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**Plasma Metabolite Profiles in Children with Current Asthma**

(Running Title: Plasma Metabolites Children with Asthma)

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40  
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66 **ABSTRACT**

67 **Background:** Identifying metabolomic profiles of children with asthma has the potential to  
68 increase understanding of asthma pathophysiology.

69 **Objective:** To identify differences in plasma metabolites between children with and without  
70 current asthma at mid-childhood.

71 **Methods:** We used untargeted mass spectrometry to measure plasma metabolites in 237 children  
72 (46 current asthma cases and 191 controls) in Project Viva, a birth cohort from eastern  
73 Massachusetts, USA. Current asthma was assessed at mid-childhood (mean age 8.0 years). The  
74 ability of a broad spectrum metabolic profile to distinguish between cases and controls was  
75 assessed using partial least squares discriminant analysis. We used logistic regression models to  
76 identify individual metabolites that were differentially abundant by case-control status. We tested  
77 significant metabolites for replication in 411 children from the VDAART clinical trial.

78 **Results:** There was no evidence of a systematic difference in the metabolome of children  
79 reporting current asthma vs. healthy controls according to partial least squares discriminant  
80 analysis. However, several metabolites were associated with odds of current asthma at a  
81 nominally significant threshold ( $p < 0.05$ ), including a metabolite of nicotinamide (N1-Methyl-2-  
82 pyridone-5-carboxamide (Odds Ratio (OR)=2.8 (95% CI 1.1 to 8.0)), a pyrimidine metabolite  
83 (5,6-dihydrothymine (OR=0.4 (95% CI 0.2 to 0.9)), bile constituents (biliverdin (OR=0.4  
84 (95%CI 0.1 to 0.9), taurocholate (OR=2.0 (95% CI 1.2 to 3.4)), two peptides likely derived from  
85 fibrinopeptide A (ORs from 1.6 to 1.7), and a gut microbiome metabolite (p-cresol sulfate  
86 OR=0.5 (95% CI 0.2 to 0.9)). The associations for N1-Methyl-2-pyridone-5-carboxamide and p-  
87 cresol sulfate replicated in the independent VDAART population(one-sided p values=0.03-0.04).

88 **Conclusions and Clinical Relevance:** Current asthma is nominally associated with altered  
89 levels of several metabolites, including metabolites in the nicotinamide pathway, and a bacterial  
90 metabolite derived from the gut microbiome.

91 **Key words:** asthma, children, metabolomics, nicotinamide synthesis, p-cresol sulfate, Bile  
92 Constituents, Pyrimidine Metabolism, Endogenous steroids, Fibrinopeptides,  
93 Glycerophospholipid metabolism

#### 94 **Abbreviations:**

95 BMI: body mass index

96 MS: mass spectrometry

97 PLS-DA: partial least square discriminant analysis

#### 98 **INTRODUCTION**

99 Asthma is the most common chronic illness in U.S. children, and results in substantial morbidity  
100 as well as health care costs, with over \$50 billion dollars spent annually. [1] Asthma  
101 exacerbations are the most common health-related cause of lost school days,[1] rendering asthma  
102 a considerable public health burden. Asthma is a heritable disease with both environmental and  
103 genetic components.[2,3] A number of molecular determinants have been identified for  
104 asthma,[4] yet much remains unknown about how these molecular variants impact the disease.  
105 Metabolomics, the systematic analysis of all metabolites in a biological system, including  
106 carbohydrates, peptides, amino acids, organic acids, nucleotides and lipids, has emerged as a  
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107 powerful tool to identify new biomarkers for several diseases, and may help identify novel  
108 pathways associated with complex disease. Metabolite fluctuations may be more accurate  
109 disease markers than transcriptional, translational or post-translational changes, as they represent  
110 an integrated pathophysiologic profile that captures genetic and environmental interactions.  
111 Changes in metabolite profiles associated with chronic disease status in epidemiological studies  
112 may highlight important metabolites or metabolic pathways to interrogate in both in vitro and in  
113 vivo studies of underlying pathophysiological mechanisms.

114 Investigators have successfully identified metabolite biomarkers for Type 2 Diabetes,  
115 Alzheimer's disease, and cardiovascular disease,[5-8] leading to the discovery of novel disease  
116 pathways for these conditions. Existing asthma metabolomic studies[9-24] have reported  
117 promising findings, identifying biologically plausible metabolites related to tricarboxylic acid  
118 (TCA) metabolism, hypermethylation, phospholipid regulation, hypoxic and oxidative stress,  
119 immune reaction and inflammation that have been associated with asthma and asthma severity.  
120 Nevertheless, the studies to date are limited by diagnostic heterogeneity, sample size, and  
121 number of metabolites, and most findings are yet to be replicated in independent populations. As  
122 such, the use of metabolomics in asthma studies remains in the early stages with much  
123 knowledge to be gained.

124 The goal of this study is to identify differences in metabolomic profiles for children with and  
125 without a current diagnosis of asthma, using an untargeted metabolomic profiling approach. We  
126 sought to determine whether children with current asthma have altered metabolic profiles as  
127 compared to controls without current asthma. Our goals were to (1) determine the ability of a  
128 broad spectrum metabolic profile to discriminate between current asthma cases vs. controls (2)  
129 identify specific individual metabolites associated with current asthma and (3) validate these  
130 findings using an independent population.

## 131 **METHODS AND MATERIALS**

### 132 **Study Population**

#### 133 *Discovery Population*

134 Project Viva is an ongoing longitudinal pre-birth cohort study of children recruited from Atrius  
135 Harvard Vanguard Medical Associates, a multispecialty group practice in eastern Massachusetts.  
136 The purpose of Project Viva is to study the effect of environmental exposures on maternal and

137 child health. Details of the study design and recruitment have been previously reported.[25] The  
138 Institutional Review Board of Harvard Pilgrim Health Care approved the study protocols. All  
139 mothers participating in the study provided written informed consent and the children provided  
140 verbal assent. A total of 1,116 mother-child pairs (with children aged 6-10 years) attended an in-  
141 person mid-childhood study visit and 648 assenting children provided fasting blood samples.  
142 Metabolomic assays were originally performed for a study of childhood obesity and  
143 metabolomics in Project Viva.[26] Given funding limitations, we selected 300 children for  
144 analysis, deliberately oversampling for maternal and child for obesity. Subjects were  
145 oversampled for obesity as the original analysis was focused on studying obesity.[27]

146 After further exclusion of 38 children with inadequate plasma volume for the metabolomic  
147 assays, 262 children had metabolomic data. Of 262, we excluded from this analysis 21 children  
148 with past asthma diagnosis, and 4 with inadequate information on current asthma. Thus the final  
149 analytic sample included 237.

150 We assessed current asthma during the mid-childhood visit (mean age 8.0 years). We defined  
151 current asthma in mid-childhood as maternal report of ever diagnosed with asthma by a  
152 healthcare professional (assessed on the mid-childhood questionnaire) plus either taking asthma  
153 medications in the past 12 months or wheezing symptoms the past 12 months. This analysis  
154 included 46 cases with current asthma, and 191 controls who had never reported a past asthma  
155 diagnosis, and who did not report wheezing or use of asthma medications in the past 12 months.  
156 We also collected information from in-person interviews and questionnaires on maternal age at  
157 enrollment, maternal education (a marker of socio-economic status), smoking during pregnancy  
158 and child age, sex, and race/ethnicity. Information on asthma medication use and asthma  
159 severity was also collected from in-person interviews and questionnaires. Height and weight  
160 data collected at the mid-childhood visit were used to compute BMI z-scores based on CDC  
161 2000 age and sex-specific reference data.[28] Allergen sensitization was defined as any specific  
162 IgE level of 0.35 IU/mL or greater to common indoor allergens (*Dermatophagoides farinae*, cat,  
163 dog, and cockroach), mold allergens (*Alternaria* or *Aspergillus* species), food allergens (egg  
164 white, milk, and soy bean), or outdoor allergens (rye grass and ragweed).

165 Plasma metabolomic profiling for Project Viva was performed at Metabolon<sup>TM</sup> for 262  
166 children based on availability of archived biospecimens for a separate study of metabolomics and  
167 obesity status.[27] Untargeted metabolomic profiling was performed using multi-platform mass  
168 spectrometry (MS); detailed descriptions of sample preparation and metabolite identification  
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169 procedures have been previously published (see Supporting Information for additional details).  
170 [29] Samples were prepared using the automated MicroLab STAR® system from Hamilton  
171 Company for both Project Viva and VDAART. Several recovery standards were added prior to  
172 the first step in the extraction process for QC purposes. The resulting extract was divided into  
173 four fractions: one for analysis by UPLC/MS/MS (positive mode), one for UPLC/MS/MS  
174 (negative mode), one for GC/MS, and one for backup. Samples were placed briefly on a  
175 TurboVap® (Zymark) to remove the organic solvent. Each sample was then frozen and dried  
176 under vacuum. Samples were then prepared for the appropriate instrument, as described below.

177 Briefly, the analysis was performed using three platforms: UPLC/MS/MS (positive  
178 mode), UPLC/MS/MS (negative mode), and GC/MS, with pooled samples included throughout  
179 the analytical run for quality control purposes. Raw data was extracted, peak-identified and  
180 quality-control processed using Metabolon Inc.® hardware and software. Semi-quantitative  
181 concentrations (expressed in arbitrary units) of 345 known biochemicals were obtained.  
182 Metabolites were identified by comparison to library entries of purified standards or recurrent  
183 unknown entities. We included both endogenous and exogenous metabolites in our analyses.  
184 Missing values were imputed with half of the minimum value for that compound. Metabolite  
185 intensities were log transformed to normalize them and *pareto*-scaled to reduce the variation in  
186 fold-change differences between the features. See supplement for full details of the metabolomic  
187 profiling procedures.

### 188 *Replication Population*

189 For replication of our findings in Project Viva, we used plasma samples of children born to  
190 mothers enrolled in the Vitamin D Antenatal Asthma Reduction Trial (VDAART) clinical trial; a  
191 two arm, double-blind, placebo controlled, randomized, clinical trial of Vitamin D, to determine  
192 whether higher vitamin D intake and levels in the pregnant mother will prevent asthma and  
193 allergy in childhood. [30] Pregnant women (who had or whose partner had allergies/asthma)  
194 were randomized (n=881) during the first trimester of pregnancy (10-18 weeks) to one of two  
195 treatment arms of a clinical trial: 4000 IU Vitamin D + prenatal vitamins or 400 IU Vitamin D +  
196 prenatal vitamins. Of the 810 live births, 411 of had non-fasting plasma samples available at age  
197 three years for metabolomics analysis (108 asthma cases, 303 controls). Metabolomic profiling  
198 for VDAART was performed at Metabolon™, using a platform similar to the one utilized for  
199 Project Viva (see supplement for details). We used physician's diagnosis of asthma to define

200 cases in VDAART at age three follow-up.[31] Parental report of physician's diagnosis of asthma  
201 was taken directly from the offspring questionnaires.

## 202 **Statistical Analysis**

203 Analyses were conducted in R.[32] Differences in maternal and child demographic  
204 characteristics between cases and controls were assessed using the t-test for continuous variables  
205 and the chi-square for categorical variables.

### 206 *Partial Least Squares Discriminant analysis*

207 Metabolite features were analyzed as measured LC-MS peak areas, which are proportional to  
208 feature concentration and can be compared for any given metabolite (relative quantitation).  
209 Partial least squares discriminant analysis (PLS-DA) was conducted using MetaboAnalyst v.2.5  
210 (<http://www.metaboanalyst.ca/>)[33] to determine the combined ability of the 345 identified  
211 metabolites to discriminate asthma cases from controls. PLS-DA identifies latent factors, or  
212 components, that best describe the relationships between the metabolites, while best predicting  
213 asthma status. A seven-fold internal cross-validation procedure was implemented to guard  
214 against model over-fitting, and the overall significance of the model's discriminatory ability was  
215 evaluated using permutation testing; specifying 'prediction accuracy during training' and 2000  
216 permutations.

### 217 *Individual Metabolite Analysis*

218 We also conducted logistic regression on individual metabolites, to determine their relationship  
219 to asthma case status. Models were adjusted for maternal age, maternal education, smoking  
220 during pregnancy, child's sex, age, BMI z-score and race/ethnicity. Associations were expressed  
221 as odds ratios (95%CI) for current asthma per unit increase in metabolite.

### 222 *Replication of significant findings*

223 For replication, metabolites significantly associated with asthma in Project Viva were tested in  
224 an independent cohort of children from the VDAART clinical trial. We performed logistic  
225 regression analysis to determine if these metabolites were associated with asthma diagnosis at  
226 age 3 years. Analyses were adjusted for child's age, sex, BMI, maternal education level and  
227 race/ethnicity. We report one-sided p values for the replication analysis.

## 228 **RESULTS**

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229 *Discovery Population*

230 Characteristics of the discovery population are presented in Table I. Of the 237 children in  
231 Project Viva, 52.3% were female and 19.4% had a mother-reported a diagnosis of current  
232 asthma. This population also included 84 obese (32.1%), 28 overweight (10.7%), and 150 normal  
233 weight (57.3%) children. The mean (+/- SD) age was 8.0 (+/- 0.9) years. The majority of  
234 children with current asthma were also sensitized to allergens. Seventy two percent had a  
235 positive specific IgE test ( $\geq 0.35$  IU/ml) to at least one of the specific indoor, outdoor or food  
236 allergens tested. In contrast, 49% of control subjects were sensitized to allergens. There were  
237 also notable differences between children with vs. without current asthma with respect to  
238 maternal age (30.3 vs. 32.5 years,  $p=0.02$ ) and education (37.8% vs. 63.9% college graduates,  
239  $p=0.002$ ) and child age (8.4 vs. 7.9 years,  $p=0.02$ ), race/ethnicity (26.1% vs. 64.4% white,  
240  $p<0.0001$ ) and BMI z-scores (1.45 v. 0.66,  $p<0.0001$ ). There were no differences between  
241 children with vs. without current asthma with respect to child's sex or maternal pregnancy  
242 smoking status.

243 We also collected information on asthma medication use and asthma severity in children with  
244 current asthma. The vast majority (80%) of current asthmatics were taking inhaled  
245 corticosteroids. In general, most current asthmatics in Project Viva showed mild to moderate  
246 disease severity. None of the current asthmatics were hospitalized in the past year for asthma.  
247 Most children with current asthma did not require emergency treatment for asthma symptoms in  
248 the past year (72%), although 17% reported one emergency room visit for asthma, 9% reported  
249 visiting the emergency room 2-3 times in the past year, with 2% reporting 3 or more visits. A  
250 little over half of the children (57%) with current asthma occasionally missed school because of  
251 asthma symptoms; 20% report one missed school day, 26% report 2-3 missed school days, and  
252 11% report more than 3 missed school days over a one year time period.

253 In VDAART, cases of current asthma were generally mild. Only 38% of children received any  
254 medication for wheezing, wheezy bronchitis or asthma since their last follow-up. Seventeen  
255 percent of current asthmatics report seeing a doctor because of wheezing, asthma, wheezing or  
256 asthmatic bronchitis since their last follow-up.

257 *Replication Population*

258 Of the 411 children with available metabolomics data in the VDAART population, 219 (53%)  
259 were male, and 108 (27%) developed asthma by age three. VDAART participants were of

260 diverse race/ethnic backgrounds, and were 48% African American, 33% white and 19% other  
261 race. Of the children with current asthma in VDAART, 58% were sensitized to at least one of  
262 the indoor, outdoor or food allergens tested, while approximately half (48%) of the control  
263 subjects were sensitized to allergens.

#### 264 *Partial Least Squares Discriminant analysis*

265 There was no evidence in these analyses that a specific profile signature derived from the 345  
266 identified metabolites differed between asthma cases and controls in Project Viva (Supplemental  
267 Figure 1). After seven-fold internal cross-validation accuracy was 0.8, but the  $R^2$  and  $Q^2$  were  
268 only 0.25 and 0.05 respectively for the first component, indicating that the model was not robust.  
269 This was confirmed by the permutation testing (permuted  $p=0.134$ ).

#### 270 *Individual Metabolite Analysis*

271 Of all of the metabolites tested in Project Viva for associations with current asthma case status,  
272 none were statistically significant after adjustment for multiple comparisons. However, ten  
273 metabolites were nominally significant at the  $p<0.05$  level (Table II). A one unit increase in N1-  
274 Methyl-2-pyridone-5-carboxamide, a metabolite in the nicotinamide pathway, was associated  
275 with an increased odd of asthma (OR=2.79, 95% CI 1.10 to 7.97). The relationship between  
276 N1-Methyl-2-pyridone-5-carboxamide and asthma case status was replicated (Table III) in the  
277 VDAART cohort (OR=1.27,  $p=0.04$  (one-sided  $p$  value)). The pyrimidine metabolite 5,6-  
278 dihydrothymine was associated with reduced odds of asthma (OR=0.44, 95% CI 0.21 to 0.86) in  
279 Project Viva (Table II), but showed the opposite direction of effect in VDAART (Table III).  
280 Metabolomics profiling also uncovered two peptides (DSGEGDFXAEGGGVR and  
281 ADSGEGDFXAEGGGVR) associated with current asthma in Project Viva. An NCBI protein  
282 blast search revealed fibrinopeptide A as the top hit (highest maximum alignment score and  
283 lowest E value) for these peptides. Bile constituents taurocholate and biliverdin were also  
284 related to asthma case status. Taurocholate, a bile salt involved in fat emulsification, was  
285 associated with current asthma (OR=2.03, 95% CI 1.22 to 3.42), while biliverdin, a bile pigment  
286 and product of heme catabolism was associated with decreased odds of current asthma (OR=0.35  
287 95% CI 0.14 to 0.87). P-cresol sulfate, a bacterial metabolite originating from the gut  
288 microbiome, was linked to decreased odds of current asthma (OR=0.47, 95% CI 0.23 to 0.94).  
289 This association was replicated in the VDAART study (Table III). Other metabolites  
290 demonstrating lower levels amongst asthmatic cases in Project Viva were cortisone (a naturally  
291 occurring glucocorticoid), tryptophan betaine (from dietary intake of legumes), and 1-  
292 docosapentaenoylglycero-phosphocholine (22:5), a lysophospholipid formed by hydrolysis  
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293 of phosphatidylcholine by phospholipase A2. Spectra for the two metabolites associated with  
294 current asthma in both the discovery and replication cohort (N1-Methyl-2-pyridone-5-  
295 carboxamide and p-cresol sulfate) are shown in supplemental figures E2 and E3.

296 Based on prior reports, we looked to see if plasma cortisone was lower in current asthmatics  
297 on ICS as compared to untreated current asthmatics in Project Viva. We did not see an  
298 association between ICS use and lower cortisone levels in current asthma cases ( $p > 0.9$  for t-test  
299 comparisons). However, the very high ICS treatment rates in current asthmatics (80%) meant  
300 that a comparison with untreated current asthmatics was likely underpowered.

## 301 DISCUSSION

302 We identified metabolites associated with asthma by profiling the plasma metabolome in  
303 children from two longitudinal birth cohort studies. In Project Viva, current asthma was  
304 associated with alterations in metabolites from the nicotinamide pathway, pyrimidine  
305 metabolism, fibrinogen-associated peptides, bile metabolites, and p-cresol sulfate, a microbial  
306 metabolite derived from the gut microbiome. Cortisone, a metabolite inversely associated with  
307 ICS use,[44] was decreased in current asthmatics. The association between asthma case status  
308 and two of the top metabolites (the bacterial metabolite p-cresol sulfate from the gut  
309 microbiome, and a nicotinoamide pathway metabolite) were replicated in a second cohort, the  
310 VDAART clinical trial.

311 Our study shows a new potential link between higher levels of plasma p-cresol sulfate and  
312 decreased odds of asthma. This association was replicated in an independent cohort. P-cresol  
313 sulfate is a microbial metabolite that may reflect gut microbiome composition.[34] Gut  
314 microbes have long been hypothesized to interact with the immune system in ways that may alter  
315 risk of allergies and asthma, and a number of publications now show compositional differences  
316 in the gut microbiome when allergic disease risk factors or allergic disease phenotypes are  
317 present. [31-34] It has been hypothesized that p-cresol sulfate may indicate a gut microbiome  
318 enterotype dominated by *Bacteroides*,[34] a genera containing multiple taxa including *B.*  
319 *Fragilis*, a species with strong experimental evidence for restoring Th1/Th2 imbalance.[35]  
320 While our observed association between p-cresol sulfate and asthma status is intriguing, further  
321 research is required to uncover the connection between the gut microbiome, the circulating  
322 metabolome, and their combined relationship to asthma.

323 We observed higher levels of the nicotinamide pathway metabolite N1-Methyl-2-pyridone-5-  
324 carboxamide in asthmatic subjects vs. controls in Project Viva, and our findings in VDAART  
325 were showed a similar relationship. The nicotinate and nicotinamide metabolic pathway has  
326 been linked to asthma phenotypes in both in vitro and population-based studies. One in vitro  
327 study of human airway smooth muscle demonstrated that cells from asthmatic donors had  
328 increased expression of *NADPH* (Nicotinamide Adenine Diphosphate) *Oxidase*, which promoted  
329 oxidative stress and smooth muscle cell contractility. [36] Other in vitro findings show  
330 increased *NADPH oxidase* expression in neutrophilic asthma, with a concomitant decrease in  
331 ciliary function of the bronchial epithelium.[37] These in vitro models provide a biological basis  
332 for our observed association between plasma nicotinamide metabolites and current asthma.  
333 While one other previous epidemiological study reported alterations in nicotinamide pathway  
334 metabolites (nicotinamide was increased in asthmatic subjects vs. controls), the sample size was  
335 small (30 subjects in total) and there were no attempts at replication.[38]

336 Metabolomic profiles of asthmatics in our study also showed increased levels of two similar  
337 peptides likely derived from fibrinopeptide A (DSGEGDFXAEGGGVR and  
338 ADSGEGDFXAEGGGVR). Fibrinopeptide A is a short amino acid sequence located within the  
339 alpha chain of soluble fibrinogen. Fibrinogen may play a role in asthma pathogenesis by  
340 enhancing inflammatory response. Huang and colleagues reported a correlation between  
341 increased plasma fibrinogen and reduced lung function. These investigators also found that  
342 obese subjects, including those with asthma, tend to have higher circulating fibrinogen levels.  
343 [39] Fibrinogen cleavage products act as TLR4 ligands, enhancing innate immune response.  
344 [40] For instance, fibrinogen cleavage products can bind Toll like receptor 4 (TLR4), priming  
345 innate immune and airway epithelial cell response to IL-13, with downstream triggering of  
346 airway inflammation.[41]

347 Two of the metabolites with altered levels in current asthmatics, taurocholate and biliverdin,  
348 are bile constituents. Taurocholate, a conjugate of taurine and cholic acid, is an emulsifier of  
349 fats. Comhair and investigators previously reported increased plasma taurocholate in asthmatics,  
350 and we replicate that finding here in a much larger cohort.[38] Yu and colleagues also report  
351 altered levels of taurocholate in a mouse model of OVA-induced asthma[42]; however, in this  
352 experimental study mice with the asthma phenotype had lower, as opposed to higher, levels of  
353 circulating taurocholate. Other experimental data suggest that activity of hemoxygenase-1, the  
354 enzyme that produces biliverdin from heme, may reduce mucous secretion in the airways. In an  
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355 in vitro study of normal human bronchial epithelial cells, over-expression of heme oxygenase-I  
356 was associated with reduced IL-13 induced goblet cell hyperplasia and decreased MUCA5  
357 secretion.[43] The lower levels of circulating biliverdin in asthma cases may reflect reduced  
358 activity of heme oxygenase I, which could potentially contribute to increased mucous production  
359 in the airways (a common feature of asthma).

360 Consistent with the findings of Reinke et al[44], we also observed decreased plasma cortisone  
361 levels in subjects with current asthma. This finding is consistent with well-known effects of ICS  
362 (inhaled corticosteroid) use, which cause suppression of the hypothalamic-pituitary axis, leading  
363 to lower production of endogenous steroids like cortisone.[45] The decreased plasma cortisone  
364 levels in asthmatics vs. controls is most likely a simple reflection of ICS use, and not an indicator  
365 of asthma pathophysiology. Since the majority of asthmatics were treated with ICS, we could  
366 not disentangle the effects of ICS use from those associated with current asthma status in our  
367 analysis.

368 Lastly, our observation that pyrimidine metabolism may be altered in asthmatics vs. controls  
369 may have relevance for asthma pathophysiology. A large metabolomics study in a cohort of  
370 childhood asthmatics demonstrated associations between pyrimidine metabolites and three  
371 phenotypic aspects of asthma (methacholine responsiveness, pre bronchodilator FEV1/FVC ratio  
372 and post bronchodilator FEV1/FVC ratio).[46] However the mechanistic connection between  
373 pyrimidine metabolism and physiological processes influencing asthma remains unclear.  
374 Findings for our discovery population and replication cohort demonstrated opposite directions of  
375 effect for the 5,6-dihydrothymine metabolite, an inconsistency that makes the associations  
376 difficult to interpret.

377 It is interesting to note that our PLS-DA analysis did not identify any global metabolite  
378 profile differences by asthma status, whereas analysis of individual metabolites did yield a  
379 number of biologically plausible associations. One interpretation of this finding is that asthma  
380 case status is associated with perturbations of few individual metabolites, rather than latent  
381 variables describing global pathway alterations (shifts in multiple correlated metabolites). An  
382 alternate explanation is that, even with hundreds of named metabolites in our analysis, we still  
383 didn't have a broad enough coverage of underlying correlated metabolites and their associated  
384 pathways to detect global metabolome shifts associated with asthma case status. Instead, we  
385 may have identified individual, surrogate metabolite markers of more global pathway  
386 perturbations, through our logistic regression analyses. As metabolite identifications continue to

387 expand, PLS-DA may be a more effective tool in identifying global shifts in the metabolome of  
388 complex diseases.

389 Our study has several strengths. First, it is one of the largest plasma metabolomic studies  
390 comparing asthma cases to control subjects to date. Second, we limited potential sources of bias  
391 by controlling for confounders, including race, BMI and maternal educational level. Third, we  
392 were able to replicate two of our top metabolites in an independent cohort. Despite the strengths  
393 of our study, a few caveats deserve mention. First, utilizing maternal report of doctor diagnosis  
394 of asthma, rather than direct physician report of asthma diagnosis may have been potential  
395 source of bias in health outcome assessment. Second, while our study is large compared to other  
396 metabolomics studies, our sample size may have been too small to identify associations that were  
397 statistically significant after accounting for multiple comparisons. An additional limitation was  
398 that the phenotypes available for our discovery and replication cohorts were different. For  
399 Project Viva, current asthma status was assessed in mid-childhood, whereas VDAART subjects  
400 were assessed at age 3, when a definitive asthma diagnosis is often difficult. Also, the  
401 racial/ethnic and socioeconomic status distributions (while accounted for in our analyses) and  
402 different geographic locations for the primary and replication populations, may have given rise to  
403 very different exposure profiles (including diet) that could not be adjusted for in our analyses.  
404 Plasma samples in Project Viva were fasting, whereas in the replication population plasma  
405 samples were non-fasting. (The use of non-fasting samples in VDAART may have had a  
406 particular influence on our ability to detect an association between current asthma status and the  
407 dietary metabolite tryptophan betaine, if recent food intake increased the variability of this  
408 metabolite.) The study designs for Project Viva and VDAART were also different; Project Viva  
409 is an observational epidemiology study based on an unselected population, while VDAART is a  
410 clinical trial of prenatal vitamin D supplementation and asthma outcomes in children with a  
411 parental history of allergies or asthma. These discrepancies in the two cohorts may have  
412 decreased our potential for replication; however, our ability to replicate two metabolites in spite  
413 of these differences suggests that our replicated associations are robust. Metabolite identification  
414 in the two populations was not identical, which meant that we did not have the opportunity to test  
415 for replication of the phospholipid 1-docosapentaenoylglycero-phosphocholine (22:5) and  
416 fibrinogen peptide findings, as these were not among the named metabolites in our replication  
417 study. It should also be noted that while many of the metabolites associated with asthma in our  
418 primary cohort are biologically plausible and supported by experimental data in the literature,  
419 none were statistically significant after adjustment for multiple comparisons.

420 In summary, our findings suggest that current asthma is associated perturbations in pathways  
421 including metabolism of nicotinamide and pyrimidines, production of bile salts, heme catabolism  
422 and metabolites generated via the gut microbiome. Asthmatics in our cohort tended to have  
423 lower levels of endogenous cortisone, which is likely an effect of inhaled corticosteroid use.  
424 Identifying a metabolomic profile characteristic of children with asthma has the potential to  
425 uncover novel candidate pathways in asthma pathophysiology. In the long term, asthma  
426 metabolomics research may identify biomarkers that improve the efficacy of therapeutic  
427 regimens for children with asthma, in turn decreasing the severity of asthma suffering and  
428 healthcare costs in the U.S. and globally.

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Table I. Demographics of Discovery Population, Project Viva (N = 237)

|                                     | Total<br>N=237     | Current asthma at mid-childhood |                     | P-value |
|-------------------------------------|--------------------|---------------------------------|---------------------|---------|
|                                     |                    | Yes<br>N=46 (19.4%)             | No<br>N=191 (80.6%) |         |
|                                     | N (%) or mean (SD) |                                 |                     |         |
| <b>Mother</b>                       |                    |                                 |                     |         |
| Age at enrollment, years            | 32.1 (5.8)         | 30.3 (5.8)                      | 32.5 (5.8)          | 0.02    |
| Smoking status, %                   |                    |                                 |                     | 0.15    |
| . Never                             | 151 (63.7)         | 30 (65.2)                       | 121 (63.4)          |         |
| . Former smoker (before pregnancy)  | 51 (21.5)          | 6 (13.0)                        | 45 (23.6)           |         |
| . Current smoker (during pregnancy) | 35 (14.8)          | 10 (21.7)                       | 25 (13.1)           |         |
| Education status, %                 |                    |                                 |                     | 0.002   |
| . Not college graduate              | 97 (41.1)          | 28 (62.2)                       | 69 (36.1)           |         |
| . College graduate                  | 139 (58.9)         | 17 (37.8)                       | 122 (63.9)          |         |
| <b>Child</b>                        |                    |                                 |                     |         |
| Sex: Female, %                      | 124 (52.3)         | 28 (60.9)                       | 96 (50.3)           | 0.25    |
| Age at mid-childhood visit, years   | 8.0 (0.9)          | 8.4 (1.2)                       | 7.9 (0.7)           | 0.02    |
| Child race/ethnicity, %             |                    |                                 |                     | <0.0001 |
| . Black                             | 56 (23.6)          | 23 (50.0)                       | 33 (17.3)           |         |
| . White                             | 135 (57.0)         | 12 (26.1)                       | 123 (64.4)          |         |
| . Other                             | 46 (19.4)          | 11 (23.9)                       | 35 (18.3)           |         |
| BMI z-score at mid-childhood        | 0.81 (1.15)        | 1.45 (1.14)                     | 0.66 (1.10)         | <0.0001 |



Allergen Sensitization\*                      114 (53)                      28 (72)                      86 (49)                      0.009

\*Allergen sensitization numbers reflect some missing data; 215 subjects out of 237 had allergen sensitization testing

Table II. Metabolites associated with Current Asthma in Project Viva.

| <b>Pathway/Grouping</b>                       | <b>Metabolite</b>                  | <b>Odds Ratio (95% CI) for Current Asthma*</b> |
|---|------------------------------------|--|
| <i>Nicotinate and Nicotinamide Metabolism</i> | N1-Methyl-2-pyridone-5-carboxamide | <b>2.79 (1.10 to 7.97)</b>                     |
| <i>Bile Constituents</i>                      | Taurocholate                       | <b>2.03 (1.22 to 3.42)</b>                     |
|   | Biliverdin                         | <b>0.35 (0.14 to 0.87)</b>                     |
| <i>Pyrimidine Metabolism</i>                  | 5,6-dihydrothymine                 | <b>0.44 (0.21 to 0.86)</b>                     |
| <i>Microbial Metabolite</i>                   | P-cresol sulfate                   | <b>0.47 (0.23 to 0.94)</b>                     |
| <i>(Putative) Fibrinopeptide A</i>            | DSGEGDFXAEGGGVR*                   | <b>1.57 (1.02 to 2.42)</b>                     |
|   | ADSGEGDFXAEGGGVR*                  | <b>1.68 (1.04 to 2.73)</b>                     |

|                                       |   |                            |
|---------------------------------------|---|----------------------------|
| <i>Dietary Metabolites</i>            | Tryptophan Betaine                                | <b>0.47 (0.28 to 0.77)</b> |
| <i>Endogenous steroids</i>            | Cortisone   | <b>0.20 (0.06 to 0.59)</b> |
| <i>Glycerophospholipid metabolism</i> | 1-docosapentaenoylglycerophosphocholine<br>(22:5) | <b>0.44 (0.22 to 0.88)</b> |

\*Odds ratios are for a 1 unit increase in metabolite level; all odds ratios are adjusted for maternal age, maternal education, smoking in pregnancy, child's BMI z-score, race/ethnicity, age and sex; Odds ratios with  $p < 0.05$  in bold

Table III. Replication of Significant Metabolites associated with Current Asthma in VDAART Population.

| <b>Pathway/Grouping</b>                       | <b>Metabolite</b>                  | <b>Odds Ratio (95% CI)<br/>for Asthma*</b> | <b>P value (one sided) for Replication</b> |
|---|------------------------------------|--|--|
| <i>Nicotinate and Nicotinamide Metabolism</i> | N1-Methyl-2-pyridone-5-carboxamide | <b>1.27 (0.97 to 1.66)</b>                 | <b>0.04</b>                                |
| <i>Bile Constituents</i>                      | Taurocholate                       | 1.03 (0.89 to 1.20)                        | 0.35                                       |
|   | Biliverdin                         | 1.08 (0.90 to 1.31)                        | 0.20                                       |
| <i>Microbial Metabolite</i>                   | P-cresol sulfate                   | <b>0.83 (0.69 to 1.00)</b>                 | <b>0.03</b>                                |
| <i>Pyrimidine Metabolism</i>                  | 5,6-dihydrothymine                 | 1.56 (1.00 to 2.40)                        | 0.02                                       |
| <i>Dietary Metabolite</i>                     | Tryptophan Betaine                 | 1.01 (0.92 to 1.12)                        | 0.38                                       |
| <i>Endogenous steroids</i>                    | Cortisone                          | 1.30 (0.97 to 1.75)                        | 0.04                                       |

\*Models adjusted for maternal age, maternal education, child's BMI, race/ethnicity, age and sex;

Odds ratio that replicates in the same direction of effect with  $p < 0.05$  is in bold

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## References

1. Barnett SB, Nurmagambetov TA. Costs of asthma in the United States: 2002-2007. *J Allergy Clin Immunol* 2011;127:145-52.
2. Rava M, Smit LA, Nadif R. Gene-environment interactions in the study of asthma in the postgenomewide association studies era. *Curr Opin Allergy Clin Immunol* 2015;15:70-8.
3. Bonnelykke K, Ober C. Leveraging gene-environment interactions and endotypes for asthma gene discovery. *J Allergy Clin Immunol* 2016;137:667-79.
4. Moffatt MF, Gut IG, Demenais F, Strachan DP, Bouzigon E, Heath S, von Mutius E, Farrall M, Lathrop M, Cookson WO, GABRIEL Consortium. A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med* 2010;363:1211-21.
5. Rhee EP, Gerszten RE. Metabolomics and cardiovascular biomarker discovery. *Clin Chem* 2012;58:139-47.
6. Wang TJ, Larson MG, Vasan RS, Cheng S, Rhee EP, McCabe E, Lewis GD, Fox CS, Jacques PF, Fernandez C, O'Donnell CJ, Carr SA, Mootha VK, Florez JC, Souza A, Melander O, Clish CB, Gerszten RE. Metabolite profiles and the risk of developing diabetes. *Nat Med* 2011;17:448-53.
7. Rhee EP, Thadhani R. New insights into uremia-induced alterations in metabolic pathways. *Curr Opin Nephrol Hypertens* 2011;20:593-8.
8. Mapstone M, Cheema AK, Fiandaca MS, Zhong X, Mhyre TR, MacArthur LH, Hall WJ, Fisher SG, Peterson DR, Haley JM, Nazar MD, Rich SA, Berlau DJ, Peltz CB, Tan MT, Kawas CH, Federoff HJ. Plasma phospholipids identify antecedent memory impairment in older adults. *Nat Med* 2014;20:415-8.
9. Carraro S, Rezzi S, Reniero F, Heberger K, Giordano G, Zanconato S, Guillou C, Baraldi E. Metabolomics applied to exhaled breath condensate in childhood asthma. *Am J Respir Crit Care Med* 2007;175:986-90.

10. Saude EJ, Skappak CD, Regush S, Cook K, Ben-Zvi A, Becker A, Moqbel R, Sykes BD, Rowe BH, Adamko DJ. Metabolomic profiling of asthma: diagnostic utility of urine nuclear magnetic resonance spectroscopy. *J Allergy Clin Immunol* 2011;127:6.
11. Sinha A, Krishnan V, Sethi T, Roy S, Ghosh B, Lodha R, Kabra S, Agrawal A. Metabolomic signatures in nuclear magnetic resonance spectra of exhaled breath condensate identify asthma. *Eur Respir J* 2012;39:500-2.
12. Mattarucchi E, Baraldi E, Guillou C. Metabolomics applied to urine samples in childhood asthma; differentiation between asthma phenotypes and identification of relevant metabolites. *Biomed Chromatogr* 2012;26:89-94.
13. Carraro S, Giordano G, Reniero F, Carpi D, Stocchero M, Sterk PJ, Baraldi E. Asthma severity in childhood and metabolomic profiling of breath condensate. *Allergy* 2013;68:110-7.
14. Gahleitner F, Guallar-Hoyas C, Beardsmore CS, Pandya HC, Thomas CP. Metabolomics pilot study to identify volatile organic compound markers of childhood asthma in exhaled breath. *Bioanalysis* 2013;5:2239-47.
15. Jung J, Kim SH, Lee HS, Choi GS, Jung YS, Ryu DH, Park HS, Hwang GS. Serum metabolomics reveals pathways and biomarkers associated with asthma pathogenesis. *Clin Exp Allergy* 2013;43:425-33.
16. Ibrahim B, Marsden P, Smith JA, Custovic A, Nilsson M, Fowler SJ. Breath metabolomic profiling by nuclear magnetic resonance spectroscopy in asthma. *Allergy* 2013;68:1050-6.
17. Ried JS, Baurecht H, Stuckler F, Krumsiek J, Gieger C, Heinrich J, Kabesch M, Prehn C, Peters A, Rodriguez E, Schulz H, Strauch K, Suhre K, Wang-Sattler R, Wichmann HE, Theis FJ, Illig T, Adamski J, Weidinger S. Integrative genetic and metabolite profiling analysis suggests altered phosphatidylcholine metabolism in asthma. *Allergy* 2013;68:629-36.
18. Loureiro CC, Duarte IF, Gomes J, Carrola J, Barros AS, Gil AM, Bousquet J, Bom AT, Rocha SM. Urinary metabolomic changes as a predictive biomarker of asthma exacerbation. *J Allergy Clin Immunol* 2014;133:5.

19. McGeachie MJ, Dahlin A, Qiu W, Croteau-Chonka DC, Savage J, Wu AC, Wan ES, Sordillo JE, Al-Garawi A, Martinez FD, Strunk RC, Lemanske RF, Jr, Liu AH, Raby BA, Weiss S, Clish CB, Lasky-Su JA. The metabolomics of asthma control: a promising link between genetics and disease. *Immun Inflamm Dis* 2015;3:224-38.
20. Chang C, Guo ZG, He B, Yao WZ. Metabolic alterations in the sera of Chinese patients with mild persistent asthma: a GC-MS-based metabolomics analysis. *Acta Pharmacol Sin* 2015;36:1356-66.
21. Smolinska A, Klaassen EM, Dallinga JW, van de Kant, K D, Jobsis Q, Moonen EJ, van Schayck OC, Dompeling E, van Schooten FJ. Profiling of volatile organic compounds in exhaled breath as a strategy to find early predictive signatures of asthma in children. *PLoS One* 2014;9:e95668.
22. Fitzpatrick AM, PhD, Park Y, PhD, Brown, Lou Ann S., PhD, Jones DP, PhD. Children with severe asthma have unique oxidative stress-associated metabolomic profiles. *Journal of Allergy and Clinical Immunology*, The 2013;133:261.e8.
23. Jung J, Kim SH, Lee HS, Choi GS, Jung YS, Ryu DH, Park HS, Hwang GS. Serum metabolomics reveals pathways and biomarkers associated with asthma pathogenesis. *Clin Exp Allergy* 2013;43:425-33.
24. Ried JS, Baurecht H, Stückler F, Krumsiek J, Gieger C, Heinrich J, Kabesch M, Prehn C, Peters A, Rodriguez E, Schulz H, Strauch K, Suhre K, Wang-Sattler R, Wichmann H- , Theis FJ, Illig T, Adamski J, Weidinger S, Simon H. Integrative genetic and metabolite profiling analysis suggests altered phosphatidylcholine metabolism in asthma. *Allergy* 2013;68:629-36.
25. Oken E, Baccarelli AA, Gold DR, Kleinman KP, Litonjua AA, De Meo D, Rich-Edwards JW, Rifas-Shiman SL, Sagiv S, Taveras EM, Weiss ST, Belfort MB, Burris HH, Camargo CA, Jr, Huh SY, Mantzoros C, Parker MG, Gillman MW. Cohort profile: project viva. *Int J Epidemiol* 2015;44:37-48.

26. Oken E, Baccarelli AA, Gold DR, Kleinman KP, Litonjua AA, De Meo D, Rich-Edwards JW, Rifas-Shiman SL, Sagiv S, Taveras EM, Weiss ST, Belfort MB, Burris HH, Camargo J, Carlos A, Huh SY, Mantzoros C, Parker MG, Gillman MW. Cohort profile: project viva. *International journal of epidemiology* 2015;44:37-48.
27. Perng W, Gillman MW, Fleisch AF, Michalek RD, Watkins SM, Isganaitis E, Patti ME, Oken E. Metabolomic profiles and childhood obesity. *Obesity (Silver Spring)* 2014;22:2570-8.
28. Kuczmarski RJ, Ogden CL, Guo SS, Grummer-Strawn LM, Flegal KM, Mei Z, Wei R, Curtin LR, Roche AF, Johnson CL. 2000 CDC Growth Charts for the United States: methods and development. *Vital Health Stat* 11 2002;display:1-190.
29. Ryals J, Lawton K, Stevens D, Milburn M. Metabolon, Inc. *Pharmacogenomics* 2007;8:863-6.
30. Litonjua AA, Lange NE, Carey VJ, Brown S, Laranjo N, Harshfield BJ, O'Connor GT, Sandel M, Strunk RC, Bacharier LB, Zeiger RS, Schatz M, Hollis BW, Weiss ST. The Vitamin D Antenatal Asthma Reduction Trial (VDAART): rationale, design, and methods of a randomized, controlled trial of vitamin D supplementation in pregnancy for the primary prevention of asthma and allergies in children. *Contemporary clinical trials* 2014;38:37-50.
31. Augusto A, Litonjua, MD, MPH, Vincent J, Carey P, Nancy Laranjo BA, Benjamin J, Harshfield, B A, Thomas F, McElrath, MD, PhD, George T, O'Connor M, MS, Megan Sandel M, MPH, Ronald E. Iverson Jr, MD, MPH, Aviva Lee-Paritz MD, Robert C. Strunk, MD, PhD, Leonard B. Bacharier, M D, George A. Macones, MD, MSCE, Robert S. Zeiger M, PhD, Michael Schatz M, MS, Bruce W. Hollis P, Eve Hornsby P, Catherine Hawrylowicz P, Ann Chen Wu M, MPH, Scott T. Weiss M, MS. Effect of Prenatal Supplementation With Vitamin D on Asthma or Recurrent Wheezing in Offspring by Age 3 Years  
The VDAART Randomized Clinical Trial. *JAMA*. 2016;315(4):362-370. 2016.
32. R Development Core Team. R: A language and environment for statistical computing. &nbsp; ;2017.

33. Xia J, Sinelnikov IV, Han B, Wishart DS. MetaboAnalyst 3.0--making metabolomics more meaningful. *Nucleic Acids Res* 2015;43:251.
34. Viaene L, Thijs L, Jin Y, Liu Y, Gu Y, Meijers B, Claes K, Staessen J, Evenepoel P. Heritability and clinical determinants of serum indoxyl sulfate and p-cresyl sulfate, candidate biomarkers of the human microbiome enterotype. *PLoS One* 2014;9:e79682.
35. Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* 2005;122:107-18.
36. Sutcliffe A, Hollins F, Gomez E, Saunders R, Doe C, Cooke M, Challiss RA, Brightling CE. Increased nicotinamide adenine dinucleotide phosphate oxidase 4 expression mediates intrinsic airway smooth muscle hypercontractility in asthma. *Am J Respir Crit Care Med* 2012;185:267-74.
37. Wan WY, Hollins F, Haste L, Woodman L, Hirst RA, Bolton S, Gomez E, Sutcliffe A, Desai D, Chachi L, Mistry V, Szyndralewicz C, Wardlaw A, Saunders R, O'Callaghan C, Andrew PW, Brightling CE. NADPH Oxidase-4 Overexpression Is Associated With Epithelial Ciliary Dysfunction in Neutrophilic Asthma. *Chest* 2016;149:1445-59.
38. Comhair SA, McDunn J, Bennett C, Fettig J, Erzurum SC, Kalhan SC. Metabolomic Endotype of Asthma. *J Immunol* 2015;195:643-50.
39. Huang F, del-Rio-Navarro BE, Alcantara ST, Ontiveros JA, Cienfuegos DR, Bello Gonzalez SA, Villafana S, Bravo G, Hong E. Plasminogen activator inhibitor-1, fibrinogen, and lung function in adolescents with asthma and obesity. *Endocr Res* 2012;37:135-44.
40. Millien VO, Lu W, Mak G, Yuan X, Knight JM, Porter P, Kheradmand F, Corry DB. Airway fibrinolysis and the initiation of allergic inflammation. *Ann Am Thorac Soc* 2014;11 Suppl 5:277.
41. Millien VO, Lu W, Shaw J, Yuan X, Mak G, Roberts L, Song LZ, Knight JM, Creighton CJ, Luong A, Kheradmand F, Corry DB. Cleavage of fibrinogen by proteinases elicits allergic responses through Toll-like receptor 4. *Science* 2013;341:792-6.



42. Yu M, Cui FX, Jia HM, Zhou C, Yang Y, Zhang HW, Ding G, Zou ZM. Aberrant purine metabolism in allergic asthma revealed by plasma metabolomics. *J Pharm Biomed Anal* 2016 Feb 20;120:181-9 doi: 10.1016/j.jpba.2015.12.018 Epub 2015;120:181-9.
43. Mishina K, Shinkai M, Shimokawaji T, Nagashima A, Hashimoto Y, Inoue Y, Inayama Y, Rubin BK, Ishigatsubo Y, Kaneko T. HO-1 inhibits IL-13-induced goblet cell hyperplasia associated with CLCA1 suppression in normal human bronchial epithelial cells. *Int Immunopharmacol* 2015;29:448-53.
44. Reinke SN, Gallart-Ayala H, Gomez C, Checa A, Fauland A, Naz S, Kamleh MA, Djukanovic R, Hinks TS, Wheelock CE. Metabolomics analysis identifies different metabotypes of asthma severity. *Eur Respir J* 2017;49:2016. Print 2017 Mar.
45. Kannisto S, Laatikainen A, Taivainen A, Savolainen K, Tukiainen H, Voutilainen R. Serum dehydroepiandrosterone sulfate concentration as an indicator of adrenocortical suppression during inhaled steroid therapy in adult asthmatic patients. *Eur J Endocrinol* 2004;150:687-90.
46. Kelly RS, Virkud Y, Giorgio R, Celedon JC, Weiss ST, Lasky-Su J. Metabolomic profiling of lung function in Costa-Rican children with asthma. *Biochim Biophys Acta* 2017; Jun;1863(6):1590-1595.

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