

The rocky road to personalized medicine in acute myeloid leukaemia

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Abstract

Acute myeloid leukaemia (AML) is a malignant disorder of the myeloid blood lineage characterized by impaired differentiation and increased proliferation of hematopoietic precursor cells. Recent technological advances have led to an improved understanding of AML biology but also uncovered the enormous cytogenetic and molecular heterogeneity of the disease. Despite this heterogeneity, AML is mostly managed by a 'one-size-fits-all' approach consisting of intensive, highly toxic induction and consolidation chemotherapy. These treatment protocols have remained largely unchanged for the past several decades and only lead to a cure in approximately 30–35% of cases. The advent of targeted therapies in chronic myeloid leukaemia and other malignancies has sparked hope to improve patient outcome in AML. However, the implementation of targeted agents in AML therapy has been unexpectedly cumbersome and remains a difficult task due to a variety of disease- and patient-specific factors. In this review, we describe current standard and investigational therapeutic strategies with a focus on targeted agents and highlight potential tools that might facilitate the development of targeted therapies for this fatal disease. The classes of agents described in this review include constitutively activated signalling pathway inhibitors, surface receptor targets, epigenetic modifiers, drugs targeting the interaction of the hematopoietic progenitor cell with the stroma and drugs that target the apoptotic machinery. The clinical context and outcome with these agents will be examined to gain insight about their optimal utilization.

Keywords: acute myeloid leukaemia • targeted therapies • drug resistance • minimal residual disease

Introduction

Personalized cancer therapy offers the hope to establish novel and more effective therapeutic standards for patients afflicted with this condition. While traditional chemotherapeutic protocols aim to

destroy rapidly dividing cells, but also affect normal ('healthy') cells, personalized medicine represents a promising concept by which patients whose cancer cells harbour pathophysiologically and therapeutically relevant molecular alterations could be treated with a biomarker-based 'targeted' therapy. In the long term, this strategy may be cost-effective, even including the required diagnostic and follow-up tests that accompany therapy (so-called companion diagnostics).

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Personalized medicine has become a synonym for the medicine of the future to which many experts ascribe a paradigm change. The overwhelming success of the tyrosine kinase inhibitor imatinib [1] and the monoclonal, CD20-targeted antibody rituximab [2] has revolutionized the care of patients with chronic myeloid leukaemia (CML) and non-Hodgkin lymphoma, respectively, and validated the use of targeted treatment strategies in the management of patients with cancer. Acute myeloid leukaemia (AML) is an aggressive form of cancer of the bone marrow (BM) and blood that is characterized by blocked differentiation and rapid proliferation of myeloid precursor cells. Despite major advances in understanding AML at the molecular level, novel treatment concepts are lacking [3]. Therapeutic concepts to manage AML have remained largely unchanged since the 1970s and frequently fail to achieve a cure, underscored by 5-year survival rates of roughly only 30% [4]. The current concept of the molecular basis of AML suggests that the disease arises in hematopoietic precursor cells and is driven by at least two types of cooperative mutations ('the two hit model'). However, novel technologies such as genome sequencing have unveiled a much more complex picture of leukemogenesis and shed further light on hitherto unknown obstacles in the way to targeted therapy for AML.

Current approaches to management of AML

Current standard treatments for AML consist of induction chemotherapy followed by several courses of consolidation chemotherapy or allogeneic stem cell transplantation (aSCT). Herein, induction protocols mostly employ the so-called 7 + 3 regimen, which entails continuous infusion cytarabine given over 7 days and 3 days of an anthracycline, typically either daunorubicin or idarubicin. Although the ideal dose of daunorubicin remains an open question, this approach has remained unchanged for the past several decades [5–7]. The combination of cytarabine and an anthracycline as intensive remission therapy produces complete remission (CR) rates of 60–80% and 40–60% in patients that are less than age 60 and age 60 or greater, respectively [8]. The lower CR rate in elderly patients is a reflection of decreased sensitivity of leukaemic cells to chemotherapy as well as a decreased tolerance to therapy and increased treatment-related mortality [9]. However, even in younger patients, standard AML induction and consolidation regimens frequently lead to complications, such as cytopenias and infections as well as gastroenteric and neurologic toxicities. In addition, only a minority of patients are cured by this approach which highlights the urgent need for novel and improved treatment concepts. Of note, CPX-351, a liposomal combination of daunorubicin and cytarabine, was recently approved by the FDA for intensive remission induction in adults with newly diagnosed therapy-related AML or AML with myelodysplasia-related changes. The approval was based on the results of a phase III clinical trial where CPX-351 significantly improved overall survival, event-free survival and response without an increase in 60-day mortality compared to standard '7 + 3' chemotherapy [10].

Heading towards targeted therapies for AML

Surface receptors

Gemtuzumab ozogamicin (GO), an anti-CD33 immunoconjugate, has the unique distinction of being the first targeted agent in AML that was approved by the FDA *via* accelerated approval in 2000 for older patients with AML in first relapse [11]. The drug was subsequently withdrawn from the U.S. market in June 2010 after a randomized study by SWOG failed to demonstrate improved efficacy, while induction mortality was increased compared to the chemotherapy alone arm [12]. To refute these findings, four subsequent randomized studies [13–16] strongly support the safety and efficacy of this agent in combination with upfront chemotherapy in AML. The addition of GO significantly reduced relapse and improved overall survival at 5 years, with this benefit being most prominent in patients with favourable or intermediate-risk cytogenetics [17]. The inferior outcomes of the SWOG study were attributed to lower anthracycline dosing in the GO arm as well higher doses of GO causing veno-occlusive disease (VOD). GO has also been combined with the hypomethylating agents (HMAs) [18, 19] based on the observation that azacitidine induces CD33 expression and decreases *P*-glycoprotein expression, with favourable response rates of 35–44%. Unfortunately, a randomized study where GO was added to low-dose cytarabine did not translate into improved survival [20]. Building on the lessons gained from GO, vadastuximab talirine (SGN-33A), another CD33-directed, antibody–drug conjugate that employs pyrrolobenzodiazepine instead of calicheamicin, was developed. A phase I study of vadastuximab in combination with an HMA (azacitidine or decitabine) [21] in untreated patients unfit for intensive therapy reported complete remission and complete remission with incomplete count recovery (CR/CRi) rates of 73% among evaluable patients. In combination with induction chemotherapy, vadastuximab produced a CR/CRi rate of 78%, with 30- and 60-day mortality of 0 and 7%, respectively [22]. While these preliminary findings are encouraging, additional studies are currently ongoing to further evaluate the role of vadastuximab in AML therapy (Table 1).

KIT

Approximately 25% of core binding factor (CBF) AML patients carry gain-of-function mutations in the KIT gene. These mutations result in a constitutively active tyrosine kinase that contributes to aggressive leukaemia growth, and is associated with unfavourable outcome [23, 24]. The German-Austrian AML Study Group (AMLSG) and the CALGB [25] conducted phase II studies that evaluated dasatinib in combination with chemotherapy followed by 1-year dasatinib maintenance in CBF AML. The CALGB 10801 study results suggest that outcome of KIT^{mut} patients approached those historically seen in KIT^{wt} patients, suggesting that dasatinib may overcome the negative prognostic effect of the KIT mutation. The AMLSG group is conducting a

Table 1 Targeted agents under clinical investigation either a single agent or in combination with acute myeloid leukaemia therapy

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/combination	Ref./identifier	Status		
Cell surface receptors	CD33	Gemtuzumab ozogamicin	III	3325 adult patients with the first course of intensive remission chemotherapy.	Combination with induction chemotherapy	[16]	Completed		
		Vadastuximab talirine	II	Patients age 60 and greater with newly diagnosed AML.	Combination with azacitidine	[17]	Completed		
Tyrosine kinase pathways	c-kit		I/II	Pre-allogeneic transplant (with conditioning regimen) OR post-allogeneic transplant (single agent) in adults >18 years	Single agent and combination	NCT02614560	Active, not recruiting		
			III	Adult patients with newly diagnosed AML	Combination with azacitidine OR decitabine	NCT02785900	Recruiting		
			I	Safety study as a single agent and in combination with HMA to determine the maximum tolerated dose in adult patients >18 years	Single agent and combination with HMA	NCT01902329	Active, not recruiting		
			Dasatinib		I	Children and adolescent patients with CBF AML to determine maximum tolerated dose	Combination with induction therapy	NCT02680951	Recruiting
					Ib/IIa	Given after induction and consolidation for maintenance therapy for 1 year in adult patients >18 years	Single agent (maintenance)	NCT00850382	Completed, results not available
				II	Given after consolidation for patients with high-risk MRD or in molecular relapse in adults age 18–60 years	Single agent (maintenance)	NCT02113319	Completed, results not available	
				III	Standard induction and consolidation therapy with or without dasatinib in adults age >18 years	Combination	NCT02013648	Recruiting	
	FLT3	Midostaurin	III	Adult patients up to age 60 with newly diagnosed FLT3-mutated AML. CR 59% versus 54%, OS 74.7 versus 25.6 months	Combination with induction, consolidation, and maintenance versus placebo	NCT00651261 [32, 115]	Completed*		

Table 1. Continued

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/combination	Ref./identifier	Status
			I	Adults age 60 and greater with newly diagnosed AML or relapsed/refractory disease	Combination with decitabine	NCT01130662	Completed, results not available
			I	Adults with newly diagnosed AML	Combination with daunorubicin and cytarabine induction	NCT00093600	Completed, results not available
			I	Adults with relapsed/refractory AML	Combination with bortezomib and cytotoxic chemotherapy	NCT01174888	Completed, results not available
			I/II	Adult patients with relapsed/refractory AML or newly diagnosed AML who are ineligible to receive intensive therapy	Combination with azacitidine	NCT01093573	Active, not recruiting
			II	Patients with AML having received allogeneic HSCT	Single agent (maintenance)	NCT02723435	Not yet open
			II/III	Patients age 60 or older with previously untreated AML	Combination with azacitidine and nivolumab	NCT03092674	Not yet open
		Sorafenib	I	Patients age 60 or older with relapsed/refractory or newly diagnosed AML who are not eligible to receive intensive therapy	Combination with bortezomib and decitabine	NCT01861314	Active, not recruiting
			IV	Patients status post-allogeneic HSCT with FLT3/ITD mutation in adults age 18-60 years	Single agent (maintenance)	NCT02474290	Recruiting
			II	Adult patients less than 60 years old with newly diagnosed AML; event-free survival 21 <i>versus</i> 9 months	Combination with standard induction therapy	NCT00893373	Completed
			II	Patients age 60 or older with newly diagnosed FLT3/ITD-mutated AML	Combination with standard induction therapy	NCT01253070	Active, not recruiting

Table 1. Continued

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/combination	Ref./identifier	Status
			I/II	Patients with newly diagnosed AML irrespective of FLT3/ITD status receiving induction therapy in adults age 18-60 years	Combination with CLAG-M induction	NCT02728050	Recruiting
			II	Patients age 60 or older with newly diagnosed AML who are ineligible for intensive therapy	Combination with azacitidine	NCT02196857	Recruiting
			I/II	Elderly patients with AML or high-risk MDS	Combination with low-dose cytarabine	NCT00516828	Completed, results not available
			I/II	Adult patients with newly diagnosed AML; 38% CR, 1-year OS 74%	Combination with standard induction therapy	NCT00542971 [32]	Completed
		Quizartinib	II	Adult patients with relapsed/refractory AML with FLT3/ITD mutation	Single agent	NCT02984995	Recruiting
			III	Adult patients with relapsed/refractory AML with FLT3/ITD mutations versus salvage chemotherapy	Single agent	NCT02039726	Recruiting
			III	Newly diagnosed AML (adults age 18-75 years) with FLT-ITD mutation receiving induction and consolidation chemotherapy, followed by maintenance	Combination with induction chemotherapy	NCT02668653	Recruiting
			I	Relapsed/refractory in adults age 18 or greater with AML irrespective of FLT3 status; 13% CR, 30% ORR	Single agent	NCT00462761	Completed
			I/II	Adult (age 18 or greater) patients with relapsed/refractory AML irrespective of FLT3 status	Combination with azacitidine or low-dose cytarabine	NCT01892371	Recruiting
		Crenolamib	II	Relapsed/refractory AML (adults age 18 or greater) with activating FLT3 mutations	Single agent	NCT01657682	Recruiting

Table 1. Continued

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/combination	Ref./identifier	Status
			II	Maintenance therapy after HSCT in FLT3-positive AML in adults age 18 or greater	Single agent (maintenance)	NCT02400255	Recruiting
			III	Adult patients with relapsed/refractory AML with FLT3 mutations receiving salvage therapy	Combination	NCT02298166	Recruiting
			I/II	Adult patients with relapsed/refractory FLT3-mutated AML receiving salvage therapy	Combination	NCT02400281	Recruiting
			II	Relapsed/refractory AML with FLT3 activating mutations in adults age 18 or greater	Single agent	NCT01522469	Completed, results not available
		Gilteritinib	III	Adult patients (age 18 or greater) with AML in CR1 following induction and consolidation	Single agent (maintenance)	NCT02927262	Recruiting
			III	FLT-3-mutated relapsed/refractory AML or CR with MRD in adults age 18 or greater	Single agent	NCT03070093	Available
			III	Maintenance therapy after allogeneic transplant in FLT-ITD-mutated AML in adults age 18 or greater	Single agent	NCT02752035	Not yet recruiting
			II/III	Azacitidine with or without gilteritinib in newly diagnosed AML age 18 or greater	Combination with azacitidine	NCT02997202	Recruiting
	RAS	Tipifarnib	II	Patients age 65 or older who are ineligible for intensive therapy	Single agent	NCT01361464	Completed, results not available
			I	Adult patients with relapsed/refractory AML or ineligible to receive intensive therapy	Single agent	NCT00101296	Completed, results not available
			II	Adult patients with poor-risk AML who have achieved a CR after induction chemotherapy	Single agent (maintenance)	NCT00045396	Completed, results not available

Table 1. Continued

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/combination	Ref./identifier	Status
			II	Adult patients 70 years or older with newly diagnosed AML who are ineligible for intensive therapy	Combination with etoposide	NCT00602771	Completed, results not available
			I/II	Adult patients with newly diagnosed AML	Combination with standard induction chemotherapy	NCT00096122	Completed, results not available
			II	Adult patients with relapsed/refractory AML	Single agent	NCT00354146	Completed, results not available
			II	Adult patients 70 years or older with newly diagnosed AML who are ineligible for intensive therapy	Single agent	NCT00093418	Completed, results not available
			II	Adult patients 60 years or older as post-consolidation therapy	Single agent (maintenance)	NCT00048503	Completed, results not available
			III	Adult patients in second or greater remission OR patients greater than 60 years old in first remission; DFS 8.87 versus 5.26 months, OS 16.36 versus 9.27 months	Single agent (maintenance)	NCT00093470	Completed
		Selumetinib	II	Adult patients with relapsed/refractory AML	Single agent	NCT00588809	Completed, results not available
		Trametinib	II	Adult patients with relapsed/refractory AML or newly diagnosed AML who are ineligible to receive intensive therapy	Combination with Akt inhibitor GSK2141795	NCT01907815	Active, not recruiting
			I	Adult patients with relapsed/refractory AML or newly diagnosed AML who are ineligible to receive intensive therapy	Combination with AMG 232 or alone	NCT02016729	Active, not recruiting
		Rigosertib	I/II	Combination with azacitidine; dose escalation, dose expansion, safety	Combination with azacitidine	NCT01926587	Recruiting

Table 1. Continued

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/combination	Ref./identifier	Status
	SYK	Entospletinib	Ib/II	Adult patients with newly diagnosed AML and relapsed/refractory disease	Combination with low- and high-intensity regimens	NCT02343939	Recruiting
	Plks	Volasertib	III	Combination with low-dose cytarabine in newly diagnosed AML age 65 and greater	Combination with low-dose cytarabine	NCT01721876	Active, not recruiting
			I/IIa	Single agent and combination with low-dose cytarabine in relapsed/refractory AML	Single agent and combination	NCT00804856	Active, not recruiting
Apoptotic targets	Bcl-2	Venetoclax	III	Adult patients with newly diagnosed AML	Combination with azacitidine	NCT02993523	Recruiting
			III	Adult patients with newly diagnosed AML who are ineligible for intensive therapy	Combination with low-dose cytarabine	NCT03069352	Not yet recruiting
			I/II	Patients 60 years and older with newly diagnosed AML who are ineligible for intensive therapy	Combination with low-dose cytarabine	NCT02287233	Active, not recruiting
Stromal targets	CXCR4 and CXCL12	Plerixafor	I	Adult patients with newly diagnosed AML receiving induction chemotherapy	Combination with induction therapy (cytarabine and daunorubicin)	NCT00990054	Completed, results not available
			I	Patients 60 years and older with newly diagnosed AML	Combination with decitabine	NCT01352650	Active, not recruiting
			I	Adults patients with relapsed/refractory AML receiving salvage therapy; CR 46%	Combination with G-CSF, mitoxantrone, etoposide, and cytarabine induction	NCT00906945 [53]	Completed
		Ulocuplumab	I/II	Combined with low-dose cytarabine in newly diagnosed AML	Combination	NCT02305563	Active, not recruiting
			I	Safety and tolerability in patients with relapsed AML	Single agent	NCT01120457	Completed, results not available
Epigenetic	Hypomethylator	Guadecitabine	III	Adult patients with relapsed/refractory AML	Single agent <i>versus</i> treatment of choice	NCT02920008	Recruiting

Table 1. Continued

Target category	Drug target	Drug	Trial phase	Patient population [Results]	Single agent/ combination	Ref./Identifier	Status
	IDH1/2	AG-221	III	AG-221 <i>versus</i> conventional care regimens in patients 60 and older with relapsed/refractory AML and IDH2 mutation	Single agent	NCT02577406	Recruiting
			I/II	Adult patients with newly diagnosed AML with IDH1/2 mutations who are ineligible to receive intensive therapy	Combination with azacitidine	NCT02677922	Recruiting
			I	Adult patients with newly diagnosed AML receiving induction therapy with IDH1/2 mutation	Combination with induction and consolidation therapy	NCT02632708	Recruiting
	Bromodomain	OTX015/ MK-8628	I	Adult patients with AML or ALL with relapsed/refractory disease	Single agent	NCT01713582	Completed, results not available
			I	Adult patients with relapsed/refractory acute leukaemias	Single agent	NCT02158858	Recruiting
			I	Adult patients with relapsed/refractory haematologic malignancies	Single agent	NCT02543879	Recruiting
	CDK	Alvociclib	II	Alvociclib and cytarabine/mitoxantrone <i>versus</i> cytarabine/mitoxantrone in adults with relapsed/refractory AML with NOXA BH3 priming of $\geq 40\%$ by mitochondrial profiling in bone marrow	Combination with induction therapy	NCT02520011	Recruiting
			I/II	Adult patients with MLL-rearranged leukaemias	Single agent	NCT02310243	Recruiting

*Landmark trial that led to the approval of midostaurin for the treatment of FLT3 mutant AML by the U.S. Food and Drug Administration.

randomized phase III study adding dasatinib to induction chemotherapy in CBF AML. A French Intergroup study showed dasatinib used as single-agent maintenance failed to prevent relapse in patients with poor molecular response or molecular recurrence following chemotherapy [26]. The disappearance of KIT mutations at relapse suggests that clonal devolution may explain the absence of efficacy observed with single-agent dasatinib.

FLT3

The negative prognostic impact of the fms-like tyrosine kinase receptor-3 internal tandem duplication mutation (FLT3/ITD) on AML outcome and its physiologic effect of constitutive signalling through a receptor tyrosine kinase make it a highly desirable drug target. Mutational burden appears to predict addiction to FLT3 signalling and thus response to FLT3 inhibition [27]. FLT3/ITD mutational burden is increased at disease progression rather than at presentation when the genomic composition of the AML is more heterogeneous [28]. In line with this finding, tumour cells derived from relapsed FLT3/ITD-mutated AML patients appear to be addicted to signalling from the constitutively activated FLT3 receptor tyrosine kinase which insinuates that less specific inhibitors may be efficacious earlier in therapy, while more specific inhibitors may be best utilized at relapse [29]. However, the optimal approach to incorporate FLT3 inhibitors into the management of newly diagnosed and relapsed/refractory FLT3-mutated AML patients remains a matter of dispute and additional, pivotal studies are needed to provide an answer to this important question. *Midostaurin* is a multikinase inhibitor that claims the unique distinction of being the first FLT3 inhibitor proven to improve overall survival (OS) in FLT3/ITD-mutated AML. As a single agent, *Midostaurin* treatment of 95 patients resulted in 1 partial and no complete remissions [30]. However, when combined with conventional chemotherapy in newly diagnosed AML patients, *midostaurin* induced high remission and survival rates in both FLT3-mutated and wild-type patients [31]. The CALGB conducted a randomized, placebo-controlled Phase III trial (RATIFY) in treatment-naïve FLT3-mutated AML patients <60 years encompassing induction chemotherapy and four consolidation cycles of high-dose cytarabine combined with placebo or *midostaurin*, followed by *midostaurin* maintenance or placebo for 1 year [32]. The median OS was 74.7 months for the group receiving *midostaurin* versus 26 months for the placebo group ($P = 0.007$). In addition, a 23% reduction in the risk of death was observed. The landmark results of this trial resulted in its FDA approval in combination with chemotherapy in AML patients younger than 60 years of age in April 2017. It is interesting to note that response rates to induction therapy did not differ significantly between treatment arms, suggesting prolonged exposure is required to benefit from the inhibitor. Moreover, patients randomized to *midostaurin* who underwent aSCT during the first remission had a survival curve plateau in the 60–70% range suggesting that aSCT remains a very relevant consideration in this population. Another interesting compound, *sorafenib*, was originally developed as an inhibitor of the serine/threonine kinase Raf but leukaemia clinical trials and physicians have capitalized on its off-target inhibition of FLT3.

Being FDA approved for hepatocellular carcinoma, it is the most widely accessible FLT3 inhibitor in clinical practice and frequently used off-label. In younger patients, the addition of *sorafenib* to chemotherapy was well tolerated and showed preferential activity in FLT3-mutated patients [33]. The phase II randomized SORAML study in younger patients bore out these results [34] with improved EFS; however, grade 3–4 toxicities were higher in the *sorafenib* arm. The Study Alliance Leukemia trial combining induction chemotherapy with *sorafenib* in a randomized trial in patients over age 60 showed no difference in the event-free survival (EFS) or OS between groups [35]. This was attributed to higher induction mortality rate due to infectious complications in the *sorafenib* arm accompanied by lower protocol adherence for post-remission therapy. These trials were FLT3 mutation agnostic and showed responses in FLT3-wt patients supporting off-target mechanisms of effect. In a Phase II study of relapsed or refractory FLT3/ITD-mutated AML, the combination of *sorafenib* and the hypomethylating agent *azacitidine* yielded response rates of 46%, [36] suggesting that the combination of the two drugs may represent a clinically valuable regimen for relapsed, FLT3/ITD-mutated AML in the elderly. The HMA backbone has the additional advantage of less up-regulation of FLT3 ligand which is normally massively up-regulated after cytotoxic chemotherapy and can compromise the efficacy of the FLT3 inhibitors. *Sorafenib* is being studied in the prevention of post-transplant relapses, with an improved 2-year progression-free survival and a reduced risk of relapse [37] but data on timing and duration of therapy is sparse. Several other more specific FLT3 inhibitors are currently undergoing clinical studies. *Quizartinib*, an exquisitely specific FLT3 inhibitor, has a significantly longer half-life than the above agents, as well as a greater capacity for inhibition of mutated FLT3 [38]. Several phase I and II studies have demonstrated encouraging activity of *quizartinib* in patients with relapsed/refractory AML [39–41]. *Crenolanib*, a drug originally developed as an inhibitor of platelet-derived growth factor receptor, has shown both activities in FLT3/ITD-mutated AML and FLT3/ITD D835-mutated AML [38]. The D835 mutation has been identified as a potent mechanism of resistance to earlier FLT3 inhibitors. *Gilteritinib*, an agent with activity against wild-type FLT3, FLT3/ITD, FLT-TKD D835 and F691, as well as Axl, has been examined for the treatment of relapsed/refractory AML in two early-phase clinical trials. In the phase I/II CHRYSALIS dose escalation trial, *gilteritinib* produced an overall response rate of 57% in FLT3-mutated patients and 63% in patients with FLT3 mutations who received a dose of 80 mg per day or greater [42]. In a follow-up study of patients with relapsed/refractory AML, where 65% of subjects received >2 lines of therapy and 23% received treatment with a TKI, the overall response rate was 55% (60% for FLT3-mutated patients and 29% for FLT3 wild-type patients) in the setting of a median overall survival of 29 weeks [43]. More recently, an exploratory analysis presented at the ASCO meeting in 2017 showed that molecular responses to *gilteritinib* in relapsed/refractory FLT3/ITD-mutated patients correlated with the clinical outcome. In this study, Altman *et al.* [44] reported that patients with an ITD signal ratio of $\leq 10^{-2}$, 10^{-3} (major molecular response), or were MRD negative demonstrated a significantly longer median overall survival compared to patients who did not achieve a molecular response, suggesting that the ITD signal ratio may serve as a predictor of durable clinical benefit of *gilteritinib*.

RAS

In AML, the RAS pathway is activated both by mutations occurring in *RAS* as well mutations and/or overexpression of upstream receptor tyrosine kinases such as *FLT3*. RAS inhibitors have had an underwhelming impact on AML. A phase 3 trial evaluating the farnesyl-transferase inhibitor tipifarnib as first-line therapy in older patients resulted in a CR rate of only 8%, and no survival benefit. A phase 2 trial of single-agent selumetinib [45] showed modest activity only in the *FLT3* wild-type subset. The oral mitogen-activated protein kinase inhibitor trametinib showed more encouraging results with selective activity in *NRAS* or *KRAS*-mutated AML and CMML [46]. Response rates of 27% were seen in CMML and the lack of activity in *RAS* wild-type leukaemias endorses the selective effect of the inhibitor. Rigosertib is a RAS-mimetic interacting with the RAS-binding domains of RAF kinases, preventing their binding to RAS and inhibiting the RAS-RAF-MEK pathway [47]. This drug is being developed mainly in the MDS arena, and a phase III multicentre randomized trial is now comparing rigosertib to best supportive care in higher risk MDS progressing on HMA. Early results from a recent phase 1b study of the MDM2 inhibitor AMG 232 in 35 relapsed/refractory AML patients showed that AMG 232 was well tolerated and exhibited promising anti-leukaemic activity (NCT02016729) [48].

Polo-like kinases

Polo-like kinases (Plks) are involved in mitotic checkpoint regulation and cell division [49]. Volasertib potently inhibits Plk1 as well as Plk2 and Plk3 blocking spindle formation and inducing cell cycle arrest in M phase. Volasertib was granted breakthrough therapy status by the FDA in 2013 for use with low-dose cytarabine in high-risk AML ineligible for standard therapy based on superior responses (31.0% *versus* 13.3%) in a randomized phase 2 study [50]. However, the phase 3 POLO-AML-2 trial in the same population failed to meet the primary end-point of superior responses [51] with an increased infection-related mortality in the volasertib arm.

Cyclin-dependent kinase inhibitors

Alvocidib, a potent inhibitor of serine-threonine Cyclin-dependent kinases (CDKs) 9, 4 and 7, has been shown to be an active agent against AML [52]. Pre-clinical studies have demonstrated that inhibition of CDK9 and CDK7 leads to down-regulation of transcripts of cyclin D1, c-MYC and MCL-1, leading to enhancement of anti-tumour effects of cell cycle-specific cytotoxic agents, such as cytarabine [53]. Alvocidib has been studied in both the newly diagnosed and relapsed/refractory AML settings. To date, several clinical studies evaluating alvocidib in conjunction with cytarabine and mitoxantrone (FLAM) in patients with newly diagnosed AML have been published with overall CR rates of approximately 68% [54–59]. Of note, patients with favourable-risk cytogenetic features such as core-binding factor AML were excluded. In patients with relapsed/refractory AML, overall CR rates for FLAM were 36% [54, 55, 57, 60]. Palbociclib, an inhibitor of both CDK

4 and 6, is currently being studied in leukaemia patients with MLL rearrangements. In a recently reported phase 1b study of six patients with relapsed/refractory leukaemia, one partial response, three disease stabilizations and two cases of the progressive disease were noted [61].

Targeting apoptosis

Dysregulation of apoptosis in AML is partly mediated by overexpression of the anti-apoptotic protein BCL-2 and related family members. Venetoclax (ABT-199) is a 'BH3-mimetic' antagonist of BCL-2. A phase 2 study of 32 patients with relapsed/refractory AML reported 5 CRs, the majority of which occurred in patients carrying *IDH1* or *IDH2* mutations. The responses, however, were short-lived [62]. Improved responses in *IDH*-mutated AML cases are attributed to 2-hydroxyglutarate-mediated inhibition of the activity of cytochrome oxidase in the mitochondrial electron transport chain, lowering the mitochondrial threshold to trigger apoptosis upon BCL-2 inhibition [63]. In a phase 1b study in treatment-naive older (≥ 65) patients with cytogenetically intermediate- or poor-risk AML ineligible for intensive chemotherapy, the combination of venetoclax with HMA yielded an overall response rate of 76% [64]. Venetoclax has been combined with low-dose cytarabine in elderly AML producing high response rates (CR/CRi of 54%), with median survival not reached among the responders [65]. This drug is garnering enthusiasm in the AML arena in combination with low-intensity therapies in elderly patients.

Targeting the stroma

Most of the progress in targeting AML–stroma interactions has been made by the development of CXCR4 inhibitors which mobilize leukaemic cells out of their protective niches by disrupting the AML–stroma interactions. These agents may also inhibit the pro-survival signals provided to the blasts *via* CXCR4/CXCL12 signalling. In a phase 2 study, 46 patients treated with plerixafor in combination with chemotherapy showed a response rate of 46% (CR+CRi) associated with twofold mobilization in leukaemic blasts into the peripheral circulation [66]. Ulocuplumab is a fully human IgG4 monoclonal antibody to CXCR4, with a half-life longer than plerixafor well tolerated with salvage chemotherapy in relapsed AML [67].

Epigenetics

Dysregulation of chromatin modifiers is a recurrent and sentinel event in oncogenesis. Strategies that target the recruitment and/or catalytic activity of these enzymes at chromatin represent an attractive therapeutic modality in leukaemia [68].

DNMT inhibitors

The HMAs 5-Azacytidine (azacitidine) and its deoxy analogue 5-aza-2'-deoxycytidine (decitabine) are the two most extensively studied

DNMT inhibitors and are approved for clinical use in haematologic malignancies in the United States. The cytidine nucleoside analogue Azacitidine which, upon cellular uptake, is in part converted into Decitabine, confers its cytotoxic effects via RNA and DNA incorporation, thereby disrupting protein and nucleic acid synthesis. DNMT inhibitors have been shown to induce response rates of 30% and more importantly prolong survival in elderly patients with AML in comparison with best available therapy for older patients [69, 70]. Predicting responsiveness to this treatment modality has been challenging due to variable methylation profiles across biologic subgroups of AML. A recent phase 2 multicentre study showed that decitabine has preferential activity in p53-mutated AML, one of the most chemotherapy resistant and unfavourable prognostic subsets of this disease. Moreover, detailed genomic analysis of the patients treated with decitabine showed robust suppression of the p53 mutant clone. These exciting data suggest an alternative up-front strategy for the treatment of this group of high-risk patients that will need to be verified in prospective trials. Guadecitabine, a dinucleotide of decitabine and deoxyguanosine and second-generation hypomethylating agent, is currently under investigation for AML patients who are ineligible to receive intensive chemotherapy [71].

IDH inhibitors

Neomorphic mutations in isocitrate dehydrogenase (*IDH1* and *IDH2*), each seen in 8–12% of AML cases result in an abnormal oncometabolite 2-hydroxyglutarate, which leads to a hypermethylated genome with a resultant block in differentiation [72]. The recently published phase 1/2 study of enasidenib (AG-221), a first-in-class IDH2 inhibitor reported response rates of 40% and median duration of response of 4.8 months [73]. This class of drugs induces differentiation of blasts rather than cytotoxicity and myeloablation. IDH differentiation syndrome was seen in 10% of patients and has also been reported with the IDH1 inhibitor ivosidenib (AG-120) [74]. While the drug potently suppresses the enzymatic activity of IDH2 and the levels of 2-HG, it does not consistently suppress the allele burden of mutant IDH2. In fact, the emergence of mutant IDH2 neutrophils supports the idea of differentiation rather than elimination of the mutant clone. Enasidenib was equally effective in IDH2 R140 and R172 mutations. Certain mutational subsets of AML such as RAS mutations are more resistant to this therapy, and the role of mutational context in predicting response will continue to be explored. The IDHENTIFY phase III clinical trial is comparing enasidenib, to the standard of care for older patients with relapsed/refractory IDH2-mutant AML. Both AG-120 and enasidenib are also being investigated in newly diagnosed AML with *IDH1* and/or *IDH2* mutations, in combination with intensive chemotherapy, as well as with azacitidine in unfit patients. The 9.3-month overall survival is also quite impressive in a pre-treated population considering the expected 3-month median survival in these patients [75]. This class of drugs offers the exciting prospect of improving current standard of care in *IDH*-mutant AML patients. Enasidenib has recently been approved by the FDA for the management of relapsed/refractory AML in patients with IDH2 mutations.

HDAC inhibitors

Histone acetylase inhibitors work by altering chromatin structure and allowing transcription factors to bind to gene promoters. Romidepsin was one of the early HDAC inhibitors studied in a multicentre phase 2 study [76] in relapsed AML and was seen to preferentially induce differentiation in core-binding factor AML cases. Vorinostat was more recently studied in combination with induction chemotherapy in a phase 3 trial, which was aborted due to lack of improvement over standard induction alone [77]. However, it has been safely combined with azacitidine and has demonstrated efficacy in MLL-rearranged AML at relapse with response rates of 35% [78] in this high-risk subset of AML patients. Other oral HDACs, including entinostat and pracinostat, are in early trials in combination with HMAs. Of note, a recent study of entinostat combined with azacitidine showed pharmacodynamic antagonism, whereas prolonged administration of the hypomethylating agent alone appeared to increase response rates when compared to standard dosing [79].

DOT1L inhibitors

Aberrant fusion proteins involving the MLL histone methyltransferase lead to recruitment of the histone methyltransferase DOT1L. Preclinical studies of DOT1L inhibition in MLL-rearranged AML showed remarkable effectiveness; however, inhibition of DOT1L in a phase I trial with the small molecule Pinemetostat (EPZ-5676) produced complete remissions in only 2 of 34 patients with an MLL-rearranged leukaemia [80]. Future studies of this agent might thus focus on combination regimens.

Bromodomain inhibitors

The BET bromodomains are transcriptional coactivators involved in chromatin-dependent signal transduction from master regulatory transcription factors to RNA polymerase II. The first direct-acting bromodomain antagonist JQ1 was reported in 2010 [81], and since then, the field has been expanding. BET recruitment is particularly relevant in MLL-rearranged [82] and NPM1-mutated AML based on proteomic studies. It has also shown synergy in combination with FLT3 inhibitors in preclinical testing in FLT3/ITD-mutated AML [83]. In a phase 1 study, the orally active BET inhibitor OTX015 was given to 41 elderly patients with relapsed/refractory acute leukaemia with five documented responses. Various other BET inhibitors have entered early clinical trials in patients with relapsed AML, including TEN-010, GSK525762, FT-1101 and GPI-0610.

AML heterogeneity and minimal residual disease

One of the major challenges to the sustained efficacy of targeted therapy is the genomic and cellular heterogeneity of AML. While bulk disease at initial diagnosis is comprised of a small number of dominant

clones [84], this belies the underlying diversity of coexisting minor subclones that share some but not all of the gene mutations and epigenetic modifications present in the dominant clones [85, 86]. Conventional cytotoxic chemotherapy or molecularly targeted agents can suppress or eradicate dominant clones leading to a complete remission but nevertheless facilitate the rise of genetically related but distinct clones either through selection of pre-existing resistant subclones or clonal evolution and subsequent development of secondary resistance in otherwise sensitive clones leading to disease relapse [28, 87, 88]. The frequency and stability at relapse of mutated genes that define the clonal architecture of AML are intimately related to its pathobiology. Pre-leukaemic and leukaemic stem cells sequentially acquire mutations and diverge into subpopulations prior to frank transformation to AML [89, 90]. Mutations in some genes, particularly those associated with epigenetic modification such as *DNMT3A* and *IDH2*, are acquired early in leukaemic development and are therefore present in nearly all clonal progeny and are almost always retained in AML at relapse [91, 92]. This contrasts with mutations in other genes such as *NRAS* and *FLT3* that are acquired late in AML pathogenesis and often lost at relapse [93–95], implying that residual pre-leukaemic or leukaemic subclones that lacked those gene mutations rise to clonal dominance at relapse. This has significant implications for the development of targeted therapy as emergence of leukaemic clones that lack the targeted mutation may become a common resistance mechanism for inhibitors of the protein products of dispensable gene mutations acquired late in AML pathogenesis. In addition to genomic diversity, the cellular heterogeneity of AML complicates the development of targeted therapies. While the bulk of AML cells are morphologically and functionally defined as myeloid blasts, pre-leukaemic and leukaemic stem and progenitor cells (LSPC) are both present during an overt clinical disease and persist in complete remission and are implicated as a source of relapse [90, 96]. Targeted therapies which effectively kill AML blasts may not have activity against LSPC due to their increased quiescence and resistance to apoptosis. Furthermore, while therapies specifically directed at LSPC are in development, the immunophenotypes that clearly delineate them from normal hematopoietic stem cells are still uncertain and significant clonal diversity exists even within the LSPC compartment, suggesting that LSPC-directed therapy may suffer from the same clonal escape that plagues treatment of bulk disease [97, 98]. Given these challenges, preclinical testing with *in vitro* systems and *in vivo* xenograft models of AML has the potential to help guide the preclinical development of targeted agents that are effective in clinical trials as well as to understand mechanisms of therapy resistance. Recent improvements in the degree and scope of immunodeficiency as well as improved engraftment conditions have enabled more clinical specimens to be used in murine xenografts for preclinical testing [99, 100]. However, despite these advances, some patient samples will fail to engraft; cells such as leukaemic blasts, progenitors and precursors that may be important in human disease cannot independently engraft in these mice which may overestimate the importance of leukaemic stem cells; and AML that does arise in these models is often restricted to a few clones that can obscure the clonal complexity or lack the most clinically relevant clones of AML in patients [100, 101]. Another tool that may improve the development of targeted therapies is the emergence

of high-sensitivity methods of detecting minimal residual disease (MRD). Measurement of leukaemia-associated aberrant immunophenotypes with multiparameter flow cytometry, gene fusion transcripts with quantitative polymerase chain reaction (qPCR) and gene mutations with qPCR, droplet digital PCR and next-generation sequencing allows precise quantitation of as few as 1 in 100,000 residual aberrant hematopoietic cells in patients in complete remission depending on the platform used. MRD detection appears to offer robust prediction of relapse risk, particularly in the traditionally favourable core binding factor leukaemias and AML with *NPM1* mutations in the absence of *FLT3*/ITD mutations [102–105], and is being further tested and validated in intermediate- and poor-risk AML both in the setting of post-induction remission assessment as well as prior to and following aSCT. Importantly, MRD measurement may be a powerful and underutilized tool for development of targeted therapies, especially in the resurgent concept of maintenance therapies during complete remission. Rather than rely on overt clinical relapse as the end-point of induction and maintenance trials, tracking MRD longitudinally may provide a surrogate marker of response and allow detection of early molecular evidence of relapse or emergence of resistance mutations. In addition, many MRD monitoring methods are amenable for use with *in vitro* and *in vivo* treatment systems with the potential to inform the assessment of the efficacy of novel agents in preclinical models. The primary drawback to MRD testing, however, is the uncertainty of which clonal hematopoietic cells are being measured. These methods detect residual disease but also measure aberrant pre-leukaemic and non-leukaemic hematopoietic cells which have unclear biological and prognostic significance [106, 107]. Further refinement of these methods will be critical to their usefulness both clinically and in pre-clinical drug development.

Conclusion

Although the tremendous progress in genetic technologies has brought more insight into the pathobiology of AML, there is still a knowledge gap with regard to the most suitable targets. The reasons for this knowledge gap are multifaceted and include the complex molecular architecture of the disease with multiple driver mutations and interconnected signal transduction pathways [108]. Additional complexity is added by host-specific factors such as the patient's age, comorbidities and psychosocial and socio-economic status [109]. However, biomarker adapted treatment protocols have already been established in several cancers but many therapies are only temporarily effective [110–112]. Drug resistance to chemotherapy and targeted agents with subsequent relapse or progression thus remains a major problem in the treatment of cancer, including AML [113]. Combination therapies offer the potential of targeting several pathways simultaneously to more effectively eliminate cancer cells and to prevent or delay the development of drug resistance. In appreciation of this concept, the 'Beat AML Master Trial', led by the Leukemia and Lymphoma Society in collaboration with several academic centres and the pharmaceutical industry, offers the hope to substantially boost the paradigm of personalized medicine in AML by utilizing companion biomarker-based treatment strategies [114]. In this trial,

patients (n = 500 +) with newly diagnosed AML will be assigned to targeted therapies after undergoing comprehensive genomic screening. Treatment arms consist of either the targeted agent alone or of the targeted agent combined with conventional therapy, such as standard '7 + 3' or an HMA. Notably, patients whose AML cells lack a targetable lesion are eligible to receive novel therapy on a marker-negative substudy. The 'Beat AML Master Trial' has enormous potential to further our understanding of the activity of currently available therapies in the treatment of AML. Despite this enthusiasm, however, it is noteworthy that, aside from expanding the boundaries of person-

alized medicine, the further development of already established FDA approved treatment protocols is critical to close our knowledge gap in optimizing the use of anti-AML agents. This requires a global effort from physicians, scientists, insurance companies, pharmaceutical industry and regulatory authorities.

Conflict of interest

The authors confirm that there are no conflicts of interest.

References

- Hochhaus A, Larson RA, Guilhot F, *et al.* Long-term outcomes of imatinib treatment for chronic myeloid leukemia. *N Engl J Med.* 2017; 376: 917–27.
- Batlevi CL, Matsuki E, Brentjens RJ, *et al.* Novel immunotherapies in lymphoid malignancies. *Nat Rev Clin Oncol.* 2016; 13: 25–40.
- Estey E. Why is progress in acute myeloid leukemia so slow? *Semin Hematol.* 2015; 52: 243–8.
- Kantarjian H, O'Brien S. Questions regarding frontline therapy of acute myeloid leukemia. *Cancer.* 2010; 116: 4896–901.
- Fernandez HF, Sun Z, Yao X, *et al.* Anthracycline dose intensification in acute myeloid leukemia. *N Engl J Med.* 2009; 361: 1249–59.
- Lowenberg B, Ossenkoppele GJ, van Putten W, *et al.* High-dose daunorubicin in older patients with acute myeloid leukemia. *N Engl J Med.* 2009; 361: 1235–48.
- Burnett AK, Russell NH, Hills RK, *et al.* A randomized comparison of daunorubicin 90 mg/m² vs 60 mg/m² in AML induction: results from the UK NCRI AML17 trial in 1206 patients. *Blood.* 2015; 125: 3878–85.
- Juliusson G, Antunovic P, Derolf A, *et al.* Age and acute myeloid leukemia: real world data on decision to treat and outcomes from the Swedish Acute Leukemia Registry. *Blood.* 2009; 113: 4179–87.
- Hiddemann W, Kern W, Schoch C, *et al.* Management of acute myeloid leukemia in elderly patients. *J Clin Oncol.* 1999; 17: 3569–76.
- Lancet JE, Uy GL, Cortes JE, *et al.* Final results of a phase III randomized trial of CPX-351 *versus* 7 + 3 in older patients with newly diagnosed high risk (secondary) AML. ASCO Annual Meeting. 2016; Abstract 7000.
- Bross PF, Beitz J, Chen G, *et al.* Approval summary: gemtuzumab ozogamicin in relapsed acute myeloid leukemia. *Clin Cancer Res.* 2001; 7: 1490–6.
- Petersdorf SH, Kopecky KJ, Slovak M, *et al.* A phase 3 study of gemtuzumab ozogamicin during induction and postconsolidation therapy in younger patients with acute myeloid leukemia. *Blood.* 2013; 121: 4854–60.
- Burnett AK, Hills RK, Milligan D, *et al.* Identification of patients with acute myeloblastic leukemia who benefit from the addition of gemtuzumab ozogamicin: results of the MRC AML15 trial. *J Clin Oncol.* 2011; 29: 369–77.
- Castaigne S, Pautas C, Terre C, *et al.* Effect of gemtuzumab ozogamicin on survival of adult patients with de-novo acute myeloid leukaemia (ALFA-0701): a randomised, open-label, phase 3 study. *Lancet.* 2012; 379: 1508–16.
- Burnett AK, Russell NH, Hills RK, *et al.* Addition of gemtuzumab ozogamicin to induction chemotherapy improves survival in older patients with acute myeloid leukemia. *J Clin Oncol.* 2012; 30: 3924–31.
- Delaunay J, Recher C, Pigneux A, *et al.* Addition of gemtuzumab ozogamicin to chemotherapy improves event-free survival but not overall survival of AML patients with intermediate cytogenetics not eligible for allogeneic transplantation. Results of the GOELAMS AML 2006 IR study. *Blood.* 2011; 118: 79.
- Hills RK, Castaigne S, Appelbaum FR, *et al.* Addition of gemtuzumab ozogamicin to induction chemotherapy in adult patients with acute myeloid leukaemia: a meta-analysis of individual patient data from randomised controlled trials. *Lancet Oncol.* 2014; 15: 986–96.
- Nand S, Othus M, Godwin JE, *et al.* A phase 2 trial of azacitidine and gemtuzumab ozogamicin therapy in older patients with acute myeloid leukemia. *Blood.* 2013; 122: 3432–9.
- Daver N, Kantarjian H, Ravandi F, *et al.* A phase II study of decitabine and gemtuzumab ozogamicin in newly diagnosed and relapsed acute myeloid leukemia and high-risk myelodysplastic syndrome. *Leukemia.* 2016; 30: 268–73.
- Burnett AK, Hills RK, Hunter AE, *et al.* The addition of gemtuzumab ozogamicin to low-dose Ara-C improves remission rate but does not significantly prolong survival in older patients with acute myeloid leukaemia: results from the LRF AML14 and NCRI AML16 pick-a-winner comparison. *Leukemia.* 2013; 27: 75–81.
- Fathi AT, Erba HP, Lancet JE, *et al.* Vadas-tuximab talirine plus hypomethylating agents: a well-tolerated regimen with high remission rate in frontline older patients with acute myeloid leukemia (AML). *Blood.* 2016; 128: 591.
- Erba HP, Levy MY, Vasu S, *et al.* A phase 1b study of vadastuximab talirine in combination with 7 + 3 induction therapy for patients with newly diagnosed acute myeloid leukemia (AML). *Blood.* 2016; 128: 211.
- Manara E, Bisio V, Masetti R, *et al.* Core-binding factor acute myeloid leukemia in pediatric patients enrolled in the AIEOP AML 2002/01 trial: screening and prognostic impact of c-KIT mutations. *Leukemia.* 2014; 28: 1132–4.
- Chen W, Xie H, Wang H, *et al.* Prognostic significance of KIT mutations in core-binding factor acute myeloid leukemia: a systematic review and meta-analysis. *PLoS ONE.* 2016; 11: 1–19.
- Marcucci G, Geyer S, Zhao W, *et al.* Adding KIT inhibitor dasatinib (DAS) to chemotherapy overcomes the negative impact of KIT mutation/over-expression in core binding factor (CBF) acute myeloid leukemia (AML): results from CALGB 10801 (Alliance). *Blood.* 2014; 124: 8.

26. **Boissel N, Renneville A, Leguay T, et al.** Dasatinib in high-risk core binding factor acute myeloid leukemia in first complete remission: a French Acute Myeloid Leukemia Intergroup trial. *Haematologica*. 2015; 100: 780–5.
27. **Pratz KW, Sato T, Murphy KM, et al.** FLT3-mutant allelic burden and clinical status are predictive of response to FLT3 inhibitors in AML. *Blood*. 2010; 115: 1425–32.
28. **Ding L, Ley TJ, Larson DE, et al.** Clonal evolution in relapsed acute myeloid leukaemia revealed by whole-genome sequencing. *Nature*. 2012; 481: 506–10.
29. **Pratz KW, Levis M.** How I treat FLT3-mutated AML. *Blood*. 2017; 129: 565–71.
30. **Fischer T, Stone RM, Deangelo DJ, et al.** Phase IIB trial of oral midostaurin (PKC412), the FMS-like tyrosine kinase 3 receptor (FLT3) and multi-targeted kinase inhibitor, in patients with acute myeloid leukemia and high-risk myelodysplastic syndrome with either wild-type or mutated FLT3. *J Clin Oncol*. 2010; 28: 4339–45.
31. **Stone RM, Fischer T, Paquette R, et al.** Phase IB study of the FLT3 kinase inhibitor midostaurin with chemotherapy in younger newly diagnosed adult patients with acute myeloid leukemia. *Leukemia*. 2012; 26: 2061–8.
32. **Stone RM, Mandrekar SJ, Sanford BL, et al.** Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. *N Engl J Med*. 2017; 377: 454–64.
33. **Ravandi F, Cortes JE, Jones D, et al.** Phase I/II study of combination therapy with sorafenib, idarubicin, and cytarabine in younger patients with acute myeloid leukemia. *J Clin Oncol*. 2010; 28: 1856–62.
34. **Rollig C, Serve H, Huttmann A, et al.** Addition of sorafenib versus placebo to standard therapy in patients aged 60 years or younger with newly diagnosed acute myeloid leukaemia (SORAML): a multicentre, phase 2, randomised controlled trial. *Lancet Oncol*. 2015; 16: 1691–9.
35. **Serve H, Krug U, Wagner R, et al.** Sorafenib in combination with intensive chemotherapy in elderly patients with acute myeloid leukemia: results from a randomized, placebo-controlled trial. *J Clin Oncol*. 2013; 31: 3110–8.
36. **Ravandi F, Alattar ML, Grunwald MR, et al.** Phase 2 study of azacytidine plus sorafenib in patients with acute myeloid leukemia and FLT-3 internal tandem duplication mutation. *Blood*. 2013; 121: 4655–62.
37. **Brunner AM, Li S, Fathi AT, et al.** Haematopoietic cell transplantation with and without sorafenib maintenance for patients with FLT3-ITD acute myeloid leukaemia in first complete remission. *Br J Haematol*. 2016; 175: 496–504.
38. **Wander SA, Levis MJ, Fathi AT.** The evolving role of FLT3 inhibitors in acute myeloid leukemia: quizartinib and beyond. *Ther Adv Hematol*. 2014; 5: 65–77.
39. **Cortes JE, Kantarjian H, Foran JM, et al.** Phase I study of quizartinib administered daily to patients with relapsed or refractory acute myeloid leukemia irrespective of FMS-like tyrosine kinase 3-internal tandem duplication status. *J Clin Oncol*. 2013; 31: 3681–7.
40. **Levis MJ, Perl AE, Dombret H, et al.** Final results of a phase 2 open-label, monotherapy efficacy and safety study of quizartinib (AC220) in patients with FLT3-ITD positive or negative relapsed/refractory acute myeloid leukemia after second-line chemotherapy or hematopoietic stem cell transplantation. *Blood*. 2012; 120: 673.
41. **Schiller GJ, Tallman MS, Goldberg SL, et al.** Final results of a randomized phase 2 study showing the clinical benefit of quizartinib (AC220) in patients with FLT3-ITD positive relapsed or refractory acute myeloid leukemia. *J Clin Oncol*. 2014; 32: 7100.
42. **Levis MJ, Perl AE, Altman JK, et al.** Results of a first-in-human, phase I/II trial of ASP2215, a selective, potent inhibitor of FLT3/Axl in patients with relapsed or refractory (R/R) acute myeloid leukemia (AML). *J Clin Oncol*. 2015; 33: 7003.
43. **Altman JK, Perl AE, Cortes JE, et al.** Antileukemic activity and tolerability of ASP2215 80 mg and greater in FLT3 mutation-positive subjects with relapsed or refractory acute myeloid leukemia: results from a phase 1/2, open-label, dose-escalation/dose-response study. *Blood*. 2015; 126: 321.
44. **Altman JK, Perl AE, Cortes JE, et al.** Deep molecular response to gilteritinib to improve survival in FLT3 mutation-positive relapsed/refractory acute myeloid leukemia. *J Clin Oncol*. 2017; 35: 7003.
45. **Jain N, Curran E, Iyengar NM, et al.** Phase II study of the oral MEK inhibitor selumetinib in advanced acute myelogenous leukemia: a University of Chicago Phase II Consortium Trial. *Clin Cancer Res*. 2014; 20: 490–8.
46. **Borthakur G, Popplewell L, Boyiadzis M, et al.** Activity of the oral mitogen-activated protein kinase kinase inhibitor trametinib in RAS-mutant relapsed or refractory myeloid malignancies. *Cancer*. 2016; 122: 1871–9.
47. **Athuluri-Divakar SK, Vasquez-Del Carpio R, Dutta K, et al.** A small molecule RAS-mimetic disrupts RAS association with effector proteins to block signaling. *Cell*. 2016; 165: 643–55.
48. **Erba HP, Becker PS, Shami PJ, et al.** Dose escalation results of a phase 1b study of the MDM2 inhibitor AMG 232 with or without trametinib in patients (Pts) with relapsed/refractory (r/r) acute myeloid leukemia (AML). *J Clin Oncol*. 2017; 35: 7027.
49. **Strebhardt K.** Multifaceted polo-like kinases: drug targets and antitargets for cancer therapy. *Nat Rev Drug Discovery*. 2010; 9: 643–60.
50. **Döhner H, Lübbert M, Fiedler W, et al.** Randomized, phase 2 trial of low-dose cytarabine with or without volasertib in AML patients not suitable for induction therapy. *Blood*. 2014; 124: 1426–33.
51. **Dohner H, Symeonidis A, Sanz MA, et al.** Phase III randomized trial of volasertib plus low-dose cytarabine (LDAC) versus placebo plus LDAC in patients aged >65 years with previously untreated AML, ineligible for intensive therapy. *Haematologica*. 2016; 101(suppl.1): 185–186.
52. **Sedlacek HH.** Mechanisms of action of flavopiridol. *Crit Rev Oncol Hematol*. 2001; 38: 139–70.
53. **Karp JE, Ross DD, Yang W, et al.** Timed sequential therapy of acute leukemia with flavopiridol: *in vitro* model for a phase I clinical trial. *Clin Cancer Res*. 2003; 9: 307–15.
54. **Karp JE, Smith BD, Levis MJ, et al.** Sequential flavopiridol, cytosine arabinoside, and mitoxantrone: a phase II trial in adults with poor-risk acute myelogenous leukemia. *Clin Cancer Res*. 2007; 13: 4467–73.
55. **Karp JE, Passaniti A, Gojo I, et al.** Phase I and pharmacokinetic study of flavopiridol followed by 1-beta-D-arabinofuranosylcytosine and mitoxantrone in relapsed and refractory adult acute leukemias. *Clin Cancer Res*. 2005; 11: 8403–12.
56. **Karp JE, Blackford A, Smith BD, et al.** Clinical activity of sequential flavopiridol, cytosine arabinoside, and mitoxantrone for adults with newly diagnosed, poor-risk acute myelogenous leukemia. *Leuk Res*. 2010; 34: 877–82.
57. **Karp JE, Smith BD, Resar LS, et al.** Phase 1 and pharmacokinetic study of bolus-infusion flavopiridol followed by cytosine arabinoside and mitoxantrone for acute leukemias. *Blood*. 2011; 117: 3302–10.
58. **Karp JE, Garrett-Mayer E, Estey EH, et al.** Randomized phase II study of two schedules of flavopiridol given as timed

- sequential therapy with cytosine arabinoside and mitoxantrone for adults with newly diagnosed, poor-risk acute myelogenous leukemia. *Haematologica*. 2012; 97: 1736–42.
59. **Zeidner JF, Foster MC, Blackford AL, et al.** Randomized multicenter phase II study of flavopiridol (alvocidib), cytarabine, and mitoxantrone (FLAM) versus cytarabine/daunorubicin (7 + 3) in newly diagnosed acute myeloid leukemia. *Haematologica*. 2015; 100: 1172–9.
 60. **Litzow MR, Wang XV, Carroll MP, et al.** A randomized phase II trial of three novel regimens for relapsed/refractory acute myeloid leukemia (AML) demonstrates encouraging results with a flavopiridol-based regimen: results of Eastern Cooperative Oncology Group (ECOG) Trial E1906. *Blood*. 2014; 124: 3742.
 61. **Frohling S, Agrawal M, Jahn N, et al.** CDK4/6 inhibitor palbociclib for treatment of KMT2A-rearranged acute myeloid leukemia: interim analysis of the AMLSG 23-14 trial. *Blood*. 2016; 128: 1608.
 62. **Konopleva M, Pollyea DA, Pottluri J, et al.** A phase 2 study of ABT-199 (GDC-0199) in patients with acute myelogenous leukemia (AML). *Blood*. 2014; 124: 118.
 63. **Chan SM, Thomas D, Corces-Zimmerman MR, et al.** Isocitrate dehydrogenase 1 and 2 mutations induce BCL-2 dependence in acute myeloid leukemia. *Nat Med*. 2015; 21: 178–84.
 64. **Pollyea DA, Dinardo CD, Thirman MJ, et al.** Results of a phase 1b study of venetoclax plus decitabine or azacitidine in untreated acute myeloid leukemia patients ≥ 65 years ineligible for standard induction therapy. *J Clin Oncol*. 2016; 34: 7009.
 65. **Wei A, Strickland SA, Roboz GJ, et al.** Safety and efficacy of venetoclax plus low-dose cytarabine in treatment-naive patients aged ≥65 years with acute myeloid leukemia. *Blood*. 2016; 128: 102.
 66. **Uy GL, Rettig MP, Motabi IH, et al.** A phase 1/2 study of chemosensitization with the CXCR4 antagonist plerixafor in relapsed or refractory acute myeloid leukemia. *Blood*. 2012; 119: 3917–24.
 67. **Becker PS, Foran JM, Altman JK, et al.** Targeting the CXCR4 pathway: safety, tolerability and clinical activity of ulocuplumab (BMS-936564), an Anti-CXCR4 antibody, in relapsed/refractory acute myeloid leukemia. *Blood*. 2014; 124: 386.
 68. **Rodriguez-Paredes M, Esteller M.** Cancer epigenetics reaches mainstream oncology. *Nat Med*. 2011; 17(3): 330–339.
 69. **Fenaux P, Mufti GJ, Hellström-Lindberg E, et al.** Azacitidine prolongs overall survival compared with conventional care regimens in elderly patients with low bone marrow blast count acute myeloid leukemia. *J Clin Oncol*. 2010; 28: 562–9.
 70. **Kantarjian HM, Thomas XG, Dmoszynska A, et al.** Multicenter, randomized, open-label, phase III trial of decitabine versus patient choice, with physician advice, of either supportive care or low-dose cytarabine for the treatment of older patients with newly diagnosed acute myeloid leukemia. *J Clin Oncol*. 2012; 30: 2670–7.
 71. **Stein EM, Tallman MS.** Emerging therapeutic drugs for AML. *Blood*. 2016; 127: 71–8.
 72. **Figueroa ME, Abdel-Wahab O, Lu C, et al.** Leukemic IDH1 and IDH2 mutations result in a hypermethylation phenotype, disrupt TET2 function, and impair hematopoietic differentiation. *Cancer Cell*. 2010; 18: 553–67.
 73. **Stein EM, DiNardo CD, Pollyea DA, et al.** Enasidenib in mutant-IDH2 relapsed or refractory acute myeloid leukemia. *Blood*. 2017; 130: 722–31.
 74. **Birendra KC, DiNardo CD.** Evidence for clinical differentiation and differentiation syndrome in patients with acute myeloid leukemia and IDH1 mutations treated with the targeted mutant IDH1 inhibitor, AG-120. *Clin Lymphoma Myeloma Leuk*. 2016; 16: 460–5.
 75. **Roboz GJ, Rosenblat T, Arellano M, et al.** International randomized phase III study of elacytarabine versus investigator choice in patients with relapsed/refractory acute myeloid leukemia. *J Clin Oncol*. 2014; 32: 1919–26.
 76. **Odenike OM, Alkan S, Sher D, et al.** Histone deacetylase inhibitor romidepsin has differential activity in core binding factor acute myeloid leukemia. *Clin Cancer Res*. 2008; 14: 7095–101.
 77. **Garcia-Manero G, Othus M, Pagel JM, et al.** SWOG S1203: a randomized phase III study of standard cytarabine plus daunorubicin (7 + 3) therapy versus idarubicin with high dose cytarabine (IA) with or without vorinostat (IA+V) in younger patients with previously untreated acute myeloid leukemia (AML). *Blood*. 2016; 128: 901.
 78. **Mims AS, Klisovic RB, Garzon R, et al.** A novel regimen for acute myeloid leukemia with MLL partial tandem duplication: results of a phase 1 study NCI 8485. *Blood*. 2016; 128: 900.
 79. **Prebet T, Sun Z, Figueroa ME, et al.** Prolonged administration of azacitidine with or without entinostat for myelodysplastic syndrome and acute myeloid leukemia with myelodysplasia-related changes: results of the US Leukemia Intergroup trial E1905. *J Clin Oncol*. 2014; 32: 1242–8.
 80. **Stein EM, Garcia-Manero G, Rizzieri DA, et al.** A phase 1 study of the DOT1L inhibitor, pinometostat (EPZ-5676), in adults with relapsed or refractory leukemia: safety, clinical activity. Exposure and target inhibition. *Blood*. 2015; 126: 2547.
 81. **Filippakopoulos P, Qi J, Picaud S, et al.** Selective inhibition of BET bromodomains. *Nature*. 2010; 468: 1067–73.
 82. **Dawson MA, Prinjha RK, Dittmann A, et al.** Inhibition of BET recruitment to chromatin as an effective treatment for MLL-fusion leukaemia. *Nature*. 2011; 478: 529–33.
 83. **Fiskus W, Sharma S, Qi J, et al.** BET protein antagonist JQ1 is synergistically lethal with FLT3 tyrosine kinase inhibitor (TKI) and overcomes resistance to FLT3-TKI in AML cells expressing FLT-ITD. *Mol Cancer Ther*. 2014; 13: 2315–27.
 84. **Cancer Genome Atlas Research Network, Ley TJ, Miller C, et al.** Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. *N Engl J Med*. 2013; 368: 2059–74.
 85. **Li S, Garrett-Bakelman FE, Chung SS, et al.** Distinct evolution and dynamics of epigenetic and genetic heterogeneity in acute myeloid leukemia. *Nat Med*. 2016; 22: 792–9.
 86. **Paguirigan AL, Smith J, Meshinchi S, et al.** Single-cell genotyping demonstrates complex clonal diversity in acute myeloid leukemia. *Sci Transl Med*. 2015; 7: 281re2.
 87. **Parkin B, Ouillette P, Li Y, et al.** Clonal evolution and devolution after chemotherapy in adult acute myelogenous leukemia. *Blood*. 2013; 121: 369–77.
 88. **Bachas C, Schuurhuis GJ, Assaraf YG, et al.** The role of minor subpopulations within the leukemic blast compartment of AML patients at initial diagnosis in the development of relapse. *Leukemia*. 2012; 26: 1313–20.
 89. **Jan M, Snyder TM, Corces-Zimmerman MR, et al.** Clonal evolution of preleukemic hematopoietic stem cells precedes human acute myeloid leukemia. *Sci Transl Med*. 2012; 4: 149ra18.
 90. **Corces-Zimmerman MR, Hong WJ, Weissman IL, et al.** Preleukemic mutations in human acute myeloid leukemia affect epigenetic regulators and persist in remission. *Proc Natl Acad Sci USA*. 2014; 111: 2548–53.
 91. **Chou WC, Lei WC, Ko BS, et al.** The prognostic impact and stability of Isocitrate

- dehydrogenase 2 mutation in adult patients with acute myeloid leukemia. *Leukemia*. 2011; 25: 246–53.
92. Hou HA, Kuo YY, Liu CY, *et al*. DNMT3A mutations in acute myeloid leukemia: stability during disease evolution and clinical implications. *Blood*. 2012; 119: 559–68.
 93. Bacher U, Haferlach T, Schoch C, *et al*. Implications of NRAS mutations in AML: a study of 2502 patients. *Blood*. 2006; 107: 3847–53.
 94. Kottaridis PD, Gale RE, Langabeer SE, *et al*. Studies of FLT3 mutations in paired presentation and relapse samples from patients with acute myeloid leukemia: implications for the role of FLT3 mutations in leukemogenesis, minimal residual disease detection, and possible therapy with FLT3 inhibitors. *Blood*. 2002; 100: 2393–8.
 95. Cloos J, Goemans BF, Hess CJ, *et al*. Stability and prognostic influence of FLT3 mutations in paired initial and relapsed AML samples. *Leukemia*. 2006; 20: 1217–20.
 96. Shlush LI, Zandi S, Mitchell A, *et al*. Identification of pre-leukaemic haematopoietic stem cells in acute leukaemia. *Nature*. 2014; 506: 328–33.
 97. Ho TC, LaMere M, Stevens BM, *et al*. Evolution of acute myelogenous leukemia stem cell properties after treatment and progression. *Blood*. 2016; 128: 1671–8.
 98. Sarry JE, Murphy K, Perry R, *et al*. Human acute myelogenous leukemia stem cells are rare and heterogeneous when assayed in NOD/SCID/IL2R γ mac-deficient mice. *J Clin Invest*. 2011; 121: 384–95.
 99. Sanchez PV, Perry RL, Sarry JE, *et al*. A robust xenotransplantation model for acute myeloid leukemia. *Leukemia*. 2009; 23: 2109–17.
 100. Wunderlich M, Chou FS, Link KA, *et al*. AML xenograft efficiency is significantly improved in NOD/SCID-IL2RG mice constitutively expressing human SCF, GM-CSF and IL-3. *Leukemia*. 2010; 24: 1785–8.
 101. Kico JM, Spencer DH, Miller CA, *et al*. Functional heterogeneity of genetically defined subclones in acute myeloid leukemia. *Cancer Cell*. 2014; 25: 379–92.
 102. Jourdan E, Boissel N, Chevret S, *et al*. Prospective evaluation of gene mutations and minimal residual disease in patients with core binding factor acute myeloid leukemia. *Blood*. 2013; 121: 2213–23.
 103. Ivey A, Hills RK, Simpson MA, *et al*. Assessment of minimal residual disease in standard-risk AML. *N Engl J Med*. 2016; 374: 422–33.
 104. Shayegi N, Kramer M, Bornhauser M, *et al*. The level of residual disease based on mutant NPM1 is an independent prognostic factor for relapse and survival in AML. *Blood*. 2013; 122: 83–92.
 105. Corbacioglu A, Scholl C, Schlenk RF, *et al*. Prognostic impact of minimal residual disease in CBFB-MYH11-positive acute myeloid leukemia. *J Clin Oncol*. 2010; 28: 3724–9.
 106. Kico JM, Miller CA, Griffith M, *et al*. Association between mutation clearance after induction therapy and outcomes in acute myeloid leukemia. *JAMA*. 2015; 314: 811–22.
 107. Debarri H, Lebon D, Roumier C, *et al*. IDH1/2 but not DNMT3A mutations are suitable targets for minimal residual disease monitoring in acute myeloid leukemia patients: a study by the Acute Leukemia French Association. *Oncotarget*. 2015; 6: 42345–53.
 108. Konig H, Santos CD. Signal transduction in acute myeloid leukemia – implications for novel therapeutic concepts. *Curr Cancer Drug Targets*. 2015; 15: 803–21.
 109. Mamdani H, Santos CD, Konig H. Treatment of acute myeloid leukemia in elderly patients – a therapeutic dilemma. *J Am Med Dir Assoc*. 2016; 17: 581–7.
 110. Trunzer K, Pavlick AC, Schuchter L, *et al*. Pharmacodynamic effects and mechanisms of resistance to vemurafenib in patients with metastatic melanoma. *J Clin Oncol*. 2013; 31: 1767–74.
 111. Stewart EL, Tan SZ, Liu G, *et al*. Known and putative mechanisms of resistance to EGFR targeted therapies in NSCLC patients with EGFR mutations-a review. *Transl Lung Cancer Res*. 2015; 4: 67–81.
 112. Rexer BN, Arteaga CL. Intrinsic and acquired resistance to HER2-targeted therapies in HER2 gene-amplified breast cancer: mechanisms and clinical implications. *Crit Rev Oncog*. 2012; 17: 1–16.
 113. Lokody I. Drug resistance: overcoming resistance in acute myeloid leukaemia treatment. *Nat Rev Cancer*. 2014; 14: 452–3.
 114. Raetz E, Kovacovics T. Personalizing therapy: the beat AML trial. *The Hematologist*. 2017; 14: 1–2.
 115. Levis M. Midostaurin approved for FLT3-mutated AML. *Blood*. 2017; 129: 3403–6.