

# Innate lymphoid cells at the human maternal-fetal interface in spontaneous preterm labor

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**Problem:** Pathological inflammation is causally linked to preterm labor and birth, the leading cause of neonatal morbidity and mortality worldwide. Our aims were to investigate whether (i) the newly described family of innate lymphoid cells (ILCs) was present at the human maternal-fetal interface and (ii) ILC inflammatory subsets were associated with the pathological process of preterm labor.

**Methods of study:** Decidual leukocytes were isolated from women with preterm or term labor as well as from gestational age-matched non-labor controls. ILCs (CD15<sup>-</sup>CD14<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD56<sup>-</sup>CD11b<sup>-</sup>CD127<sup>+</sup> cells) and their subsets (ILC1, T-bet<sup>+</sup> ILCs; ILC2, GATA3<sup>+</sup> ILCs; and ILC3, RORγt<sup>+</sup> ILCs) and cytokine expression were identified in the decidual tissues using immunophenotyping.

**Results:** (i) The proportion of total ILCs was increased in the decidua parietalis of women with preterm labor; (ii) ILC1s were a minor subset of decidual ILCs during preterm and term gestations; (iii) ILC2s were the most abundant ILC subset in the decidua during preterm and term gestations; (iv) the proportion of ILC2s was increased in the decidua basalis of women with preterm labor; (v) the proportion of ILC3s was increased in the decidua parietalis of women with preterm labor; and (vi) during preterm labor, ILC3s had higher expression of IL-22, IL-17A, IL-13, and IFN-γ compared to ILC2s in the decidua.

**Conclusion:** ILC2s were the most abundant ILC subset at the human maternal-fetal interface during preterm and term gestations. Yet, during preterm labor, an increase in ILC2s and ILC3s was observed in the decidua basalis and decidua parietalis,

respectively. These findings provide evidence demonstrating a role for ILCs at the maternal-fetal interface during the pathological process of preterm labor.

#### KEYWORDS

cytokine, decidua, inflammation, innate immunity, interleukin, mucosal immunity, parturition, pregnancy, tolerance

## 1 | INTRODUCTION

Preterm birth, defined as birth prior to 37 weeks of gestation, is one of the most common obstetrical syndromes<sup>1-3</sup> and the leading cause of perinatal morbidity and mortality worldwide.<sup>4-8</sup> In 2013, 11.39% of all births in the USA were diagnosed as preterm.<sup>9</sup> Premature neonates are at an increased risk of short- and long-term morbidities that represent a substantial burden for society and the healthcare system.<sup>10-13</sup> Approximately 70% of all preterm births are preceded by spontaneous preterm labor<sup>1,14</sup> with multiple pathological processes involved.<sup>15</sup> Therefore, it is essential to determine the mechanisms implicated in spontaneous preterm labor and to develop novel therapies and strategies to prevent this syndrome.

Inflammation is implicated in the pathological process of spontaneous preterm labor.<sup>15-42</sup> Pathological inflammation can result from the activation of innate immunity<sup>43-55</sup> by microorganisms<sup>29,56-59</sup> or endogenous signals derived from necrosis or cellular stress,<sup>48,51,52,60-65</sup> termed damage-associated molecular patterns<sup>66</sup> or alarmins.<sup>67</sup> Moreover, it has been demonstrated that activation of the adaptive immune system can also lead to pathological inflammation.<sup>68</sup> Hence, characterization of innate and adaptive immune cells and their mediators may provide an understanding into the mechanisms that lead to spontaneous preterm labor.

Recently, a new family of immune cells that belong to the lymphoid lineage without expressing antigen-specific receptors was described and termed innate lymphoid cells (ILCs).<sup>69,70</sup> Such cells are defined by 3 main features: (i) the absence of recombination activating gene (RAG)-dependent rearranged antigen receptors; (ii) a lack of myeloid cell and dendritic cell phenotypical markers; and (iii) their lymphoid morphology.<sup>69,70</sup> Despite lacking antigen recognition capabilities, ILCs exhibit a functional diversity, which resembles that of T cells.<sup>71</sup> Two prototypical members of the ILC family have been previously described: natural killer (NK) cells<sup>72</sup> and lymphoid tissue-inducer (LTi) cells.<sup>73</sup> These 2 cell types, while distinct, are related through a shared requirement of the common cytokine receptor  $\gamma$ -chain (IL-2R $\gamma$ ) and the transcriptional repressor inhibitor of DNA binding 2 (ID2) for development.<sup>70,74</sup>

Distinct ILC subsets have since been described, which rely on signaling through the IL-7 receptor- $\alpha$  (IL-7R $\alpha$  or CD127) in addition to the abovementioned markers.<sup>70</sup> These new members of the ILC family were classified based on their functional similarities to T-cell subsets.<sup>70</sup> Group 1 ILCs (ILC1) are based on expression of the transcription factor T-bet and include NK cells as well as other IFN- $\gamma$ -producing Th1-like ILCs.<sup>75</sup> Group 2 ILCs (ILC2) are characterized by Th2-like expression of the cytokines IL-5 and IL-13 and are dependent on the transcription

factors GATA-binding protein 3 (GATA3) and retinoic acid receptor-related orphan receptor- $\alpha$  (ROR $\alpha$ ) for development.<sup>76,77</sup> Finally, group 3 ILCs (ILC3) include cells that produce Th17-like cytokines, eg, IL-17A and IL-22, and depend on expression of ROR $\gamma$ t.<sup>78</sup> Yet, there is plasticity among ILC subsets, which makes their characterization and identification challenging.<sup>79,80</sup>

Innate lymphoid cell subsets have been identified in the human decidua during early pregnancy,<sup>81-86</sup> however, whether such cells are present at the human maternal-fetal interface (decidua basalis and decidua parietalis<sup>87</sup>) during preterm and term gestations and are implicated in the pathological process of preterm labor is unknown.

The aims of this study were (i) to determine whether ILCs are present in the decidua of women with preterm and term gestations; (ii) to investigate whether the proportions of ILC subsets in the decidua are altered in women who underwent spontaneous preterm labor; and (iii) to characterize the cytokine signature of decidual ILCs in the pathological process of preterm labor.

## 2 | MATERIALS AND METHODS

### 2.1 | Human subjects, clinical specimens, and definitions

Human placental basal plate and chorioamniotic membrane samples were collected from patients within 30 minutes after delivery at Hutzel Women's Hospital in the Detroit Medical Center, Detroit, MI, USA, in partnership with Wayne State University School of Medicine and the Perinatology Research Branch, an intramural program of the Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, U S Department of Health and Human Services (NICHD/NIH/DHHS), Detroit, MI, USA. The collection and utilization of biological materials for research purposes were approved by the Institutional Review Boards of Wayne State University and NICHD. All participating women provided written informed consent. The study groups included women who delivered at term with (TIL) or without (TNL) labor and women who delivered preterm with (PTL) or without (PTNL) labor. Two separate sets of samples were utilized in this study: exploratory and confirmatory. The demographic and clinical characteristics of the study populations are shown in Tables 1 and 2. Preterm birth was defined as delivery before 37 weeks of gestation. Labor was defined by the presence of regular uterine contractions at a frequency of at least 2 contractions every 10 minutes with cervical changes resulting in delivery. For each case, several tissue sections of the

**TABLE 1** Demographic and clinical characteristics of the study population (exploratory set of samples)

	TNL (n = 8)	TIL (n = 25)	PTNL (n = 8)	PTL (n = 17)	P value
Age (y; median [IQR]) <sup>a</sup>	26 (23-32)	24 (22-27)	31.5 (24.3-34.5)	22 (21-25)	NS
Body mass index (kg/m <sup>2</sup> ; median [IQR]) <sup>a</sup>	28.5 (22.5-28.5)	29.2 (24.4-34)	22.5 (22.1-33.2)	27.3 (20.2-32.1)	NS
Gestational age at delivery (wk; median [IQR]) <sup>a</sup>	39 (38.6-39.3)	39.3 (38.3-40)	34.4 (31.3-36.5)	33.9 (31.3-34.9)	<.001
Race (n [%]) <sup>b</sup>					NS
African American	8 (100%)	25 (100%)	7 (87.5%)	15 (88.2%)	
Caucasian	0 (0%)	0 (0%)	1 (12.5%)	1 (5.9%)	
Other	0 (0%)	0 (0%)	0 (0%)	1 (5.9%)	
Primiparity (n [%]) <sup>b</sup>	0 (0%)	5 (20%)	2 (25%)	1 (5.9%)	NS
Cesarean section (n [%]) <sup>b</sup>	8 (100%)	2 (8%)	8 (100%)	4 (23.5%)	<.001
Acute chorioamnionitis (n [%]) <sup>b</sup>					
Acute subchorionitis/ chorionitis	0/8 (0%)	4/25 (16%)	0/8 (0%)	3/17 (17.6%)	NS
Acute chorioamnionitis	0/8 (0%)	7/25 (28%)	1/8 (12.5%)	2/17 (11.8%)	NS
Necrotizing chorioamnionitis	0/8 (0%)	0/25 (0%)	0/8 (0%)	3/17 (17.6%)	NS
Umbilical cord pathology (n [%]) <sup>b</sup>					
Umbilical phlebitis	0/8 (0%)	9/25 (36%)	0/8 (0%)	4/17 (23.5%)	NS
Umbilical arteritis	0/8 (0%)	1/25 (4%)	1/8 (12.5%)	1/17 (5.9%)	NS
Necrotizing funisitis	0/8 (0%)	0/25 (%)	0/8 (0%)	0/17 (0%)	NS

IQR, interquartile range.

<sup>a</sup>Kruskal-Wallis test.

<sup>b</sup>Fisher's exact test.

chorioamniotic membranes, umbilical cord, and placental disk were evaluated by pathologists blinded to the clinical outcome, according to the published criteria.<sup>88</sup> Patients with neonates having congenital or chromosomal abnormalities were excluded from this study.

## 2.2 | Decidual leukocyte isolation

Decidual leukocytes were isolated from the decidual tissue of patients from each study group as previously described.<sup>87</sup> Briefly, the decidua basalis was collected from the basal plate of the placenta, and the decidua parietalis was separated from the chorioamniotic membranes. The decidual tissues were homogenized in StemPro Accutase Cell Dissociation Reagent (Life Technologies, Grand Island, NY, USA) using a gentleMACS Dissociator (Miltenyi Biotec, San Diego, CA, USA). Homogenized tissues were incubated in Accutase for 45 minutes at 37°C with gentle agitation. After incubation, tissues were washed in 1× phosphate-buffered saline (PBS; Life Technologies) and filtered through a 100-µm cell strainer (Fisher Scientific, Durham, NC, USA). The resulting cell suspensions were centrifuged at 300 g for 10 minutes at 4°C. The decidual mononuclear cells were then separated using a density gradient (Ficoll-Paque Plus; GE Healthcare Biosciences, Piscataway, NJ, USA) by following the manufacturer's instructions. The cells collected from the mononuclear layer of the density gradient were washed with 1× PBS and immediately used for immunophenotyping.

## 2.3 | Immunophenotyping of decidual innate lymphoid cells

Mononuclear cell suspensions from decidual tissues were stained with BD Horizon Fixable Viability Stain 510 dye (BD Biosciences, San Jose, CA, USA) prior to immunophenotyping. Mononuclear cell suspensions were then washed with FACS staining buffer (CAT#554656; BD Biosciences) and incubated with 20 µL of human FcR Blocking Reagent (CAT#130-059-901; Miltenyi Biotec) in 80 µL of FACS staining buffer (BD Biosciences) for 10 minutes at 4°C. The cells were incubated with extracellular fluorochrome-conjugated anti-human monoclonal antibodies for 30 minutes at 4°C in the dark (Table S1). After extracellular staining, the cells were fixed and permeabilized using the Foxp3/Transcription Factor Staining Buffer Set (eBioscience, San Diego, CA, USA) prior to staining with intracellular and intranuclear antibodies (Table S1). Stained cells were washed and resuspended in 0.5 mL of FACS staining buffer and acquired using an LSRFortessa flow cytometer and FACSDiva 6.0 software (BD Biosciences). Data were analyzed using FlowJo software version 10 (TreeStar, Ashland, OR, USA).

## 2.4 | Statistics

Statistical analyses were conducted using SPSS software version 19.0 (IBM Corporation, Armonk, NY, USA). The Mann-Whitney *U*-test was

**TABLE 2** Demographic and clinical characteristics of the study population (confirmatory set of samples)

	TNL (n = 11)	TIL (n = 39)	PTNL (n = 12)	PTL (n = 28)	P value
Age (y; median [IQR]) <sup>a</sup>	26 (23-31)	24 (20-26)	31 (27.8-32.5)	27 (22.5-30)	.002
Body mass index (kg/m <sup>2</sup> ; median [IQR]) <sup>a</sup>	35.1 (29.2-39)	26.1 (23.1-30.7)	35.1 (29.4-36.3)	31.1 (25.4-39.6)	.009
Gestational age at delivery (wk; median [IQR]) <sup>a</sup>	39.1 (39-39.3)	39.3 (38.6-40.2)	34.1 (31.5-36.5)	34.6 (33.6-35.8)	<.001
Birth weight (g; median [IQR]) <sup>a</sup>	3370 (3125-3705)	3190 (2960-3352.5)	2017.5 (1393.8-2760)	2223 (1760-2420)	<.001
Race (n [%]) <sup>b</sup>					NS
African American	9 (81.8%)	35 (89.7%)	11 (91.7%)	20 (71.4%)	
Caucasian	2 (18.2%)	2 (5.1%)	1 (8.3%)	5 (17.9%)	
Other	0 (0%)	2 (5.1%)	0 (0%)	3 (10.7%)	
Primiparity (n [%]) <sup>b</sup>	0 (0%)	4 (10.3%)	1 (8.3%)	6 (21.4%)	NS
Cesarean section (n [%]) <sup>b</sup>	11 (100%)	2 (5.1%)	12 (100%)	12 (42.9%)	<.001
Acute chorioamnionitis (n [%]) <sup>b</sup>					
Acute subchorionitis/ chorionitis	1/11 (9.1%)	13/38 (34.2%) <sup>c</sup>	1/11 (9.1%) <sup>c</sup>	4/28 (14.3%)	NS
Acute chorioamnionitis	0/11 (0%)	9/38 (23.7%) <sup>c</sup>	0/11 (0%) <sup>c</sup>	4/28 (14.3%)	NS
Necrotizing chorioamnionitis	0/11 (0%)	0/38 (0%) <sup>c</sup>	0/11 (0%) <sup>c</sup>	1/28 (3.6%)	NS
Umbilical cord pathology (n [%]) <sup>b</sup>					
Umbilical phlebitis	0/11 (0%)	10/38 (26.3%) <sup>c</sup>	0/11 (0%) <sup>c</sup>	1/28 (3.6%)	.01
Umbilical arteritis	0/11 (0%)	2/38 (5.3%) <sup>c</sup>	0/11 (0%) <sup>c</sup>	4/28 (14.3%)	NS
Necrotizing funisitis	0/11 (0%)	0/38 (0%) <sup>c</sup>	0/11 (0%) <sup>c</sup>	2/28 (7.1%)	NS

IQR, interquartile range.

<sup>a</sup>Kruskal-Wallis test.

<sup>b</sup>Fisher's exact test.

<sup>c</sup>Calculated based on the available placental pathology information.

used for comparisons between study groups or different samples, and the Wilcoxon's signed-rank paired test was used for comparisons of different subpopulations from the same samples. For patient demographics, the Kruskal-Wallis test was performed for continuous variables and the Fisher's exact test for nominal variables. A *P*-value <.05 was considered statistically significant.

### 3 | RESULTS

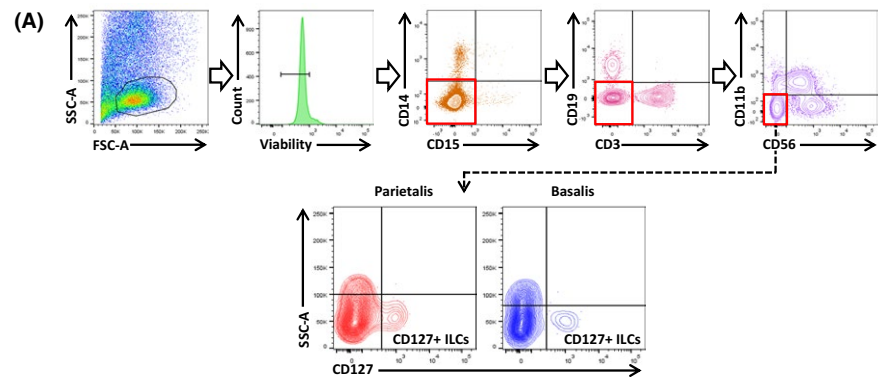
#### 3.1 | The proportion of ILCs is increased in the decidua parietalis of women with spontaneous preterm labor

We first conducted an exploratory study to determine the proportions and phenotypes of decidual ILCs in women with spontaneous term or preterm labor and non-labor gestational age-matched controls (Table 1). The gating strategy used to identify ILCs in the decidua parietalis and decidua basalis is shown in Figure 1A. ILCs were identified as CD15<sup>-</sup>CD14<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD56<sup>-</sup>CD11b<sup>-</sup>CD127<sup>+</sup> cells within the viability gate (Figure 1A). A higher proportion of total ILCs was observed in the decidua parietalis from women who underwent

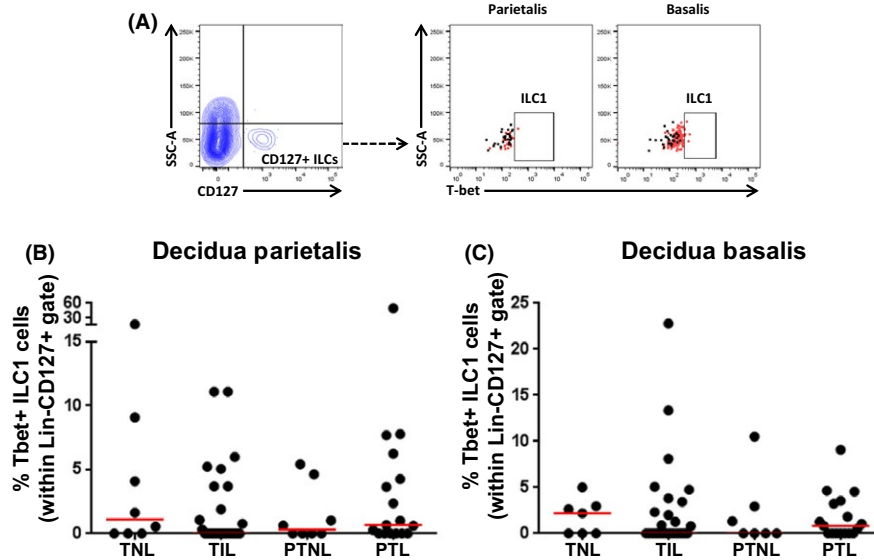
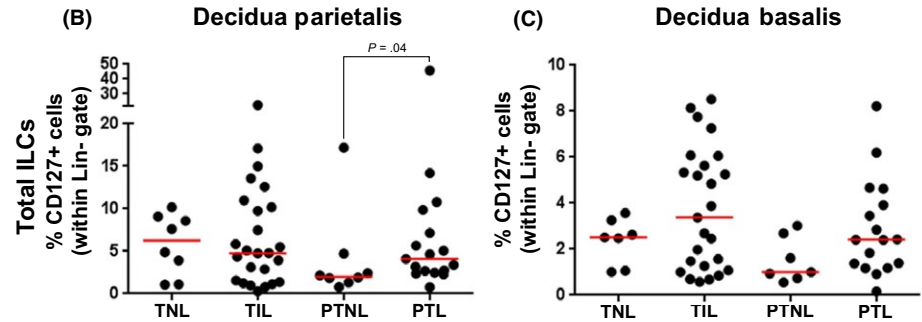
spontaneous preterm labor when compared to patients who delivered preterm without labor (Figure 1B). However, no significant differences were observed in ILCs in the decidua parietalis between women who underwent spontaneous labor at term compared to those who delivered at term in the absence of labor (Figure 1B). No significant differences were observed in the proportion of total ILCs in the decidua parietalis between women who underwent spontaneous preterm labor and those with labor at term (Figure 1B). There were no differences in the proportion of total ILCs in the decidua basalis among the study groups; yet, ILCs tended to be more abundant in the decidua basalis of women with preterm labor than in those who delivered preterm in the absence of labor (Figure 1C). These results show that ILCs are present at the human maternal-fetal interface during preterm and term gestations, and an increase in these cells is associated with spontaneous preterm labor.

#### 3.2 | ILC1s are a minor subset of decidual ILCs during preterm and term gestations

We continued our exploratory study by characterizing the populations of ILC1s, ILC2s, and ILC3s in the decidua parietalis and decidua



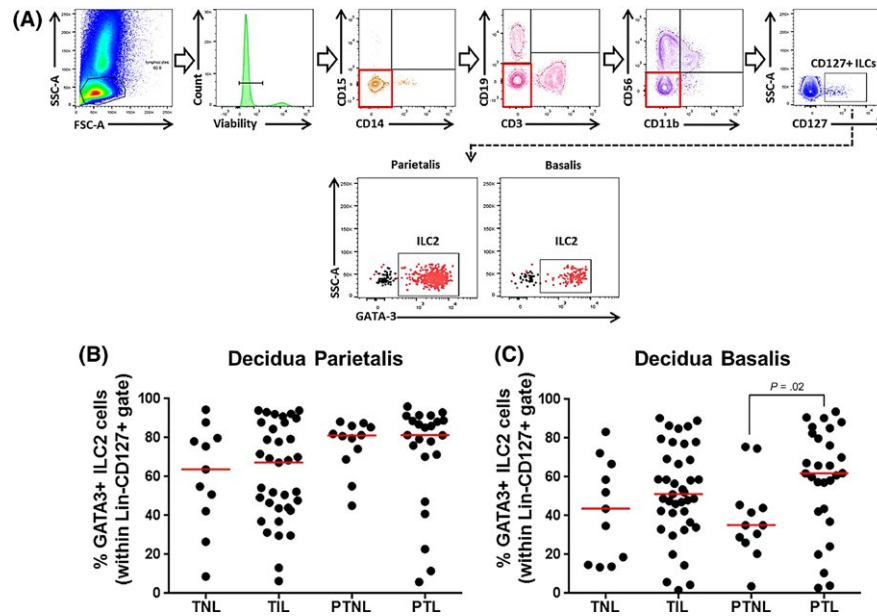
**FIGURE 1** Innate lymphoid cells (ILCs) are present at the human maternal-fetal interface. (A) Mononuclear cells were isolated from the decida parietalis and decida basalis. Flow cytometry gating strategy used for immunophenotyping of ILCs. ILCs (CD127<sup>+</sup>) were initially gated within the viability gate and lineage-negative (Lin<sup>-</sup>; CD15<sup>-</sup>CD14<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD56<sup>-</sup>CD11b<sup>-</sup>) gate. Red boxes represent the lineage-negative populations. Representative flow cytometry contour plots show the expression of CD127 by ILCs from the decida parietalis and decida basalis. The proportion of total ILCs in the decida parietalis (B) and decida basalis (C) of women who underwent spontaneous preterm (PTL) or term (TIL) labor and those who delivered preterm (PTNL) or term (TNL) without labor. n = 8-25 per group



**FIGURE 2** ILC1s are a minor population in the decida. (A) Mononuclear cells were isolated from the decida parietalis and decida basalis. Flow cytometry gating strategy used for immunophenotyping of ILC1s. ILCs (CD127<sup>+</sup>) were initially gated within the viability gate and lineage-negative (Lin<sup>-</sup>; CD15<sup>-</sup>CD14<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD56<sup>-</sup>CD11b<sup>-</sup>) gate. Representative flow cytometry contour plots show the expression of T-bet by ILC1s from the decida parietalis and decida basalis (red dots). Isotype controls are shown as black dots. The proportion of ILC1s in the decida parietalis (B) and decida basalis (C) of women who underwent spontaneous preterm (PTL) or term (TIL) labor and those who delivered preterm (PTNL) or term (TNL) without labor. n = 8-25 per group

basalis (Table 1). ILC1s were distinguished by the expression of the ILC1-associated transcription factor T-bet (Figure 2A). A very small proportion of ILC1s was identified in both the decida parietalis (median < 2%) and decida basalis (median < 3%) (Figure 2B,C). No significant differences in the proportion of decidual ILC1s were

found among the study groups (Figure 2B,C). These data indicate that ILC1s may not have a significant role in the decida during preterm and term gestations. Due to the small proportion of ILC1s detected in the decida, we did not pursue further examination of this population.



**FIGURE 3** ILC2s are the most abundant ILC subset in the decidua. (A) Mononuclear cells were isolated from the decidua parietalis and decidua basalis. Flow cytometry gating strategy used for immunophenotyping of ILC2s. ILCs (CD127<sup>+</sup>) were initially gated within the viability gate and the lineage-negative (Lin<sup>-</sup>; CD15<sup>-</sup>CD14<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD56<sup>-</sup>CD11b<sup>-</sup>) gate. Red boxes represent the lineage-negative populations. Representative flow cytometry contour plots show the expression of GATA3 by ILC2s from the decidua parietalis and decidua basalis (red dots). Isotype controls are shown as black dots. The proportion of ILC2s in the decidua parietalis (B) and decidua basalis (C) of women who underwent spontaneous preterm (PTL) or term (TIL) labor and those who delivered preterm (PTNL) or term (TNL) without labor. n = 11-39 per group

### 3.3 | ILC2s are the most abundant ILC subset in the decidua during preterm and term gestations

Our exploratory study revealed that ILC2s were the most abundant ILC subset in the human decidua (data not shown); therefore, we conducted a subsequent confirmatory study using a different and larger set of samples (Table 2). Decidual ILC2s were determined by the expression of GATA3 on CD127<sup>+</sup> ILCs (Figure 3A). In this second cohort, the ILC2 subset was also the most abundant population of ILCs in both the decidua parietalis and decidua basalis (Figure 3B,C). The proportion of ILC2s was higher in the decidua parietalis (median 60%-80%) than in the decidua basalis (median 35%-60%; Figure 3B,C). No differences in the proportion of ILC2s were observed in the decidua parietalis among the study groups; yet, ILC2s tended to be more abundant in preterm than in term gestations (Figure 3B). However, a higher proportion of ILC2s was found in the decidua basalis of women who underwent spontaneous preterm labor compared to non-labor controls (Figure 3C). There were no differences in the proportion of ILC2s in the decidua basalis of women who underwent spontaneous labor at term compared to those who delivered at term in the absence of labor or those who underwent spontaneous preterm labor (Figure 3C). These data show that ILC2s are the dominant ILC population in the decidua parietalis and decidua basalis and that an increase in these cells in the decidua basalis is associated with spontaneous preterm labor.

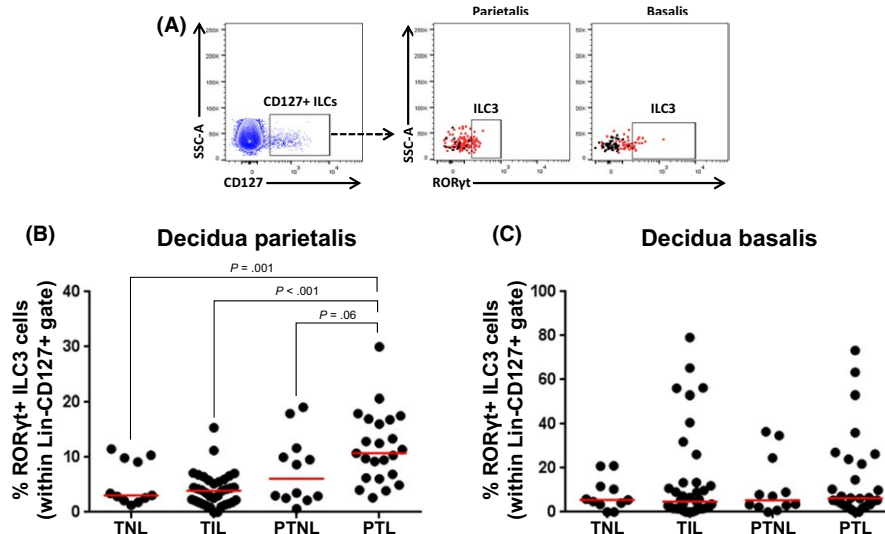
### 3.4 | The proportion of ILC3s is increased in the decidua parietalis of women who underwent spontaneous preterm labor

Next, we evaluated the presence of ILC3s in the decidual tissues (Table 2). The gating strategy used to determine the proportion of decidual ILC3s by the expression of ROR $\gamma$ t is shown in Figure 4A. The proportion of ILC3s was significantly increased in the decidua parietalis of women who underwent spontaneous preterm labor compared to that of women who delivered preterm in the absence of labor or those who delivered at term (Figure 4B). The proportion of ILC3s in the decidua basalis did not vary among the study groups (Figure 4C). These findings indicate that an increase of ILC3s in the decidua parietalis is associated with spontaneous preterm labor.

### 3.5 | Decidual ILC3s express high levels of IL-22, IL-17A, IL-13, and IFN- $\gamma$ in women with spontaneous preterm labor

To further characterize decidual ILC2s and ILC3s from women with spontaneous preterm labor, we evaluated the expression of cytokines associated with the 3 ILC subsets (Figure 5A). The mean fluorescence intensity (MFI) of IL-22 (Figure 5B,C), IL-13 (Figure 5F,G), and IFN- $\gamma$  (Figure 5H,I) was higher for ILC3s from the decidua parietalis and decidua basalis compared to that of ILC2s. The MFI of IL-17A was





**FIGURE 4** ILC3s are increased in the decidua parietalis of women who underwent spontaneous preterm labor. (A) Mononuclear cells were isolated from the decidua parietalis and decidua basalis. Flow cytometry gating strategy used for immunophenotyping of ILC3s. ILCs (CD127<sup>+</sup>) were initially gated within the viability gate and lineage-negative (Lin<sup>-</sup>; CD15<sup>-</sup>CD14<sup>-</sup>CD3<sup>-</sup>CD19<sup>-</sup>CD56<sup>-</sup>CD11b<sup>-</sup>) gate. Representative flow cytometry contour plots show the expression of RORγt by ILC3s from the decidua parietalis and decidua basalis (red dots). Isotype controls are shown as black dots. The proportion of ILC3s in the decidua parietalis (B) and decidua basalis (C) of women who underwent spontaneous preterm (PTL) or term (TIL) labor and those who delivered preterm (PTNL) or term (TNL) without labor. n = 11-39 per group

increased solely for ILC3s in the decidua parietalis compared to that of ILC2s (Figure 5D). No differences were observed in the expression of IL-5 between decidual ILC2s and ILC3s (data not shown). Together, these data show that ILC3s expressed higher levels of IL-22, IL-17A, IL-13, and IFN-γ than ILC2s in the decidua during the pathological process of preterm labor.

## 4 | DISCUSSION

### 4.1 | Principal findings of the study

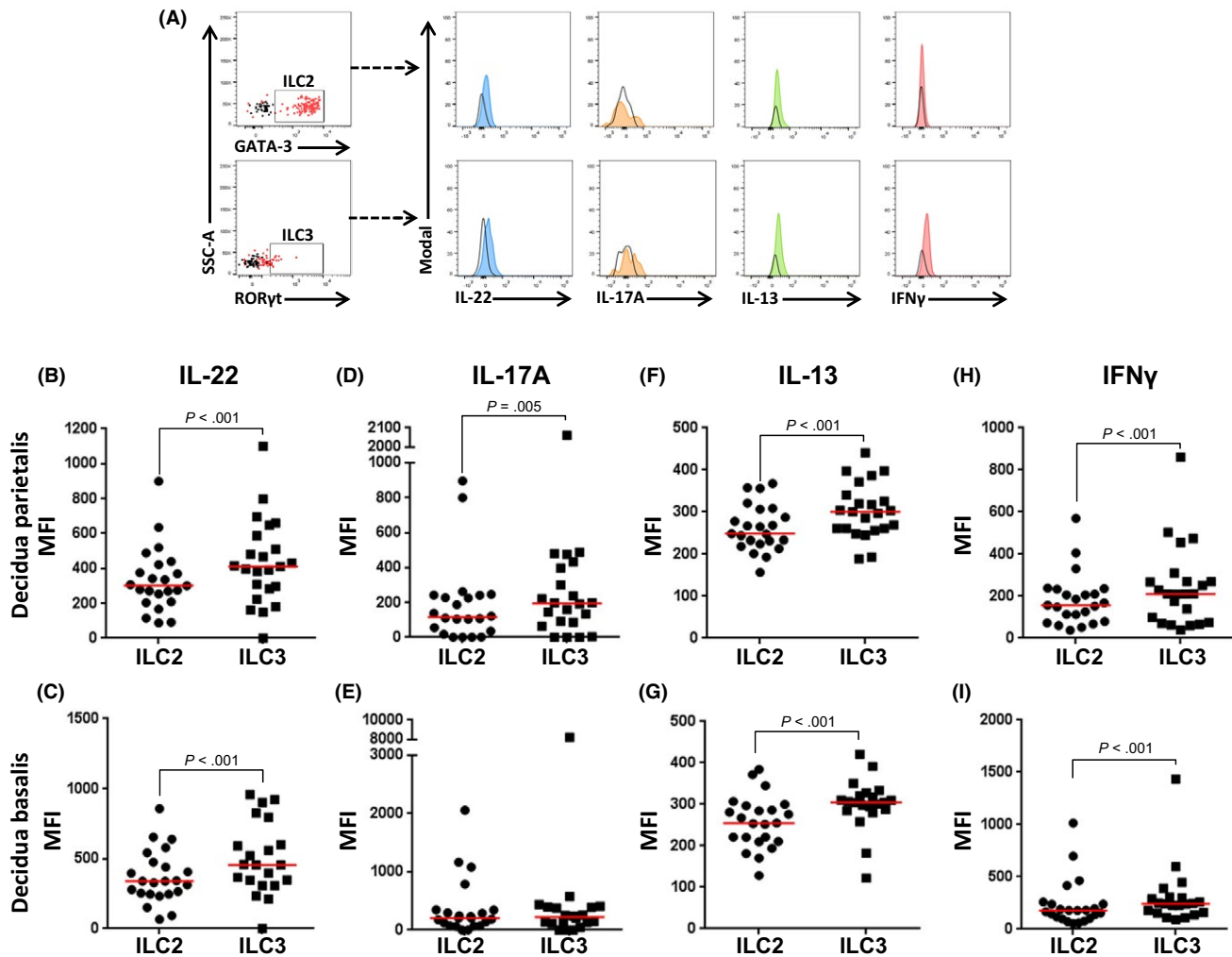
(i) The proportion of total CD127<sup>+</sup> ILCs was increased in the decidua parietalis of women who underwent spontaneous preterm labor; (ii) ILC1s were a minor subset of decidual ILCs during preterm and term gestations; (iii) ILC2s were the most abundant ILC subset in the decidua during preterm and term gestations; (iv) the proportion of ILC2s was increased in the decidua basalis of women who underwent spontaneous preterm labor; (v) the proportion of ILC3s was increased in the decidua parietalis of women who underwent spontaneous preterm labor; (vi) ILC3s had higher expression of IL-22, IL-17A, IL-13, and IFN-γ compared to ILC2s in the decidua of women who underwent spontaneous preterm labor; and (vii) decidual ILC2s and ILC3s had similar expression of IL-5 in the decidua of women who underwent spontaneous preterm labor. Collectively, these findings show that although ILC2s are the most abundant ILC subset at the human maternal-fetal interface during preterm and term gestations, an increase in ILC2s and ILC3s in the decidua basalis and decidua parietalis is observed during the pathological process of preterm labor.

### 4.2 | ILCs at the human maternal-fetal interface during preterm and term gestations

Innate lymphoid cells have been described in mucosal tissues, such as the lung, where they contribute to asthma and allergy-related processes,<sup>89-91</sup> and the gastrointestinal tract, where they provide defense against parasitic and microbial infections.<sup>92-95</sup> The discovery of enriched ILCs in mucosal tissues suggests that these cells are implicated in chronic intestinal inflammatory disorders, (e.g. Crohn's disease).<sup>96</sup> The association between ILCs and inflammatory diseases has led to the search for these cells at other sites of mucosal immunity, such as the reproductive tissues.<sup>81</sup> The 3 conventional ILC subsets have been described in the murine uterus during early and mid-gestation, although there is controversy as to which ILC subset is dominant during this period.<sup>81,82</sup> Indeed, ILC subsets have also been identified in the non-pregnant state in mice<sup>97</sup> and humans.<sup>82</sup> During early pregnancy, ILC1s and ILC3s are present at the human maternal-fetal interface, where such lymphoid cells crosstalk with neutrophils to modulate their migration and function.<sup>81,85</sup> In the study herein, we extended these observations by demonstrating that ILCs are present at the human maternal-fetal interface during term and preterm gestations (ie, third trimester). Together, these results indicate that ILCs are present at the human maternal-fetal interface; yet, their subsets are dynamically changing throughout gestation and with the onset of preterm labor.

### 4.3 | A role for ILC2s at the human maternal-fetal interface during the third trimester and in preterm labor

ILC2s are a distinct subset of ILCs that bear a functional resemblance to Th2 cells and were first described in a mouse model of helminth



**FIGURE 5** Cytokines expressed by decidual ILC2s and ILC3s in women who underwent spontaneous preterm labor. (A) Mononuclear cells were isolated from the decidua parietalis and decidua basalis. Representative flow cytometry histograms show the mean fluorescence intensity (MFI) expression of IL-22 (blue histograms), IL-17A (orange histograms), IL-13 (green histograms), and IFN- $\gamma$  (red histograms) by decidual ILC2s and ILC3s (red dots). Isotype controls are shown as black outline histograms or as black dots. The MFI of IL-22 (B and C), IL-17A (D and E), IL-13 (F and G), and IFN- $\gamma$  (H and I) expression by ILC2s and ILC3s in the decidua parietalis (upper row) and the decidua basalis (bottom row) of women who underwent spontaneous preterm labor.  $n = 23$

parasitic infection as novel producers of the Th2-like cytokines IL-4, IL-5, and IL-13.<sup>92</sup> ILC2s have been identified in the mesenteric lymph nodes, spleen, liver, intestines, and airways.<sup>98</sup> More recently, ILC2s were abundantly found in the murine uterus during early pregnancy,<sup>97</sup> where their presence may be regulated by female sex hormones (eg, estrogen).<sup>99</sup> Herein, we show that ILC2s are the most abundant ILC subset at the human maternal-fetal interface (decidua basalis and decidua parietalis). Tissue ILC2s display homeostatic functions through the secretion of tissue repair factors, such as amphiregulin and IL-13,<sup>100-102</sup> whose properties resemble those exhibited by M2 decidual macrophages in term and preterm gestations.<sup>50</sup> Therefore, we suggest that ILC2s, as well as M2 macrophages,<sup>103</sup> display homeostatic roles at the human maternal-fetal interface during the third trimester.

In addition to displaying homeostatic functions, ILC2s also exhibit proinflammatory functions. For example, ILC2s can contribute to the pathogenesis of ulcerative colitis, a chronic disease characterized by

elevated concentrations of Th2 cytokines, such as IL-4, IL-5, and IL-13.<sup>104,105</sup> Herein, we found that the proportion of ILC2s was increased in the decidua basalis of women who underwent spontaneous preterm labor. Interestingly, preterm labor is associated with chronic inflammatory lesions of the placenta<sup>106</sup> (eg, chronic deciduitis, infiltration of lymphocytes or plasma cells into the basal plate of the placenta<sup>107</sup>), which provides evidence that preterm labor can also be a chronic inflammatory disease. These data indicate that ILC2s may participate in the chronic inflammatory microenvironment that accompanies the pathological process of preterm labor in the decidua basalis.

#### 4.4 | A role for ILC3s at the human maternal-fetal interface during preterm labor

ILC3s were first described in the small intestine as unique innate cells that express the transcription factor ROR $\gamma$ t and the cytokine



IL-22<sup>108,109</sup> and were later shown to produce IL-17A.<sup>110,111</sup> ILC3s have been studied primarily in the context of inflammatory bowel disorders and other gastrointestinal diseases due to their presence in the gut mucosa and interactions with commensal bacteria.<sup>93,112</sup> Moreover, ROR $\gamma$ t<sup>+</sup> ILC3s can express MHC class II molecules and process and present microbial antigens to gut T cells.<sup>113</sup> This presentation of microbial peptides in the gut results in diminished commensal bacteria-specific T-cell responses.<sup>113</sup> In addition, IL-23-responsive ILC3s producing IL-17A and IL-22 have been implicated in the development of colitis in mouse models and in human studies.<sup>96</sup> In the current study, ILC3s were enriched in the decidua parietalis (decidua attached to the chorioamniotic membranes<sup>87</sup>) of women who underwent spontaneous preterm labor. Such ILCs expressed high levels of IL-22 and IL-17A, suggesting that this subset is implicated in the localized inflammatory milieu that accompanies the pathological process of preterm labor.

Herein, we found that decidual ILC3s expressed a high level of IL-13 (a cytokine mainly produced by ILC2s<sup>91,92</sup>) during the process of preterm labor. In line with this observation, previous studies have demonstrated that IL-13 is expressed by the placental<sup>114</sup> and decidual tissues.<sup>115,116</sup> IL-13 promotes the activation and migration of dendritic cells to the draining lymph nodes, leading to the differentiation of Th2 cells.<sup>117</sup> This cytokine is also important for tissue repair responses, reduction of ILC3-mediated inflammation, and defense against parasitic infections.<sup>101,118</sup> Together, these results allow us to propose that decidual ILC3s express a high level of the anti-inflammatory cytokine IL-13 to regulate the inflammatory responses exhibited by such cells.

Interestingly, ILC3s are also capable of expressing IFN- $\gamma$  when exposed to IL-12, IL-18, and/or IL-1 $\beta$  by upregulating T-bet, suggesting that a small proportion of ILC1s are derived from ILC3s.<sup>79</sup> In the current study, we found that decidual ILC3s expressed a high level of IFN- $\gamma$ , besides expressing ILC3 cytokines. Together, these results indicate that decidual ILC3s express ILC1 and ILC2 cytokines, supporting the concept that immune cells at the maternal-fetal interface display unique phenotypical characteristics.<sup>119</sup>

## 5 | SUMMARY

In the current study, we provide evidence that ILCs are present at the human maternal-fetal interface; yet, their subsets are dynamically changing throughout late gestation and with the onset of preterm labor. First, we found that ILC2s are the most abundant ILC subset at the human maternal-fetal interface and that their proportion in the decidua basalis (decidua attached to the placenta) increased in women who underwent spontaneous preterm labor. Next, we showed that ILC3s are enriched in the decidua parietalis (decidua attached to the chorioamniotic membranes) in women who underwent spontaneous preterm labor. Lastly, we demonstrated that ILC3s expressed high levels of IL-22, IL-17A, IL-13, and IFN- $\gamma$  at the human maternal-fetal interface during preterm labor. Collectively, these data provide the first evidence demonstrating a

role for ILCs at the human maternal-fetal interface during the pathological process of preterm labor.

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## CONFLICT OF INTEREST

The authors declare no financial interests in any of the work submitted here.

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## REFERENCES

- Romero R, Mazor M, Munoz H, Gomez R, Galasso M, Sherer DM. The preterm labor syndrome. *Ann NY Acad Sci.* 1994;734:414-429.
- Romero R, Espinoza J, Kusanovic JP, et al. The preterm parturition syndrome. *BJOG.* 2006;113(Suppl 3):17-42.
- Gotsch F, Gotsch F, Romero R, et al. The preterm parturition syndrome and its implications for understanding the biology, risk assessment, diagnosis, treatment and prevention of preterm birth. *J Matern Fetal Neonatal Med.* 2009;22(Suppl 2):5-23.
- Muglia LJ, Katz M. The enigma of spontaneous preterm birth. *N Engl J Med.* 2010;362:529-535.
- Blencowe H, Cousens S, Oestergaard MZ, et al. National, regional, and worldwide estimates of preterm birth rates in the year 2010 with time trends since 1990 for selected countries: a systematic analysis and implications. *Lancet.* 2012;379:2162-2172.
- Hamilton BE, Hoyert DL, Martin JA, Strobino DM, Guyer B. Annual summary of vital statistics: 2010-2011. *Pediatrics.* 2013;131:548-558.
- Monier I, Ancel PY, Ego A, et al. Fetal and neonatal outcomes of preterm infants born before 32 weeks of gestation according to antenatal vs postnatal assessments of restricted growth. *Am J Obstet Gynecol.* 2017;216:516.e1-516.e10.
- Travers CP, Carlo WA, McDonald SA, et al. Mortality and pulmonary outcomes of extremely preterm infants exposed to antenatal corticosteroids. *Am J Obstet Gynecol.* 2017;218:130.e1-130.e13.
- Martin JA, Hamilton BE, Osterman MJ, Curtin SC, Matthews TJ. Births: final data for 2013. *Natl Vital Stat Rep.* 2015;64:1-65.

10. Lubow JM, How HY, Habli M, Maxwell R, Sibai BM. Indications for delivery and short-term neonatal outcomes in late preterm as compared with term births. *Am J Obstet Gynecol.* 2009;200:e30-e33.
11. Mwaniki MK, Atieno M, Lawn JE, Newton CR. Long-term neurodevelopmental outcomes after intrauterine and neonatal insults: a systematic review. *Lancet.* 2012;379:445-452.
12. Manuck TA, Rice MM, Bailit JL, et al. Preterm neonatal morbidity and mortality by gestational age: a contemporary cohort. *Am J Obstet Gynecol.* 2016;215:103.e1-103.e14.
13. Chevallier M, Debillon T, Pierrat V, et al. Leading causes of preterm delivery as risk factors for intraventricular hemorrhage in very preterm infants: results of the EPIPAGE 2 cohort study. *Am J Obstet Gynecol.* 2017;216:518.e1-518.e12.
14. Goldenberg RL, Culhane JF, Iams JD, Romero R. Epidemiology and causes of preterm birth. *Lancet.* 2008;371:75-84.
15. Romero R, Dey SK, Fisher SJ. Preterm labor: one syndrome, many causes. *Science.* 2014;345:760-765.
16. Romero R, Sirtori M, Oyarzun E, et al. Infection and labor. V. Prevalence, microbiology, and clinical significance of intraamniotic infection in women with preterm labor and intact membranes. *Am J Obstet Gynecol.* 1989;161:817-824.
17. Romero R, Avila C, Santhanam U, Sehgal PB. Amniotic fluid interleukin 6 in preterm labor. Association with infection. *J Clin Invest.* 1990;85:1392-1400.
18. Romero R, Quintero R, Nores J, et al. Amniotic fluid white blood cell count: a rapid and simple test to diagnose microbial invasion of the amniotic cavity and predict preterm delivery. *Am J Obstet Gynecol.* 1991;165:821-830.
19. Andrews WW, Hauth JC, Goldenberg RL, Gomez R, Romero R, Cassell GH. Amniotic fluid interleukin-6: correlation with upper genital tract microbial colonization and gestational age in women delivered after spontaneous labor versus indicated delivery. *Am J Obstet Gynecol.* 1995;173:606-612.
20. Romero R, Gomez R, Chaiworapongsa T, Conoscenti G, Kim JC, Kim YM. The role of infection in preterm labour and delivery. *Paediatr Perinat Epidemiol.* 2001;15(Suppl 2):41-56.
21. Yoon BH, Chang JW, Romero R. Isolation of *Ureaplasma urealyticum* from the amniotic cavity and adverse outcome in preterm labor. *Obstet Gynecol.* 1998;92:77-82.
22. Yoon BH, Jun JK, Park KH, Syn HC, Gomez R, Romero R. Serum C-reactive protein, white blood cell count, and amniotic fluid white blood cell count in women with preterm premature rupture of membranes. *Obstet Gynecol.* 1996;88:1034-1040.
23. Yoon BH, Romero R, Kim M, et al. Clinical implications of detection of *Ureaplasma urealyticum* in the amniotic cavity with the polymerase chain reaction. *Am J Obstet Gynecol.* 2000;183:1130-1137.
24. Jacobsson B, Mattsby-Baltzer I, Andersch B, et al. Microbial invasion and cytokine response in amniotic fluid in a Swedish population of women in preterm labor. *Acta Obstet Gynecol Scand.* 2003;82:120-128.
25. Shim SS, Romero R, Hong JS, et al. Clinical significance of intra-amniotic inflammation in patients with preterm premature rupture of membranes. *Am J Obstet Gynecol.* 2004;191:1339-1345.
26. Yoon BH, Romero R, Moon JB, et al. Clinical significance of intra-amniotic inflammation in patients with preterm labor and intact membranes. *Am J Obstet Gynecol.* 2001;185:1130-1136.
27. Cobo T, Palacio M, Martinez-Terron M, et al. Clinical and inflammatory markers in amniotic fluid as predictors of adverse outcomes in preterm premature rupture of membranes. *Am J Obstet Gynecol.* 2011;205:126.e1-126.e8.
28. Lee SE, Romero R, Jung H, Park CW, Park JS, Yoon BH. The intensity of the fetal inflammatory response in intraamniotic inflammation with and without microbial invasion of the amniotic cavity. *Am J Obstet Gynecol.* 2007;197:294.e1-294.e6.
29. Romero R, Espinoza J, Goncalves LF, Kusanovic JP, Friel L, Hassan S. The role of inflammation and infection in preterm birth. *Semin Reprod Med.* 2007;25:21-39.
30. Romero R, Miranda J, Chaiworapongsa T, et al. A novel molecular microbiologic technique for the rapid diagnosis of microbial invasion of the amniotic cavity and intra-amniotic infection in preterm labor with intact membranes. *Am J Reprod Immunol.* 2014;71:330-358.
31. Romero R, Miranda J, Chaiworapongsa T, et al. Sterile intra-amniotic inflammation in asymptomatic patients with a sonographic short cervix: prevalence and clinical significance. *J Matern Fetal Neonatal Med.* 2014;28:1-17.
32. Romero R, Miranda J, Chaiworapongsa T, et al. Prevalence and clinical significance of sterile intra-amniotic inflammation in patients with preterm labor and intact membranes. *Am J Reprod Immunol.* 2014;72:458-474.
33. Combs CA, Gravett M, Garite TJ, et al. Amniotic fluid infection, inflammation, and colonization in preterm labor with intact membranes. *Am J Obstet Gynecol.* 2014;210:125.e1-125.e15.
34. Romero R, Miranda J, Chaemsaitong P, et al. Sterile and microbial-associated intra-amniotic inflammation in preterm prelabor rupture of membranes. *J Matern Fetal Neonatal Med.* 2015;28:1394-1409.
35. Gervasi MT, Chaiworapongsa T, Naccasha N, et al. Phenotypic and metabolic characteristics of maternal monocytes and granulocytes in preterm labor with intact membranes. *Am J Obstet Gynecol.* 2001;185:1124-1129.
36. Gervasi MT, Chaiworapongsa T, Naccasha N, et al. Maternal intravascular inflammation in preterm premature rupture of membranes. *J Matern Fetal Neonatal Med.* 2002;11:171-175.
37. Kacerovsky M, Musilova I, Andrys C, et al. Prelabor rupture of membranes between 34 and 37 weeks: the intraamniotic inflammatory response and neonatal outcomes. *Am J Obstet Gynecol.* 2014;210:325.e1-325.e10.
38. Yoneda N, Yoneda S, Niimi H, et al. Polymicrobial amniotic fluid infection with mycoplasma/ureaplasma and other bacteria induces severe intra-amniotic inflammation associated with poor perinatal prognosis in preterm labor. *Am J Reprod Immunol.* 2016;75:112-125.
39. Oh KJ, Kim SM, Hong JS, et al. Twenty-four percent of patients with clinical chorioamnionitis in preterm gestations have no evidence of either culture-proven intraamniotic infection or intraamniotic inflammation. *Am J Obstet Gynecol.* 2017;216:604.e1-604.e11.
40. Romero R, Chaemsaitong P, Chaiyasit N, et al. CXCL10 and IL-6: markers of two different forms of intra-amniotic inflammation in preterm labor. *Am J Reprod Immunol.* 2017;78:e12685.
41. Gomez-Lopez N, Romero R, Xu Y, et al. Are amniotic fluid neutrophils in women with intraamniotic infection and/or inflammation of fetal or maternal origin? *Am J Obstet Gynecol.* 2017;217:693.e1-693.e16.
42. Tarca AL, Fitzgerald W, Chaemsaitong P, et al. The cytokine network in women with an asymptomatic short cervix and the risk of preterm delivery. *Am J Reprod Immunol.* 2017;78:e12686.
43. Kim YM, Romero R, Chaiworapongsa T, et al. Toll-like receptor-2 and -4 in the chorioamniotic membranes in spontaneous labor at term and in preterm parturition that are associated with chorioamnionitis. *Am J Obstet Gynecol.* 2004;191:1346-1355.
44. Koga K, Cardenas I, Aldo P, et al. Activation of TLR3 in the trophoblast is associated with preterm delivery. *Am J Reprod Immunol.* 2009;61:196-212.
45. Cardenas I, Means RE, Aldo P, et al. Viral infection of the placenta leads to fetal inflammation and sensitization to bacterial products predisposing to preterm labor. *J Immunol.* 2010;185:1248-1257.
46. Cardenas I, Mulla MJ, Myrtolli K, et al. Nod1 activation by bacterial iE-DAP induces maternal-fetal inflammation and preterm labor. *J Immunol.* 2011;187:980-986.
47. Romero R, Chaiworapongsa T, Alpay Savasan Z, et al. Damage-associated molecular patterns (DAMPs) in preterm labor with intact

- membranes and preterm PROM: a study of the alarmin HMGB1. *J Matern Fetal Neonatal Med.* 2011;24:1444-1455.
48. Vaisbuch E, Romero R, Erez O, et al. Activation of the alternative pathway of complement is a feature of pre-term parturition but not of spontaneous labor at term. *Am J Reprod Immunol.* 2010;63:318-330.
  49. Jaiswal MK, Agrawal V, Mallers T, Gilman-Sachs A, Hirsch E, Beaman KD. Regulation of apoptosis and innate immune stimuli in inflammation-induced preterm labor. *J Immunol.* 2013;191:5702-5713.
  50. Xu Y, Romero R, Miller D, et al. An M1-like macrophage polarization in decidual tissue during spontaneous preterm labor that is attenuated by rosiglitazone treatment. *J Immunol.* 2016;196:2476-2491.
  51. St Louis D, Romero R, Plazyo O, et al. Invariant NKT cell activation induces late preterm birth that is attenuated by rosiglitazone. *J Immunol.* 2016;196:1044-1059.
  52. Gomez-Lopez N, Romero R, Arenas-Hernandez M, et al. In vivo activation of invariant natural killer T cells induces systemic and local alterations in T-cell subsets prior to preterm birth. *Clin Exp Immunol.* 2017;189:211-225.
  53. Anders AP, Gaddy JA, Doster RS, Aronoff DM. Current concepts in maternal-fetal immunology: recognition and response to microbial pathogens by decidual stromal cells. *Am J Reprod Immunol.* 2017;77:e12623.
  54. Gomez-Lopez N, Romero R, Xu Y, et al. A role for the inflammasome in spontaneous preterm labor with acute histologic chorioamnionitis. *Reprod Sci.* 2017;24:1382-1401.
  55. Negishi Y, Shima Y, Takeshita T, Takahashi H. Distribution of invariant natural killer T cells and dendritic cells in late pre-term birth without acute chorioamnionitis. *Am J Reprod Immunol.* 2017;77:e12658.
  56. Thaxton JE, Nevers TA, Sharma S. TLR-mediated preterm birth in response to pathogenic agents. *Infect Dis Obstet Gynecol.* 2010;2010:pii378472.
  57. Racicot K, Kwon JY, Aldo P, et al. Type I interferon regulates the placental inflammatory response to bacteria and is targeted by virus: mechanism of polymicrobial infection-induced preterm birth. *Am J Reprod Immunol.* 2016;75:451-460.
  58. Gomez-Lopez N, Romero R, Xu Y, et al. Neutrophil extracellular traps in the amniotic cavity of women with intra-amniotic infection: a new mechanism of host defense. *Reprod Sci.* 2017;24:1139-1153.
  59. Gomez-Lopez N, Romero R, Garcia-Flores V, et al. Amniotic fluid neutrophils can phagocytize bacteria: a mechanism for microbial killing in the amniotic cavity. *Am J Reprod Immunol.* 2017;78:e12723.
  60. Friel LA, Romero R, Edwin S, et al. The calcium binding protein, S100B, is increased in the amniotic fluid of women with intra-amniotic infection/inflammation and preterm labor with intact or ruptured membranes. *J Perinat Med.* 2007;35:385-393.
  61. Chaiworapongsa T, Erez O, Kusanovic JP, et al. Amniotic fluid heat shock protein 70 concentration in histologic chorioamnionitis, term and preterm parturition. *J Matern Fetal Neonatal Med.* 2008;21:449-461.
  62. Romero R, Espinoza J, Hassan S, et al. Soluble receptor for advanced glycation end products (sRAGE) and endogenous secretory RAGE (esRAGE) in amniotic fluid: modulation by infection and inflammation. *J Perinat Med.* 2008;36:388-398.
  63. Lee SE, Park IS, Romero R, Yoon BH. Amniotic fluid prostaglandin F2 increases even in sterile amniotic fluid and is an independent predictor of impending delivery in preterm premature rupture of membranes. *J Matern Fetal Neonatal Med.* 2009;22:880-886.
  64. Gomez-Lopez N, Romero R, Plazyo O, et al. Intra-amniotic administration of HMGB1 induces spontaneous preterm labor and birth. *Am J Reprod Immunol.* 2016;75:3-7.
  65. Plazyo O, Romero R, Unkel R, et al. HMGB1 induces an inflammatory response in the chorioamniotic membranes that is partially mediated by the inflammasome. *Biol Reprod.* 2016;95:130.
  66. Lotze MT, Deisseroth A, Rubartelli A. Damage associated molecular pattern molecules. *Clin Immunol.* 2007;124:1-4.
  67. Oppenheim JJ, Yang D. Alarmins: chemotactic activators of immune responses. *Curr Opin Immunol.* 2005;17:359-365.
  68. Gomez-Lopez N, Romero R, Arenas-Hernandez M, et al. In vivo T-cell activation by a monoclonal alphaCD3epsilon antibody induces preterm labor and birth. *Am J Reprod Immunol.* 2016;76:386-390.
  69. Spits H, Cupedo T. Innate lymphoid cells: emerging insights in development, lineage relationships, and function. *Annu Rev Immunol.* 2012;30:647-675.
  70. Spits H, Artis D, Colonna M, et al. Innate lymphoid cells—a proposal for uniform nomenclature. *Nat Rev Immunol.* 2013;13:145-149.
  71. Klose CS, Artis D. Innate lymphoid cells as regulators of immunity, inflammation and tissue homeostasis. *Nat Immunol.* 2016;17:765-774.
  72. Kiessling R, Klein E, Pross H, Wigzell H. "Natural" killer cells in the mouse. II. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Characteristics of the killer cell. *Eur J Immunol.* 1975;5:117-121.
  73. Mebius RE, Rennert P, Weissman IL. Developing lymph nodes collect CD4+ CD3- LTbeta+ cells that can differentiate to APC, NK cells, and follicular cells but not T or B cells. *Immunity.* 1997;7:493-504.
  74. Yokota Y, Mansouri A, Mori S, et al. Development of peripheral lymphoid organs and natural killer cells depends on the helix-loop-helix inhibitor Id2. *Nature.* 1999;397:702-706.
  75. Powell N, Walker AW, Stolarczyk E, et al. The transcription factor T-bet regulates intestinal inflammation mediated by interleukin-7 receptor+ innate lymphoid cells. *Immunity.* 2012;37:674-684.
  76. Mjosberg J, Bernink J, Golebski K, et al. The transcription factor GATA3 is essential for the function of human type 2 innate lymphoid cells. *Immunity.* 2012;37:649-659.
  77. Wong SH, Walker JA, Jolin HE, et al. Transcription factor RORalpha is critical for nuocyte development. *Nat Immunol.* 2012;13:229-236.
  78. Eberl G, Marmon S, Sunshine MJ, Rennert PD, Choi Y, Littman DR. An essential function for the nuclear receptor RORgamma(t) in the generation of fetal lymphoid tissue inducer cells. *Nat Immunol.* 2004;5:64-73.
  79. Bernink JH, Peters CP, Munneke M, et al. Human type 1 innate lymphoid cells accumulate in inflamed mucosal tissues. *Nat Immunol.* 2013;14:221-229.
  80. Klose CS, Kiss EA, Schwierzeck V, et al. A T-bet gradient controls the fate and function of CCR6-RORgammat+ innate lymphoid cells. *Nature.* 2013;494:261-265.
  81. Vacca P, Montaldo E, Croxatto D, et al. Identification of diverse innate lymphoid cells in human decidua. *Mucosal Immunol.* 2015;8:254-264.
  82. Doisne JM, Balmas E, Boulenouar S, et al. Composition, development, and function of uterine innate lymphoid cells. *J Immunol.* 2015;195:3937-3945.
  83. Zhang J, Dunk C, Croy AB, Lye SJ. To serve and to protect: the role of decidual innate immune cells on human pregnancy. *Cell Tissue Res.* 2016;363:249-265.
  84. Boulenouar S, Doisne JM, Sferruzzi-Perri A, et al. The residual innate lymphoid cells in NFIL3-deficient mice support suboptimal maternal adaptations to pregnancy. *Front Immunol.* 2016;7:43.
  85. Croxatto D, Micheletti A, Montaldo E, et al. Group 3 innate lymphoid cells regulate neutrophil migration and function in human decidua. *Mucosal Immunol.* 2016;9:1372-1383.
  86. Gaynor LM, Colucci F. Uterine natural killer cells: functional distinctions and influence on pregnancy in humans and mice. *Front Immunol.* 2017;8:467.
  87. Xu Y, Plazyo O, Romero R, Hassan SS, Gomez-Lopez N. Isolation of leukocytes from the human maternal-fetal interface. *J Vis Exp.* 2015;99:e52863.
  88. Kim CJ, Romero R, Chaemsaitong P, Chaiyasit N, Yoon BH, Kim YM. Acute chorioamnionitis and funisitis: definition,

- pathologic features, and clinical significance. *Am J Obstet Gynecol.* 2015;213:S29-S52.
89. Halim TY, Krauss RH, Sun AC, Takei F. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. *Immunity.* 2012;36:451-463.
  90. Kim HY, Chang YJ, Subramanian S, et al. Innate lymphoid cells responding to IL-33 mediate airway hyperreactivity independently of adaptive immunity. *J Allergy Clin Immunol.* 2012;129:216-227.e1-216-227.e16.
  91. Klein Wolterink RG, Kleinjan A, van Nimwegen M, et al. Pulmonary innate lymphoid cells are major producers of IL-5 and IL-13 in murine models of allergic asthma. *Eur J Immunol.* 2012;42:1106-1116.
  92. Fallon PG, Ballantyne SJ, Mangan NE, et al. Identification of an interleukin (IL)-25-dependent cell population that provides IL-4, IL-5, and IL-13 at the onset of helminth expulsion. *J Exp Med.* 2006;203:1105-1116.
  93. Cella M, Fuchs A, Vermi W, et al. A human natural killer cell subset provides an innate source of IL-22 for mucosal immunity. *Nature.* 2009;457:722-725.
  94. Gladiator A, Wangler N, Trautwein-Weidner K, LeibundGut-Landmann S. Cutting edge: IL-17-secreting innate lymphoid cells are essential for host defense against fungal infection. *J Immunol.* 2013;190:521-525.
  95. Klose CSN, Flach M, Mohle L, et al. Differentiation of type 1 ILCs from a common progenitor to all helper-like innate lymphoid cell lineages. *Cell.* 2014;157:340-356.
  96. Geremia A, Arancibia-Carcamo CV. Innate lymphoid cells in intestinal inflammation. *Front Immunol.* 2017;8:1296.
  97. Li M, Gao Y, Yong L, et al. Molecular signature and functional analysis of uterine ILCs in mouse pregnancy. *J Reprod Immunol.* 2017;123:48-57.
  98. Walker JA, Barlow JL, McKenzie AN. Innate lymphoid cells—how did we miss them? *Nat Rev Immunol.* 2013;13:75-87.
  99. Bartemes K, Chen CC, Iijima K, Drake L, Kita H. IL-33-responsive group 2 innate lymphoid cells are regulated by female sex hormones in the uterus. *J Immunol.* 2018;200:229-236.
  100. Monticelli LA, Sonnenberg GF, Abt MC, et al. Innate lymphoid cells promote lung-tissue homeostasis after infection with influenza virus. *Nat Immunol.* 2011;12:1045-1054.
  101. Molofsky AB, Nussbaum JC, Liang HE, et al. Innate lymphoid type 2 cells sustain visceral adipose tissue eosinophils and alternatively activated macrophages. *J Exp Med.* 2013;210:535-549.
  102. Nussbaum JC, Van Dyken SJ, von Moltke J, et al. Type 2 innate lymphoid cells control eosinophil homeostasis. *Nature.* 2013;502:245-248.
  103. Ning F, Liu H, Lash GE. The role of decidual macrophages during normal and pathological pregnancy. *Am J Reprod Immunol.* 2016;75:298-309.
  104. Fuss IJ, Neurath M, Boirivant M, et al. Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease. Crohn's disease LP cells manifest increased secretion of IFN-gamma, whereas ulcerative colitis LP cells manifest increased secretion of IL-5. *J Immunol.* 1996;157:1261-1270.
  105. Fuss IJ, Heller F, Boirivant M, et al. Nonclassical CD1d-restricted NK T cells that produce IL-13 characterize an atypical Th2 response in ulcerative colitis. *J Clin Invest.* 2004;113:1490-1497.
  106. Edmondson N, Bocking A, Machin G, Rizek R, Watson C, Keating S. The prevalence of chronic deciduitis in cases of preterm labor without clinical chorioamnionitis. *Pediatr Dev Pathol.* 2009;12:16-21.
  107. Kim CJ, Romero R, Chaemsaihong P, Kim JS. Chronic inflammation of the placenta: definition, classification, pathogenesis, and clinical significance. *Am J Obstet Gynecol.* 2015;213:S53-S69.
  108. Satoh-Takayama N, Vosshenrich CA, Lesjean-Pottier S, et al. Microbial flora drives interleukin 22 production in intestinal NKp46+ cells that provide innate mucosal immune defense. *Immunity.* 2008;29:958-970.
  109. Luci C, Reynders A, Ivanov II, et al. Influence of the transcription factor RORgammat on the development of NKp46+ cell populations in gut and skin. *Nat Immunol.* 2009;10:75-82.
  110. Takatori H, Kanno Y, Watford WT, et al. Lymphoid tissue inducer-like cells are an innate source of IL-17 and IL-22. *J Exp Med.* 2009;206:35-41.
  111. Cupedo T, Crellin NK, Papazian N, et al. Human fetal lymphoid tissue-inducer cells are interleukin 17-producing precursors to RORC+ CD127+ natural killer-like cells. *Nat Immunol.* 2009;10:66-74.
  112. Sonnenberg GF, Monticelli LA, Alenghat T, et al. Innate lymphoid cells promote anatomical containment of lymphoid-resident commensal bacteria. *Science.* 2012;336:1321-1325.
  113. Hepworth MR, Monticelli LA, Fung TC, et al. Innate lymphoid cells regulate CD4+ T-cell responses to intestinal commensal bacteria. *Nature.* 2013;498:113-117.
  114. Dealtry GB, Clark DE, Sharkey A, Charnock-Jones DS, Smith SK. Expression and localization of the Th2-type cytokine interleukin-13 and its receptor in the placenta during human pregnancy. *Am J Reprod Immunol.* 1998;40:283-290.
  115. Higuma-Myojo S, Sasaki Y, Miyazaki S, et al. Cytokine profile of natural killer cells in early human pregnancy. *Am J Reprod Immunol.* 2005;54:21-29.
  116. Sharma S, Godbole G, Modi D. Decidual control of trophoblast invasion. *Am J Reprod Immunol.* 2016;75:341-350.
  117. Halim TY, Steer CA, Matha L, et al. Group 2 innate lymphoid cells are critical for the initiation of adaptive T helper 2 cell-mediated allergic lung inflammation. *Immunity.* 2014;40:425-435.
  118. Allen JE, Sutherland TE. Host protective roles of type 2 immunity: parasite killing and tissue repair, flip sides of the same coin. *Semin Immunol.* 2014;26:329-340.
  119. Vazquez J, Chavarria M, Li Y, Lopez GE, Stanic AK. Computational flow cytometry analysis reveals a unique immune signature of the human maternal-fetal interface. *Am J Reprod Immunol.* 2018;79:e12774.

## SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

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