 and PF4). In a similar fashion to IL-8, all of the C-X-C chemokines that contained the ELR motif demonstrated significant endothelial cell chemotactic activity, whereas the endothelial cell chemotactic activity induced by either IP-10 or PF4 were similar to control (background). In addition, ENA-78, GCP-2, GRO α , GRO β , GRO γ , and NAP-2 in a similar concentration to IL-8 demonstrated significant corneal neovascularization without any evidence for inflammatory cellular infiltration.

To delineate whether IP-10 (C-X-C chemokine lacking the ELR motif) could modulate C-X-C chemokine (ELR motif present)-induced endothelial cell chemotactic activity, IL-8, GRO α , CTAP-III, and NAP-2 were assessed for endothelial cell chemotaxis in the presence of IP-10. The presence of IP-10 significantly attenuated endothelial cell chemotactic activity in response to C-X-C chemokines containing the ELR motif. In addition, PF4 in a similar fashion to IP-10 also attenuated C-X-C chemokine (ELR motif present)-induced endothelial cell chemotactic activity. Furthermore, by using the corneal micropocket model of neovascularization, IP-10 in equivalent molar concentrations was found to attenuate IL-8-, ENA-78-, GCP-2-, and GRO α -induced angiogenesis. These findings support the premise that members of the C-X-C chemokine family may be acting as either angiogenic or

TABLE 1. C-X-C Chemokines Display Disparate Angiogenic Activity

Angiogenic C-X-C chemokines
IL-8
ENA-78
GCP-2
GRO α
GRO β
GRO γ
CTAP-III
β TG
NAP-2
Angiostatic C-X-C chemokines
PF4
IP-10
MIG

angiostatic factors (Table 1). The net biological balance of these C-X-C chemokines, in the context of tumorigenesis, may play an important role in modulating NSCLC neovascularization.

PRESENCE OF C-X-C CHEMOKINES IN NSCLC

The evidence that C-X-C chemokines could function as either angiogenic or angiostatic factors, depending on the presence of the ELR motif, lead to our investigation as to whether C-X-C chemokines were present in natural human NSCLC and whether they contributed to overall NSCLC tumor-derived angiogenic activity. IL-8, ENA-78, and GRO α were found in a 4-, 3-, and 2.5-fold excess, respectively, in tumor tissue compared with normal lung tissue normalized to total protein (TP). Normal lung tissue contained 2.6 ± 0.7 , 15 ± 6 , and 6 ± 2.5 ng/mg TP of IL-8, ENA-78, and GRO α compared with 11.9 ± 3.8 , 43.6 ± 10 , and 11.5 ± 2.5 ng/mg TP for tumor specimens, respectively. There were similar elevations of IL-8 and ENA-78 from both adenocarcinomas and squamous cell carcinomas, whereas GRO α was found to be especially elevated in squamous cell (17.6 ± 7.8 ng/mg TP) carcinomas. In contrast, PF4 levels were found in the carcinoma tissue homogenates to be equivalent to normal lung tissue (592.4 ± 127.8 ng/mg TP and 579.1 ± 195.3 ng/mg TP for tumors and normal lung tissue, respectively). Although IP-10 levels trended toward being greater in tumors than in normal lung tissue homogenates, IP-10 was significantly lower in adenocarcinomas (0.9 ± 0.5 ng/mg TP) than in normal lung tissue (3.6 ± 2 ng/mg TP). Importantly, these specific findings may be reflected in the different clinical behaviors of squamous cell carcinoma and adenocarcinoma NSCLCs. The more aggressive course of adenocarcinomas could be related to their capacity to generate a greater angiogenic compared with angiostatic C-X-C chemokine signal. To further substantiate that IP-10 may be acting as an endogenous angiostatic C-X-C chemokine to balance the effect of angiogenic factors in the context of NSCLC, we assessed squamous cell carcinoma tissue homogenates for angiogenesis in the presence of neutralizing IP-10 or control antibodies. By using either endothelial cell chemotaxis or a corneal micropocket model of neovascularization, we found that neutralizing IP-10 antibodies (without evidence of lipopolysaccharide contamination) significantly aug-

mented tumor-derived angiogenic activity by twofold. These findings support the presence of an imbalance in ELR-C-X-C compared with XXX-C-X-C chemokines, with the balance favoring a greater presence of angiogenic C-X-C chemokines in NSCLC tumors.

HUMAN NSCLC/SEVERE COMBINED IMMUNODEFICIENT (SCID) MOUSE CHIMERA AS A MODEL FOR HUMAN TUMORIGENESIS

Although neoplastic transformation is dependent on multiple genetic and epigenetic events [5], the success of tumorigenesis is dependent on the complex biological interplay between the neoplastic cells and the resident and recruited host-responding cells. For example, in the absence of local neovascularization these neoplasms could not grow beyond 1–2 mm³ [6–8]. Thus, to effectively study the complex biology of human solid tumors, the use of human tumor xenografts in immunodeficient mice have provided significant insight into the biology of tumor growth and metastasis [111–120]. Our preliminary data supports the notion that specific members of the C-X-C chemokine family are important cytokines involved in orchestrating a significant portion of the angiogenic response during tumorigenesis and that other members may be important in attenuating this response. To characterize the qualitative and quantitative presence of these angiogenic and angiostatic C-X-C chemokines, as well as determine their net contribution to human NSCLC tumorigenesis in vivo, we have transplanted either intact natural human NSCLC or NSCLC cell lines (A549, adenocarcinoma and Calu-1, squamous cell carcinoma) into SCID mice to create a human NSCLC/SCID mouse chimera. The mice were evaluated before xenograft to have a murine serum Ig concentration of <1 μ g/ml by ELISA. Freshly isolated intact NSCLC (adenocarcinoma) were cut into 1 mm³ and placed subcutaneously into the bilateral flanks of SCID mice and allowed to grow for 4 weeks. At this time point the tumors measured 4–9 mm in diameter and the mice were killed. Immunohistochemistry for IL-8 and ENA-78 demonstrated the immunolocalization of these C-X-C chemokines within the viable NSCLC cells, stromal cells, and mononuclear cells, whereas the control antibodies failed to show nonspecific staining. These findings suggested that natural human NSCLC remained viable in SCID mice and that they continue to express both IL-8 and ENA-78 that may have allowed them to generate angiogenic activity and survive in the SCID mouse host.

Because our central hypothesis is that overall tumor-derived angiogenesis is dependent on an imbalance in the production of members of the C-X-C chemokine family that act as angiogenic rather than angiostatic factors, we next assessed in vitro whether human NSCLC cell lines (A549 and Calu-1) produced disparate levels of angiogenic (IL-8, ENA-78, and GRO α) and angiostatic (PF4 and IP-10) C-X-C chemokines. Both A549 and Calu-1 human NSCLC cell lines were found to constitutively produce high levels of angiogenic C-X-C chemokines. Interestingly, the A549 cells were found to constitutively produce greater levels of angiogenic C-X-C chemokines, and when grown in SCID for 4 weeks, were found to induce tumors twofold greater in size than the Calu-1 NSCLC cell line. These findings corroborate the previous findings that A549 cells display greater tumorigenicity and spontaneous metastasis than Calu-1 cells in vivo [119–121]. Although

these *in vitro* studies were important to establish whether an imbalance in the production of angiogenic compared with angiostatic C-X-C chemokines existed in NSCLC cell lines, we next assessed whether an imbalance in their production occurred *in vivo* during tumorigenesis of A549 NSCLC cells in a SCID mouse host.

SCID mice received 10^6 A549 cells suspended in 100 μ l of phosphate-buffered saline to each flank. The mice were killed in a time-dependent manner at 2, 3, 4, 5, 6, and 7 weeks postxenograft and tumors were analyzed for tumor growth, and tumor-derived and plasma levels of C-X-C chemokines standardized to either TP or volume (biopsy). Immunolocalization of IL-8 from A549 tumors at week 7 demonstrated a heterogeneous expression pattern with $35 \pm 6\%$ of the cells expressing IL-8 protein by image analysis, whereas IL-8 expression from spontaneous A549 tumor metastasis to the lungs of SCID mice at 7–8 weeks demonstrated a more homogenous pattern of immunolocalization of IL-8, with more than 80% of the cells expressing IL-8 protein by image analysis. Moreover, when A549 tumors in SCID mice and plasma from these mice were analyzed for the presence of C-X-C chemokines, we found a significant correlation of the temporal expression of angiogenic compared with angiostatic C-X-C chemokines during tumorigenesis. These studies substantiated that the production of angiogenic C-X-C chemokines paralleled tumor growth and supported our contention that an imbalance in angiogenic and angiostatic C-X-C chemokines exist during tumorigenesis. However, these studies did not demonstrate whether these angiogenic C-X-C chemokines directly contributed to an imbalance favoring net angiogenesis leading to tumor growth or tumor survival. To assess whether IL-8 directly contributed to tumor growth and survival, human NSCLC (A549) tumorigenesis was allowed to occur in SCID for a period of 4 weeks before the initiation of *in vivo* passive immunization with neutralizing antibodies to IL-8 (every other day during weeks 5, 6, and 7 of tumor growth). We found the tumors (at 8 weeks) to demonstrate markedly more central necrosis in animals that had received neutralizing antibodies to IL-8 compared with animals receiving control antibodies. In addition, morphological analysis, thresholded to NSCLC cell nuclear size, showed a significant reduction in cellularity of the tumors that were exposed to neutralizing IL-8 antibodies. The above findings provide evidence that supports our contention that the ELR motif containing C-X-C chemokines are important angiogenic molecules during tumorigenesis, and that an imbalance in their expression compared with angiostatic C-X-C chemokines, dictates their role in mediating net tumor-derived angiogenesis during tumorigenesis. Furthermore, the use of the human NSCLC/SCID mouse model has allowed us to validate our central hypothesis and examine the biology of C-X-C chemokines as they relate to angiogenesis *in vivo*. This model of human tumorigenesis has allowed the unique opportunity to examine the temporal magnitude, cellular sources, and contribution of the angiogenic and angiostatic C-X-C chemokines to the evolution of tumorigenesis and spontaneous metastasis.

CYTOKINE NETWORKS THAT MAY PROMOTE ANGIOGENIC C-X-C CHEMOKINES

The above preliminary data coupled with what is known regarding the role of IFNs in wound repair, suggest that

the magnitude of IFN expression would be a pivotal event in locally regulating both angiogenic (through negative feedback) and angiostatic (through positive feedback) C-X-C chemokine production. IFNs are pleiotropic cytokines that exert a broad range of immunomodulatory and inflammatory effects, however, they share a salient feature of cellular growth inhibition. IFN- α and IFN- β are produced primarily by mononuclear phagocytes and fibroblasts, respectively, whereas IFN- γ is produced by CD4⁺ and CD8⁺ T cells and natural killer cells [109, 110]. These IFNs are all known inhibitors of wound repair, specifically they appear to have a significant inhibitory influence on endothelial cell biology [37, 90, 95, 102–110]. This supports the notion that IFNs can modulate suppression of neovascularization through both direct and indirect pathways. Although IFN- α , IFN- β , and IFN- γ can inhibit the cellular expression of angiogenic C-X-C chemokines, these IFNs are important for the induction of angiostatic C-X-C chemokines (IP-10 and MIG). The net effect of IFN stimulation would be to shift the biological balance in favor of angiostatic C-X-C chemokines. This potential mechanism could be operative during the later stages of normal wound repair, where new capillary blood vessel formation is rapid, under strict control, and undergoes marked regression to a physiological steady-state level. In contrast, angiogenesis during neoplastic transformation is exaggerated [7, 8]. This perpetual neovascularization during tumorigenesis suggests two possible pathological mechanisms, neither of which are mutually exclusive: (1) tumorigenesis is associated with an increase in the synthesis and/or elaboration of angiogenic mediators and (2) tumorigenesis is associated with reduced levels of substances that inhibit neovascularization. Because our preliminary data supported the former, we also postulated that these neoplasms may be associated with equivalent or reduced levels of IFN- γ compared with normal lung tissue, which would support the latter. To test this hypothesis, we analyzed human tissue homogenates of both normal and NSCLC for the presence of IFN- γ by using an IFN- γ specific ELISA. IFN- γ concentrations from tumor samples were found to be similar to normal lung tissue. These findings corroborated the study by Vitolo et al. [122] who found that cytokine expression of mRNA (TH-1 profile) from tumor-infiltrating lymphocytes was unexpectedly reduced within solid tumors compared with other chronic inflammatory disorders. The above findings may reflect the reduced immunogenicity of tumor-associated antigens, however, an alternative explanation may be the ability of the tumor or host-responding cells to produce immunosuppressive factors that may directly impact on the local production of IFN- γ .

IL-10 is a recently characterized cytokine that demonstrates varied immunosuppressive bioactivity. Since its initial isolation by Mosmann et al. [123–126] in 1988, investigations have elucidated many of the immunological properties of this cytokine. Originally identified as a product of CD4⁺ T cells, IL-10 is also produced by monocytes, macrophages, B cells, certain populations of CD8⁺ T cells, and Epstein-Barr virus-transformed lymphoblastoid cells lines [123, 124, 126–130]. Recent work has demonstrated that epidermal cells may also elaborate IL-10 [131]. Functional studies reveal that IL-10 has profound effects on monocytes, resulting in alterations in cell morphology and cytotoxicity, down-regulation of the expression of major histocompatibility complex class II antigens, and

inhibition of proinflammatory cytokine production [132-137]. Furthermore, IL-10 also exerts direct effects on the growth and function of T cells, B cells, and mast cells [123, 124]. These specific actions result in the capacity for IL-10 to attenuate a wide range of effector immune responses, including T cell cytokine (i.e., IFN- γ) production and antigen-specific proliferation, B cell Ig synthesis, and the elaboration of tumor necrosis factor- α and IFN- γ by natural killer cells [123, 124]. IL-10 may play an important role in homeostasis under normal circumstances; however, IL-10, in the context of tumorigenesis, may be a major immunosuppressive factor that attenuates the local generation of IFN- γ . To test this premise, we investigated the capacity of NSCLC to produce IL-10 [138]. We found increased levels of antigenic IL-10 in tissue homogenates of NSCLC (13.7 ± 2.8 ng/mg TP) compared with normal lung tissue (5.8 ± 0.8 ng/mg TP). No significant difference in IL-10 levels were seen between the squamous cell carcinoma (15.4 ± 3.7 ng/mg TP) and adenocarcinoma (12.3 ± 4.2 ng/mg TP). To determine the cellular source of IL-10 in the NSCLCs, immunohistochemical staining of NSCLC showed primary localization of antigenic IL-10 to individual NSCLC cells [138]. In addition, immunolocalization using HAM56, (Enzo Diagnostics, Inc., Farmingdale, NY), a murine monoclonal antibody against human mononuclear cells, confirmed that tumor cells, rather than immune cells, were the primary cellular source of antigenic IL-10 [138]. Because IL-10 had previously been demonstrated to be produced by epidermal cells [131], we analyzed the conditioned medium of several unstimulated human NSCLC cell lines (A549, A427, and Calu-6) for the constitutive production of IL-10. These cells produced 6.3 ± 1 ng/ml, 1.9 ± 0.9 ng/ml, and 7.6 ± 1.1 ng/ml of IL-10 after 24 h of culture, respectively [138]. These findings demonstrate that NSCLC cell lines can elaborate IL-10. Thus, IL-10 may play an important role in impairing immune cell effector function and enable the NSCLC to evade host immune defenses; however, IL-10 suppression of IFN- γ may play an equally important role in perpetuating tumor-associated neovascularization.

Our data demonstrates and supports the potential presence of an imbalance in the expression of angiogenic and angiostatic C-X-C chemokines during tumorigenesis that favors the tumor-derived angiogenesis. In addition, the presence of augmented levels of IL-10 in NSCLC may favor the notion that IL-10 may promote angiogenesis via a direct role in attenuating IFN- γ and therefore, indirectly inhibiting angiostatic C-X-C chemokines during tumorigenesis. The further elucidation of the biology of these cytokines will provide new insight into the specific aspects of C-X-C chemokine biology responsible for NSCLC angiogenesis and subsequent tumorigenesis and metastasis.

ACKNOWLEDGMENTS

Supported in part by National Institutes of Health grants HL50057, CA66180, HL02401, and 1P50HL46487 (R.M.S.), HL39926 (P.J.P.), and HL31693 and HL35276 (S.L.K.); and the A.S.L.K. Cancer Foundation, Belgium (G.O. and J.V.D.).

REFERENCES

- Garfinkel, A. (1991) Cancer statistics and trends. In *American Cancer Society Textbook of Clinical Oncology* (A.I. Holleb, D.J. Fink, G.P. Murphy, eds.), American Cancer Society, Atlanta, GA, 2-9.
- Faber, L.P. (1991) Lung cancer. In *American Cancer Society Textbook of Clinical Oncology* (A.I. Holleb, D.J. Fink, G.P. Murphy, eds.), American Cancer Society, Atlanta, GA, 194-212.
- Brown, C.C., Kessler, L.G. (1988) Projections of lung cancer mortality in the United States: 1985-2025. *J. Natl. Cancer Inst.* **80**, 43-51.
- Whalen, G.F. (1990) Solid tumours and wounds: transformed cells misunderstood as injured tissue? *Lancet* **336**, 1489-1492.
- Shields, P.G., Harris, C.C. (1993) Genetic predisposition to cancer. In *Lung Cancer* (J.A. Roth, J.D. Cox, W.K. Hong, eds.), Blackwell Scientific, Boston, MA, 3-19.
- Folkman, J., Cotran, R. (1976) Relation of vascular proliferation to tumor growth. *Int. Rev. Exp. Pathol.* **16**, 207-248.
- Folkman, J. (1985) Tumor angiogenesis. *Adv. Cancer Res.* **43**, 175-203.
- Bouck, N. (1990) Tumor angiogenesis: the role of oncogenes and tumor suppressor genes. *Cancer Cells* **2**, 179-185.
- Atherton, A. (1977) Growth stimulation of endothelial cells by simultaneous culture with sarcoma 180 cells in diffusion chambers. *Cancer Res.* **37**, 3619-3622.
- Fenselau, A., Mello, R.J. (1976) Growth stimulation of cultured endothelial cells by tumor cell homogenates. *Cancer Res.* **36**, 3269-3273.
- Zetter, B.R. (1980) Migration of capillary endothelial cells is stimulated by tumour-derived factors. *Nature* **285**, 41-43.
- Auerbach, R. (1981) Angiogenesis-inducing factors: a review. *Lymphokines*, Vol. 4, Academic Press, New York, 69-88.
- Polverini, P.J. Macrophage-induced angiogenesis: a review. *Cytokines*, Vol. 1, S. Karger, Basel, 54-73.
- Folkman, J., Klagsbrun, M. (1987) Angiogenic factors. *Science* **235**, 442-447.
- Leibovich, S.J., Weisman, D.M. (1988) Macrophages, wound repair and angiogenesis. *Prog. Clin. Biol. Res.* **266**, 131-145.
- Engerman, R.L., Pfaffenbach, D., Davis, M.D. (1967) Cell turnover of capillaries. *Lab. Invest.* **17**, 738-743.
- Tannock, I.F., Hayashi, S. (1972) The proliferation of capillary and endothelial cells. *Cancer Res.* **32**, 77-82.
- Folkman, J., Watson, K., Ingber, D., Hanahan, D. (1989) Induction of angiogenesis during the transition from hyperplasia to neoplasia. *Nature* **339**, 58-61.
- Maiorana, A., Gullino, P.M. (1978) Acquisition of angiogenic capacity and neoplastic transformation in the rat mammary gland. *Cancer Res.* **38**, 4409-4414.
- Herlyn, M., Clark, W.H., Rodeck, U., Mancianti, M.L., Jambrosic, J., Koprowski, H. (1987) Biology of tumor progression in human melanocytes. *Lab. Invest.* **56**, 461-474.
- Weidner, N., Semple, J.P., Welch, W.R., Folkman, J. (1991) Tumor angiogenesis and metastasis-correlation in invasive breast carcinoma. *N. Eng. J. Med.* **324**, 1-8.
- Weidner, N., Carroll, P.R., Flax, J., Blumenfeld, W., Folkman, J. (1993) Tumor angiogenesis correlates with metastasis in invasive prostate carcinoma. *Am. J. Pathol.* **143**, 401-409.
- Macchiarini, P., Fontanini, G., Hardin, M.J., Squartini, F., Angeletti, C.A. (1992) Relation of neovascularization to metastasis of non-small cell lung cancer. *Lancet* **340**, 145-146.
- Eisenstein, R., Kuettner, K.E., Neopolitan, C., Sobel, L.W., Sorgente, N. (1975) The resistance of certain tissues to invasion. III. Cartilage extracts inhibit the growth of fibroblasts and endothelial cells in culture. *Am. J. Pathol.* **81**, 337-347.
- Sorgente, N., Kuettner, K.E., Sobel, L.W., Eisenstein, R. (1975) The resistance of certain tissues to invasion. II. Evidence for extractable factors in cartilage which inhibit invasion by vascularized mesenchyme. *Lab. Invest.* **32**, 217-222.
- Brem, H., Folkman, J. (1975) Inhibition of tumor angiogenesis mediated by cartilage. *J. Exp. Med.* **141**, 427-439.
- Lee, A., Langer, R. (1983) Shark cartilage contains inhibitors of tumor angiogenesis. *Science* **221**, 1185-1187.
- Langer, R., Conn, H., Vacanti, J., Haudenschild, C.C., Folkman, J. (1980) Control of tumor growth in animals by infusion of an antiangiogenesis inhibitor. *Proc. Natl. Acad. Sci. USA* **77**, 4331-4335.
- Brem, S., Preis, I., Langer, R., Brem, H., Folkman, J., Patz, A. (1977) Inhibition of neovascularization by an extract derived from vitreous. *Am. J. Ophthalmol.* **84**, 323-328.
- Lutty, G.A., Thompson, D.C., Gallup, J.Y., Mello, R.J., Fenselau, A. (1983) Vitreous: an inhibitor of retinal extract-induced neovascularization. *Inv. Ophthalmol. Visual Sci.* **24**, 52-56.

31. Madri, J.A., Pratt, B.M., Tucker, A.M. (1988) Phenotypic modulation of endothelial cells by transforming growth factor-beta depends upon the composition and organization of the extracellular matrix. *J. Cell Biol.* **106**, 1375-1384.
32. Ingber, D.E., Madri, J.A., Folkman, J. (1986) A possible mechanism for inhibition of angiogenesis by angiostatic steroids: induction of capillary basement membrane dissolution. *Endocrinology* **119**, 1768-1775.
33. Ingber, D.E., Folkman, J. (1988) Inhibition of angiogenesis through modulation of collagen metabolism. *Lab. Invest.* **59**, 44-51.
34. Maragoudakis, M.E., Sarmonika, M., Panoutsacopoulou, M. (1988) Inhibition of basement membrane biosynthesis prevents angiogenesis. *J. Pharmacol. Exp. Ther.* **244**, 729-733.
35. Shapiro, R., Vallee, B.L. (1987) Human placental ribonuclease inhibitor abolishes both angiogenic and ribonucleolytic activities of angiogenin. *Proc. Natl. Acad. Sci. USA* **84**, 2238-2241.
36. Sato, N., Fukuda, K., Nariuchi, H., Sagara, N. (1987) Tumor necrosis factor inhibits angiogenesis in vitro. *J. Natl. Cancer Inst.* **79**, 1383-1391.
37. Sidky, Y.A., Borden, E.C. (1987) Inhibition of angiogenesis by interferons: effects on tumor- and lymphocyte-induced vascular responses. *Cancer Res.* **47**, 5155-5161.
38. Peterson, H.-I. (1986) Tumor angiogenesis inhibition by prostaglandin synthetase inhibitors. *Anticancer Res.* **6**, 251-254.
39. Homandberg, G.A., Kramer-Bjerke, J., Grant, D., Christianson, G., Eisenstein, R. (1986) Heparin-binding fragments of fibronectin are potent inhibitors of endothelial cell growth: structure and function correlates. *Biochem. Biophys. Acta* **874**, 61-71.
40. Taylor, S., Folkman, J. (1982) Protamine is an inhibitor of angiogenesis. *Nature* **297**, 307-312.
41. Crum, R., Szabo, S., Folkman, J. (1985) A new class of steroids inhibits angiogenesis in the presence of heparin or a heparin fragment. *Science* **230**, 1375-1378.
42. Polverini, P.J., Novak, R.F. (1986) Inhibition of angiogenesis by the antineoplastic agents mitoxantrone and bisantrene. *Biochem. Biophys. Res. Commun.* **140**, 901-907.
43. Lee, K., Erturk, E., Mayer, R., Cockett, A.T.K. (1987) Efficacy of antitumor chemotherapy in C3H mice enhanced by the antiangiogenesis steroid, cortisone acetate. *Cancer Res.* **47**, 5021-5024.
44. Maione, T.E., Gray, G.S., Petro, J., Hunt, A.J., Donner, A.L., Bauer, S.I., Carson, H.F., Sharpe, R.J. (1990) Inhibition of angiogenesis by recombinant human platelet factor-4. *Science* **247**, 77-79.
45. Harris, Jr., E.D. (1976) Recent insights into the pathogenesis of the proliferative lesion in rheumatoid arthritis. *Arthritis Rheum.* **19**, 68-72.
46. Koch, A.E., Polverini, P.J., Leibovich, S.J. (1986) Stimulation of neovascularization by human rheumatoid synovial tissue macrophages. *Arthritis Rheum.* **29**, 471-479.
47. Koch, A.E., Polverini, P.J., Kunkel, S.L., Harlow, L.A., DiPietro, L.A., Elner, V.M., Elner, S.G., Strieter, R.M. (1992) Interleukin-8 (IL-8) as a macrophage-derived mediator of angiogenesis. *Science* **258**, 1798-1801.
48. Koch, A.E., Litvak, M.A., Burrows, J.C., Polverini, P.J. (1992) Decreased monocyte-mediated angiogenesis in scleroderma. *Clin. Immunol. Immunopathol.* **64**, 153-160.
49. Rastinejad, F., Polverini, P.J., Bouck, N.P. (1989) Regulation of the activity of a new inhibitor of angiogenesis by a cancer suppresser gene. *Cell* **56**, 345-355.
50. Good, D.J., Polverini, P.J., Rastinejad, F., LeBeau, M.M., Lemons, R.S., Frazier, W.A., Bouck, N.P. (1990) A tumor suppresser-dependent inhibitor of angiogenesis is immunologically and functionally indistinguishable from a fragment of thrombospondin. *Proc. Natl. Acad. Sci. USA* **87**, 6624-6428.
51. Tolsma, S.S., Volpert, O.V., Good, D.J., Frazier, W.A., Polverini, P.J., Bouck, N. (1993) Peptides derives from two separate domains of the matrix protein thrombospondin-1 have antiangiogenic activity. *J. Cell Biol.* **122**, 497-511.
52. DiPietro, L.A., Polverini, P.J. (1993) Angiogenic macrophages produce the angiogenic inhibitor thrombospondin 1. *Am. J. Pathol.* **143**, 678-684.
53. Baggiolini, M., Dewald, B., Walz, A. (1992) Interleukin-8 and related chemotactic cytokines. In *Inflammation: Basic Principles and Clinical Correlates* (J.I. Gallin, I.M. Goldstein, R. Snyderman, eds.), Raven Press, Ltd., New York.
54. Baggiolini, M., Walz, A., Kunkel, S.L. (1989) Neutrophil-activating peptide-1/interleukin 8, a novel cytokine that activates neutrophils. *J. Clin. Invest.* **84**, 1045-1049.
55. Matsushima, K., Oppenheim, J.J. (1989) Interleukin 8 and MCAF: novel inflammatory cytokines inducible by IL-1 and TNF. *Cytokine* **1**, 2-13.
56. Oppenheim, J.J., Zachariae, O.C., Mukaida, N., Matsushima, K. (1991) Properties of the novel proinflammatory supergene "intercrine" cytokine family. *Annu. Rev. Immunol.* **9**, 617-648.
57. Miller, M.D., Krangel, M.S. (1992) Biology and biochemistry of the chemokines: a family of chemotactic and inflammatory cytokines. *Crit. Rev. Immunol.* **12**, 17-46.
58. Farber, J.M. (1993) HuMIG: a new member of the chemokine family of cytokines. *Biochem. Biophys. Res. Commun.* **192**, 223-230.
59. Proost, P., De Wolf-Peters, C., Conings, R., Opdenakker, G., Billiau, A., Van Damme, J. (1993) Identification of a novel granulocyte chemotactic protein (GCP-1) from human tumor cells: in vitro and in vivo comparison with natural forms of GRO α , IP-10, and IL-8. *J. Immunol.* **150**, 1000-1010.
60. Walz, A., Burgener, R., Car, B., Baggiolini, M., Kunkel, S.L., Strieter, R.M. (1991) Structure and neutrophil-activating properties of a novel inflammatory peptide (ENA-78) with homology to interleukin-8. *J. Exp. Med.* **174**, 1355-1362.
61. Deutsch, E., Kain, W. (1961) Studies on platelet factor 4. In *Blood Platelets* (S.A. Jonson, R.W. Monto, J.W. Rebusck, R.C. Horn, eds.), Little-Brown, Boston, MA, 337.
62. Kaplan, G., Luster, A.D., Hancock, G., Cohn, Z. (1987) The expression of a ginterferon-induced protein (IP-10) in delayed immune responses in human skin. *J. Exp. Med.* **166**, 1098-1108.
63. Gusella, G.L., Musso, T., Bosco, M.C., Espinoza-Delgado, I., Matsushima, K., Varesio, L. (1993) IL-2 up-regulates but IFN-g suppresses IL-8 expression in human monocytes. *J. Immunol.* **151**, 2725-2732.
64. Anisowicz, A., Zajchowski, D., Stenman, G., Sager, R. (1988) Functional diversity of GRO gene expression in human fibroblasts and mammary epithelial cells. *Proc. Natl. Acad. Sci. USA* **85**, 9645-9649.
65. Anisowicz, A., Bardwell, L., Sager, R. (1987) Constitutive overexpression of a growth-regulated gene in transformed Chinese hamster and human cells. *Proc. Natl. Acad. Sci. USA* **84**, 7188-7192.
66. Richmond, A., Thomas, H.G. (1988) Melanoma growth stimulatory activity: isolation from human melanoma tumors and characterization of tissue distribution. *J. Cell. Biochem.* **36**, 185-198.
67. Yoshimura, T., Matsushima, K., Oppenheim, J.J., Leonard, E.J. (1987) Neutrophil chemotactic factor produced by LPS-stimulated human blood mononuclear leukocytes: partial characterization and separation from interleukin-1. *J. Immunol.* **139**, 788-794.
68. Matsushima, K., Morishita, K., Yoshimura, T., Lavu, S., Obayashi, Y., Lew, W., Appella, E., Kung, H.F., Leonard, E.J., Oppenheim, J.J. (1988) Molecular cloning of a human monocyte-derived neutrophil chemotactic factor (MDNCF) and the induction of MDNCF mRNA by interleukin-1 and tumor necrosis factor. *J. Exp. Med.* **167**, 1883-1893.
69. Strieter, R.M., Kunkel, S.L., Showell, H.J., Marks, R.M. (1988) Monokine-induced gene expression of human endothelial cell-derived neutrophil chemotactic factor. *Biochem. Biophys. Res. Commun.* **156**, 1340-1345.
70. Strieter, R.M., Kunkel, S.L., Showell, H., Remick, D.G., Phan, S.H., Ward, P.A., Marks, R.M. (1989) Endothelial cell gene expression of a neutrophil chemotactic factor by TNF- α , LPS, and IL-1 β . *Science* **243**, 1467-1469.
71. Strieter, R.M., Phan, S.H., Showell, H.J., Remick, D.G., Lynch, J.P., Genard, M., Raiford, C., Eskandari, M., Marks, R.M., Kunkel, S.L. (1989) Monokine-induced neutrophil chemotactic factor gene expression in human fibroblasts. *J. Biol. Chem.* **264**, 10621-10626.
72. Thornton, A.J., Strieter, R.M., Lindley, I., Baggiolini, M., Kunkel, S.L. (1990) Cytokine-induced gene expression of a neutrophil chemotactic factor/interleukin-8 by human hepatocytes. *J. Immunol.* **144**, 2609-2613.
73. Elner, V.M., Strieter, R.M., Elner, S.G., Baggiolini, M., Lindley, I., Kunkel, S.L. (1990) Neutrophil chemotactic factor (IL-8) gene expression by cytokine-treated retinal pigment epithelial cells. *Am. J. Pathol.* **136**, 745-750.
74. Strieter, R.M., Chensue, S.W., Basha, M.A., Standiford, T.J., Lynch, J.P., Kunkel, S.L. (1990) Human alveolar macrophage gene expression of interleukin-8 by TNF- α , LPS and IL-1 β . *Am. J. Respir. Cell. Mol. Biol.* **2**, 321-326.
75. Standiford, T.J., Kunkel, S.L., Basha, M.A., Chensue, S.W., Lynch, J.P., Toews, G.B., Strieter, R.M. (1990) Interleukin-8 gene expression by a pulmonary epithelial cell line: a model for cytokine networks in the lung. *J. Clin. Invest.* **86**, 1945-1953.
76. Strieter, R.M., Kasahara, K., Allen, R., Showell, H.J., Standiford, T.J., Kunkel, S.L. (1990) Human neutrophils exhibit disparate chemotactic factor gene expression. *Biochem. Biophys. Res. Commun.* **173**, 725-730.
77. Brown, Z., Strieter, R.M., Chensue, S.W., Ceska, P., Lindley, I., Niell, G.H., Kunkel, S.L., Westwick, J. (1991) Cytokine activated human

- mesangial cells generate the neutrophil chemoattractant–interleukin 8. *Kidney Int.* **40**, 86–90.
78. Rolfe, M.W., Kunkel, S.L., Standiford, T.J., Chensue, S.W., Allen, R.M., Evanoff, H.L., Phan, S.H., Strieter, R.M. (1991) Pulmonary fibroblast expression of interleukin-8: a model for alveolar macrophage-derived cytokine networking. *Am. J. Respir. Cell. Mol. Biol.* **5**, 493–501.
 79. Nickoloff, B.J., Karabin, G.D., Barker, J.N.W.N., Giffiths, C.E.M., Sarma, V., Mitra, R.S., Elder, J.T., Kunkel, S.L., Dixit, V.M. (1991) Cellular localization of interleukin-8 and its inducer, tumor necrosis factor-alpha in psoriasis. *Am. J. Pathol.* **138**, 129–140.
 80. Strieter, R.M., Kasahara, K., Allen, R.M., Standiford, T.J., Rolfe, M.W., Becker, F.S., Chensue, S.W., Kunkel, S.L. (1992) Cytokine-induced neutrophil-derived interleukin-8. *Am. J. Pathol.* **141**, 397–407.
 81. Hotta, K., Hayashi, K., Ishikawa, J., Tagawa, M., Hashimoto, K., Mizuno, S., Suzuki, K. (1990) Coding region structure of interleukin-8 gene of human lung giant cell carcinoma LU65C cells that produce LUCT/interleukin-8: homogeneity in interleukin-8 genes. *Immunol. Lett.* **24**, 165–169.
 82. VanMeir, E., Ceska, M., Effenberger, F., Walz, A., Grouzmann, E., Desbaillets, I., Frei, K., Fontana, A., deTribolet, N. (1992) Interleukin-8 is produced in neoplastic and infectious diseases of the human central nervous system. *Cancer Res.* **52**, 4297–4305.
 83. Abruzzo, L.V., Thornton, A.J., Liebert, M., Grossman, H.B., Evanoff, H., Westwick, J., Strieter, R.M., Kunkel, S.L. (1992) Cytokine-induced gene expression of interleukin-8 in human transitional cell carcinomas and renal cell carcinomas. *Am. J. Pathol.* **140**, 365–373.
 84. Strieter, R.M., Kunkel, S.L., Elnor, V.M., Martonyi, C.L., Koch, A.E., Polverini, P.J., Elnor, S.G. (1992) Interleukin-8: a corneal factor that induces neovascularization. *Am. J. Pathol.* **141**, 1279–1284.
 85. Hu, D.E., Hori, Y., Fan, T.P.D. (1993) Interleukin-8 stimulates angiogenesis in rats. *Inflammation* **17**, 135–143.
 86. Sharpe, R.J., Byers, H.R., Scott, C.F., Bauer, S.I., Maione, T.E. (1990) Growth inhibition of murine melanoma and human colon carcinoma by recombinant human platelet factor 4. *J. Natl. Cancer Inst.* **82**, 848–853.
 87. Maione, T.E., Gray, G.S., Hunt, A.J., Sharpe, R.J. (1991) Inhibition of tumor growth in mice by an analogue of platelet factor 4 that lacks affinity for heparin and retains potent angiostatic activity. *Cancer Res.* **51**, 2077–2083.
 88. Hebert, C.A., Vitangcol, R.V., Baker, J.B. (1991) Scanning mutagenesis of interleukin-8 identifies a cluster of residues required for receptor binding. *J. Biol. Chem.* **266**, 18989–18994.
 89. Clark-Lewis, I., Dewald, B., Geiser, T., Moser, B., Baggiolini, M. (1993) Platelet factor 4 binds to interleukin 8 receptors and activates neutrophils when its N terminus is modified with Glu-Leu-Arg. *Proc. Natl. Acad. Sci. USA* **90**, 3574–3577.
 90. Kasahara, T., Mukaido, N., Yamashita, K., Yagisawa, H., Akahoshi, T., Matsushima, K. (1991) IL-1 and TNF-alpha induction of IL-8 and monocyte chemotactic and activating factor (MCAF) mRNA expression in a human astrocytoma cell line. *Immunology* **74**, 60–67.
 91. Kaashoek, J.G., Mout, R., Falkenburg, J.H., Willemze, R., Fibbe, W.E., Landegent, J.E. (1991) Cytokine production by the bladder carcinoma cell line 5637: rapid analysis of mRNA expression levels using a cDNA-PCR procedure. *Lymphokine Cytokine Res.* **10**, 231–235.
 92. Zetter, B.R. (1988) Angiogenesis: state of the art. *Chest* **93**, 159S–166S.
 93. Smith, D.R., Polverini, P.J., Kunkel, S.L., Orringer, M.B., Whyte, R.I., Burdick, M.D., Wilke, C.A., Strieter, R.M. (1994) IL-8 mediated angiogenesis in human bronchogenic carcinoma. *J. Exp. Med.* **179**, 1409–1415.
 94. Schwigerer, L. (1988) Basic fibroblast growth factor and its relation to angiogenesis in normal and neoplastic tissue. *Klin. Wochenschr.* **66**, 340–345.
 95. McKay, I.A., Leigh, I.M. (1991) Epidermal cytokines and their role in cutaneous wound healing. *Br. J. Dermatol.* **124**, 513–518.
 96. Clark, R.A.F. (1993) Basics of cutaneous wound repair. *J. Dermatol. Surg. Oncol.* **19**, 693–670.
 97. Nickoloff, B.J. (1991) The cytokine network in psoriasis. *Arch. Dermatol.* **127**, 871–884.
 98. Koch, A.E., Kunkel, S.L., Burrows, J.L., Evanoff, H.L., Haines, G.K., Pope, R.M., Strieter, R.M. (1991) The synovial tissue macrophage as a source of the chemotactic cytokine interleukin-8. *J. Immunol.* **147**, 2187–2195.
 99. Koch, A.E., Kunkel, S.L., Pearce, W.H., Shaw, M., Parikh, D., Evanoff, H.L., Haines, G.K., Burdick, M.D., Strieter, R.M. (1993) Enhanced production of the chemotactic cytokines interleukin-8 (IL-8) and monocyte chemoattractant protein-1 (MCP-1) in human abdominal aortic aneurysms. *Am. J. Pathol.* **142**, 1423–1431.
 100. Mulliken, J., Glowacki, J. (1982) Hemangiomas and vascular malformations in infants and children: a classification based on endothelial characteristics. *Plastic Reconstr. Surg.* **69**, 412–420.
 101. Brown, L.F., Dvorak, A.M., Dorak, H.F. (1989) Leaky vessels, fibrin deposition, and fibrosis: a sequence of events common to solid tumors and to many types of disease. *Am. Rev. Respir. Dis.* **140**, 1104–1107.
 102. Symington, F.W. (1989) Lymphotoxin, tumor necrosis factor and gamma interferon are cytostatic for normal human keratinocytes. *J. Invest. Dermatol.* **92**, 798–805.
 103. Yaar, M., Karassik, R.L., Schipper, L.E., Gilchrist, B.A. (1985) Effect of alpha and beta interferon on cultured human keratinocytes. *J. Invest. Dermatol.* **85**, 70–74.
 104. Nickoloff, B.J., Varani, J., Mitra, S. (1991) Modulation of keratinocyte biology by gamma interferon: relevance to cutaneous wound healing. In *Clinical Experimental Approaches to Dermal and Epidermal Repair: Normal and Chronic Wounds*, Wiley-Liss, Inc., New York, 141–154.
 105. Shipley, G.D., Pittelkow, M.R., Wille, J.J., Jr., Scott, R.E., Moses, H.L. (1986) Reversible inhibition of human prokeratinocyte proliferation by type beta transforming growth factor inhibitor in serum free medium. *Cancer Res.* **46**, 2068–2071.
 106. Nickoloff, B.J., Mitra, R.S. (1989) Inhibition of 125I-epidermal growth factor binding to cultured keratinocytes by antiproliferative molecules gamma interferon, cyclosporine A, and transforming growth factor-beta. *J. Invest. Dermatol.* **93**, 799–803.
 107. Klagsbrun, M., D'Amore, M. (1991) Regulators of angiogenesis. *Annu. Rev. Physiol.* **53**, 217–239.
 108. Pober, J.S., Cotran, R.S. (1990) Cytokines and endothelial cell biology. *Pathol. Rev.* **70**, 427–451.
 109. Stout, A.J., Gresser, I., Thompson, D. (1993) Inhibition of wound healing in mice by local interferon-alpha/b injection. *Int. J. Exp. Pathol.* **74**, 79–85.
 110. Demaeyer, E., Demaeyer-Guignard, J. (1988) *Interferons and Other Regulatory Cytokines*. Wiley, New York.
 111. Mette, S.A., Pilewski, J., Buck, C.A., Albelda, S.M. (1993) Distribution of integrin cell adhesion receptors on normal bronchial epithelial cells and lung cancer cells in vitro and in vivo. *Am. J. Respir. Cell. Mol. Biol.* **8**, 562–572.
 112. Mueller, B.M., Reisfeld, R.A. (1991) Potential of the SCID mouse as a host for human tumors. *Cancer Metastasis Rev.* **10**, 193–200.
 113. Wang, X., Fu, X., Hoffman, R.M. (1992) A new patient-like metastatic model of human lung cancer constructed orthotopically with intact tissue via thoracotomy in immunodeficient mice. *Int. J. Cancer* **51**, 992–995.
 114. McLemore, T.L., Eggleston, J.C., Shoemaker, R.H., Abbott, B.J., Bohlan, M.E., Liu, M.C., Fine, D.L., Mayo, J.G., Boyd, M.R. (1988) Comparison of intrapulmonary, percutaneous intrathoracic, and subcutaneous models for the propagation of human pulmonary and nonpulmonary cancer cell lines in athymic nude mice. *Cancer Res.* **48**, 2880–2886.
 115. Bankert, R.B., Umamoto, T., Sugiyama, Y., Chen, F.A., Repasky, E., Yokota, S. (1989) Human lung tumors, patient's peripheral blood lymphocytes and tumor infiltrating lymphocytes propagated in SCID mice. *Curr. Top. Microbiol. Immunol.* **152**, 201–210.
 116. Phillips, R.A., Jewett, M.A.S., Gallie, B.L. (1989) Growth of human tumors in immune-deficient SCID and nude mice. *Curr. Top. Microbiol. Immunol.* **152**, 260–263.
 117. Hendrickson, E.A. (1993) The SCID mouse: relevance as an animal model system for studying human disease. *Am. J. Pathol.* **143**, 1511–1522.
 118. Rendt, K.E., Barry, T.S., Jones, D.M., Richter, C.B., McCachren, Haynes, B.F. (1993) Engraftment of human synovium into severe combined immune deficient mice. *J. Immunol.* **151**, 7324–7336.
 119. Caamano, J., Ruggeri, B., Momiki, S., Sickler, A., Zhang, S.Y., Klein-Szanto, A.J.P. (1991) Detection of p53 in primary lung tumors and nonsmall cell lung carcinoma cell lines. *Am. J. Pathol.* **139**, 839–845.
 120. Zucker, S., Lysik, R.M., Malik, M., Bauer, B.A., Caamano, J., Klein-Szanto, A.J.P. (1992) Secretion of gelatinases and tissue inhibitors of metalloproteinases by human lung cancer cell lines and revertant cell lines: not an invariant correlation with metastasis. *Int. J. Cancer* **52**, 366–371.
 121. Momiki, S., Baba, M., Caamano, J., Iizasa, T., Nakajima, M., Yamaguchi, Y., Klein-Szanto, A. (1991) In vivo and in vitro invasiveness of human lung carcinoma cell lines. *Invasion Metastasis* **11**, 66–75.
 122. Vitolo, D., Zerbe, T., Kanbour, A., Dahl, C., Herberman, R.B., Whiteside, T.L. (1992) Expression of mRNA for cytokines in tumor-infiltrating mononuclear cells in ovarian adenocarcinoma and invasive breast cancer. *Int. J. Cancer* **51**, 573–580.

123. de Waal Malefyt, R., Yssel, H., Roncarolo, M.G., Spits, H., de Vries, J.E. (1992) Interleukin-10. *Curr. Opin. Immunol.* **4**, 314-320.
124. Howard, M., O'Garra, A., Ishida, H., de Waal Malefyt, R., de Vries, J. (1992) Biological properties of interleukin 10. *J. Clin. Immunol.* **12**, 239-247.
125. Mosmann, T.R., Cherwinski, H., Bond, M.W., Giedlin, M.A., Coffman, R.L. (1986) Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J. Immunol.* **136**, 2348-2357.
126. Fiorentino, D.F., Bond, M.W., Mosmann, T.R. (1989) Two types of mouse helper T cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. *J. Exp. Med.* **170**, 2081-2095.
127. Yssel, H., Waal-Malefyt, R.D., Roncarolo, M.G., Abrahms, J.S., Spits, H., DeVries, J.E. (1992) Interleukin-10 is produced by subsets of human CD4⁺ T cell clones and peripheral blood T cells. *J. Immunol.* **149**, 2378-2384.
128. Vieira, P., Waal-Malefyt, R.D., Dang, M.N., Johnson, K.E., Kastelein, R., Fiorentino, D.F., DeVries, J.E., Roncarolo, M.G., Mosmann, T.R., Moore, K.W. (1991) Isolation and expression of human cytokine synthesis inhibitory factor cDNA clones: homology to Epstein-Barr virus open reading frame BCRF1. *Proc. Natl. Acad. Sci. USA* **88**, 1172-1176.
129. de Waal Malefyt, R., Abrahms, J., Bennet, B., Figdor, C.G., deVries, J.E. (1991) Interleukin-10 (IL-10) inhibits cytokine synthesis by human monocytes-an autoregulatory role of IL-10 produced by monocytes. *J. Exp. Med.* **174**, 1209-1220.
130. O'Garra, A., Chang, R., Go, N., Hastings, R., Haughton, G., Howard, M. (1992) Ly-1 B (B-1) cells are the main source of B cell-derived interleukin 10. *Eur. J. Immunol.* **22**, 711-717.
131. Rivas, J.M., Ullrich, S.E. (1989) Keratinocyte-derived IL-10. *J. Invest. Dermatol.* **98**, 578-581.
132. Moore, K.W., Vieira, P., Fiorentino, D.F., Trounstein, M.L., Khan, T.A., Mosmann, T.R. (1990) Homology of cytokine synthesis inhibitory factor (IL-10) to the Epstein-Barr virus gene BCRF1. *Science* **248**, 1230-1234.
133. Fiorentino, D.F., Zlotnik, A., Mosmann, T.R., Howard, M., O'Garra, A. (1991) IL-10 inhibits cytokine production by activated macrophages. *J. Immunol.* **147**, 3815-3822.
134. de Waal Malefyt, R., Abrams, J., Bennett, B., Figdor, C.G., DeVries, J.E. (1991) Interleukin-10 inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. *J. Exp. Med.* **174**, 1209-1220.
135. Bogdan, C., Vodovotz, Y., Nathan, C. (1991) Macrophage deactivation by interleukin 10. *J. Exp. Med.* **174**, 1549-1555.
136. Ralph, P., Nakoinz, I., Sampson-Johannes, A., Fong, S., Lowe, D., Min, H.Y., Lin, L. (1992) IL-10, T lymphocyte inhibitor of human blood cell production of IL-1 and tumor necrosis factor. *J. Immunol.* **148**, 808-814.
137. de Velde, A.A., de Waal-Malefyt, R., Huijbens, R.J.F., deVries, J.E., Figdor, C. (1992) IL-10 stimulates monocyte Fc gamma R surface expression and cytotoxic activity: distinct regulation of antibody-dependent cellular cytotoxicity by IFN-gamma, IL-4, and IL-10. *J. Immunol.* **149**, 4048-4052.
138. Smith, D.R., Kunkel, S.L., Burdick, M.D., Wilke, C.A., Orringer, M.B., Whyte, R.I., Strieter, R.M. (1994) The production of interleukin-10 by human bronchogenic carcinoma. *Am. J. Pathol.* **145**, 18-25.