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## Supporting Information

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Branched Poly(ethylene imine)s as Anti-algal and Anti-cyanobacterial Agents with Selective Flocculation Behavior to Cyanobacteria over Algae

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# **Branched Poly(ethylene imine)s as Anti-algal and Anti-cyanobacterial Agents with Selective Flocculation Behavior to Cyanobacteria over Algae**

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**Supporting Information.** Polymer characteristics (**Supplementary Table S1**), additional characteristics of studied phototrophic microorganisms (**Supplementary Table S2**), summary of IC<sub>50</sub> values for growth-inhibitory effect (**Supplementary Table S3**), cidal, aggregation and flocculation activities of B-PEIs (**Supplementary Figure S1-5**) (PDF).

**TABLES***Supplementary Table S1.* Structural characteristics of B-PEIs used in this study.

B-PEIs <sup>a)</sup>	MW <sup>b)</sup>	$\bar{M}_n$ <sup>c)</sup> (GPC)	$\bar{M}_w$ <sup>d)</sup> (GPC)	$\bar{M}_w / \bar{M}_n$	pKa1 <sup>e)</sup>	pKa2 <sup>e)</sup>
<b>B-PEI<sub>0.5</sub></b>	600	470	2,100	4.5	9.4	6.2
<b>B-PEI<sub>1.1</sub></b>	1,800	1,100	1,400	1.3	9.6	6.2
<b>B-PEI<sub>12</sub></b>	10,000	12,000	19,000	1.6	9.0	5.8

<sup>a)</sup> See the text for denotation; <sup>b)</sup> MW and volume reported by a supplier; <sup>c)</sup> The number average molecular weight; Gibney *et al.* (2012)<sup>[1]</sup>; <sup>d)</sup> The weight average molecular weight; Gibney *et al.* (2012)<sup>[1]</sup>; <sup>e)</sup> Gibney *et al.* (2012)<sup>[1]</sup>

*Supplementary Table S2.* Growth-inhibitory effect of low MW PPI-DEN and B-PEIs on freshwater green algae *Chlamydomonas reinhardtii* (*CR*) and *Desmodesmus quadricauda* (*DQ*) and cyanobacteria *Synechococcus elongatus* (*SE*) and *Microcystis aeruginosa* (*MA*) based on monitoring optical density ( $\lambda = 680$  nm). IC<sub>50</sub> values are expressed for comparison in  $\mu\text{g mL}^{-1}$  (**A**) and in  $\mu\text{M}$  (**B**).

**A**

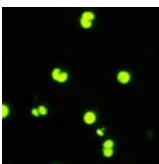
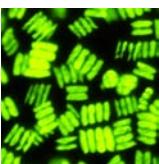
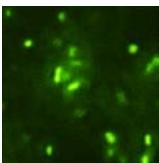
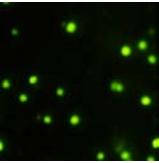
POLYMER	GREEN ALGAE						CYANOBACTERIA				
	<i>CR</i>			<i>DQ</i>			<i>SE</i>			<i>MA</i>	
	IC <sub>50</sub> <sup>a)</sup> ( $\mu\text{g mL}^{-1}$ )			IC <sub>50</sub> ( $\mu\text{g mL}^{-1}$ )			IC <sub>50</sub> ( $\mu\text{g mL}^{-1}$ )			IC <sub>50</sub> ( $\mu\text{g mL}^{-1}$ )	
	24 h <sup>b)</sup>	48 h	72 h	48 h	72 h	96 h	24 h	48 h	72 h	72 h	96 h
PPI-DEN	<b>0.22</b> (0.09-0.55)	<b>0.22</b> (0.09-0.74)	<b>0.24<sup>aA</sup></b> (0.21-0.36)	<b>3.04</b> (1.27-7.30)	<b>2.72<sup>aB</sup></b> (1.63-4.55)	<b>2.21</b> (1.46-3.34)	<b>0.23</b> (0.10-0.49)	<b>0.21</b> (0.12-0.36)	<b>0.21<sup>aA</sup></b> (0.14-0.33)	<b>0.28<sup>aA</sup></b> (0.10-0.49)	<b>0.33</b> (0.23-0.50)
B-PEI <sub>0.5</sub>	<b>0.35</b> (0.23-0.56)	<b>0.18*</b> (0.08-0.38)	<b>0.14*<sup>abA</sup></b> (0.13-0.14)	<b>3.39</b> (1.94-5.90)	<b>2.59<sup>aB</sup></b> (1.42-4.74)	<b>2.43</b> (0.80-7.38)	<b>0.35</b> (0.11-1.17)	<b>0.33</b> (0.19-0.56)	<b>0.36<sup>bC</sup></b> (0.30-0.45)	<b>0.22<sup>aAC</sup></b> (0.10-0.47)	<b>0.26</b> (0.16-0.43)
B-PEI <sub>1.1</sub>	<b>0.32</b> (0.16-0.65)	<b>0.21</b> (0.17-0.27)	<b>0.21<sup>ba</sup></b> (0.14-0.32)	<b>4.06</b> (2.91-5.65)	<b>3.36<sup>aB</sup></b> (1.92-5.87)	<b>2.71</b> (1.09-6.75)	<b>0.23</b> (0.19-0.29)	<b>0.24</b> (0.19-0.29)	<b>0.27<sup>abA</sup></b> (0.21-0.34)	<b>0.27<sup>aA</sup></b> (0.17-0.43)	<b>0.26</b> (0.20-0.34)
B-PEI <sub>12</sub>	<b>0.74</b> (0.35-1.53)	<b>0.56</b> (0.45-0.70)	<b>0.58<sup>cA</sup></b> (0.48-0.69)	<b>~<sup>c</sup>2.5-40</b> (40-44 %)	<b>2.5-40<sup>aB</sup></b> (40-47 %)	<b>~2.5-40</b> (40-47 %)	<b>0.39</b> (0.19-1.13)	<b>0.35</b> (0.27-0.50)	<b>0.36<sup>bA</sup></b> (0.27-0.55)	<b>0.24<sup>aA</sup></b> (0.15-0.39)	<b>0.19</b> (0.09-0.40)

**B**

PPI-DEN	<b>0.70</b> (0.28-1.74)	<b>0.68</b> (0.28-2.34)	<b>0.74<sup>aA</sup></b> (0.67-1.14)	<b>9.60</b> (4.01-23.06)	<b>8.59<sup>aB</sup></b> (5.15-14.37)	<b>6.98</b> (4.61-10.55)	<b>0.73</b> (0.32-1.55)	<b>0.66</b> (0.38-1.14)	<b>0.66<sup>aA</sup></b> (0.44-1.04)	<b>0.88<sup>aA</sup></b> (0.32-1.55)	<b>1.04</b> (0.73-1.58)
B-PEI <sub>0.5</sub>	<b>0.74</b> (0.49-1.19)	<b>0.38*</b> (0.17-0.81)	<b>0.30*<sup>ba</sup></b> (0.28-0.30)	<b>7.21</b> (4.13-12.55)	<b>5.51<sup>abB</sup></b> (3.02-10.09)	<b>5.17</b> (1.70-15.70)	<b>0.74</b> (0.23-2.49)	<b>0.70</b> (0.40-1.19)	<b>0.77<sup>aC</sup></b> (0.64-0.96)	<b>0.47<sup>bAC</sup></b> (0.21-1.00)	<b>0.455</b> (0.34-0.91)
B-PEI <sub>1.1</sub>	<b>0.29</b> (0.15-0.59)	<b>0.19</b> (0.15-0.25)	<b>0.19<sup>cA</sup></b> (0.13-0.29)	<b>3.69</b> (1.62-3.14)	<b>3.05<sup>bb</sup></b> (1.75-5.34)	<b>2.46</b> (0.99-5.14)	<b>0.21</b> (0.17-0.26)	<b>0.22</b> (0.17-0.26)	<b>0.25<sup>ba</sup></b> (0.19-0.31)	<b>0.25<sup>cA</sup></b> (0.15-0.24)	<b>0.24</b> (0.18-0.31)
B-PEI <sub>12</sub>	<b>0.06</b> (0.03-0.13)	<b>0.05</b> (0.04-0.06)	<b>0.05<sup>dA</sup></b> (0.04-0.06)	<b>~<sup>c</sup>0.2-3.3</b> (40-44 %)	<b>~0.2-3.3<sup>bb</sup></b> (40-47 %)	<b>~0.2-3.3</b> (40-47 %)	<b>0.03</b> (0.02-0.09)	<b>0.03</b> (0.02-0.04)	<b>0.03<sup>cc</sup></b> (0.02-0.05)	<b>0.02<sup>dA</sup></b> (0.01-0.03)	<b>0.02</b> (0.01-0.03)

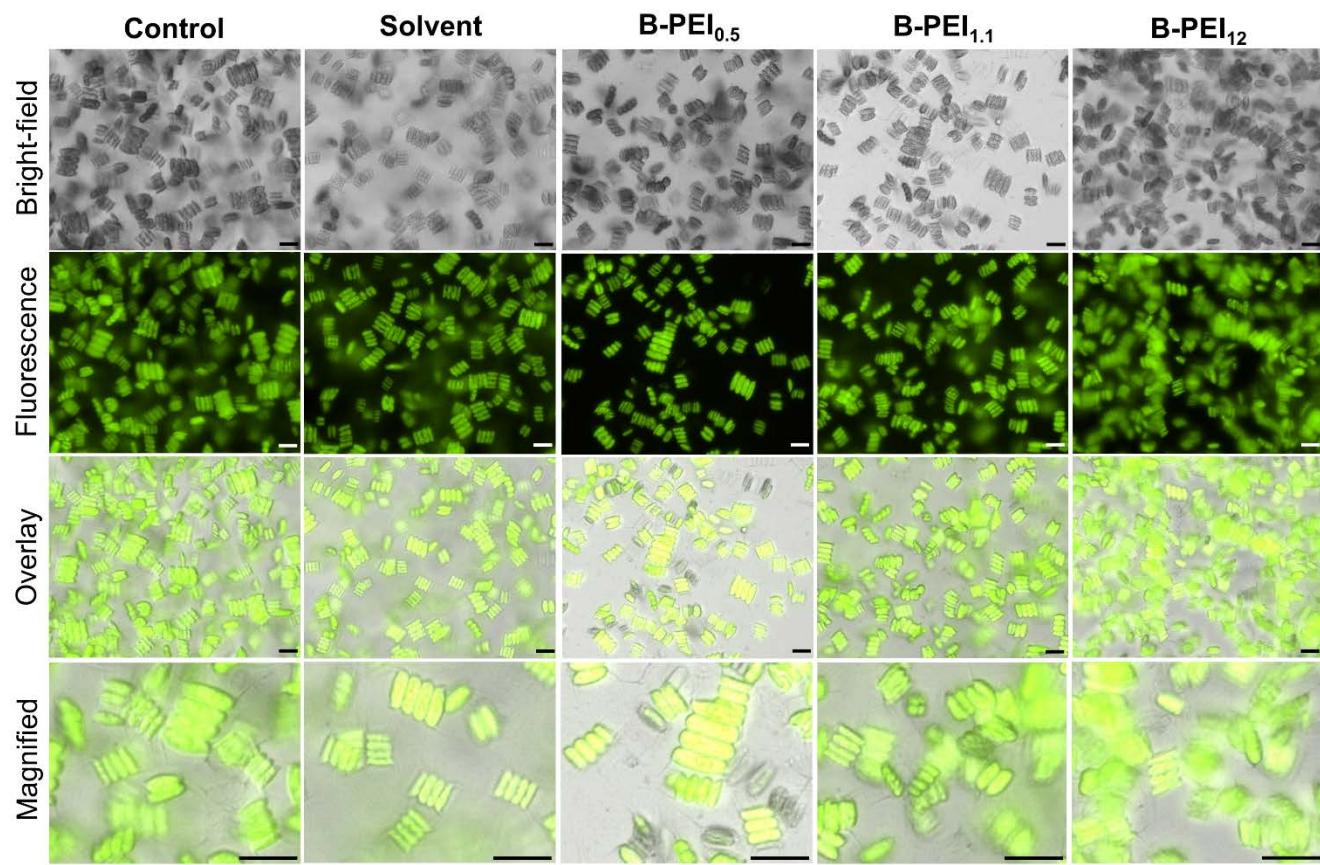
<sup>a)</sup> The concentration of polymer causing 50% inhibition of the algal or cyanobacterial growth. The IC<sub>50</sub> value was determined for each independent experiment and the geometric mean with 95% confidence limits (CI) was calculated (n≥3); <sup>b)</sup> Exposure time; <sup>c)</sup> A clear concentration-dependent effect at the concentrations of 0.3-2.5  $\mu\text{g mL}^{-1}$  (0.03-0.2  $\mu\text{M}$ ) was followed by a plateau value of growth inhibition of about 40-47% observed along the concentration range from 5 to 40  $\mu\text{g mL}^{-1}$  (0.4-3  $\mu\text{M}$ ). Asterisk denotes the significant difference from the IC<sub>50</sub> value after 24 (48) h ( $P \leq 0.05$ ; one-way ANOVA with Dunnett's method or unpaired two-tailed t-test). Values with different lowercase letter superscripts differ significantly ( $P \leq 0.05$ ; one-way ANOVA with Tukey's method) among the polymers within the same microorganism tested. Values with different italic uppercase letter superscripts differ significantly ( $P \leq 0.05$ ; one-way ANOVA with Tukey's method) among different microorganisms tested within the same polymer. B-PEI, branched polyethylenimine; PPI-DEN, poly(propylenimine)-dendrimer.

*Supplementary Table S3.* Characteristics of studied freshwater phototrophic microorganisms

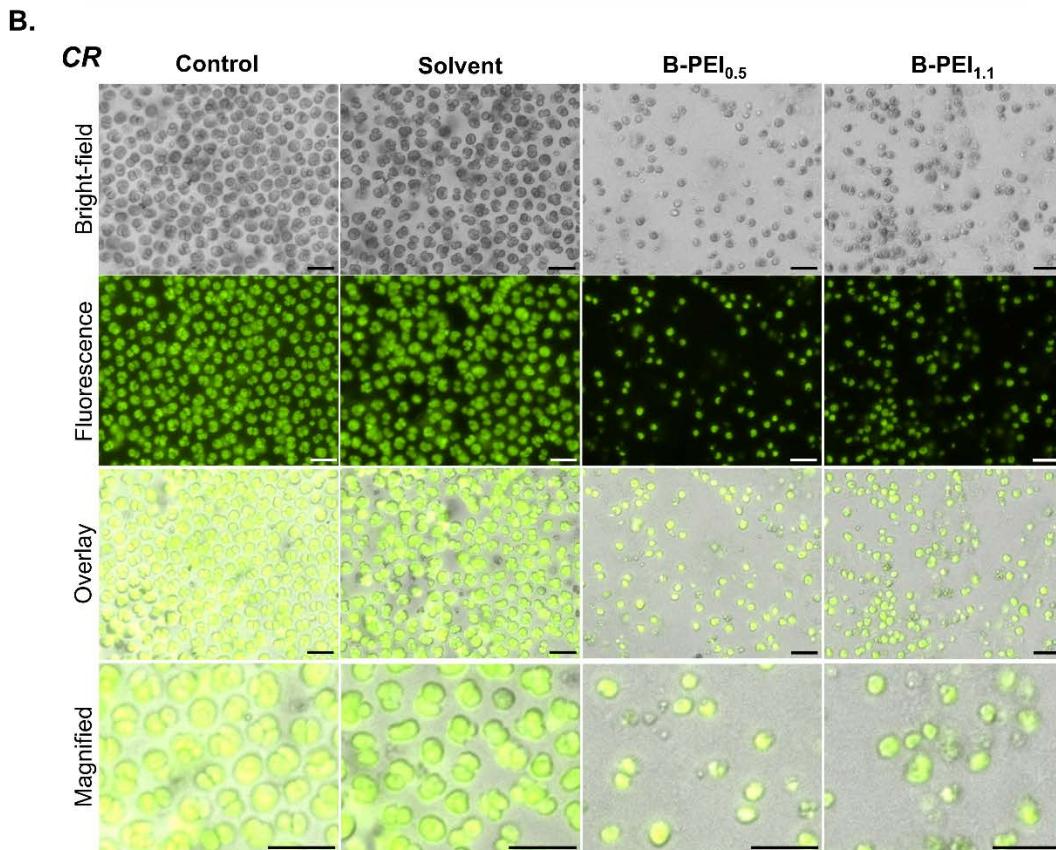
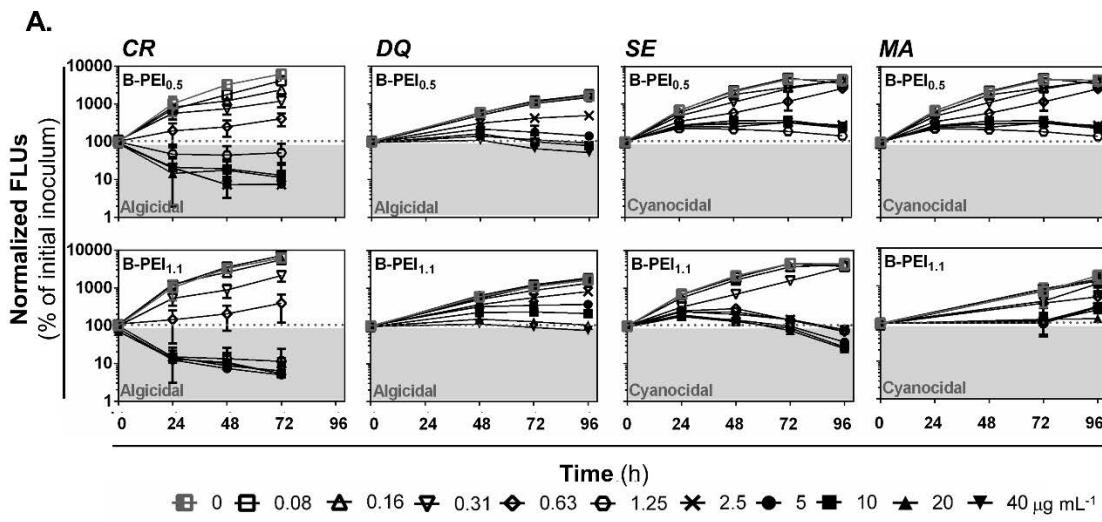
Genus	Species	Photo	Cell shape <sup>[2]</sup>	Length/ width ( $\mu\text{m}$ ) <sup>a)</sup>	Volume <sup>b)</sup> ( $\mu\text{m}^3$ )	Dt <sup>c)</sup> (h)	Characteristics & impact	Negative impact	Cell envelope (cell wall & membrane)
Green algae – eukaryotic cell type	<i>CR</i>		Sphere	6-12	113-904	19	<ul style="list-style-type: none"> <li>• ubiquitous algae</li> <li>• common species of extensive water blooms</li> <li>• model organisms for antialgal tests and for biofouling</li> </ul>	<ul style="list-style-type: none"> <li>• multilayers composed of high-molecular-weight glycoproteins rich in hydroxyproline, arabinose and galactose (40-100 nm thick)<sup>[3]</sup></li> <li>• plasma membrane - mainly phospholipids PG and PE, <math>\beta</math>-carotene, betaine lipid DGTS<sup>[4]</sup></li> </ul>	
	<i>DQ</i>		Prolate spheroid	10-18/ 7	3.5- 64-462	26			
Cyanobacteria – prokaryotic cell type	<i>SE</i>		Rod-shaped ⇒ Sphere model <sup>[2]</sup>	0.6-1.6	0.1-2	14	<ul style="list-style-type: none"> <li>• common cyanobacteria in freshwater sources</li> <li>• a model freshwater cyanobacterium for anticyanobacterial tests</li> </ul>	<ul style="list-style-type: none"> <li>• external surface layers (slime or sheath)</li> <li>• outer membrane (neutral or low negative charge) - phospholipids, lipopolysaccharide (LPS), glycolipids, carotenoids and porins<sup>[6]</sup></li> <li>• peptidoglycan (multilayers - thicker and with a higher degree of cross-linking between PG chains than those in other Gram-negative bacteria)<sup>[6]</sup></li> <li>• plasma membrane - mainly glycolipids MGDG, DGDG and SQDG, and phospholipid PG<sup>[4b,7]</sup></li> </ul>	
	<i>MA</i>		Sphere	3.5-6	22-113	27	<ul style="list-style-type: none"> <li>• produces cyanotoxins microcystins</li> <li>• presence of gas vesicles</li> <li>• causes harmful water blooms of health, economic and ecological importance</li> </ul>		

<sup>a)</sup> Protist Information Server, URL: <http://protist.i.hosei.ac.jp/>; <sup>b)</sup> Calculated based on geometric models for phytoplankton<sup>[2]</sup>; <sup>c)</sup> Doubling time (Dt) under the conditions of this study. DGDG, digalactosyldiacylglycerol; DGTS, Diacylglyceryltrimethylhomoserine; MGDG, monogalactosyldiacylglycerol; SQDG, sulfoquinovosyldiacylglycerol; PC, phosphatidylcholine; PE, phosphatidylethanolamine; PG, phosphatidylglycerol.

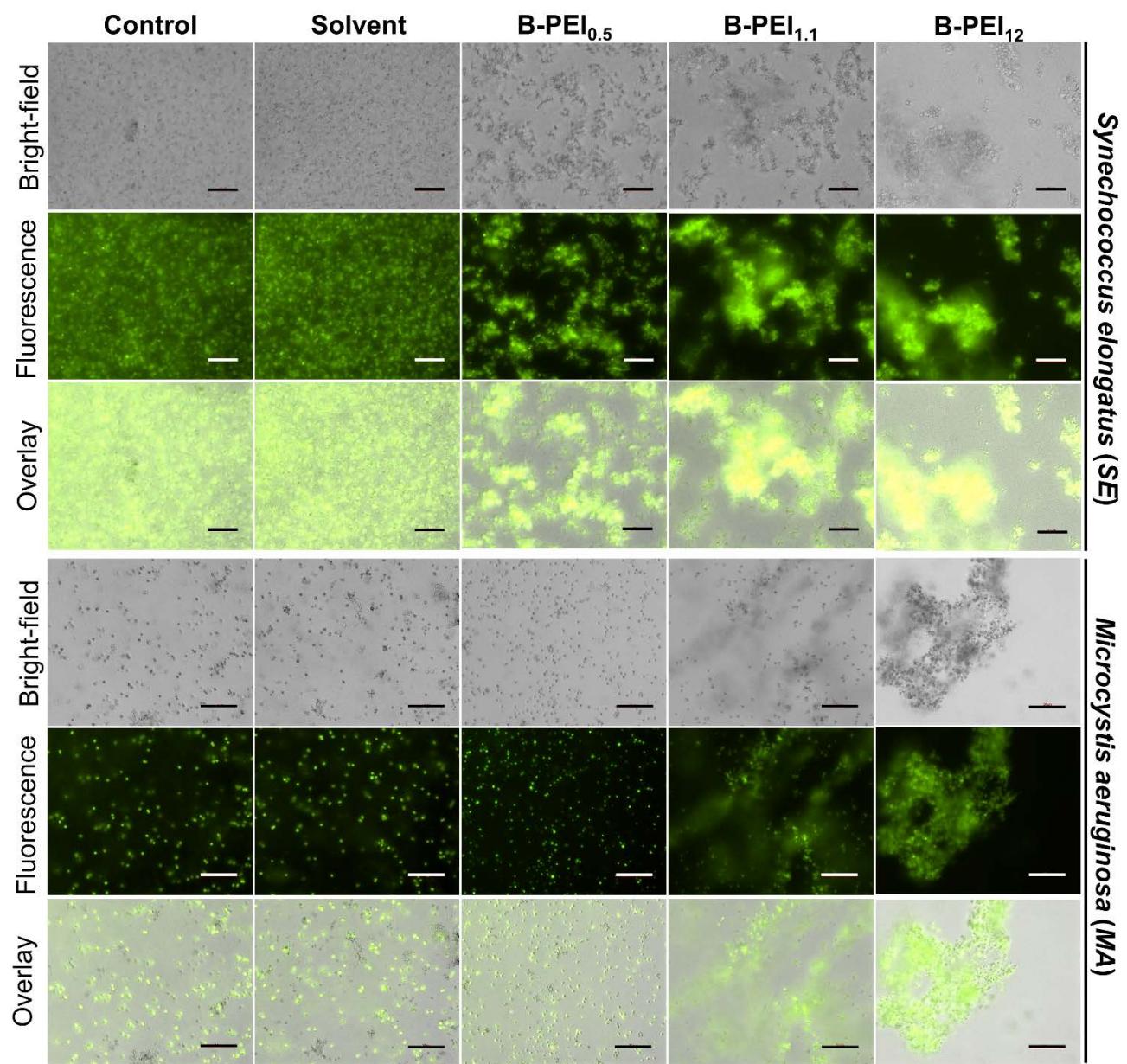
## FIGURES



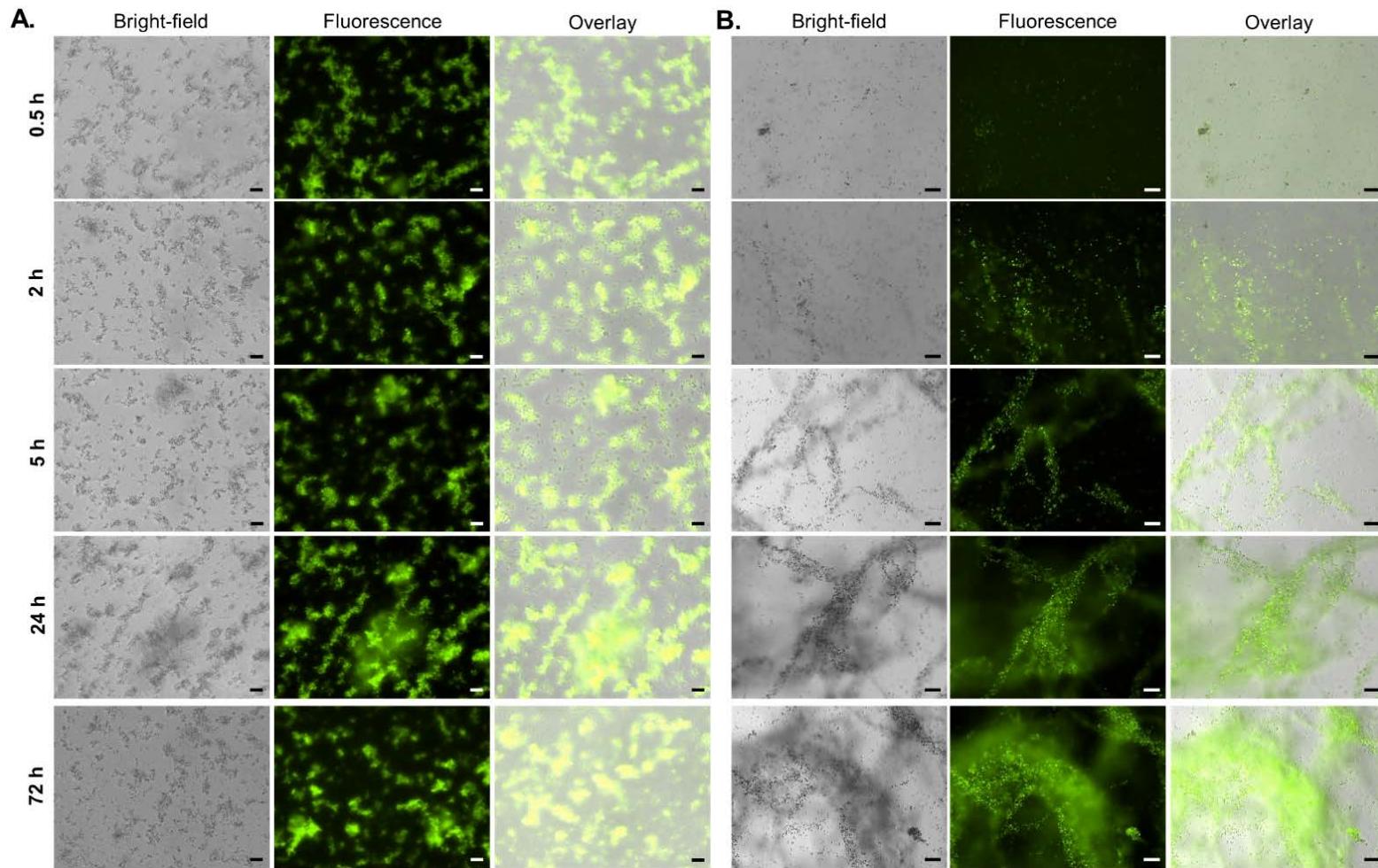
**Figure S1. Freshwater green algae DQ after treatment with B-PEIs.** Representative bright-field, fluorescence, overlay and magnified images of non-treated cells (naïve and solvent controls) and cells treated with B-PEI<sub>0.5</sub>, B-PEI<sub>1.1</sub> and B-PEI<sub>12</sub> at the concentration of 20 µg mL<sup>-1</sup> for 72 h. *DQ*, *Desmodesmus quadricauda*. Scale bar = 20 µm.



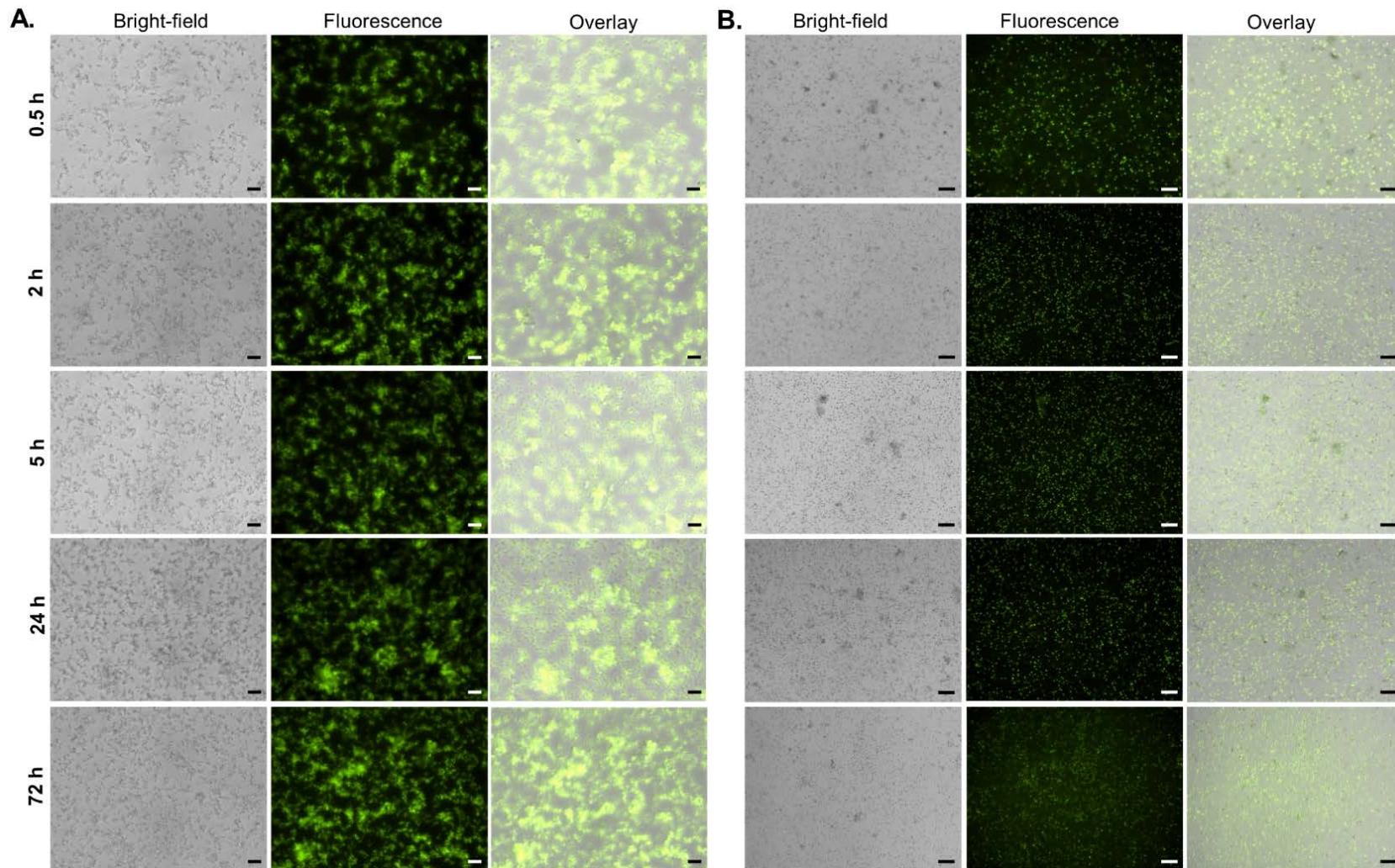
**Figure S2. Cidal activity of B-PEIs to algae CR, but not to algae DQ and cyanobacteria SE and MA. (A)** B-PEI<sub>0.5</sub> and B-PEI<sub>1.1</sub> against *Chlamydomonas reinhardtii* (CR) and *Desmodesmus quadricauda* (DQ) and cyanobacteria *Synechococcus elongatus* (SE) and *Microcystis aeruginosa* (MA) as assessed by autofluorescence ( $\lambda_{\text{ex}} = 485 \text{ nm}/\lambda_{\text{em}} = 675 \text{ nm}$ ) at different time points (24–96 h). B-PEIs concentrations: 0–40  $\mu\text{g mL}^{-1}$ . Data are expressed as a percentage of fluorescence units (FLUs) for initial inoculum and presented as means (SD; n = 3–5). **(B)** CR cells after treatment with B-PEI<sub>0.5</sub> or B-PEI<sub>1.1</sub>. Representative bright field, fluorescence and overlaid images of non-treated CR cells (naïve and solvent controls) and CR cells treated with B-PEI<sub>1.1</sub> at the concentration of 20  $\mu\text{g mL}^{-1}$  for 72 h. Scale bar = 20  $\mu\text{m}$ .



**Supplementary Figure S3. Cyanobacteria SE and MA after treatment with B-PEIs.** Representative bright-field, fluorescence and overlaid images of non-treated cells (naïve and solvent controls) and cells treated with B-PEI<sub>0.5</sub>, B-PEI<sub>1.1</sub> and B-PEI<sub>12</sub> at the concentration of 20 µg mL<sup>-1</sup> for 72 h. B-PEI, branched polyethylenimine; MA, *Microcystis aeruginosa*; SE, *Synechococcus elongatus*. Scale bar = 20 µm (SE)/ 50 µm (MA).



**Supplementary Figure S4. Time-dependent aggregation and flocculation activities of B-PEI<sub>1.1</sub> inducing clustering of both cyanobacterial cell types.** Representative bright-field, fluorescence and overlay images of cyanobacteria *Synechococcus elongatus* (SE: A) or *Microcystis aeruginosa* (MA: B) treated with B-PEI<sub>1.1</sub> at the concentration of 20  $\mu\text{g mL}^{-1}$  for 0.5, 2, 5, 24, and 72 h. The aggregated cells (flocs) were still autofluorescent, suggesting the presence of live cells with intact photosynthetic pigments in the aggregates. Scale bar = 20  $\mu\text{m}$  (SE)/ 50  $\mu\text{m}$  (MA).



**Supplementary Figure S5. Time-dependent aggregation and flocculation activities of B-PEI<sub>0.5</sub> inducing clustering of cyanobacterial SE cells, but not MA cells.** Representative bright-field, fluorescence and overlaid images of cyanobacteria *Synechococcus elongatus* (SE: A) or *Microcystis aeruginosa* (MA: B) treated with B-PEI<sub>0.5</sub> at the concentration of 20 µg mL<sup>-1</sup> for 0.5, 2, 5, 24 and 72 h. The aggregated cells (flocs) of SE were still autofluorescent, suggesting the presence of live cells with intact photosynthetic pigments in the aggregates. Scale bar = 20 µm (SE)/ 50 µm (MA).

## REFERENCES

- [1] K. A. Gibney, I. Sovadinova, A. I. Lopez, M. Urban, Z. Ridgway, G. A. Caputo, K. Kuroda, *Macromol. Biosci.* **2012**, *12*, 1279.
- [2] J. Sun, D. Liu, *J. Plankton Res.* **2003**, *25*, 1331.
- [3] a) D. S. Domozych, M. Ciancia, J. U. Fangel, M. D. Mikkelsen, P. Ulvskov, W. G. T. Willats, *Front. Plant Sci.* **2012**, *3*, 82; b) J. Voigt, J. Woestemeyer, R. Frank, *J. Biol. Chem.* **2007**, *282*, 30381.
- [4] a) I. Domonkos, M. Kis, Z. Gombos, *Acta Biol. Szeged.* **2015**, *59*, 83; b) D. R. Janero, R. Barrnett, *J. Lipid Res.* **1981**, *22*, 1126.
- [5] D. Umysova, M. Vitova, I. Douskova, K. Bisova, M. Hlavova, M. Cizkova, J. Machat, J. Doucha, V. Zachleder, *BMC Plant Biol.* **2009**, *9*, 58.
- [6] a) M. F. Fiore, J. T. Trevors, *Biometals* **1994**, *7*, 83; b) U. J. Jürgens, C. Martin, J. Weckesser, *FEMS Microbiol. Lett.* **1989**, *65*, 47; c) E. Hoiczyk, A. Hansel, *J. Bacteriol.* **2000**, *1825*, 1191; d) E. Gantt, in *The Molecular Biology of Cyanobacteria*, (Ed: D. A. Bryant), *Science* **2006**, p. 119.
- [7] H. Wada, N. Murata, in *Lipids in Photosynthesis: Structure, Function and Genetics. Advances in Photosynthesis and Respiration* (Eds: S. Paul-André, M. Norio), Springer, Dordrecht **2006**, Vol. 16, p. 65.